Intersectional Gene Expression in Zebrafish Using the Split KalTA4 System

Citation for published version:
Almeida, RG & Lyons, DA 2015, 'Intersectional Gene Expression in Zebrafish Using the Split KalTA4 System' Zebrafish. DOI: 10.1089/zeb.2015.1086

Digital Object Identifier (DOI):
10.1089/zeb.2015.1086

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Zebrafish

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Intersectional Gene Expression in Zebrafish Using the Split KalTA4 System

Rafael Gois Almeida¹–³ and David Anthony Lyons¹–³

Abstract

In this study, we describe the adaptation of the split Gal4 system for zebrafish. The Gal4-UAS system is widely used for expression of genes-of-interest by crossing driver lines expressing the transcription factor Gal4 (under the control of the promoter of interest) with reporter lines where upstream activating sequence (UAS) repeats (recognized by Gal4) drive expression of the genes-of-interest. In the Split Gal4 system, hemi-drivers separately encode the DNA-binding domain (DBD) and the activation domain (AD) of Gal4. When encoded under two different promoters, only those cells in the intersection of the promoters’ expression pattern and in which both promoters are active reconstitute a functional Gal4 and activate expression from a UAS-driven transgene. We split the zebrafish-optimized version of Gal4, KalTA4, and generated a hemi-driver encoding the KalTA4 DBD and a hemi-driver encoding KalTA4’s AD. We show that split KalTA4 domains can assemble in vivo and transactivate a UAS reporter transgene and that each hemi-driver alone cannot transactivate the reporter. Also, transactivation can happen in several cell types, with similar efficiency to intact KalTA4. Finally, in transient mosaic expression assays, we show that when hemi-drivers are preceded by two distinct promoters, they restrict the expression of an UAS-driven reporter from a broader pattern (sox10) to its constituent smaller neuronal pattern. The Split KalTA4 system should be useful for expression of genes-of-interest in an intersectional manner, allowing for more refined manipulations of cell populations in zebrafish.

Introduction

Ectopic expression of genes-of-interest (GOI) in cells is a fundamental method used to understand the function of genes and the biology of cells. Driving expression of a fluorescent protein, for instance, illuminates the target cells’ morphology, origin, and behavior.¹ Expression of optogenetic tools permits interrogation or manipulation of the physiological state of the target cell.² Expressing wild-type or mutant alleles of GOI helps elucidate their role in the specification, development, or function of target cells or tissues.³ In recent efforts to systematically generate mutations in each protein-coding gene of the zebrafish genome, it was determined that over 80% of nonsense or essential splice mutations, representing over 1200 genes, have no overt morphological or behavioral phenotype within 5 dpf.⁴ Functional data for many of these genes may arise from studies resorting to ectopic expression of their wild-type and mutant forms.⁵ One manner in which to direct gene expression to target cells is to use upstream regulatory sequence fragments, typically from promoters, known to be expressed in the desired cell type. Such constructs are typically injected into fertilized eggs, resulting in transient, mosaic expression in the target cells. These animals can be raised and their offspring assessed for genome integration of the construct, which results in stable, nonmosaic expression in all target cells in the animal.⁶ A number of approaches greatly increase the efficiency of genome insertion (see Refs.⁶,⁷). The pattern of gene expression in such stable transgenic lines needs to be compared to that of the endogenous gene, as artifacts in the pattern of gene expression can be caused by the incomplete promoter fragments used and presence of vector sequences,⁸,⁹ or by positional effects of transgene insertion into the genome.⁸,¹⁰ Researchers can also target gene expression by resorting to the bipartite Gal4/UAS system, amply used in Drosophila³,¹¹

¹Centre for Neuroregeneration, University of Edinburgh, Edinburgh, United Kingdom.
²MS Society Centre for Translational Research, University of Edinburgh, Edinburgh, United Kingdom.
³Euan MacDonald Centre for Motor Neurone Disease Research, University of Edinburgh, Edinburgh, United Kingdom.

