Genome Wide Association Identifies Common Variants at the SERPINA6/SERPINA1 Locus Influencing Plasma Cortisol and Corticosteroid Binding Globulin

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Abstract

Variation in plasma levels of cortisol, an essential hormone in the stress response, is associated in population-based studies with cardio-metabolic, inflammatory and neuro-cognitive traits and diseases. Heritability of plasma cortisol is estimated at 30–60% but no common genetic contribution has been identified. The CORtisol NETwork (CORNET) consortium undertook genome wide association meta-analysis for plasma cortisol in 12,597 Caucasian participants, replicated in 2,795 participants. The results indicate that <1% of variance in plasma cortisol is accounted for by genetic variation in a single region of chromosome 14. This locus spans SERPINA6, encoding corticosteroid binding globulin (CBG, the major cortisol-binding protein in plasma), and SERPINA1, encoding α1-antitrypsin (which inhibits cleavage of the reactive centre loop that releases cortisol from CBG). Three partially independent signals were identified within the region, represented by common SNPs; detailed biochemical investigation in a nested sub-cohort showed all these SNPs were associated with variation in total cortisol binding activity in plasma, but some variants influenced total CBG concentrations while the top hit (rs12589136) influenced the immunoreactivity of the reactive centre loop of CBG. Exome chip and 1000 Genomes imputation analysis of this locus in the CROATIA-Korcula cohort identified missense mutations in SERPINA6 and SERPINA1 that did not account for the effects of common variants. These findings reveal a novel common genetic source of variation in binding of cortisol by CBG, and reinforce the key role of CBG in determining plasma cortisol levels. In turn this genetic variation may contribute to cortisol-associated degenerative diseases.
Introduction

The adrenal steroid hormone cortisol plays a vital role in adaptation to environmental stress. In response to stressors such as starvation, infection or injury, cortisol secretion is elevated by activation of the hypothalamic-pituitary-adrenal (HPA) axis. Cortisol acts predominantly through glucocorticoid receptors to induce a wide range of physiological responses, including liberating fuel (by facilitating gluconeogenesis and lipolysis), maintaining cardiovascular homeostasis (by inducing sodium retention and vasoconstriction), altering mood and memory (in favour of focusing on ‘fight or flight’ responses), and acting as a ‘brake’ on the innate immune response (preventing bystander damage from unrestrained inflammation) [1]. Chronic elevations in cortisol, however, may be maladaptive, as exemplified in patients with tumours of the pituitary or adrenal gland causing Cushing’s syndrome; here, elevated plasma cortisol is responsible for obesity, type 2 diabetes, hypertension, dyslipidaemia, depression, memory loss, impaired wound healing, osteoporosis, myopathy, and many other features.

Epidemiological data suggest that subtle activation of the HPA axis associates with many of these traits within the population, in people who do not harbour the tumours which cause overt Cushing’s syndrome. In these studies higher plasma cortisol concentration, measured in the morning, provided a robust marker of the activation of the HPA axis which accompanies high blood pressure, hyperglycaemia and dyslipidaemia [2–5].
age-associated cognitive dysfunction [6], and low mood [7]. Conversely, lower cortisol associates with immunological abnormalities [8], post-traumatic stress disorder (PTSD) [9], and obesity [1] (the inverse association with obesity is likely due to increased metabolic clearance of cortisol and confounds the positive association of cortisol with other cardiovascular risk factors, explaining some inconsistencies in the associations of cortisol with ‘metabolic syndrome’ [1]). Mechanisms underlying these associations remain uncertain, with most investigators suggesting abnormal central control of the HPA axis [1,10,11]. A high proportion of cortisol in plasma is protein bound, mostly to corticosteroid binding globulin (CBG). Although variations in total CBG concentrations have been associated with features of metabolic syndrome [12,13], this does not account entirely for associations of total plasma cortisol with other quantitative traits [5,14,15].

Morning plasma cortisol has a heritability of 30–60% [16–18]. Identifying genetic variants which contribute to variation in morning cortisol values could provide key insights into the mechanism of HPA axis activation associated with common quantitative traits, and an opportunity to dissect causality using Mendelian randomisation [19]. Attempts to identify these genetic variants to date have been limited to small candidate gene studies [18]. We therefore established the CORtisol NETwork (CORNET) consortium with the initial aim of identifying genetic determinants of inter-individual variation in HPA axis function.

