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The Glycosylation Status of PrP°C Is a Key Factor in Determining Transmissible Spongiform Encephalopathy Transmission between Species

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ABSTRACT

The risk of transmission of transmissible spongiform encephalopathies (TSE) between different species has been notoriously unpredictable because the mechanisms of transmission are not fully understood. A transmission barrier between species often prevents infection of a new host with a TSE agent. Nonetheless, some TSE agents are able to cross this barrier and infect new species, with devastating consequences. The host PrP°C misfolds during disease pathogenesis and has a major role in controlling the transmission of agents between species, but sequence compatibility between host and agent PrP°C does not fully explain host susceptibility. PrP°C is posttranslationally modified by the addition of glycan moieties which have an important role in the infectious process. Here, we show in vivo that glycosylation of the host PrP°C has a significant impact on the transmission of TSE between different host species. We infected mice carrying different glycosylated forms of PrP°C with two human agents (sCJDMM2 and vCJD) and one hamster strain (263K). The absence of glycosylation at both or the first PrP°C glycosylation site in the host results in almost complete resistance to disease. The absence of the second site of N-glycan has a dramatic effect on the barrier to transmission between host species, facilitating the transmission of sCJDMM2 to a host normally resistant to this agent. These results highlight glycosylation of PrP°C as a key factor in determining the transmission efficiency of TSEs between different species.

IMPORTANCE

The risks of transmission of TSE between different species are difficult to predict due to a lack of knowledge over the mechanisms of disease transmission; some strains of TSE are able to cross a species barrier, while others do not. The host protein, PrP°C, plays a major role in disease transmission. PrP°C undergoes posttranslational glycosylation, and the addition of these glycans may play a role in disease transmission. We infected mice that express different forms of glycosylated PrP°C with three different TSE agents. We demonstrate that changing the glycosylation status of the host can have profound effects on disease transmission, changing host susceptibility and incubation times. Our results show that PrP°C glycosylation is a key factor in determining risks of TSE transmission between species.

Transmissible spongiform encephalopathies (TSE), or prion diseases, are fatal neurodegenerative diseases that can be sporadic, genetic, or acquired by infection (1). These diseases are characterized by a distinct pathology in the central nervous system (CNS), with neuronal loss, spongiform degeneration, and gliosis (2). Numerous mammalian species are susceptible to infection with TSE agents, such as scrapie in sheep and goats, bovine spongiform encephalopathy (BSE) in cattle, Creutzfeldt-Jakob disease (CJD) in humans, and chronic wasting disease (CWD) in cervids.

The host cellular protein PrP°C has been shown to have a key role in the transmission of disease (3, 4). During the disease process, PrP°C misfolds from the normal conformation to an aberrant form (PrPSc), which is partially resistant to proteases. The prion hypothesis proposes that PrPSc is the infectious agent responsible for disease transmission and that it is able to self-propagate and induce TSE disease in a new host in the apparent absence of any nucleic acid (5).

Transmission of TSE between different species often is limited by a species barrier to infection (6, 7). In experimental models of disease, the species barrier is characterized by an inefficient primary infection with low susceptibility and long incubation times in the new host. Adaptation to the new host then usually occurs in subsequent passages with an increased attack rate and shorter incubation time (6, 8). In naturally occurring TSE, the species barrier prevents transmission of certain agents between different species. However, some agents have been shown to be able to cross this barrier and cause devastating epidemics in a new host. For example, BSE in cattle can be transmitted to humans via the oral...
route to cause variant CJD (vCJD) (9, 10). BSE also was able to naturally infect a number of different species, such as goats, nyala, kudu, and domestic or captive wild cats (11–13). Understanding how the species barrier is regulated is important, so that the zoonotic potential of a TSE in other animal populations transmitting to humans can be assessed. This is particularly important for newly emergent strains of TSE in both farmed and wild animals (8, 14).

Despite many studies in recent decades, the mechanisms regulating the species barrier to TSE transmission still are elusive. It has been proposed that sequence identity between host and donor PrPSc is important to determine the barrier to transmission. In particular, evidence suggests that sequence homology between host PrP and PrPSc leads to high susceptibility and shorter incubation time, whereas sequence differences between these two proteins can lead to lower susceptibility of the host (6, 15, 16). However, this is not always the case (17–19), and it becomes difficult to predict the transmissibility of a strain in a new recipient based solely on sequence identity between host and donor. It is likely that other factors should be taken in account to understand and predict the species barrier.

