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Surface Co-Expression of Two Different PfEMP1 Antigens on Single *Plasmodium falciparum*-Infected Erythrocytes Facilitates Binding to ICAM1 and PECAM1

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Abstract

The *Plasmodium falciparum* erythrocyte membrane protein 1 (PfEMP1) antigens play a major role in cytoadhesion of infected erythrocytes (IE), antigenic variation, and immunity to malaria. The current consensus on control of variant surface antigen expression is that only one PfEMP1 encoded by one var gene is expressed per cell at a time. We measured var mRNA transcript levels by real-time Q-PCR, analysed var gene transcripts by single-cell FISH and directly compared these with PfEMP1 antigen surface expression and cytoadhesion in three different antibody-selected *P. falciparum* 3D7 sub-lines using live confocal microscopy, flow cytometry and in vitro adhesion assays. We found that one selected parasite sub-line simultaneously expressed two different var genes as surface antigens, on single IE. Importantly, and of physiological relevance to adhesion and malaria pathogenesis, this parasite sub-line was found to bind both CD31/PECAM1 and CD54/ICAM1 and to adhere twice as efficiently to human endothelial cells, compared to infected cells having only one PfEMP1 variant on the surface. These new results on PfEMP1 antigen expression indicate that a re-evaluation of the molecular mechanisms involved in *P. falciparum* adhesion and of the accepted paradigm of absolutely mutually exclusive var gene transcription is required.

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Introduction

*Plasmodium falciparum* is the most pathogenic human malaria parasite and a major cause of morbidity and mortality in Africa. Its pathogenesis is closely associated with the adhesive properties of the variable erythrocyte surface antigens, PfEMP1, encoded by the var gene family [1]. These adhesins force erythrocytes infected with the parasite to bind to host receptors such as CD36, CD31/PECAM1 and CD54/ICAM1 on the endothelial lining of venular capillaries –2–5]. The more mature, replicating stages of the parasite thus leave the peripheral circulation, perhaps to avoid elimination by the reticuloendothelial system of the host spleen. Their sequestration damages the host, as occlusion and inflammation around capillaries results in damage to vital organs and sometimes leads to potentially fatal complications such as cerebral malaria [1,6,7].

The highly polymorphic var genes are concentrated at the 28 chromosome telomeres, with a minority of loci in more central regions [8]. Switching expression of PfEMP1 antigens with alternative antigenic and cytoadherent properties allows *P. falciparum* to evade acquired immune responses and maintain infections despite antibody-mediated immune pressure [1,9].

Based on distinct types of promoter sequences and their internal versus telomeric chromosomal location, the 60 member var gene family of the sequenced 3D7 clone can be classified into related groups [10,11]. There are three major groups, (A, B, and C), two intermediate groups (B/A and B/C) and the more distantly related var1 and var2csa genes. Mutation and recombination have generated a vast repertoire of polymorphic variants and how this antigenic variation system operates during infection is a major question in malaria biology and clinical research.

The interpretation of experiments testing the relationship between var gene transcription and the antigenic and adhesive properties of IE has been that a limited number of var mRNAs, possibly only one, is ultimately expressed as surface antigen on individual infected erythrocytes [12–17]. Such “mutually exclusive” var gene expression would limit exposure of antigens to the immune system and thus serve to extend infection periods. This seems advantageous to a vector-borne parasite whose transmission is governed by unpredictable environmental conditions and host
**Author Summary**

*Plasmodium falciparum* is the most pathogenic human malaria parasite and its virulence has been linked to its capacity to express different adhesion proteins that enable the developing parasitized erythrocyte to bind to capillaries of the host, thereby avoiding removal by the spleen. Each parasite has approximately 60 genes encoding different versions of this adhesion protein, and a switch in surface display of these proteins enables the parasite to evade the immune system. Here we show that different variants of these binding proteins can be found expressed simultaneously on single infected red blood cells mediating binding to different endothelial receptors.

CD31/PECAM1 and showed a markedly increased binding of IE to human umbilical vein endothelial cells (HUVEC). This binding is reminiscent of the multiple receptor interactions shown to be important in other vascular adhesion interactions [32,33].

Interestingly, the severity of malaria disease has previously been associated with adhesion of IE to multiple receptors rather than one and clinical isolates of *P. falciparum* will bind several receptors simultaneously [34–36]. The explanation previously put forward to explain this has been that distinct domains of a single PfEMP1 molecule are interacting with multiple host receptors [37]. To our knowledge, this is the first study to demonstrate that multi-receptor binding can be mediated by different PfEMP1 antigens that are co-expressed on the same infected erythrocyte surface.

**Results**

Surface expression of PFD1235w and PF11_0008 on erythrocytes infected with selected 3D7 parasites

The expression of particular PfEMP1 surface antigens may switch during the infection and is proposed to be mutually exclusive in the sense that any single infected erythrocyte expresses only one variant antigen at a time, on the IE surface [14,16,17,38]. We have shown that 3D7 parasites in culture, pre-selected using IgG from semi-immune children are transcribing several var genes. However, human IgG selected parasitized erythrocytes predominantly expressed one Group A PfEMP1 antigen (PFD1235w) on the IE surface [39]. Using new rat and rabbit antisera specific for the proteins encoded by two different var genes, annotated as PFD1235w and PF11_0008, we re-examined var gene and PfEMP1 surface antigen expression in three different antibody-selected sub-lines of 3D7.