© Rafael Gois Almeida, et al. 2015; Published by Mary Ann Liebert, Inc. This Open Access article is distributed under the terms of the Creative Commons License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.
A striking limitation of both approaches is that gene expression is ultimately under the control of a single promoter, which may not achieve the required cell type specificity. In fact, existing Drosophila or zebrafish Gal4 driver lines, whether enhancer-trapped or genetically engineered to drive Gal4 expression by a known promoter, rarely confine expression to a single cell type.\textsuperscript{19,20} This may preclude the analysis of the cell autonomy of the effects of gene expression, for instance, or may not enable visualization of a particular cell type with the necessary specificity, if related or nearby cell types also express a reporter protein.

To overcome this limitation, ternary systems such as the Split Gal4 system have been developed (Fig. 1B).\textsuperscript{21,22} This system takes advantage of the modularity of Gal4 domains, whereby the DNA-binding domain (DBD) can be encoded separately from the activation domain (AD) in so called hemi-drivers. Each domain is not functional on its own, but when coexpressed reconstitute Gal4 function and activate expression of UAS-driven transgenes. When each hemi-driver is encoded under the control of a different promoter, only those cells in which both promoters are simultaneously active synthesize both hemi-drivers. Each hemi-driver contains a heterodimerizing leucine zipper that ensures that where coexpression happens, the two hemi-drivers dimerize and reconstitute a functional Gal4, which can bind UAS and initiate transcription.\textsuperscript{21} This allows gene expression in the intersection of two expression patterns, when a single promoter does not provide enough specificity, allowing dissection of complex expression patterns by iterative combinations with more specific promoters. The split Gal4 system has been recently used to refine gene expression to specific neuron subsets and correlate them to a particular function or behavior (see examples in Drosophila studies here\textsuperscript{23-29}).

In this study, we describe an adaptation of the Split Gal4 system for use in zebrafish, which we term Split KalTA4, since we based it on the optimized version of Gal4, KalTA4.\textsuperscript{30} Compared to the original Gal4 constructs, KalTA4 shows robust zebrafish expression due to a number of optimal modifications: (1) it includes a strong Kozak sequence and a rabbit \(\beta\)-globin intronic sequence that increase expression levels; (2) codon usage is optimized throughout for translation in zebrafish; (3) the entire VP16 AD is replaced with attenuated repeats of the VP16 core sequence, TA4, which is similarly potent in transgene activation, but less toxic. We design KalTA4-based hemi-drivers and confirm that they work in the developing zebrafish, and we provide an example where the use of split KalTA4 allowed the dissection of a complex expression pattern by restricting reporter expression to a subset of cells in that pattern.

**Materials and Methods**

**Fish husbandry**

The following wild-type and transgenic zebrafish lines were used: AB; Tg(sox10(7.2):mRFP)\textsuperscript{31}; Tg(olig2:EGFP)\textsuperscript{32}, Tg(mbp:EGFP)\textsuperscript{33}; and Tg(UAS:kaede).\textsuperscript{18} For this study, we generated the transgenic line Tg(sox10(7.2):KalTA4GI). Throughout the text and figures, we refer to Tg(sox10(7.2):
mRFP) as Tg(sox10:mRFP) and Tg(sox10(7.2):KalTA4) as Tg(sox10:KalTA4) for simplicity. All animals were maintained in accordance with the United Kingdom Home Office guidelines.

**Generation of DBD hemi-driver**

The coding sequence for the original DBD hemi-driver\(^\text{21}\) encodes a heterodimerizing leucine zipper (Zip), a decaglycine linker (gly), and the DBD of Gal4 (Gal4DBD, residues 1–147), as depicted in Figure 1C. Residues 1–147 are identical in Gal4 and KalTA4, but the coding sequence has been optimized for zebrafish expression in KalTA4.\(^\text{30}\) We polymerase chain reaction (PCR)-amplified KalTA4’s DBD coding sequence from pCSKalTA4\(^\text{30}\) using Phusion High-Fidelity DNA Polymerase (New England Biolabs) and added the remaining features by consecutive extension PCRs, using the primers described in Table 1. Following each round, we purified the PCR product of the expected size using the QIAquick Gel Extraction Kit (Qiagen) and used it as a template in the subsequent round. In Table 1, italics denote KalTA4’s DBD coding sequence, bold denotes the ‘Zip’ sequence, and underlined is the ‘gly’ sequence. DBD, DNA-binding domain; PCR, polymerase chain reaction.