PrPSc is variably glycosylated at two highly conserved sites (amino acid positions 180 and 196 in mice). N-glycan attachment to these sites results in four glycosylated forms (glycotypes) of PrP, diglycosylated, monoglycosylated at position 180, monoglycosylated at position 196, and unglycosylated. While the ratios of diglycosylated, monoglycosylated, and unglycosylated PrPSc remain reasonably constant in uninfected brains, the ratios of PrPSc are highly variable in brains infected with different TSE agents.

In vitro studies have demonstrated that the choice of mutation at the glycosylation sites can have profound effects on PrP trafficking, preventing infection of the resulting mutant mice (20). In order to define the role of glycosylation in the transmission of disease, glycosylation-deficient mice have been produced by three laboratories, and inoculations with a number of TSE agents derived from the same species have been carried out (20–22). Conventional transgenic mice expressing hamster PrP in which the first N-glycan site (T183A) or both the first and second site (T183A, T199A) were disrupted were resistant to hamster-passaged prion strains Sc237 and 139H. Mice devoid of the second site also were resistant to 139H but susceptible to Sc237, with extended incubation periods compared to those of the controls (21). In contrast, transgenic mice expressing a 3F4-tagged murine PrP devoid of the first site (T182N) showed no evidence of resistance to any of the murine agents ME7, 139A, and murine-passaged BSE (301C) (20). Mice lacking the second site (T198A) also were susceptible to these strains with extended incubation times (20). Cancellotti et al. produced three inbred lines of gene-targeted mice in which the first (G1; N180T), second (G2; N196T), or both (G3; N180T and N196T) N-glycan attachment sites were disrupted. The homozygous transgenic mice produce partially or unglycosylated PrPSc under the control of the endogenous mouse Prnp promoter (22). Mice lacking the first N-glycan site were susceptible to 79A, but unlike those of Neuendorf et al., they were resistant to ME7. Mice lacking the second site were susceptible to a number of TSE strains, including ME7, 79A, and 301C, similar to those of Neuendorf et al. However, extended incubation periods were observed for all strains in Neuendorf et al. but only in the case of 301C in Tuzi et al. (20, 25). The differences between the lines of mice can be attributed to many variables, including the different point mutations used to disrupt the glycosylation sites, the species of origin of the PrP gene, different gene constructs, the addition of epitopes, copy numbers, and integration sites, and strain differences of the inocula. The gene-targeted mice, however, remove at least some of the variability observed with the conventional lines of mice.

Multiple strains of TSE agents have been identified. Strains differ in their disease characteristics, such as their transmissibility to other host species or the degree of pathology induced in the brain of the host (24–26). These different strains are proposed to result from different conformations of PrPSc (27–29). The ratio of the glycotypes of PrPSc also differs between TSE strains (30, 31). For example, PrPSc associated with vCJD is predominately diglycosylated, whereas that associated with sporadic CJD (sCJD) is mostly monoglycosylated (32). The N-glycans attached to PrP may influence the conformational flexibility of the protein and could influence its misfolding (33, 34). Conformational flexibility may be particularly important during interspecies transmission of TSE, as the infectious agent must adapt to the novel source of PrPSc. Indeed, in vitro interspecies misfolding of PrPSc induced by PrPSc is inhibited specifically by glycosylation of the protein (35). Moreover, the replication of a given strain may require the misfolding of a precise combination of PrPSc glycotypes to replicate the glycoform (36). For example, the replication of a strain in which PrPSc is predominately monoglycosylated may be favored in a host that produces an elevated level of monoglycosylated PrPSc.

Here, we have tested the effect of host PrPSc glycosylation on the TSE species barrier. This is the first in vivo study of the role of glycosylation of PrPSc on the transmission of TSE agents between species. Previous in vitro and in vivo studies to investigate this issue have provided inconsistent results (35, 37, 38) or used mouse-passaged TSE agents (20, 21). Some studies have suggested that the glycosylation of PrPSc impedes the transmission between host species (35), whereas others have shown no such effect (20, 38).

We challenged three lines of gene-targeted glycosylation mutants (22) with three nonmurine TSE agents: human vCJD, human sCJD (sCJDMM2), and hamster scrapie 263K. These agents differ in their PrPSc glycoform ratios and their relative transmissibility to wild-type mice (9, 39, 40). Two of the agents used in this study (263K and vCJD) have similar PrPSc glycoform ratios in which PrPSc is predominately diglycosylated. The sCJDMM2 agent has an amino acid sequence identical to that of the vCJD agent and a cleavage site similar to that of proteinase K (PK); however, it has a significantly different PrPSc glycoform ratio, as the PrPSc associated with this agent is predominately monoglycosylated. By comparing the relative transmissibility of these TSE agents to our transgenic models, we have established the importance of host PrPSc glycosylation in determining the transmissibility of a TSE strain across a species barrier.