Results

Genome-wide association meta-analysis

We conducted a discovery meta-analysis of genome-wide association studies (GWAMA) of morning plasma cortisol levels, investigating ~2.5 million SNPs in 12,597 men and women, aged 14–102 years, of European origin (Table S1 for participant characteristics). There was very little inflation of test statistics (λG = 1.005, Table S2). The −log10P values by chromosome for age- and sex-adjusted cortisol z-scores are shown in Figure 1a. A quantile–quantile plot (Figure 1b) showed marked departure from the null for SNPs with low P values (0.04,0.10; P = 3.1×10−5), −0.04 (−0.07,−0.01; P = 0.012), and −0.03 (−0.07,0.00; P = 0.037), respectively. The SERPINA6 gene encodes corticosteroid binding globulin (CBG). The neighbouring (upstream) gene, SERPEN1, encodes α1-antitrypsin, the inhibitor of neutrophil elastase which cleaves and inactivates CBG [21].

![Figure 1. Meta-analysis of genome wide association studies for morning plasma cortisol. A) Manhattan plot of −log10P values by chromosome. The red horizontal line indicates genome-wide significance (P<5×10−8) and the blue horizontal line indicates moderate significance (P<5×10−6). The lead SNP rs12589136 (chr14:94,793,686; b37) in red is genome-wide significant. SNPs within 50 kb of cortisol-related candidate genes (listed in Table S6) are highlighted in colours. B) Quantile-quantile plot of −log10P, comparing the distribution of observed −log10P-values and that expected by chance. doi:10.1371/journal.pgen.1004747.g001](Image 315x551 to 551x596)
Conditional analysis

A quantile-quantile plot after removal of the SERPINA6/SERPINA1 region (chr14: 94,768,859–94,843,565; Genome Reference Consortium build 37) showed no evident inflation of test statistics (not shown). Conditional analyses adjusting for each of the partially independent genome-wide significant variants (rs12589136, rs11621961, rs2749527) in a subset of the meta-analysis population reduced the significance of all other SNPs to $P \leq 1 \times 10^{-5}$ for an association with plasma cortisol.

Gene-centric associations

We used a gene-centric approach to analyse the combined effect of all SNPs within a gene, rather than individual SNP associations, using VEGAS [22]. This produced a gene-based test statistic from meta-analysis results, allowing identification of genes containing multiple SNPs that individually did not reach genome-wide significance (Table S4). Only SERPINA6 and SERPINA1 were identified as gene-wide significant ($P < 3 \times 10^{-6}$); both included rs12509136 in the gene boundary.

Candidate gene analysis

A list of 61 candidate genes thought likely to influence plasma cortisol was collated by a panel of experts. A Manhattan plot of the $-\log_{10} P$ values highlighted for SNPs in these candidate genes showed that only SERPINA6 reached genome-wide significance (Figure 1a). Using gene-based p-values from VEGAS, after adjusting for multiple testing, only SERPINA6 was associated with plasma cortisol (Table S5). SERPINA1 was not included in the candidate gene list.

Replication

In 2,795 participants in additional cohort studies, the association with plasma cortisol was replicated for the lead SNP rs12509136 ($P = 0.0002$), rs11621961 ($P = 0.003$) and rs2749529 (used as proxy for rs2749527, $P = 0.019$)(Table 1; Table S1 for participant characteristics; Table S6 for results in each cohort).

Functional consequences of genetic variation in the SERPINA6/SERPINA1 region

We explored the associations between rs12589136, rs11621961, rs2749527 with cortisol and CBG phenotypes in more detail in 316 subjects from the CROATIA-Korcula cohort (Table 2). Together these three SNPs explained 0.54% of the variance in total plasma cortisol in CROATIA-Korcula. However, there were
Table 1. Association with morning plasma cortisol of SNPs representing signals in the SERPINA6/SERPINA1 region from meta-analyses of discovery genome-wide association studies and of replication studies.

<table>
<thead>
<tr>
<th>SNP ID</th>
<th>Chr</th>
<th>Position (cM)</th>
<th>Alleles</th>
<th>Effect on other</th>
<th>Beta (95%CI)</th>
<th>P</th>
<th>Beta (95%CI)</th>
<th>Effects</th>
<th>P</th>
<th>Beta (95%CI)</th>
<th>Effects</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs12589136</td>
<td>14</td>
<td>94,793,686</td>
<td>T/G</td>
<td>0.22</td>
<td>0.21</td>
<td>0.12</td>
<td>0.19</td>
<td>0.16</td>
<td>0.09</td>
<td>0.12</td>
<td>0.03</td>
</tr>
<tr>
<td>rs12589137</td>
<td>14</td>
<td>94,793,686</td>
<td>T/G</td>
<td>0.49</td>
<td>0.45</td>
<td>0.07</td>
<td>0.06</td>
<td>0.07</td>
<td>0.04</td>
<td>0.06</td>
<td>0.01</td>
</tr>
<tr>
<td>rs11621961</td>
<td>14</td>
<td>94,827,068</td>
<td>T/C</td>
<td>0.49</td>
<td>0.45</td>
<td>0.07</td>
<td>0.06</td>
<td>0.07</td>
<td>0.04</td>
<td>0.06</td>
<td>0.01</td>
</tr>
</tbody>
</table>