**Generation of TA4 hemi-driver**

The coding sequence for the original Gal4 AD or VP16 hemi-drivers\(^\text{21}\) encodes a long SV40 nuclear localization signal (“nls”); the transcription-AD of Gal4 or VP16 (AD); a decaglycine linker (gly); and a leucine zipper complementary to that in the DBD hemi-driver (Zip), as depicted in Figure 1C. In KalTA4, which has been optimized for zebrafish,\(^\text{30}\) the AD consists only of the core repeats of the VP16 AD, TA4. We PCR-amplified the sequence encoding TA4 (residues 154–192) from pCSKalTA4\(^\text{30}\) using Phusion High-Fidelity DNA Polymerase and added the remaining features by consecutive extension PCRs, using the primers described in Table 2. Following each round, we gel-purified the PCR product of the expected size and used it as a template in the subsequent round. In Table 2, italics denote the TA4 coding sequence, bold denotes the Zip sequence, and underlined is the gly sequence. Round 1 adds a short SV40 nuclear localization signal (delimited by dashes). Round 2 adds a start codon and strong Kozak sequence from KalTA4. In the last round of PCR we added attB1 and attB2R sequences (lowercase) and used BP clonase to recombine it with pDONR221 to generate a Gateway middle-entry vector, pME-TA4.

During primer design, codon usage was optimized for zebrafish expression by using the most frequently used codon for each particular residue according to Yarden.\(^\text{34}\) Constructs were verified by Sanger sequencing (Source BioScience).  

![Table 1](https://example.com/table1.png)

<table>
<thead>
<tr>
<th>Round</th>
<th>Primers (5'-3')</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F: GCTACGGCCCTCCTGGGCGGCGGCGGAAGGCTAGCGGAGGAGGCCGGCGCGGGAGGC GGGACACTAGTATAGAAAACTGCTCTCATTCCAG&lt;br&gt;R: ggggacatcggccacagagctggcTTACGAAACAGTACAGTCTCTGTCTC</td>
</tr>
<tr>
<td>2</td>
<td>F: CCGAGGGCGAGCTGAGGACAGAGGTAGGAGGCCGGCTTGGAGAAACGGAGGTAGGAGCCA&lt;br&gt;GTACGAGCGCTACGCGCCCTGGGCGGCGGC&lt;br&gt;R: same as Round 1</td>
</tr>
<tr>
<td>3</td>
<td>F: GCCGCACACATGCTAGAATCCCGCGCGCTTCCTGGCGCGAGCGCAACACCGCCTG&lt;br&gt;CGACCGAGGTGGGCGAGCGGAGG&lt;br&gt;R: same as Round 1</td>
</tr>
<tr>
<td>4</td>
<td>F: ggggacatcggccacagagctggcTTACGAAACAGTACAGTCTCTGTCTC&lt;br&gt;R: same as Round 1</td>
</tr>
</tbody>
</table>

**Table 2. Primers Used for Generating TA4 Hemi-Driver by Extension PCRs**

<table>
<thead>
<tr>
<th>Round</th>
<th>Primers (5'-3')</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F: CCACCATGCCATCCAAAGAAGAGAAGGCTAGGTAAGCT-GCCCGAGCCCGCTTG&lt;br&gt;R: CCAGGAAGCGGCGCTCGATCCAG&lt;br&gt;GCTAGGCAGAGTTGCTAGTCTCG</td>
</tr>
<tr>
<td>2</td>
<td>F: ggggacatggtgatcaaaagagcgcctGCCCGACATAGGAA&lt;br&gt;R: GGCAGGCGCTGCAAGCCGGCTG&lt;br&gt;TCGCCAGTCCAGGAAAGCGCGCTG</td>
</tr>
<tr>
<td>3</td>
<td>F: same as Round 2&lt;br&gt;R: TTAACGTGCGCCGCGCGCAGGGGCGTG&lt;br&gt;GGCGGCTGACGCGGCTGG</td>
</tr>
<tr>
<td>4</td>
<td>F: same as Round 2&lt;br&gt;R: ggggacatcggccacagagctggcTTACGAAACAGTACAGTCTCTGTCTC</td>
</tr>
</tbody>
</table>

Italics denote the TA4 coding sequence, bold denotes the ‘Zip’ sequence, and underlined is the ‘gly’ sequence.
LifeSciences), and are available upon request, as are their sequences.