MATERIALS AND METHODS

Preparation of TSE inocula and intracerebral injection. Inocula were prepared from brain tissue from a patient with pathologically confirmed sCJDMM2 (0.1% [wt/vol] in physiological saline), the National Institute for Biological Standards and Control (NIBSC) variant CJD reference case (0.1% [wt/vol] in physiological saline), and a hamster with clinical scrapie, strain 263K (1% [wt/vol] in physiological saline). Consent for the use of these materials for research was obtained with ethics approval by the Lothian National Health Service Ethics Committee (reference no. 2000/4/157). G1 (N180T), G2 (N196T), and G3 (N180T, N196T)
transgenic and wild-type mice were genotyped by mismatched PCR as previously described (22) and coded prior to intracerebral (i.c.) injection with 20 μl of inoculum. All groups were age and sex matched.

The second passage was carried out in a similar manner using brain material from selected G2 and wild-type mice showing evidence of TSE vacuolation and/or PrP deposition. A second passage was not carried out in G1 or G3 mice due to extremely low numbers of mice exhibiting TSE vacuolation and/or PrP deposition. Incuba
cation time for mice with clinical signs and evidence of TSE vacuolation. Studies were maintained to ~700 dpi for mice with no clinical signs of TSE disease.

TABLE 1 Sporadic CJDMM2 inoculation into glycosylation-deficient mice at first and seconda passages

<table>
<thead>
<tr>
<th>Mouse line</th>
<th>Inoculum</th>
<th>Incubation timea (days) ± SEM (range)</th>
<th>Clinical disease (no. positive/total no.)</th>
<th>TSE vac and/or PrP depositiona</th>
<th>Survival time (days) of mice with TSE vac and/or PrP depositiona</th>
</tr>
</thead>
<tbody>
<tr>
<td>G1</td>
<td>sCJDMM2</td>
<td>404 ± 8 (378–430)</td>
<td>0/18</td>
<td>0/18</td>
<td>367 ± 50 (245–494)</td>
</tr>
<tr>
<td>G2</td>
<td>sCJDMM2</td>
<td>404 ± 8 (378–430)</td>
<td>7/20</td>
<td>11/20</td>
<td></td>
</tr>
<tr>
<td>G3</td>
<td>sCJDMM2</td>
<td>0/8</td>
<td>0/18</td>
<td>0/18</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>sCJDMM2</td>
<td>0/41</td>
<td>3/41</td>
<td></td>
<td>658 ± 49 (559–707)</td>
</tr>
<tr>
<td>G2</td>
<td>G2sCJDa</td>
<td>129 ± 3</td>
<td>11/11</td>
<td>11/11</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2sCIDa</td>
<td>430 ± 20</td>
<td>12/12</td>
<td>12/12</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>G2sCIDb</td>
<td>147 ± 3</td>
<td>10/11</td>
<td>11/11</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2sCIDb</td>
<td>439 ± 22</td>
<td>10/10</td>
<td>10/10</td>
<td></td>
</tr>
</tbody>
</table>

a Brain material from two sCJDMM2-inoculated G2 mice with pathological evidence of disease were further passaged into G2 and wild-type mice.

b Incubation time for mice with clinical signs and evidence of TSE vacuolation. Studies were maintained to ~700 dpi for mice with no clinical signs of TSE disease.

c TSE vacuolation (vac) and/or PrP deposition were used as evidence of transmission for primary passage due to low numbers of mice with clinical signs and vacuolation present. Limited confirmatory immunohistochemistry for PrP deposition was carried out in the second passage due to high numbers of mice with clinical signs and vacuolation present.

d Survival time of mice with no clinical signs and evidence of vacuolation and/or PrP deposition.

Reactions were terminated by the addition of phenylmethylsulfonyl fluoride. If required, PrPSc then was concentrated by sodium phosphotungstic acid (NaPTA)-mediated precipitation (43). Samples (brain homogenate/NaPTA-concentrated PrPSc) were analyzed for PrPSc by Western blotting using the anti-PrP antibody 8H4 (1/10,000) (44). PrPSc could not be easily detected in vCJD-challenged G2 mice. In order to detect PrPSc in G2 mice, NaPTA-mediated precipitation was carried out on 10% (wt/vol) brain homogenates such that the equivalent of 5 μg of total brain was run per well. Equal PrP signal then was achieved by diluting wild-type PrPSc 10-fold prior to electrophoresis.