Adjusted for age and sex, \( P_{\text{adj}} = 0.017 \) compared with MAF = 0.0046 in dbSNP. After adjusting for age, sex, and accounting for kinship, participants who were heterozygote for the Leuven variant had lower total cortisol and markedly lower total cortisol binding activity, but normal CBG immunoreactivity (Table 2).

Discussion

These results clearly attribute inter-individual differences in morning plasma cortisol amongst Europeans to genetic variation within a region on chromosome 14 containing the SERPINA6 and SERPINA1 genes. The association of this region with plasma cortisol was consistent across multiple cohorts and was observed not only in genome-wide meta-analysis of individual SNPs, but also in gene-based hypothesis-free analysis, and in a candidate gene analysis. Investigation of the functional consequences of genetic variation in this region in a genetic isolate population in

distinct patterns of association of ‘high cortisol’ alleles with CBG. After adjusting for age and sex, although all three variants were associated with differences in total cortisol binding activity, measured by the binding of \([\text{H}]\)-cortisol, there were different associations with CBG immunoreactivity. The T allele at rs2749527 was associated with higher ‘total’ CBG concentration by radioimmunoassay, and there were similar, but weaker associations of total CBG immunoreactivity with variation at rs11621961. Differences in calculated free plasma cortisol reflected these differences in total CBG immunoreactivity, which is used in the calculation of free cortisol. In contrast, however, the minor (T) allele at rs12589136 was not associated with ‘total’ CBG immunoreactivity but was strongly associated with the proportion of CBG bound by a monoclonal antibody against an epitope in the reactive centre loop of CBG [23]. None of these SNPs representing signals in the SERPINA6/SERPINA1 region was associated with \(2\)-antitrypsin concentrations in blood. However, \(2\)-antitrypsin levels were negatively correlated with plasma total cortisol (\(P = 0.17 \quad 95\% \text{CI} = -0.28, -0.06\); \(P = 0.002\) and calculated free cortisol (\(P = 0.13 \quad 95\% \text{CI} = -0.24, -0.02\); \(P = 0.021\), although they did not correlate with ratio of intact/total CBG 0.02 \quad 95\% \text{CI} = -0.09, 0.13; \(P = 0.715\).
Croatia indicates that the effects of variation at SERPINA1 on plasma cortisol are likely to be mediated through neutrophil elastase and resultant reconfiguration of the reactive centre loop of CBG. Since the process of CBG cleavage by unrecognised variability in the immunoreactivity of the reactive CBG concentrations, and in part in association with a previously globulin (CBG). In part, this is determined by differences in total alterations in total cortisol binding by corticosteroid binding

<table>
<thead>
<tr>
<th>rs11621961</th>
<th>TT</th>
<th>CT</th>
<th>CC</th>
<th>Beta (95%CI)</th>
<th>(P^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total cortisol(^a)</td>
<td>655 (618,694)</td>
<td>664 (644,685)</td>
<td>675 (654,696)</td>
<td>-0.05 (-0.14,0.05)</td>
<td>0.305</td>
</tr>
<tr>
<td>Calculated free cortisol</td>
<td>42 (35,051)</td>
<td>47 (42,522)</td>
<td>50 (46,553)</td>
<td>-0.14 (-0.28,0.00)</td>
<td>0.049</td>
</tr>
<tr>
<td>Measured free cortisol(^b)</td>
<td>14 (11,17)</td>
<td>14 (11,16)</td>
<td>13 (10,16)</td>
<td>0.01 (-0.2,0.22)</td>
<td>0.933</td>
</tr>
<tr>
<td>Total CBG</td>
<td>0.90 (0.82,0.99)</td>
<td>0.90 (0.85,0.94)</td>
<td>0.96 (0.92,1.01)</td>
<td>-0.12 (-0.26,0.02)</td>
<td>0.103</td>
</tr>
<tr>
<td>RCL/total CBG</td>
<td>0.76 (0.71,0.80)</td>
<td>0.77 (0.74,0.79)</td>
<td>0.76 (0.73,0.78)</td>
<td>0.01 (-0.14,0.15)</td>
<td>0.940</td>
</tr>
<tr>
<td>Cortisol binding activity</td>
<td>503 (473,532)</td>
<td>492 (474,511)</td>
<td>531 (512,550)</td>
<td>-0.17 (-0.32,0.03)</td>
<td>0.016</td>
</tr>
<tr>
<td>(\alpha1)-antitrypsin</td>
<td>2.74 (2.37,3.16)</td>
<td>2.74 (2.51,2.99)</td>
<td>2.66 (2.43,2.91)</td>
<td>0.02 (-0.14,0.17)</td>
<td>0.840</td>
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</table>