Generation of DBD and TA4 synthetic mRNAs

The Kozak and coding sequences for the DBD and TA4 hemi-drivers were PCR-amplified using Phusion High-Fidelity DNA Polymerase and cloned into pCS2+ digested with BamHI and Klenow blunted. Constructs were verified by Sanger sequencing. Synthetic mRNA was produced by linearizing the templates with NorI and transcribing from the SP6 promoter using SP6 mMESSAGE mMACHINE (Ambion).

Gateway recombination of split KalTA4 hemi-drivers

We used LR clonase II Plus (Life Technologies) to recombine 5′-entry vectors containing a fragment of the h2z (Kwan et al.), sox10 (Kirby et al.), or elavl3 (formerly huC) (Park et al.) promoters; middle-entry vectors pME-DBD, pME-TA4, or pME-KalTA4GI; a 3′-entry vector containing a poly adenylation signal from the Tol2kit and destination vectors pDestTol2pA2 or pDestTol2CG2 (for sox10(7.2):KalTA4GI line) to generate the constructs used in this study. All entry vectors were verified by Sanger sequencing and all recombined constructs were screened by at least three restriction enzyme digestions.

Microinjection

For DNA injections, 2–10 pg of DNA of each construct with 30–50 pg of tol2 transposase mRNA were microinjected in a volume of 1 nL into the cell(s) of fertilized UAS:kaede eggs (1–4 cells stage). For synthetic mRNA injections, 100 pg of each mRNA in water was microinjected in a volume of 1 nL into the yolks of fertilized Tg(UAS:kaede) eggs (eggs injected with both hemi-drivers contained a total amount of 200 pg mRNA). The injection solution contained 0.05% Phenol red for visualization.

For the generation of the stable transgenic line Tg(sox10(7.2):KalTA4GI), plasmid DNA encoding sox10:KalTA4 and tol2 transposase mRNA were injected as described in one-cell stage AB fertilized eggs. Successful integration of sox10:KalTA4 into the genome was detected by cmcl2:EGFP expression in the founder with germ-line transmission of sox10:KalTA4 was found and the stability of a genetic transmission was confirmed by germ-line injection.

Results

Design of split KalTA4 hemi-drivers

To generate a conditional expression system in zebrafish where GOI are expressed in the intersection of expression patterns, we adapted the split Gal4 system developed for Drosophila, where the DBD and the AD of the transcription factor Gal4 are encoded in separate hemi-drivers. To facilitate assembly of a functional transcription factor when hemi-drivers are coexpressed, Luan et al. fused a heterodimerizing leucine zipper to each hemi-driver (Fig. 1B, C). We designed hemi-drivers based on the original sequences, splitting instead KalTA4, a version of Gal4 optimized for zebrafish (see Fig. 1C for construct design).

We used the sequence encoding residues 1–147 of KalTA4 as a starting point for the DBD hemi-driver, and residues 154–192 as a starting point for the TA4 hemi-driver, and added the remaining features by extension PCR (see Materials and Methods section for details). In addition to the decaglycine linker and the heterodimerizing leucine zippers, we changed the original nuclear localization signal (nls) sequence of the AD hemi-driver to the shorter SV40 nls sequence. Similar to the KalTA4 constructs, we also included a strong Kozak sequence in both hemi-drivers, and also optimized codon usage in all features for expression in zebrafish. In our constructs, only the plasmids encoding intact KalTA4 (positive control) included the rabbit β-globin intron downstream of the coding sequence, which is a modification reported to increase the expression originally included in the design of KalTA4. All constructs were verified by Sanger sequencing.