The 3D7 PFD1235w sub-line was selected from 3D7Dodowa1 [31,39] using an antisera targeting the DBL4γ domain of PFD1235w. The 3D7 PFD1235w/PF11_0008 and 3D7PF11_0008 sub-lines were selected from 3D7 using antisera targeting the CIDR1α domain of PFD1235w and the CIDR2β domain of PF11_0008, respectively. Each was grown in culture for only 3–5 cycles after the end of the selection period prior to the assay. We immunostained parasites for flow cytometry using antisera targeting DBL1α-CIDR1α, CIDR1β, DBL3β, DBL4γ, DBL5δ, DBL5δ-CIDR2β of PFD1235w, DBL4β and CIDR2β of PF11_0008, and DBL5ε and DBL5ε-DLB6ε of VAR2CSA and also examined individual live un-fixed single invaded IE using confocal microscopy.

The majority of erythrocytes infected with the 3D7 PFD1235w sub-line stained positively in flow cytometry with all PFD1235w antisera (Figure S1A1–A8) without expression of any PF11_0008 (Figure S1A9 and S1A10). Live confocal microscopy of individual IE showed the characteristic punctate pattern of PfEMP1 antigen staining, indicating surface expression of the PFD1235w-encoded var gene (Figure 1A1, A3, A4, A6 and Figure 2A1–A8). No PF11_0008 (Figure 1A2, A3, A5, A7 and Figure 2A1–A8) or VAR2CSA (Figure 1A4, A5, A8) antigen staining was observed.

Similarly the majority of erythrocytes infected with the 3D7PF11_0008 sub-line stained positively with antisera targeting DBL4β and CIDR2β of PF11_0008 (Figure S1B9–10) without expression of PFD1235w (Figure S1B1–B8). Confocal microscopy agreed with the flow cytometric data, indicating expression of the PF11_0008 encoded var gene (Figure 1B2, B3, B5, B7 and Figure 2B1–B8) without PFD1235w (Figure 1B1, B3–B4, B6 and Figure 2B1–B8) or VAR2CSA (Figure 1B4, B5, and B8) antigen staining.

Availability. Northern blots, nuclear run-on assays and some single-cell RT-PCR experiments clearly demonstrated transcription of several var genes in ring stages, sometimes [16–18], but not always showing a reduction in the extent of polygenic var transcription as rings develop into trophozoites [19,20]. This has been explained as the result of mRNA extraction from cell populations rather than individual cells and by indications that there is a trend, as intra-erythrocytic development progresses, for “loose” multi-var locus transcription to decrease, and a single mRNA type to become predominant, this dominant transcript becoming the sole translated and exported PfEMP1 [16,17,21]. Recent nuclear run-on data using A4 parasites, antibody-selected for expression of the A4varICAM1 PfEMP1, and also parasites preselected for adhesion to CSA or CD36 have somewhat reinforced the experimental support for the existence of predominant transcripts and mutually exclusive processes of transcriptional control [22,23].

Strictly exclusive expression is most strongly supported by the demonstration that transfection of *P. falciparum* with plasmids transcribing var promoters is followed by a shut down of all endogenous var transcription [24–28]. These experiments elegantly demonstrate that there must be some form of epigenetic memory of transcriptional status. However, although transcriptional memory is likely to be fundamental to the control of the *P. falciparum* antigenic variation system, it is less clear how well transcription mimics in vivo regulation of var gene transcription in wild-type cultures. Nor is it clear how mutually exclusive expression is naturally established e.g in parasites which have not been expressing var genes such as those emerging into the bloodstream from the liver.

A monoclonal antibody (Bc6), detecting the antigen encoded by the A4varICAM1 gene on the surface of A4 infected erythrocytes, has been used to correlate this var gene expression with its adhesion phenotype in several studies [14,15,18,29,30]. However, efforts to link other var transcripts to the surface expression of particular PfEMP1 on single infected erythrocytes have been hampered by difficulties in generating a broad repertoire of specific, surface-reactive anti-PfEMP1 antibodies.

In this work, we have generated a number of such antisera after immunising with recombinant-produced PfEMP1 domains. We have combined these with in vitro antibody-selected *P. falciparum* lines [31] to identify simultaneous surface expression of PFD1235w and PF11_0008 PfEMP1 antigens on single infected erythrocytes. Given the potential importance of dual PfEMP1 expression in vivo, we also assessed the adhesion to endothelial cells of IE with either one or two different PfEMP1 variants on the cell surface. Parasites with dual expression of both the PFD1235w and PF11_0008 antigens bound to both CD54/ICAM1 and CD31/PECAM1 and showed a markedly increased binding of IE to human umbilical vein endothelial cells (HUVEC). This binding is reminiscent of the multiple receptor interactions shown to be important in other vascular adhesion interactions [32,33].
The 3D7/PFD1235w/PF11_0008 sub-line co-expresses PFD1235w and PF11_0008 on single infected erythrocytes

Contradicting the ‘mutually exclusive’ expression model [16,17,26,27], single and single-infected erythrocytes infected by the 3D7/PFD1235w/PF11_0008 sub-line of parasites stained positively with both of the differentially labelled antisera known to bind two different PfEMP1 antigens. In confocal microscopy the PFD1235w-DBL4c and the CIDR2b PF11_0008 antisera showed an erythrocyte surface double staining pattern with punctate fluorescence, which showed limited or no co-localization (Figure 1C3 and Video S1).