<table>
<thead>
<tr>
<th>rs12589136</th>
<th>TT</th>
<th>GT</th>
<th>GG</th>
<th>Beta (95%CI)</th>
<th>(P^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total cortisol(^a)</td>
<td>706 (656,760)</td>
<td>681 (657,706)</td>
<td>655 (638,673)</td>
<td>0.13 (0.02,0.24)</td>
<td>0.017</td>
</tr>
<tr>
<td>Calculated free cortisol</td>
<td>47 (41,541)</td>
<td>54 (45,654)</td>
<td>46 (42,502)</td>
<td>0.13 (-0.01,0.26)</td>
<td>0.073</td>
</tr>
<tr>
<td>Measured free cortisol(^b)</td>
<td>13 (11,16)</td>
<td>n/a</td>
<td>13 (12,15)</td>
<td>0.03 (-0.15,0.2)</td>
<td>0.777</td>
</tr>
<tr>
<td>Total CBG</td>
<td>1.01 (0.95,1.07)</td>
<td>0.91 (0.82,1.02)</td>
<td>0.91 (0.87,0.95)</td>
<td>0.06 (-0.07,0.20)</td>
<td>0.371</td>
</tr>
<tr>
<td>RCL/total CBG</td>
<td>0.68 (0.64,0.71)</td>
<td>0.69 (0.65,0.73)</td>
<td>0.80 (0.78,0.82)</td>
<td>-0.49 (-0.64,0.34)</td>
<td>9.4 \times 10^{-11}</td>
</tr>
<tr>
<td>Cortisol binding activity</td>
<td>557 (532,581)</td>
<td>552 (513,591)</td>
<td>491 (477,505)</td>
<td>0.30 (0.16,0.44)</td>
<td>2.3 \times 10^{-3}</td>
</tr>
<tr>
<td>(\alpha1)-antitrypsin</td>
<td>2.68 (2.36,3.03)</td>
<td>2.77 (2.43,3.16)</td>
<td>2.69 (2.50,2.90)</td>
<td>0.06 (-0.09,0.22)</td>
<td>0.421</td>
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<table>
<thead>
<tr>
<th>rs2749527</th>
<th>CC</th>
<th>GT</th>
<th>TT</th>
<th>Beta (95%CI)</th>
<th>(P^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total cortisol(^a)</td>
<td>659 (635,684)</td>
<td>668 (649,687)</td>
<td>674 (643,707)</td>
<td>0.03 (-0.06,0.12)</td>
<td>0.495</td>
</tr>
<tr>
<td>Calculated free cortisol</td>
<td>49 (43,55)</td>
<td>45 (41,50)</td>
<td>48 (42,55)</td>
<td>0.02 (-0.11,0.16)</td>
<td>0.717</td>
</tr>
<tr>
<td>Measured free cortisol(^b)</td>
<td>16 (14,19)</td>
<td>11 (8,15)</td>
<td>13 (11,15)</td>
<td>-0.11 (-0.29,0.07)</td>
<td>0.246</td>
</tr>
<tr>
<td>Total CBG</td>
<td>0.87 (0.82,0.92)</td>
<td>0.92 (0.87,0.98)</td>
<td>1.00 (0.96,1.05)</td>
<td>0.17 (0.03,0.30)</td>
<td>0.014</td>
</tr>
<tr>
<td>RCL/total CBG</td>
<td>0.76 (0.73,0.79)</td>
<td>0.76 (0.74,0.79)</td>
<td>0.76 (0.73,0.79)</td>
<td>-0.02 (-0.16,0.13)</td>
<td>0.824</td>
</tr>
<tr>
<td>Cortisol binding activity</td>
<td>486 (468,504)</td>
<td>507 (486,528)</td>
<td>545 (524,566)</td>
<td>0.24 (0.11,0.38)</td>
<td>4.3 \times 10^{-4}</td>
</tr>
<tr>
<td>(\alpha1)-antitrypsin</td>
<td>2.58 (2.31,2.88)</td>
<td>2.86 (2.62,3.12)</td>
<td>2.65 (2.38,2.94)</td>
<td>0.08 (-0.06,0.23)</td>
<td>0.268</td>
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<table>
<thead>
<tr>
<th>rs113418909 (Leuven)</th>
<th>AA</th>
<th>AT</th>
<th>Beta (95%CI)</th>
<th>(P^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total cortisol</td>
<td>673 (658,688)</td>
<td>562 (500,631)</td>
<td>-0.61 (-1.01,-0.25)</td>
<td>0.001</td>
</tr>
<tr>
<td>Calculated free cortisol</td>
<td>48 (45,52)</td>
<td>38 (27,50)</td>
<td>-0.43 (-1.06,0.20)</td>
<td>0.182</td>
</tr>
<tr>
<td>Measured free cortisol</td>
<td>14 (12,15)</td>
<td>13 (8,20)</td>
<td>-0.04 (-0.92,0.83)</td>
<td>0.924</td>
</tr>
<tr>
<td>Total CBG</td>
<td>0.93 (0.90,0.96)</td>
<td>0.94 (0.82,1.07)</td>
<td>0.05 (-0.58,0.68)</td>
<td>0.870</td>
</tr>
<tr>
<td>RCL/total CBG</td>
<td>0.76 (0.74,0.78)</td>
<td>0.82 (0.78,0.87)</td>
<td>0.51 (-0.18,1.19)</td>
<td>0.146</td>
</tr>
<tr>
<td>Cortisol binding activity</td>
<td>512 (500,524)</td>
<td>347 (305,389)</td>
<td>-1.54 (-2.17,-0.91)</td>
<td>1.9 \times 10^{-6}</td>
</tr>
<tr>
<td>(\alpha1)-antitrypsin</td>
<td>2.67 (2.51,2.84)</td>
<td>2.40 (1.91,3.03)</td>
<td>-0.12 (-0.83,0.58)</td>
<td>0.732</td>
</tr>
</tbody>
</table>