Immunohistochemistry. Coronal brain sections were stained using the 6H4 antibody (1/20,000; Prionics) to detect PrP. Antigen retrieval by autoclaving at 121°C for 15 min and a 5-min formic acid (98%) treatment was used. Sections then were blocked with normal rabbit serum prior to incubation with the primary antibody. Antibody binding was detected with either the catalyzed signal amplification system (Dako) or Vector ABC kit (Vector Laboratories) and visualized with 3,3’-diaminobenzidine chromogen. All sections were counterstained with hematoxylin. In all experiments, normal brain homogenate-inoculated mouse and PrPSc mouse (NPU, Edinburgh, United Kingdom) sections were used as a negative control (4).

Biochemical assessment of PrPSc in uninfected glycosylation-deficient transgenic mice. Brains from uninfected G1, G2, G3, and wild-type mice were used to prepare 10% (wt/vol) homogenates. An α-tubulin loading control was included to determine variation in the amount of protein being loaded. Samples were analyzed by Western blotting using seven primary anti-PrP antibodies (7A12/epitope, conformation epitope; DE3/epitope, aa146-153; FH10/epitope, aa202-210; AE11/epitope, aa140-145; BH1/epitope, aa143/154; FD12/epitope, unknown at 0.1 μg/ml; and BC6/epitope, aa146-156 at 0.01 mg/ml) (45, 46) and species-specific peroxi-
dase-conjugated AffiniPure antibodies (Stratech) at 0.05 μg/ml diluted in 0.5% blocking solution for 30 min at room temperature. Bound second-
ary antibody was detected by light emission from SuperSignal West Dura chemiluminescent substrate using a Kodak 440 Image Station with typical exposures of 1, 5, and 10 min. Immunoblots were imaged using the Kodak Image Station and Kodak MI software. Protein bands were selected man-
ually by drawing around them and the sum intensity calculated. A blank region also was selected to measure the background intensity of the blot.

RESULTS
Absence of the first and both PrPSc glycosylation-deficient sites limits transmission with all three agents. Infection of G1 and G3 mice with vCJD and sCJDMM2 resulted in no pathologically confirmed clinical disease and no evidence of TSE vacuolation and/or PrP deposition in the brain (Tables 1 and 2). While there were a num-

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TABLE 2 Variant CJD inoculation into glycosylation-deficient mice at first and second passages

<table>
<thead>
<tr>
<th>Mouse line</th>
<th>Inoculum</th>
<th>Incubation time (days)</th>
<th>Clinical disease (no. positive/total no.)</th>
<th>TSE vac and/or PrP deposition</th>
<th>Survival time (days) of mice with TSE vac and/or PrP deposition</th>
</tr>
</thead>
<tbody>
<tr>
<td>G1</td>
<td>vCJD</td>
<td>0/17</td>
<td></td>
<td>0/17</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>vCJD</td>
<td>0/17</td>
<td></td>
<td>0/17</td>
<td></td>
</tr>
<tr>
<td>G3</td>
<td>vCJD</td>
<td>0/17</td>
<td></td>
<td>0/17</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>vCJD</td>
<td>0/17</td>
<td></td>
<td>0/17</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>G2vCJDa</td>
<td>10/11</td>
<td></td>
<td>11/11</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2vCJDa</td>
<td>12/12</td>
<td></td>
<td>12/12</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>G2vCJDb</td>
<td>12/12</td>
<td></td>
<td>12/12</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2vCJDb</td>
<td>12/12</td>
<td></td>
<td>12/12</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>WtvCJD</td>
<td>13/13</td>
<td></td>
<td>13/13</td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>WtvCJD</td>
<td>12/12</td>
<td></td>
<td>12/12</td>
<td></td>
</tr>
</tbody>
</table>

a Brain material from two vCJD-inoculated G2 mice and one vCJD inoculated wild-type mouse with pathological evidence of disease were further passaged into G2 and wild-type mice.
b Incubation time for mice with clinical signs and evidence of TSE vacuolation. Studies were maintained to ~700 dpi for mice with no clinical signs of TSE disease.
c TSE vacuolation and/or PrP deposition were used as evidence of transmission for primary passage due to low numbers of mice with clinical signs and vacuolation present. Limited confirmatory immunohistochemistry for PrP deposition was carried out in the second passage due to high numbers of mice with clinical signs and vacuolation present.
d Survival time of mice with no clinical signs and evidence of TSE vacuolation and/or PrP deposition.