The PFD1235w-DBL4c, PFD1235-CIDR1a and PF11_0008-CIDR2b antisera used for selection do not cross-react with trypsin-resistant parasite surface antigens, as prior trypsinization of the IE abolished reactivity with all the antisera used in flow cytometry (Figure S2A–C). Blocking surface binding of these antisera with excess homologous antigen prior to staining of the sub-lines also indicated that neither antisera cross-reacted with other surface antigens on IE (Figure S2D–F).

To further exclude the possibility that the double staining of single IE was an artefact resulting from cross-reactivity with other surface antigens, we used various combinations of antisera with known specificity for different domains of the PfEMP1 encoded by the PFD1235w gene (DBL1a-CIDR1a, CIDR1a*, DBL3β, DBL5β, DBL5β-CIDR2β and the PF11_0008 gene (CIDR2β and DBL4β). All of the anti-PFD1235w and anti-PF11_0008 sera were specific for the 3D7/PFD1235w and the 3D7/PF11_0008 sub-line, respectively (Figure S1). In combination with αPF11_0008-CIDR2β, each anti-PFD1235w antiserum also double stained the 3D7/PFD1235w (A1, A3, A4, A6) and 3D7/PFD1235w/PF11_0008 (C1, C3, C4, C6) sub-lines was detected using a secondary antibody labelled with Alexa 488 (green). Antiserum staining of PF11_0008 expressed by the 3D7/PF11_0008 (B2, B3, B5, B7) and 3D7/PFD1235w/PF11_0008 (C2, C3, C5, C7) was detected using a secondary antibody labelled with Alexa 568 (red). Staining of VAR2CSA expressed by NF54/VAR2CSA is red (D4, D8) and green (D5, D8).

DAPI staining of DNA in the nuclei is blue. Scale bar 5 μm.

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Figure 1. Surface expression of PfEMP1 on single 3D7 infected erythrocytes. (A) erythrocytes infected with a 3D7/PFD1235w, (B) a 3D7/PF11_0008, (C) a 3D7/PFD1235w/PF11_0008, and (D) a NF54/VAR2CSA sub-line. Localisation of PfEMP1 by confocal microscopy was done using (A1–D1) rat PFD1235w-DBL4c antisera and (A2–D2) rabbit PF11_0008-CIDR2β antisera. Double surface staining was done using the following combinations (A3–D3) rat PFD1235w-DBL4c and rabbit PF11_0008-CIDR2β antisera; (A4–D4) rat PFD1235w-DBL4c and mouse VAR2CSA-DBL5c antisera; (A5–D5) rabbit PF11_0008-CIDR2β and mouse VAR2CSA-DBL5c antisera; (A6–D6) rat PFD1235w-DBL4c and rabbit PFD1235w-CIDR1a; (A7–D7) rabbit PF11_0008-CIDR2β and rat PF11_0008-DBL4c; (A8–D8) rabbit VAR2CSA-DBL5c-DBL4c and mouse VAR2CSA-DBL5c antisera. Antiserum staining of PFD1235w expressed by the 3D7/PFD1235w (A1, A3, A4, A6) and 3D7/PFD1235w/PF11_0008 (C1, C3, C4, C6) sub-lines was detected using a secondary antibody labelled with Alexa 488 (green). Antiserum staining of PF11_0008 expressed by the 3D7/PF11_0008 (B2, B3, B5, B7) and 3D7/PFD1235w/PF11_0008 (C2, C3, C5, C7) was detected using a secondary antibody labelled with Alexa 568 (red). Staining of VAR2CSA expressed by NF54/VAR2CSA is red (D4, D8) and green (D5, D8).
fluorescently labelled antibodies, a rat antiserum raised against the DBL4\textsubscript{c} domain and rabbit antiserum raised against CIDR1\textsubscript{b} domain of the same PFD1235w antigen also showed dual staining, with limited co-localisation and distinctly labelled spots of erythrocyte surface fluorescence (Figure 1A6 and C6). Similarly, staining of the 3D7PF11_0008 sub-line using differentially labelled antibodies targeting PF11_0008 DBL4\textsubscript{b} (rat) and CIDR2\textsubscript{b} (rabbit) (Figure 1B7 and C7) and staining of the 3D7VAR2CSA sub-line using antibodies targeting DBL5\textsubscript{e} (mouse) and DBL5\textsubscript{e}-DBL6\textsubscript{e} (rabbit) of VAR2CSA also showed the dual staining, without significant co-localization phenotype (Figure 1D8).