\(^a^\)Data for total cortisol is from the whole Croatia-Korcula sample, \(n = 898\).
\(^b^\)Samples selected for measured free cortisol assay were age and sex matched homozygotes at rs12589136.
\(^c^\)adjusted for age, sex, and first three principal components, using kinship matrix derived from GWAS data.

*Functional consequences of variants in the SERPINA6/A1 locus significantly associated with morning plasma cortisol in GWAMA, and of the Leuven variant, in CROATIA-Korcula.*

Cortisol values are nmol/L, CBG concentration nmol/L, and \(\alpha1\)-antitrypsin g/L. RCL = reactive centre loop.

Data are geometric mean (95% CI)[n]. Cortisol values are nmol/L, CBG concentration nmol/L, and \(\alpha1\)-antitrypsin g/L. RCL = reactive centre loop.

Table 2. Functional consequences of variants in the SERPINA6/A1 locus significantly associated with morning plasma cortisol in GWAMA, and of the Leuven variant, in CROATIA-Korcula.
cohort studies as many other phenotypes. This may reflect the perceived difficulty of obtaining samples at a fixed time of day and in un-stressed conditions, to avoid confounding effects. The CORNET consortium had to decline samples from many cohorts in which time of sampling was inadequately controlled, and even then there was high variability in plasma cortisol. Thus, although the variants we identified in the SERPINA6/SERPINA1 region of chromosome 14 accounted for <1% of the variance in plasma cortisol, this signal may be obscured by substantial unmeasured confounding and measurement error and may comprise a considerable component of the estimated 30–60% heritability of plasma cortisol [16–18]. We identified 3 SNPs with partially independent effects on plasma cortisol. There may be a small degree of linkage disequilibrium between these SNPs, but they also show different associations with CBG biochemistry, suggesting that they represent independent effects. None of these 3 SNPs appears directly to affect CBG function; although rs12589136 is close to a consensus estrogen response element, there was no gender difference in its association with plasma cortisol.