Expression of both hemi-drivers is required to transactivate a reporter transgene in vivo

We first determined whether the modifications we inserted into the system worked in vivo. We sought to ubiquitously express both hemi-drivers in zebrafish embryos. We injected synthetic mRNA of each hemi-driver in isolation or combined, and of intact KalTA4, into fertilized Tg(UAS:kaede) eggs. mRNA injection results in ubiquitous expression in the early embryo, and if the split KalTA4 hemi-driver proteins assemble into a functional KalTA4, it will bind the UAS fluorescence from 24 hpf in a Nikon SMZ1500 stereomicroscope equipped with a Nikon Intensilight Illuminator. Flourescence intensity measurements were performed in Fiji. Briefly, two to four optical sections of the z-stack encompassing the soma of randomly selected muscle or neuronal cells were z-projected. A threshold was applied to select the soma of the cells, and the measure tool was used to determine the Mean Gray Value, reflecting the average fluorescence intensity across the entire soma area.

Live imaging

The mRNA-injected embryos were screened for Kaede fluorescence from 24 hpf in a Nikon SMZ1500 stereomicroscope equipped with a Nikon Intensilight Illuminator. Plasmid-injected embryos/larvae were embedded in 1.5% low melting point agarose in 10 mM HEPES-buffered E3 embryo medium with tricaine (168 μg/mL, 3-amino-benzoic acid ethyl ester) and screened for Kaede+ fluorescence using a Zeiss Axio Imager A1 microscope equipped with a 20× objective. A check for Kaede photoconversion from green to red fluorescence was performed using UV illumination for 30–60 s. Select embryos/larvae were live-imaged using either a Zeiss Axio Imager Z1 equipped with an Apotome.2 unit and a 10× objective, or a Zeiss LSM710 confocal microscope equipped with a 20× objective, or a Zeiss 780 confocal microscope equipped with a 63× water immersion objective. All fluorescent spinal cord images represent a lateral view, anterior to the left and dorsal to the top. Image processing was kept to a minimum and included only cropping and global changes of brightness and contrast, and was performed in Fiji software (a distribution of ImageJ). Figure panels were produced using Adobe Illustrator CS8.
sequence and drive expression of the fluorescent protein Kaede in the embryo. When mRNA for each hemi-driver was injected alone, expression of Kaede was not detected at any stage analyzed (up to 4 dpf; Fig. 2A; DBD: 0/40 embryos; TA4: 0/60 embryos). When mRNAs for both hemi-drivers were co-injected, clear expression of Kaede was detected from at least 24 hpf (Fig. 2A, at 48 hpf, 20/31 embryos). Injection of intact KalTA4 mRNA similarly results in the activation of Kaede expression throughout the embryo (Fig. 2A, 28/41 embryos). Thus, our modifications to the split Gal4 system, which we term split KalTA4, are functional in vivo, and recognition of the UAS sequence and activation of downstream gene transcription requires the presence of both split KalTA4 hemi-drivers.

Split KalTA4 functions in a variety of cell types and with similar efficiency to intact KalTA4

We then generated DNA constructs where each hemi-driver’s coding sequence is preceded by a fragment of the histone 2-a promoter, which drives the expression in many cell types (Kwan et al.7). Similarly to the mRNA microinjections, only co-injection of h2a:DBD with h2a:TA4 or injection of the intact h2a:KalTA4 resulted in activation of the UAS:kaede reporter (h2a:DBD: 0/20 embryos; h2a:TA4: 0/20; h2a:DBD and h2a:TA4: 29/35 embryos; h2a:KalTA4: 23/45 embryos). The use of the quasi-ubiquitous h2a promoter and of DNA constructs resulted in sparser expression than mRNA injections and allowed us to identify by morphology and location the cell types expressing the reporter fluorescent protein Kaede. Similar to intact KalTA4, co-injections of the split KalTA4 hemi-drivers resulted in Kaede expression in both central and peripheral neurons, muscle cells (Fig. 2B), skin cells, and notochord cells. Thus, the hemi-drivers can heterodimerize and transactivate the reporter transgene in many cell types.