Simultaneous surface expression of PFD1235w and PF11_0008 in clones of 3D7PFD1235w/PF11_0008 single infected erythrocytes

To obtain clonal lines of 3D7PFD1235w/PF11_0008 IE, we carried out a limiting dilution cloning exercise, following which the resulting clones were grown for 7–10 cycles, prior to flow cytometry and confocal microscopy. Simultaneous expression of PfEMP1 was monitored by flow cytometry (dot plots) and confocal microscopy (photomicrographs) (Figure 2A). The 3D7PFD1235w/PF11_0008 sub-lines was detected using a secondary antibody labelled with Alexa 488 (green). Rabbit antiserum staining of PF11_0008 expressed by the 3D7PF11_0008 was detected using a secondary antibody labelled with Alexa 568 (red). Flow cytometry settings and capture parameters for confocal microscopy were identical for all images. DAPI staining of DNA in the nuclei is blue. Scale bar 5 \textmu m.

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Luminex, ELISA, flow cytometry and confocal microscopy testing of reagents used for the surface staining of IE

Affinity purified antisera raised in rats against PFD1235w-DBL4\textsubscript{c} and in rabbits against CIDR1\textsubscript{a} of PF11_0008, as well as rabbit antiserum against PFD1235w-CIDR1\textsubscript{a}, were depleted of any anti-V5-HIS fusion protein reactivity. The specificity of the resulting antibodies was then assessed by both ELISA and a bead-based Luminex assay [40]. We found the three depleted antibody preparations to be specific for the homologous immunizing protein (Figure S4) as none cross-reacted with any of the 48 heterologous PfEMP1 domains in the Luminex (Figure S4D–F). The Luminex assay and ELISA both included the CIDR1\textsubscript{a} domain of PF08_0103, i.e. representing a var gene transcript known to be highly abundant in the 3D7PFD1235w/PF11_0008 IE tested (Figure 4).

Having tested the specificity of the antisera and of the affinity purified and depleted antibodies, we repeated the surface staining experiments presented in Figure 1 and Figure S1 and the double staining experiments shown in Figure 1 and Figure 2, on aliquots of the same batch of parasites, with the affinity purified and tag-depleted antibodies (Figure S5). The tag-depleted
PFD1235w antibodies surface stained the 3D7\textit{PFD1235w} and the 3D7\textit{PFD1235w/PF11_0008} sub-lines, but not the 3D7\textit{PF11_0008} line. Similarly, the depleted PF11_0008 antibodies surface stained the 3D7\textit{PF11_0008} and the 3D7\textit{PFD1235w/PF11_0008} sub-lines, but not the 3D7\textit{PFD1235w} line. All antibody purifications showing identical punctate PfEMP1 surface staining patterns to those observed with the complete antisera used in Figure 1 and Figure 2. Likewise, flow cytometry data using purified and depleted antibodies gave the same results as obtained using crude antisera (Figure S1 and Figure S5).

Transcription of \textit{var} genes in the 3D7 sub-lines

In parallel to the antibody-based PfEMP1 surface antigen detection studies, we measured \textit{var} gene transcript levels using primer sets targeting each of the 58 active \textit{var} genes and two pseudo \textit{var} genes of the 3D7 genome [41–43]. Only the NF54\textit{VAR2CSA} sub-line culture (Figure 4D1), showed exclusive transcription of a single \textit{var} gene (\textit{var2csa}). All other sub-lines showed the presence of several major transcripts during the ring-stages, the most transcriptionally active stages for \textit{var} gene expression (Figure 4A1–C1). The 3D7\textit{PFD1235w} sub-line culture transcribed three different \textit{var} genes in similar quantities; PFD1235w (Group A), MAL6P1.316 (Group A/B), and PFD0625c (Group C). The three most abundant \textit{var} transcripts in the 3D7\textit{PF11_0008} sub-line cultures were PF11_0008 (Group A), PF07_0050 (Group B/C), and PFD0625c (Group C). In the 3D7\textit{PFD1235w/PF11_0008} sub-line cultures the three most abundant transcripts were PFD1235w (Group A), PF11_0008 (Group A), and PF08_0103 (Group B/C).

PFD1235w and PF11_0008 constituted 19% and 1%, respectively, of the total \textit{var} transcripts in the 3D7\textit{PFD1235w} sub-line