Previous investigations of the genetic determinants of plasma cortisol [reviewed in [18]] have been underpowered candidate gene studies, including some which included a tandem repeat in intron 1 of SERPINA6 [27,28]. Interestingly, many of the genetic variants previously associated with cortisol, eg for glucocorticoid [1] and mineralocorticoid [29,30] receptors, showed no signal whatsoever in the adequately powered GWAMA and candidate gene analysis conducted here.

Rare mutations in SERPINA6 have been described which cause absent CBG protein or, more often, reduced affinity of CBG for cortisol [31–35]. Afflicted individuals have low total plasma cortisol but normal free plasma cortisol. However, they also have abnormal pulsatility of plasma cortisol, and non-specific symptoms including fatigue which are unresponsive to cortisol supplementation; features which have been attributed to abnormal function of CBG in delivering cortisol to target tissues, including in brain regions involved in negative feedback regulation of the HPA axis [21]. Although one of these mutations, A51V, has been found to be surprisingly prevalent (MAF>5%) amongst Chinese subjects [34], it has not been found in non-Asian populations and we did not find Caucasians carrying this mutation when tested by exome chip analysis. In cohort studies, plasma CBG concentrations have been associated with features of the metabolic syndrome [12,13,15] and one previous candidate gene study with >900 participants showed that SNPs in SERPINA6, including some identified as being associated with plasma cortisol in this GWAMA, were predictive of somatic symptoms [36]. We found evidence that genetic variation in the SERPINA6/SERPINA1 region influences total plasma cortisol not only through changes in total CBG concentrations, but also in association with alterations in the immunoreactivity of the reactive centre loop of the CBG protein.

Cleavage of the reactive centre loop (RCL) of CBG by neutrophil elastase and inhibition of elastase activity by α1-antitrypsin has been recognised for more than 20 years [26]. However, the recent development of monoclonal antibodies which recognise the intact RCL of CBG has allowed this process to be studied in vivo for the first time [23]. Using these tools in samples from Croatia-Korcula has provided the novel insight that immunoreactivity of the RCL of CBG is variable in the population, and further that this is explained in part by genetic variations in the SERPINA6/SERPINA1 region. It remains to be determined whether this difference in immunoreactivity of the RCL represents altered susceptibility to CBG cleavage. We show that a common variant (rs12589136) associated with impaired RCL antibody binding was associated with higher total plasma cortisol and higher cortisol binding activity. These observations are inconsistent with the interpretation that impaired RCL antibody binding represents enhanced RCL cleavage [23], given that cleaved CBG has a lower affinity than intact CBG for cortisol binding [37]. Alternatively, the altered immunoreactivity of the RCL epitope of CBG is associated with impaired negative feedback of the HPA axis due to reduced tissue delivery of cortisol by CBG, analogous with findings in CBG knockout mice [38]. Although we could not confirm associated elevation in free plasma cortisol concentrations, these measurements are notoriously unreliable, for example being similarly unhelpful in dissecting the consequences of CBG deficiency described above.

We found further evidence for the importance of CBG using exome chips in the genetic isolate population of Korcula in Croatia, where we discovered an unusually high prevalence of heterozygotes for the Leuven mutation in SERPINA6 [24]. These individuals have lower total plasma cortisol despite normal total CBG concentrations, and we confirmed substantial reductions in total cortisol binding activity, without any difference in RCL antibody binding. The presence of the Leuven variant, however, did not account for the association of the top hit SNPs identified by GWAMA with plasma cortisol or CBG RCL antibody binding.

It is possible that a combination of alterations in CBG substrate as well as in neutrophil elastase level and/or activity may determine cleavage of CBG and tissue delivery of cortisol, especially in local sites of inflammation [21]. Intriguingly, we found inverse associations between levels of α1-antitrypsin, the inhibitor of neutrophil elastase, and plasma cortisol concentrations, consistent with instability of CBG resulting in HPA axis activation as proposed above; however, we could not identify a genetic influence on this relationship, or confirm its association with CBG RCL immunoreactivity. Specifically, we did not identify independent signals for SERPINA6 and SERPINA1 in conditional analysis, and the rare variant Leuven mutation was in linkage disequilibrium with the only rare variant we identified in SERPINA1. Recent studies have identified variants in SERPINA1 that are associated with coronary artery calcification [39] and serum lipid profile [40], the latter represented by rs1303 which is in linkage disequilibrium with the top hit rs12589136 identified by GWAMA (r² = 0.35). These findings are consistent with variation in the SERPINA6/SERPINA1 locus affecting downstream actions of cortisol, but it remains unclear if an interaction exists between the variants at these two genes. Mutations in SERPINA1 can cause the syndrome of α1-antitrypsin deficiency, but we are not aware of any investigations of CBG or cortisol in these patients, and their HPA axis may be disturbed anyway by un-restrained neutrophil-mediated tissue damage. Rare variants in SERPINA1 (notably rs112635299) have been associated with α1-globulin plasma protein levels, of which α1-antitrypsin is a major constituent, using GWAS with 1000 Genomes imputation [41]. However, in the CROATIA-Korcula cohort neither rs1303 (Table S7) nor SNPs imputed from 1000 Genomes (including rs112635299) were associated with plasma cortisol. More detailed phenotyping amongst participants with contrasting genotypes at the SERPINA6/SERPINA1 region will be required to clarify the basis for altered interaction between the two gene products.