We then wondered if reconstituted split KalTA4 activated the reporter transgene with the same efficacy as intact KalTA4. We considered fluorescence intensity as a proxy for the level of UAS:kaede expression and transactivation efficiency. We measured the average fluorescence intensity of the soma area of randomly selected Kaede+ neurons or muscle cells. Intensity in Kaede+ muscle cells driven by split KalTA4 was similar to intensity in Kaede+ muscle cells driven by intact KalTA4 (34 ± 23 AU in 24 h2a:DBD + h2a:TA4 cells versus 31 ± 15 AU in 21 h2a:KalTA4 cells, p = 0.8482 in Mann–Whitney test). The intensity in Kaede+ neurons driven by split KalTA4 was slightly reduced compared to the intensity in Kaede+ neurons driven by intact KalTA4 (84 ± 42 AU in 20 h2a:DBD + h2a:TA4 neurons versus 109 ± 27 AU in 20 h2a:KalTA4 neurons, p = 0.0122 in Mann–Whitney test). This suggests that the efficiency of
Split KalTA4 transactivation of the transgene may be slightly variable between cell types, but can readily reach similar levels to intact KalTA4 in vivo.

**Split KalTA4 restricts sox10 expression pattern to smaller subsets in vivo**

One potential application of split Gal4/KalTA4 systems is to dissect complex gene expression patterns to its constituents, driving GOI expression in increasingly specific subsets of cells. The sox10 gene encodes an important transcription factor that regulates the development of many cells at several stages, including some neuronal subtypes, neural crest, its derivatives, and myelinating glia.35–38 Fragments of the promoter of the zebrafish sox10 gene drive the expression of reporter genes in several of these cell types.39 For instance, in the Tg(sox10:mRFP) line, a 7.2 kbp fragment of the sox10 promoter drives the expression of mRFP in central neurons, glia, and neural crest-derived cells.40

At 2 dpf, we find that a subpopulation of sox10:mRFP+ dorsal neurons is evident in the embryonic spinal cord (Fig. 3A, arrowheads), in addition to oligodendrocyte precursor cells (OPCs). Unlike the dorsal neurons, OPCs also express the transcription factor Olig2, and are labeled by EGFP in the Tg(olig2:EGFP) line (Fig. 3A, asterisk). Later, at 4 dpf, fewer dorsal neurons are apparent, suggesting downregulation of sox10 expression in these neurons. Many OPCs have terminally differentiated into myelinating oligodendrocytes (OLs), which retain expression of sox10 and initiate expression of myelin basic protein (mbp), becoming labeled in the Tg(mbp:EGFP) line (Fig. 3B, spinal cord). In the peripheral nervous system, myelinating Schwann cells in the pLL also express sox10 and mbp (Fig. 3B, pLL).

To exclude the possibility that such an expression pattern is specific to this Tg(sox10:mRFP) line (for instance, due to positional effects of original transgene insertion), we generated a different stable transgenic line using the same sox10 promoter fragment, driving intact KalTA4, Tg(sox10:KalTA4). When crossed with the reporter Tg(UAS:kaede), sox10+ cells are sparsely labeled in the offspring, permitting visualization of their morphology and unambiguous identification. Such sparse, variegated labeling in stable reporter UAS transgenic lines is thought to be due to epigenetic silencing of the repetitive UAS sequences,41 but in this case is useful to identify isolated cells and more completely visualize their individual morphology. Similar to the Tg(sox10: mRFP) line, analysis of the Tg(sox10:KalTA4) line at 3 and 4 dpf confirmed that within the spinal cord, sox10 is expressed in some dorsal neurons (Fig. 3C, cell a), mature OLs (Fig. 3C, cell b), and OPCs (Fig. 3C, cell c).