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**Figure 3. Simultaneous surface expression of PfEMP1 on 3D7\textit{PFD1235w/PF11_0008} clone 3 infected erythrocytes.** Double surface staining and localisation of PfEMP1 by flow cytometry (dotplots) and confocal microscopy (photo inserts) was done using (A) buffer, (B) rat and rabbit pre-bleeds, (C) rat sera against PFD1235w-DBL4\textit{c} and rabbit sera against PF11_0008-CIDR2\textit{b}, (D) rabbit sera against PFD1235w-DBL4\textit{c} and rabbit sera against PF11_0008-DBL4\textit{γ}, (E) rat sera against PFD1235w-DBL4\textit{γ} and rabbit pre-bleed, (F) rat pre-bleed and rabbit sera against PFD1235w-DBL5\textit{α}-CIDR2\textit{b} and rabbit sera against PF11_0008-CIDR2\textit{b}. For confocal microscopy rat antisera staining of PFD1235w (C, E, G) and of PF11_0008 expressed by the 3D7\textit{PFD1235w/PF11_0008} clone 3 (D) was detected using a secondary antibody labelled with Alexa 488 (green). Similarly rabbit antisera staining of PFD1235w (D) and PF11_0008 (C, F, G) was detected using a secondary antibody labelled with Alexa 568 (red). Flow cytometry plots include both infected and uninfected erythrocytes (see Materials and Methods). Flow cytometry settings and capture parameters for confocal microscopy were identical for all images. DAPI staining of DNA in the nuclei is blue. Scale bar 5 µm.

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**Figure 4. Individual \textit{var} gene transcripts relative to the total \textit{var} transcript copy number in ring-stage IE.** The analysis was done on cultures of the (A1) 3D7\textit{PFD1235w}, (B1) 3D7\textit{PF11_0008}, (C1) 3D7\textit{PFD1235w/PF11_0008}, and (D1) 3D7\textit{VAR2CSA} sub-lines. A similar analysis was done on Dynabeads antibody enriched 3D7\textit{PFD1235w/PF11_0008} (A2) IE prior to enrichment, (B2) IE enriched using rabbit PFD1235w-DBL4\textit{γ} antisera, (C2) IE enriched using rabbit PFD1235w-COR1\textit{α} antisera, (D2) IE enriched using rabbit PF11_0008-CIDR2\textit{b} antisera. In another experiment we used unsorted (A3) 3D7\textit{PFD1235w/PF11_0008} IE and (B3) FACS sorted 3D7\textit{PFD1235w/PF11_0008} IE stained using rabbit PF11_0008-CIDR2\textit{b} and rat PFD1235w-DBL4\textit{γ} antisera followed by FITC- and Alexa Fluor 610-R-PE conjugated secondary antibodies as described in Materials and Methods. The 3D7\textit{PFD1235w/PF11_0008} sub-line was cloned by limiting dilution and the transcript profile of nine different clones was analysed. The transcript profiles of three representative clones are shown (A4-C4).

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similar to that seen in mRNA extracted from ring-stages of the above.

To verify that the 3D7PF11_0008 culture is a homogenous population we did different experiments to enrich for PFD1235w and PF11_0008 double positive IE. Populations of 3D7PF1235w/PF11_0008 were enriched using antisera (zpFD1235w-DBL4γ, zPFD1235w-CIDR1γ, and zPF11_0008-CIDR2β) bound to Protein-A coupled Dynabeads. FACS sorted using zPFD1235w-DBL4γ and zPF11_0008-CIDR2β simultaneously in addition to cloning by limiting dilution as described above.

The Dynabeads-enriched and FACS sorted IE were grown for less than one cycle and RNA was extracted from the ring stage. Extracted mRNA from these stages showed the presence of several major var transcripts (Figure 4B2–D2 and Figure 4B3), a profile similar to that seen in mRNA extracted from ring-stages of the unsorted 3D7PF1235w/PF11_0008 IE in Figure 4C1, 4A2, and 4A3.

Three different representative 3D7PF1235w/PF11_0008 clones (1, 3, and 5) originating from the limiting dilution experiment showed an almost identical transcript profile with the presence of several major transcripts (Figure 4A4–C4). This indicates the unsorted 3D7PF1235w/PF11_0008 culture largely is a homogenous population, a conclusion also supported by the flow cytometry data (Figure 2C1–C8).

**Full length transcription of PFD1235w and PF11_0008**

Full length transcription of 3D7PF1235w and PF11_0008 and all the var genes listed in Table S2 was tested using specific cross-intron primers. In real-time quantitative PCR assays with cDNA synthesised from total RNA extracted from the three 3D7 sub-lines and our clones we obtained similar Ct-values, whether using primers targeting exon I, or primers spanning the intron regions (Figure S6). This, in addition to Northern blotting (Figure 5K and 5L), indicates that these genes are being transcribed into full length mRNA species by the parasites. In addition, sequencing of the introns of both PFD1235w and of PF11_0008 (using genomic DNA from selected sub-line and clonal IE as template) showed they were intact and identical to the genomic sequences available at http://plasmodb.org/plasmo/. Sequencing of cDNA also showed the introns of PFD1235w, PF11_0008, and the introns of the var genes in Table S2, were correctly spliced out of the mature mRNA.

**FISH and Northern blotting analysis of PFD1235w and PF11_0008 transcripts**

Single-cell PFD1235w and PF11_0008 transcription of the two var genes located on chromosome 4 and 11 was further analysed by using RNA-FISH in situ hybridization to var gene mRNA using appropriate probes. The FISH indicates that 65–80% of single ring-stage nuclei of parasites taken from cultures of the 3D7PF1235w and 3D7PF11_0008 sub-lines transcribe either PFD1235w (Figure 5A–C) or PF11_0008 (Figure 5D–F). None of the 3D7PF1235w IE showed staining with the PF11_0008 probe and vice versa.