TABLE 3 263K inoculation into glycosylation-deficient mice at first and second passages

<table>
<thead>
<tr>
<th>Mouse line</th>
<th>Inoculum</th>
<th>Incubation time (days)</th>
<th>Clinical disease (no. positive/total no.)</th>
<th>TSE vac and/or PrP deposition</th>
<th>Survival time (days) of mice with TSE vac and/or PrP deposition</th>
</tr>
</thead>
<tbody>
<tr>
<td>G1</td>
<td>263K</td>
<td>0/14</td>
<td>4/14</td>
<td>579 ± 35 (507–674)</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>263K</td>
<td>0/18</td>
<td>11/18</td>
<td>552 ± 18 (461–629)</td>
<td></td>
</tr>
<tr>
<td>G3</td>
<td>263K</td>
<td>0/16</td>
<td>0/16</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>263K</td>
<td>0/18</td>
<td>11/18</td>
<td>612 ± 18 (503–698)</td>
<td></td>
</tr>
<tr>
<td>G2</td>
<td>G2–263Ka</td>
<td>12/12</td>
<td>12/12</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2–263Ka</td>
<td>10/12</td>
<td>9/12</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>G2–263Kb</td>
<td>4/10</td>
<td>6/10</td>
<td></td>
<td>343 ± 6 (282–408)</td>
</tr>
<tr>
<td>G2</td>
<td>Wt263K</td>
<td>11/12</td>
<td>12/12</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wt</td>
<td>Wt263K</td>
<td>9/10</td>
<td>10/10</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

a Brain material from two 263K-inoculated G2 mice and one 263K-inoculated wild-type mouse with pathological evidence of disease were further passaged into G2 and wild-type mice.
b Incubation time for mice with clinical signs and evidence of TSE vacuolation. Studies were maintained to ~700 dpi for mice with no clinical signs of TSE disease.
c TSE vacuolation and/or PrP deposition were used as evidence of transmission for primary passage due to low numbers of mice with clinical signs and vacuolation present. Limited confirmatory immunohistochemistry for PrP deposition was carried out in the second passage due to high numbers of mice with clinical signs and vacuolation present.
d Survival time of mice with no clinical signs and evidence of TSE vacuolation and/or PrP deposition.
showed a similar trend in vacuolation intensity throughout the brain (Fig. 1A and B). The lack of the second glycosylation site in the host appears to greatly facilitate the infection with sCJD, suggesting that the diglycosylated form of host PrP is important in maintaining the species barrier of a predominantly monoglycosylated strain.

Diglycosylated PrPSc in the host influences incubation time in the interspecies transmission of vCJD. Variant CJD PrPSc has

![Image of lesion profile analysis](image1)

**FIG 1** Lesion profile analysis of wild-type (Wt) and G2 mice after intracerebral inoculation with sCJDMM2 (A), second passage of sCJDMM2 (B), vCJD (C), and second passage of vCJD (D). The second passage was carried out from selected G2 and wild-type mice showing TSE vacuolation and/or PrP. Group size, n ≥ 6 (± standard errors of the means). Gray matter scoring regions are labeled G1 to G9: G1, dorsal medulla; G2, cerebellar cortex; G3, superior colliculus; G4, hypothalamus; G5, thalamus; G6, hippocampus; G7, septum; G8, retrosplenial cortex; G9, cingulate and adjacent motor cortex. White-matter scoring regions are labeled W1 to W3: W1, cerebellar white matter; W2, mesencephalic tegmentum; W3, pyramidal tract.

![Image of PrP deposition](image2)

**FIG 2** PrP deposition in the brains of wild-type (Wt) and G2 mice after intracerebral inoculation with sCJDMM2, vCJD, or 263K agent. (A) G2 mouse inoculated with sCJDMM2; (B) G2 mouse inoculated with vCJD; (C) G2 mouse inoculated with 263K; (D) wild-type mouse inoculated with sCJDMM2; (E) wild-type mouse inoculated with vCJD; (F) wild-type mouse inoculated with 263K. Arrows indicate examples of PrP accumulation in the form of plaque-like deposits in panels A and B and examples of fine punctate PrP accumulation in panels C, E, and F. No PrP accumulation was detected in panel D. PrP was detected with 6H4 antibody. dg, dentate gyrus; cc, corpus callosum. Scale bars, 500 μm.
Glycosylation and Cross-Species TSE Transmission