**FIG. 3.** Reporter transgenes indicate expression of sox10 in several cell types. (A) Lateral view of the spinal cord of 2 dpf Tg(olig2:EGFP; sox10:mRFP) larva. Arrowheads: examples of sox10:mRFP+ dorsal neurons; asterisk: example of olig2:EGFP+ sox10:mRFP+ OPC. (B) Lateral view of the spinal cord (top) and pLL (bottom) of 4 dpf Tg(mbp:EGFP; sox10:mRFP) larva. Many OPCs become mbp:EGFP+ mature OLs by 4 dpf in the spinal cord, and are associated with sox10:mRFP myelin sheaths, as are mbp:EGFP+ Schwann cells in the pLL. (C) Lateral view of the spinal cord of 3 dpf Tg(sox10:KalTA4; UAS:kaede) larva. Cell a: dorsal neuron; cell b: OL; cell c: OPC. (D) Venn Diagram of the expression patterns of sox10, olig2, mbp, elavl3, and cntn1b, indicating some of the cell types found in the intersection of each subdomain. All scale bars: 20 μm. OL, oligodendrocyte; OPC, oligodendrocyte precursor cell. Color images available online at www.liebertpub.com/zeb
We then used the split KalTA4 system to restrict expression of the reporter transgene to subtypes of sox10+ cells, in transient expression assays. Most newly differentiated neurons also express elavl3, which encodes an RNA-binding protein used as a pan-neuronal marker.42 Thus, we sought to label cells only in the intersection of the sox10 and elavl3 expression patterns (Fig. 3D); we generated hemi-driver constructs preceded by fragments of the sox10 and elavl3 promoters, and injected different combinations of the hemi-drivers in the reporter UAS:kaede line. As is typical of injected DNA constructs, these transient expression assays resulted in sparse, mosaic Kaede expression. We identified Kaede+ cells in multiple injected embryos for each combination of constructs, to establish a consensus expression pattern for each combination. Table 3 shows the number of injected larvae screened at 4 dpf with Kaede+ cells of several cell types.

First, injection of each hemi-driver alone, preceded by either the elavl3 promoter fragment or the sox10 promoter

![Fig. 4](image)

**Fig. 4.** Split KalTA4 system restricts sox10 expression pattern to each component cell type. (A) Coinjection of elavl3:DBD and elavl3:TA4 activate reporter expression in neurons. (i) Overview of two spinal cord segments showing kaede+ neurons (arrowheads indicate examples of neuron somata). (ii) High-resolution view of spinal interneurons. (iii–iv) A pLL neuron and its axon innervating neuromasts. (B) Coinjection of sox10:DBD and sox10:TA4 activate reporter expression in neurons and glia. (i, ii) Spinal cord overviews showing kaede+ neurons (arrowheads) and OLs (arrows) forming myelin sheaths (brackets). (iii) High-resolution view of ventral OL in (i). (iv) A Schwann cell in the pLL. (v) High-resolution view of a Rohon-Beard cell in the dorsal spinal cord. (C) Coinjection of sox10:DBD and elavl3:TA4 activate reporter expression only in neurons. (i, ii) Spinal cord overviews showing kaede+ neurons (arrowheads). (iii) High-resolution view of a circumferential descending interneuron. (iv) High-resolution view of Rohon-Beard neuron and of a dorsal interneuron in (ii). All panels are lateral views at 4 dpf; all scale bars: 20 µm.

<table>
<thead>
<tr>
<th>DBD</th>
<th>TA4</th>
<th>Neurons</th>
<th>Glia</th>
<th>Zero</th>
<th>No. of embryos</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>all</td>
<td>pLL</td>
<td>RB</td>
<td>B</td>
<td>RS</td>
</tr>
<tr>
<td>elavl3</td>
<td>—</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>—</td>
<td>elavl3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>sox10</td>
<td>elavl3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>—</td>
<td>sox10</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>sox10</td>
<td>sox10</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>14</td>
</tr>
<tr>
<td>elavl3</td>
<td>elavl3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>sox10</td>
<td>sox10</td>
<td>7</td>
<td>7</td>
<td>3</td>
<td>27</td>
</tr>
<tr>
<td>sox10</td>
<td>cntn1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

all, anterior lateral line; B, brain neurons; FP, floorplate cells; IN, spinal interneurons; M, muscle; Mn, motoneurons; N, notochord; OL, oligodendrocytes; pLL, posterior lateral line; RB, Rohon-Beard neurons; RS, reticulospinal neurons; SC, Schwann cells.
Conclusions