In agreement with the antibody-mediated PiEMP1 antigen surface detection data (Figures 1–3, Figure S1, Figure S3 and Figure S5) and the real-time quantitative PCR data (Figure 4 and Figure S6) PFD1235w and PF11_0008 could both be detected as hybridising mRNA species being transcribed in 99 of 100 different single nucleated cells of the 3D7PF1235w/PF11_0008 clone 3 (Figure 5G–I). The specificity of the probes for the two differently sized PiEMP1 mRNA species was further confirmed by Northern blotting (Figure 5K and 5L) and the identified length of mRNA agrees with that predicted as the transcript length from their respective sequenced genes (http://plasmodb.org/plasmo/). Control slides pre-treated with RNAse prior to hybridization were all negative.

When co-expressed on single IE, the PFD1235w and PF11_0008 PiEMP1 antigens mediate binding to two different endothelial receptors

Having established simultaneous expression of two species of PiEMP1 on the IE surface, we then analysed whether this dual expression affects the binding phenotype of the infected erythrocytes (Figure 6). 3D7PF1235w infected erythrocytes showed specific binding to CD54/ICAM1 transfected CHO cells (Figure 6A) and no binding to CHO-CD36 cells (Figure 6B) or wild type CHO cells (Figure 6C). The observed binding was specifically inhibited by anti-CD54/ICAM1 (15.2 and My13) antibodies (Figure 6A) which have been shown to inhibit binding of PiEMP1 to ICAM1/CD54 [44].

Erythrocytes infected with 3D7PF11_0008 did not bind CD54/ ICAM1 (Figure 6A). The 3D7PF1235w/PF11_0008 sub-line, which expressed both PFD1235w and PF11_0008, bound as strongly to CD54/ICAM1 as the PFD1235w infected cells which express only this ICAM1–binding antigen (Figure 6A). The 3D7PF11_0008 sub-line bound specifically to the un-stimulated HUVEC cells which constitutively express PECAM1/CD31 and von Willebrand factor. This binding was markedly reduced in the presence of antibodies to CD31/PECAM1 (Figure 6D), indicating that CD31/PECAM1 was important for PiEMP1-mediated binding in this assay. Cells expressing PFD1235w also showed very slight binding to un-stimulated HUVEC cells, probably explicable by a low level expression of CD54/ICAM1 on these cells as this binding was slightly reduced by CD54/ICAM1 antibodies (Figure 6D).

However cells expressing both PFD1235w and PF11_0008 bound strongly to the HUVEC cells, with binding levels 2–3 times higher than the binding of the cells expressing PF11_0008 only. This binding could only be partially inhibited by anti-CD31/PECAM1 and CD54/ICAM1 antibodies. This result indicates that cells co-expressing PFD1235w and PF11_0008 can bind both CD54/ICAM1 and CD31/PECAM1, whereas IE having only one of the two PiEMP1 species on the surface bind to one or another of the two receptors, but not both. Interestingly the binding of the 3D7PF1235w/PF11_0008 sub-line to HUVEC cells (Figure 6D) was stronger than the binding of the 3D7PF11_0008 sub-line. This could reflect an additive effect of PiEMP1 co-expression.

**Discussion**

The complete P. falciparum 3D7 genome sequence and real-time quantitative RT-PCR enable accurate measurement of the relative amounts of var gene transcripts in intra-erythrocytic malaria parasite populations. To further analyse the relationship between var gene transcripts and the individual infected erythrocyte’s PiEMP1 antigen surface expression and cytoadhesion phenotype, we have combined mRNA quantification and FISH analysis with FACs and single-cell live confocal microscopy using PiEMP1 antisera of demonstrated specificity in addition to in vitro adhesion assays, in antibody-selected, but otherwise genetically unmodified parasites. The primary intention was to test the ‘mutually exclusive expression’ hypothesis for the P. falciparum PiEMP1 mediated antigenic variation system.

A handful of studies have investigated this relationship between var transcripts and its eventual outcome in terms of surface
PfEMP1 expression and adhesion. These have used parasites selected for rosetting [16,45], adhesion to ICAM1 or CSA [15,17,19] or selected using the A4varICAM1 monoclonal antibody Bc6 [15,46]. Data supporting exclusive antigen expression on IE has been obtained using 'pan-reactive' antibodies targeting the conserved Acidic Terminal Sequence of PfEMP1, which detected single PfEMP1-sized bands in immuno-precipitations and Western blots [16]. However, similar antibodies have also shown protein expression of several differently sized PfEMP1 bands in other experiments with adhesion-selected clonal populations [19] and