These findings emphasise the biological importance of plasma protein binding for steroid hormones, and are analogous to recent findings that a common variant in sex hormone binding globulin contributes to variation in total testosterone levels [42]. Given the
excluded SNPs with obvious input errors (e.g., discrepancies in effect allele frequencies). The results from analysis with or without genomic control were nearly identical, as expected with \(\lambda_{GC} = 1.005\). The genome-wide significance threshold for the meta-analysis was \(P < 5 \times 10^{-8}\). Percentage variation of cortisol was calculated from meta-analysis results as \((2*\text{effect allele frequency}^2*\text{effect allele frequency}^\text{beta}^2/\text{sd}^2)\). A regional plot was generated using LocusZoom [45], and heat map using snp.plotter [46] in R version 2.15.2. Joint analysis of meta-analysis was performed with the GCTA program [47].

**Clumping analysis.** To detect independent top SNPs on the basis of empirical estimates of linkage disequilibrium between the SNPs, we used the clumping function as implemented in PLINK [20]. All the SNPs with a \(P\)-value \(< 5 \times 10^{-8}\) in meta-analysis were used for clumping. We grouped the SNPs within 500 kb of the index SNP that have \( r^2 > 0.2 \) with the index SNP.

**Replication genotyping and analysis.** Genes identified in the meta-analysis were evaluated in the Raine Study, MrOS-Sweden, and Edinburgh type 2 Diabetes Study (ET2DS). Raine Study and MrOS-Sweden had GWAS data so we extracted the replication SNP results, and ET2DS was genotyped at the Wellcome Trust Clinical Research Facility Genetics Core Laboratory in Edinburgh using the OpenArray genotyping platform. rs2749527 failed manufacture for the OpenArray, and was replaced with rs2749529 (\( r^2 = 0.905, D' = 1.0\)) in all replication cohorts. Genotypic association analysis in these studies followed the same methods as those described above for the discovery meta-analysis, adjusting for age and sex.

**Conditional & sex-specific analysis.** We performed meta-analyses of sex-specific GWAS and conditional GWAS in a subset of populations, using single marker association tests of \(z\)-scores of log-transformed cortisol. For the sex-specific analysis, each study adjusted for age in both men (\(n = 3,546\)) and women (\(n = 5,956\)). For the conditional analysis, each study adjusted for age, sex, and each of the SERPINA16 loci SNPs rs12589136 (\(n = 9,308\), \(r^2 = 0.16\)), rs11621961 (\(n = 7,687\)), and rs2749527 (\(n = 9,307\)) individually. We then did fixed effects meta-analysis using GWAMA program.

**Gene-based analysis.** We used the Versatile Gene-based Association Study (VEGAS) program [22] to perform gene-centric analysis. This used individual SNP \(p\)-values derived from the meta-analysis results to compute a gene-based \(p\)-value. We used two methods: all SNPs within a gene, or a subset of the 10% most significant SNPs in each gene boundary. VEGAS accounted for linkage disequilibrium between SNPs using the HapMap phase 2 population (CEU). SNPs were assigned to \(\sim 18,000\) genes based on positions in build 36 (hg18), with gene boundaries of \(\pm 50\) kb of the UTR. Bonferroni corrected threshold for gene-wide significance was \(5 \times 10^{-8}\). The overlap of SNPs included in the gene boundaries in our results indicates this is likely to be an overly conservative correction factor [22].

A list of 61 candidate genes with known biological function in the regulation of cortisol was compiled by a panel of experts in the field. All SNPs within and \(\pm 50\) kb of these genes were examined in the GWAMA results, and gene-based \(p\) values were inspected in VEGAS results.