FIG 3  Biochemical analysis of PrPSc from the brains of G2 and wild-type (WT) mice inoculated with sCJDMM2, vCJD, and 263K. (A) PrPSc was isolated from four sCJD-challenged G2 mouse, one vCJD-challenged G2 mouse (assayed in duplicate), and one vCJD-challenged wild-type mouse (assayed in duplicate) by standard PK digestion. PrPSc was concentrated by NaPTA precipitation. The amount of PrPSc in the vCJD-challenged G2 sample was too low to detect. Uninfected mice and a 263K-challenged wild-type mouse were included as controls. (B) PrPSc was isolated from vCJD-challenged G2 mice by standard PK digestion followed by NaPTA precipitation. The amount of PrPSc in the vCJD-challenged G2 sample was too low to detect. Uninfected mice were included as controls. (C) PrPSc was isolated from three 263K-challenged G2 mice and three wild-type mice by standard PK digestion. Uninfected mice and a 263K-challenged hamster were included as controls. The different isoforms of PrP are denoted Di (for diglycosylated), Mono (monoglycosylated), and Un (unglycosylated). PrPSc was detected with 8H4.

To further investigate the effect of the second site on vCJD transmission, we inoculated brain material from two G2 mice (G2-vCJD with monoglycosylated PrPSc) and a wild-type mouse (WT-vCJD with fully glycosylated PrPSc) previously infected with vCJD into G2 and wild-type recipients. Wild-type mice expressing fully glycosylated PrPSc developed clinical disease within 192 ± 2 days and 193 ± 3 days of infection with two independent G2-vCJD isolates. Two groups of G2 mice that were challenged with these isolates developed clinical disease later than the wild-type controls (283 ± 3 days and 315 ± 3) (Table 2). The same pattern also was observed when mice were challenged with the WT-vCJD brain material; G2 mice developed a clinical disease later (334 ± 6 days) than wild-type mice (156 ± 1 days) (Table 2). Despite changes in incubation times, the pattern of TSE vacuolation appeared to be similar between G2 and wild-type mice and between primary and secondary passages of vCJD (Fig. 1C and D).

Glycosylation compatibility between the hamster strain 263K and host PrPSc reduces the species barrier. Primary transmission was carried out with the hamster TSE strain 263K. PrPSc associated with the 263K agent is predominately diglycosylated. In contrast to vCJD, wild-type mice do not develop a clinical disease with this TSE strain but do show evidence of PK-resistant PrPSc in the brain following inoculation (39, 47). After challenge with 263K, no wild-type or G2 mice exhibited clinical disease, consistent with previous data. However, both G2 and wild-type mice demonstrated pathological signs of a subclinical infection (G2, 11/18; wild type, 11/18) (Table 3 and Fig. 2C and F). PK-resistant PrPSc also was detected in the brains of these mice by Western blotting (Fig. 3C). The PrPSc glycoprofile in these samples is consistent with the majority of PrPSc being monoglycosylated in G2 recipients and fully glycosylated in wild types.

G2 and wild-type mice were challenged by an i.c. route with brain homogenate from two 263K-inoculated G2 mice (G2-263K with monoglycosylated PrPSc) and a 263K-infected wild-type mouse (WT-263K with fully glycosylated PrPSc). In contrast to the vCJD transmissions, G2 mice had a higher susceptibility to disease after challenge with G2-263K than wild-type mice; all G2 mice succumbed to disease after challenge, whereas 10/12 (with the first G2 brain) and 4/10 (with the second G2 brain) wild-type mice developed clinical signs of TSE disease after G2-263K infection (Table 3). Moreover, G2 mice developed disease more quickly (284 ± 7 days and 320 ± 7 days) than wild-type mice (437 ± 32 days and 536 ± 6 days) after challenge with G2-263K (Table 3). The differences in the attack rate and incubation periods elicited by the two isolates of G2-263K may occur because the two animals from which they were derived were at different stages of subclinical infection; hence, they had different titters of infectivity within their brains. The G2 mice challenged with WT-263K developed disease later (365 ± 0 days) than wild-type mice (172 ± 5 days) (Table 3). Thus, 263K transmits with similar efficiency in the G2 mice and the wild-type mice in the primary passage, but unlike vCJD on the second passage following G2 transmission, this agent was considerably faster in the G2 host than in the wild-type host.