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Figure 5. Nuclear transcripts of \textit{PFD1235w} and \textit{PF11_0008} in single erythrocytes. FISH analysis was done on single ring stage parasites in 100 single-cells of (A–C) 3D7\textit{PFD1235w} IE, (D–F) 3D7\textit{PF11_0008} IE and (G–I) 3D7\textit{PFD1235w/PF11_0008} clone 3 IE using a (B, E, H) \textit{PFD1235w} (green) and (C, F, I) \textit{PF11_0008} specific probe (red). The overlaying of the two colours identified transcripts of \textit{PFD1235w} in (A) 3D7\textit{PFD1235w} IE, \textit{PF11_0008} in (D) 3D7\textit{PF11_0008} IE, and simultaneous transcription of \textit{PFD1235w} and \textit{PF11_0008} in (G) 3D7\textit{PFD1235w/PF11_0008} clone 3 IE. Ethidium bromide staining (J) and Northern blotting (K and L) verification of the specificity of the probes used for FISH was done on RNA purified from 3D7 sub-lines transcribing \textit{PFD1235w} (J1, K1, L1), \textit{PF11_0008} (J2, K2, L2), co-transcribing \textit{PFD1235w} and \textit{PF11_0008} (J3, K3, L3), and FCR3 transcribing var2CSA (J4, K4, L4). The blots were probed with (K) a \textit{PFD1235w} and (L) a \textit{PF11_0008} specific probe. M: RNA molecular weight marker; < \textit{PFD1235w} transcript (10.66 kb); < < \textit{PF11_0008} 8.98 kb transcript. R: ribosomal band. Scale bar 5 μm.
immune sera-selected cultures are also known to express several species of PfEMP1 [39]. Additionally, a more recent study using a transfected A4 parasite line showed surface co-expression of miniPfEMP1 protein with endogenous PfEMP1 [47]. Direct demonstration of PfEMP1 using gel electrophoretic methods is technically difficult due to the low amounts of PfEMP1 on parasitized erythrocytes and the fact that Western blot-based typing is rarely possible because typing sera usually recognize conformational epitopes and not denatured blotted PfEMP1. However, flow cytometry and confocal microscopy can detect native PfEMP1 conformations on live cells, at both cell and population level. We used this to analyse PfEMP1 surface expression on antibody-selected IE populations known to predominantly, but not exclusively, transcribe the PFD1235w and PF11_0008 var genes.

The ring-stages of antibody selected 3D7-derived lines showed polygenic var transcription with several abundant var mRNAs (Figure 4A1–C1, Figure 4A2, and Figure 4A3). Dynabeads sorted (Figure 4B2–D2), and FACS sorted lines (Figure 4B3) also showed polygenic var gene transcription. The real-time Q-PCR data are also supported by RNA-FISH and Northern blot analysis showing co-transcription of full-length PFD1235w and PF11_0008 var genes associated with single-cell nuclei of 3D7 PFD1235w/PF11_0008 (Figure 5). By contrast, the NF54VAR2CSA sub-line had a single dominant gene, var2csa, whose transcript was 98% of total var mRNA (Figure 4D1). The dominant, exclusive var2csa transcription of repeatedly CSA selected parasites [41,42,48] is the best demonstration of ‘mutually exclusive’ var transcription. However, it is also to some extent a special case. The var2csa gene has its own promoter, upsE [10], and an unique upstream open reading frame involved in the control of var2csa expression [49] and thus its regulation has certain non-standard features [42]. Strictly monomorphic or unilocular expression of the var2csa gene has however, also been questioned by recent publications showing transcription of duplicated var2csa genes [50], which appears to be simultaneous in

Figure 6. Infected erythrocytes bind CD54/ICAM1, CD31/PECAM1 or both receptors. Binding of (1) 3D7PFD1235w, (2) 3D7PF11_0008, (3) 3D7PFD1235w/PF11_0008, (4) 3D7PFD1235w mixed 1:1 with the 3D7PF11_0008 sub-line and (5) non selected knob positive 3D7 (3D7k+) to (A) CHO expressing CD54/ICAM1, (B) CHO expressing CD36, or (C) wildtype CHO cells was done in the absence or presence of anti-CD54/ICAM1 (clone 15.2 and My13) and anti-CD36 antibodies. Similarly binding of the different 3D7 sub-lines to (D) human umbilical vein endothelial cells (HUVEC) was done in the absence or presence of anti-CD54/ICAM1 (clone 15.2 and My13), anti-CD36 or anti-CD31/PECAM1 antibodies. The data represents the mean of at least three independent experiments except for the αICAM1 (My13) experiment in B, C, and D which was only done once. Error bars are ± SD. Buffer: background control with no IE added; no Ab: binding assay done using IE, but without addition of antibodies.
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individual parasites [51]. However, dual expression of both VAR2CSA variants on the erythrocyte surface has not been shown.

We observe neither monoallelic mRNA transcription, nor monomorphic antigen expression at the erythrocyte surface in individual 3D7/PFD1235w/PF11_0008 IE. These cells clearly reacted with more than one specific anti-PiEMP1 antiserum. Other selected sub-lines reacted exclusively with either antiserum (Figure 1, Figure 2, Figure S1 and Figure S5). In our assays these antisera were highly specific for the immunizing antigen (Figure S1, S2, and S4). Our experiments appear to exclude the possibility that these antisera are non-specifically reacting with other surface expressed parasite antigens. We consider that these novel results indicate that, contrary to the current consensus, at least two different PiEMP1 antigens can in some circumstances be expressed on the surface of a single 3D7 IE.