**Exome chip and 1000 Genomes imputed data analysis**
Genotypes for the SERPINA6/SERPINA1 gene region (chr14:94,770,585–94,857,029, build 37) in the CROATIA-Korcula samples (\(n = 898\)) were extracted from an Illumina Exome Chip v1 analysis. Genotypes were called in GenomeStudio (Illumina) using the CHARGE Consortium joint calling cluster file.
Detailed biochemical studies

More detailed phenotyping was undertaken in samples from 316 participants in the CROATIA-Korcula cohort, comprising 158 age- and sex-matched homozygotes at the top hit variant rs12589136 (53 T/T, 106 G/G; however, one T/T sample had insufficient sample for CBG measurement resulting in 52 T/T and 106 G/G). An additional 160 randomly selected samples had CBG measured, however two samples failed genotyping resulting in an additional 47 T/G and 111 G/G.

Total plasma cortisol was measured with a commercial radioimmunoassay (MP Biomedicals, UK). Total CBG was also measured in CROATIA-Korcula samples using a commercial radioimmunoassay (DiaSource, Louvain-la-Neuve, Belgium). Total cortisol binding capacity was measured using a ligand-saturation assay, with [3H]-cortisol (PerkinElmer Life Sciences, Waltham, MA) as the labelled ligand and dextran-coated charcoal to separate the CBG-bound [3H]-cortisol, as previously described [52]. α1-Antitrypsin was measured with a commercial ELISA (Genway Biotech, San Diego, USA). Unbound free plasma cortisol was calculated from measured total plasma cortisol and total CBG values using Coolens’ equation [53]. Free cortisol was also measured by ELISA following equilibrium dialysis. Briefly, dialysis tubing (12–14 kD, Medicell, London, UK) was heated to 80°C for 30 min in 2% Na bicarbonate and 1 mM EDTA before overnight dialysis of plasma into phosphate buffered saline containing 1% gelatin at 37°C and measurement of dialysed free cortisol by commercial ELISA (Salimetrics Europe Ltd, Newmarket, UK). CBG were also measured, as previously described [29], by ELISAs using monoclonal antibodies 12G2 and 9G12. Antibody 9G12 binds to an epitope in the reactive centre loop (RCL), the elastase cleavage site on CBG, and has been used to infer intact ( uncleaved) CBG, whereas antibody 12G2 binds to a distant epitope and measures total ( cleaved and uncleaved) CBG. As CROATIA-Korcula is a population isolate, we used the polygenic and mnscore functions in GenABEL [51]. All regression equations included the first three principal components of 1000 Genomes imputation was performed using genotypes from Illumina HumanHap370CNV after quality control (Individual Call Rate 97%, SNP Call Rate 98%, MAF 0.01, HWE 1×10⁻⁶); prephasing was performed using ShapeIt v2 [49] and imputation using IMPUTE2 [50] and the ALL (Phase 1 integrated release v3, April 2012) reference panel. Associations with plasma cortisol were analysed in GenABEL [51].

Supporting Information

Table S1 Characteristics of participants in cohorts included in genome-wide association meta-analysis and replication for morning plasma cortisol.

Table S2 Genome-wide data characteristics for discovery cohorts used in meta-analysis.

Table S3 All SNPs with p-values <5×10⁻⁵ in discovery genome wide association meta-analysis for morning plasma cortisol.

Table S4 Top 10 genes identified in VEGAS as associated with morning plasma cortisol from genome wide association meta-analysis with adjustment for age and sex.

Table S5 VEGAS results for candidate genes based on genome wide association meta-analysis results adjusted for age and sex.

Table S6 Replication results in individual cohorts and by meta-analysis for association with morning plasma cortisol of SNPs representing independent signals in the SERPINA6/SERPINA1 region discovered in genome-wide association meta-analysis.

Table S7 Associations with plasma cortisol of variants in SERPINA6/A1 locus on chromosome 14 identified by exome chip in n = 808 subjects from CROATIA-Korcula.

Text S1 Supporting methods.

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Genome Wide Association Studies of Plasma Cortisol

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Author Contributions

Conceived and designed the experiments: BRW HC AW HT. Performed the experiments: JLB CH ND JGL GLH LAH AA JH FPV JL JGE MK NTJ TT PDvH AMa CML LNA Je JFP. Analyzed the data: JLB CH ND HT BRW. Contributed reagents/materials/analysis tools: JFW HC IR AW NH SHW AH AGU KR EK EW AP MRJ GDS SMR DME BDP YM SB LFJGMR SJLB NV RPFD AMo LL EJ CEP SJL SGM DM CO MWJS KMR HT. Wrote the paper: JLB CH ND HT BRW.

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