PrPSc expression is reduced in glycosylation-deficient mice. To ascertain whether the protein levels of PrPSc in any of the lines of mice could influence the incubation times in the transmissions described above, we undertook analysis of the PrPSc levels in all mouse lines. Immunoblot analysis showed the expected banding...
of PrPC than wild-type controls. Therefore, this enhanced susceptibility to TSE disease, with reduced levels of the protein resulting in longer incubation periods observed in this model after challenge with vCJD, the infected monoglycosylated PrPSc, allowing replication of the infective agent. This is the first time that glycosylation-deficient transgenic mice have shown an enhanced susceptibility to TSE infection compared to that of wild-type mice. This suggests that glycosylation at the second glycosylation site can protect against transmission both between and within species.

Experimental transmissions from wild-type or G2 mice infected with the 263K strain provide additional evidence that similar glycosylation statuses of host PrPSc and the PrPSc in the inoculated strain can greatly accelerate TSE incubation periods. Indeed, the incubation period in G2 recipients was almost half that of wild-type mice after challenge with the G2-263K strain.

In both primary and secondary passages of vCJD, incubation periods were shorter in wild-type mice than in mice in which the second PrPSc N-glycan attachment site was disrupted. The shorter incubation periods were observed irrespective of the glycosylation status of the second site in the infecting PrPSc. While these differences in incubation time can be explained on the basis of lower PrPSc expression levels in the G2 mice, we cannot discount the possibility that it indicates a preference of this strain for a PrPSc diglycosylated host irrespective of the passage history of the strain. This may explain the ability of this agent to infect a large number of host species and its transmissibility across many species barriers.

G1 and G3 mice showed little susceptibility to infection throughout this study. Indeed, these transgenic mice did not develop any pathologically confirmed clinical TSE disease after inoculation with any of the three agents used, although asymptomatic infection in the form of PrP deposition was detected in extremely low numbers. This may be linked to an inability of this particular host PrPSc to propagate nonmurine strains; previous experiments performed with a number of mouse-adapted scrapie strains by several routes have highlighted an intrinsic resistance of both G1 and G3 mice to infection (23, 52). Therefore, it is more likely that the resistance observed in G1 and G3 mice in this study is linked to a more general mechanism rather than an effect of the species barrier. Why the absence of the first glycosylation site should lead to such a dramatic loss of host susceptibility may be related to the conversion efficiency of PrPSc to PrPSc. Some in vitro conversion assays have previously suggested that glycosylation inhibits the conversion activity (30). However, such in vitro systems have not revealed the complexity of the glycosylation issue observed in these in vivo studies. The resistance observed in the G3 mice is linked to the absence of the G1 glycosylation. However, G3 mice also show more C1-truncated PrPSc upon biochemical analysis than G1, G2, and wild-type mice. Previous in vitro studies have shown that higher levels of C1 PrPSc are associated with resistance to TSE infection (53). In addition, G3 mice show the lowest PrPSc expression of the three glycosylation mutants and a different PrPSc localization (22). All of these factors might contribute to the resistance to TSE infection of this specific line of mice.

The absence of glycans at the second site may alter the biology of PrPSc or PrPSc interaction in a very different way than that of the first glycosylation site. A number of biochemical properties and the cellular localization of PrPSc in the G2 mice resemble that ob-

**DISCUSSION**

Expression of PrPSc is known to influence incubation times of a TSE disease, with reduced levels of the protein resulting in longer incubation periods (48). Earlier studies showed conflicting results over whether PrPSc expression levels are altered within the glycosylation transgenic mice (22). This most likely was due to the epitope recognition of the antibodies used and detection of only a subset of isoforms. Our expanded studies here, using a range of monoclonal antibodies within the C-terminal and central region of PrPSc, are able to detect all isoforms of PrPSc, demonstrating that G1, G2, and G3 mice do have lower levels of PrPSc expression than wild-type mice. However, while lower levels of PrPSc in the G1 and G3 mice may contribute to longer incubation times, the levels observed in these mice are not likely to explain the resistance to TSE disease observed here. Studies have shown that mice heterozygous for PrPSc expression and with a level of PrPSc expression similar to that of the G1 mice are fully susceptible to TSE disease, albeit with incubation periods of almost twice that of wild-type mice (48–50). Our studies were maintained to approximately 700 dpi, almost twice the incubation period of sCJD in G2 mice, which also show 50% PrPSc expression. Thus, factors other than a reduction in PrPSc expression level are likely to contribute to the resistance of these mice to TSE disease. While the lower expression of PrPSc in G2 mice may contribute to the longer incubation period observed in this model after challenge with vCJD, the G2 mice are more susceptible to infection with the sCJDMM2- and G2-passaged 263K TSE agents despite expressing lower levels of PrPSc than wild-type controls. Therefore, this enhanced susceptibility can be directly attributed to the altered glycosylation status of the host.