We have adapted an intersectional expression system, Split Gal4, for use in zebrafish, and named it Split KalTA4. In this ternary system, two distinct promoters drive expression of a hemi-driver each, and in those subsets of cells in which both promoters are active, hemi-drivers reconstitute the transcription factor KalTA4, resulting in activation of a UAS-driven GOI in the intersection of two expression patterns. We showed that our modifications to the system function in living zebrafish. These principally include zebrafish-specific adaptations that were part of the original KalTA4 optimization.

Careful fluorescent analysis of stable transgenic sox10 fluorescent reporters showed that in the embryonic spinal cord, a subset of dorsally located neurons also express sox10. We provided a specific example for the use of the split KalTA4 system, in which we restricted the broad sox10 expression pattern to distinct components, neuronal or glial. Researchers wishing to analyze the OL lineage cell population should bear in mind that imaging solely the sox10:mRFP line at early stages will, therefore, additionally include neuronal cell profiles. The split KalTA4 system allowed us to restrict reporter expression to the neuronal population, by combining sox10 and elav3 hemi-drivers. Furthermore, the combination of sox10- and cntn1b-driven hemi-drivers also restricted UAS-driven reporters solely to neurons, indicating the sox10+ dorsal neurons also express contactin1b. Thus, the split KalTA4 system may aid in finely resolving the expression pattern of promoter fragments.

The functionality of the Split KalTA4 system may be further expanded by generating hemi-drivers preceded by the heat-shock inducible promoter hsp70, which would additionally allow temporal control over expression of GOI. Furthermore, the hemi-drivers themselves may be further optimized: for instance, in the TA4 hemi-driver, the TA4 configuration of the core VP16 activation repeats can be replaced with configurations less efficient (TA6) or more efficient (TA2), providing an additional level of control of GOI expression. Generating hemi-drivers based on the tamoxifen-inducible Gal4-ERT protein may also allow temporal control of gene expression, while permitting the use of two specific promoters to drive expression of split KalTA4 hemi-drivers.

We urge researchers wishing to use the split KalTA4 system to establish full, stable transgenic hemi-driver lines, which should circumvent both the ectopic expression from the hemi-driver vectors in muscle and notochord cells, and the labeling mosaicism typical of transient DNA injections. Furthermore, use of UAS reporter lines with fewer non-repetitive UAS sequences should help circumvent heritable methylation-based silencing and further reduce mosaicism. Finally, we recommend that researchers screen multiple potential founders and establish stable lines only from those founder animals showing faithful expression of the transgene.

We are happy to provide middle-entry vectors pME-DBD and pME-TA4 to researchers wishing to establish their own transgenic hemi-driver lines by modular recombination with relevant promoters to target their cell populations of interest. We hope the Split KalTA4 system will be useful to dissect complex expression patterns and refine genetic manipulations in zebrafish.

Acknowledgments

The authors thank Carl Tucker, Paul Wright, John Arthur, Charli Corcoran, and Patricia Smart for fishroom support.
The authors would also like to thank Cerrar hotels for helping to provide microscope access. They thank Dr. Katy Cole for assistance with live imaging. Special thanks go to the Chien laboratory for the Tol2kit. The authors are grateful to members of the Lyons laboratory for critical reading of the article. They are grateful to Reinhard Koester for the kind gift of the KalTA4 constructs. This work was supported by a David Phillips Fellowship from the BBSRC, a Research Prize from the Lister Institute, a Senior Research Fellowship from the Wellcome Trust and an International Reintegration Grant (to D.A.L.), and a FCT doctoral studentship (to R.G.A.).

Disclosure Statement

No competing financial interests exist.

References

43. Czopka T, Ffrench-Constant C, Lyons DA. Individual oligodendrocytes have only a few hours in which to generate new myelin sheaths in vivo. Dev Cell 2013;25:599–609.