The monoglycosylated sCJDMM2 agent was transmitted to a normally resistant host (51) by removal of the glycans at PrP residue 196 (as removed in G2 mice). Moreover, sCJDMM2 became adapted in the G2 host and produced very short incubation times on the second pass. The data suggest that the presence of glycans at PrP residue 196 (as present in G1 or wild-type mice) is responsible for the sCJDMM2 transmission barrier; removal of this site may facilitate the interaction between host monoglycosylated PrPSc and the infective monoglycosylated PrPSc, allowing replication of the infective agent. This is the first time that glycosylation-deficient transgenic mice have shown an enhanced susceptibility to TSE infection compared to that of wild-type mice. This suggests that glycosylation at the second glycosylation site may protect against transmission both between and within species.

The absence of glycans at the second site may alter the biology of PrPSc or PrPSc interaction in a very different way than that of the first glycosylation site. A number of biochemical properties and the cellular localization of PrPSc in the G2 mice resemble that ob-

**FIG 4** PrPSc expression levels in the brains of glycosylation-deficient transgenic mice. Representative Western blots showing the different PrPSc isoforms in wild-type (Wt), G1, G2, and G3 mice using BC6 (A) and BH1 (B) antibodies. Western blots underwent densitometry to measure levels of PrPSc. α-Tubulin was used as a loading control. The different isoforms of PrPSc are denoted Di (for diglycosylated), Mono (monoglycosylated), Un (unglycosylated), and C1 frag. (C1 fragments).
served in wild-type and G1 mice (22). However, the presence/absence of carbohydrates in a specific portion of PrPSc may influence other characteristics, such as the ultrastructural localization of PrPSc (e.g., localization in a different portion of the cell membrane) or its conformation, and this may dictate the different susceptibility to infection of the G2 mice compared to that of the G1 mice.

We have argued that altered glycosylation status of PrPSc alters the host susceptibility. An alternative explanation is that the point mutations inserted in order to modify the N-linked glycosylation sites on PrP are the cause of this change (22). Previous transmission studies performed by us (23) and Neuendorf et al. (20) have shown similar results upon primary passage of both ME7 and mouse BSE strains with prolonged incubation periods in mice deficient at the first glycosylation site despite utilizing different amino acid substitutions and expressing different levels of PrP. In addition, Ikeda et al. (54) showed that substitution of Asn residues to abolish glycosylation sites does not prevent conversion of PrPc to PrPSc. In this study, the differences between the wild-type and G2 hosts in susceptibility to primary passage with two human agents, vCJD and sCJDMM2 (characterized by an identical PrPSc sequence and PK cleavage pattern but a different glycoprofile), further argues for the glycosylation status being the main determinant of host susceptibility rather than the change in amino acid sequence.

The deposition of PrP in the brains of G2 mice infected with vCJD differed from that observed in wild-type mice infected with the same agent. First, the total amount of PrP that accumulated by disease endpoint appeared to be lower in G2 mice. This could be due to less PrPSc being available for replication, or it could mean that the rates of misfolding, clearance, and/or toxicity of PrP are changed in the absence of glycosylation at the second site. In addition, small PrP aggregates were observed in G2 mice infected with vCJD, in contrast to the diffuse PrP deposition observed in wild-type mice. Large aggregated deposits of PrP also were observed in G2 mice challenged with sCJDMM2. These data suggest that PrPSc that lacks the second glycosylation site has altered misfolding or clearance kinetics, which also may have an important effect on host susceptibility.

In summary, we propose that the transmission of TSE agents across different species can be profoundly influenced by post-translational events in both PrPc and PrPSc. In particular, we have demonstrated that the glycosylation status of host PrPSc (55, 56) can dramatically alter cross-species transmission characteristics and likely is important for this protein to act as a receptor for the incoming TSE agent.

On the other hand, the prevalence of certain PrPSc glycotypes in an infectious inoculum may determine its conformation and the ability to interact with the host and cause a TSE infection. This combination may lead to the binding between PrPSc and PrPc occurring through direct interactions between the glycan residues and/or different PrP regions that have been recently suggested to be important for TSE transmission between different species (57) or by interactions with a number of conversion cofactors previously suggested, such as host proteins or nucleic acids (58–60).

The dramatic effects in altered host susceptibility, in particular the resistance of the G1 and G3 mice to infection, suggests this mechanism provides an important focus for blocking the disease process and protecting the infected individual from neurodegeneration.

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