A puzzling phenomenon associated with PiEMP1 surface staining with antigen and domain specific antisera remains. This is the observation that differentially labelled antisera raised against separate domains (Figure 1) or identical domains (unpublished data) of the same PiEMP1 protein consistently show non-colocalizing patterns of punctate surface fluorescence. This could be the result of some PiEMP1 topological phenomena, or more likely an artificial outcome of the experimental setup with cross-linking of primary via secondary species-specific antibodies [52].

Disregarding our flow cytometry and single-cell microscopy data the presence of several abundant full-length var mRNAs in the three antibody selected sub-lines (Figure 4A–C1, Figure 4A2, Figure 4A3) could be explained by that fact that mRNA was extracted from in vitro cultured cell populations rather than individual cells. To exclude this possibility we did limiting dilution of the 3D7/PFD1235w/PF11_0008 sub-line to obtain a clonal population of cells. From this experiment nine clones were obtained of which seven double positive clones show a slightly reduced complexity, but still multiplicity of var gene transcripts (compare Figure 4A4–C4 and Figure 4C1). Thus, the multiple var transcripts seen (Figure 4C1, Figure 4A2, and Figure 4A3) can not be explained by our use of a heterogeneous cell population.

At the genetic level, several DNA sequences required for appropriate var gene control have been reported e.g. the pairing of a 5′var promoter with an intronic promoter at the 3′end of the same var gene [24]. We therefore screened for potential confounding defects in our parasite lines and in clone 3. Using cross-intron primers for the abundantly transcribed var genes we found correct splicing of exon I to exon II. In addition, the real time Q-PCR for each abundant var gene transcript indicated similar transcript levels of several full-length var mRNAs in the early stage IE (Figure S6). Sequencing the intronic regions of the PFD1235w and PF11_0008 var genes located on two different chromosomes revealed that these were identical to the sequences available on PlasmoDB. Thus, constitutive transcription by single 3D7/PFD1235w/PF11_0008 IE due to some defect in their respective intron sequences does not explain our dual surface expression data. In addition as shown by flow cytometry, two of the clones switched surface expression during culturing with a minor population still co-expressing PFD1235w and PF11_0008 and a majority expressing unidentified PiEMP1s (Figure S3). This strongly indicates a shut-down of PFD1235w and PF11_0008 PiEMP1 surface expression and further indicates constitutive transcription does not explain our dual surface staining data.

Whether the simultaneous expression of more than one PiEMP1 occurs in single IE during natural infections is not known as we currently have a better understanding of malaria antigen variation in vitro than in vivo. In natural infections, P. falciparum sequesters in deep tissue prior to the onset of cell division and under physiological blood flow sequestration is mediated by avidity-dependent binding to multiple host-receptors, mimicking the process of leukocyte recruitment [53–55]. Antibodies to CD36 have been shown to reduce rolling and adhesion of IE, residual rolling being further inhibited by antibody to ICAM1 [55]. These receptors operate synergistically to mediate strong cytoadherence when coexpressed on endothelial cells [56].

PiEMP1 proteins have multiple domains [8] and most CIDR-α type domains bind CD36 [7], while some DBLβ2 domains bind ICAM1 [58–61] and some single PiEMP1 species have been shown to mediate multiple independent interactions with a diverse set of host receptors including CD31/PECAM-1, the blood group A antigen, normal nonimmune IgM, heparan sulfate-like glucosaminoglycan, and CD36 [62].

A study has suggested that the ability of parasites to bind to multiple receptors is correlated with disease severity [63]. In addition, several lines of evidence have implicated CD54/ICAM1 [64–67], CD31/PECAM1 [68] as well as PFD1235w and PF11_0008 [39,69,70] as having a role in severe disease. Interestingly, we found the PFD1235w and PF11_0008 surface co-expressed on the 3D7/PFD1235w/PF11_0008 sub-line to mediate binding to CD54/ICAM1 and CD51/PECAM1, respectively (Figure 6). To our knowledge this is the first study to demonstrate dual receptor binding in malaria cytoadherence being mediated by two different PiEMP1 molecules on the surface of single IE. A potential mechanism for the two PiEMP1 interactions could involve a role for PFD1235w as a primary ligand for rolling and CD54/ICAM1 binding on the endothelium thus enabling further contacts with CD31/PECAM1 which could be mediated by PF11_0008, when both are present on the IE surface. Adhesion would then be improved through simultaneous binding to several receptors as indicated by our data in Figure 6. This process may be similar to that of lymphocyte rolling based on carbohydrate-receptor interactions [71].

If more than one PiEMP1 antigen is expressed on individual IE in vivo infections it may be most advantageous during the immediately post-hepatocytic establishment phase. The first post-hepatocytic generation lacks epigenetic memory of var gene transcription and translation. Relaxed transcription [72] and translation may thus be initially unavoidable. It may also ensure the highest avidity binding interaction possible and rapid sequestration. Transcription of those var genes expressed by successfully sequestering survivors of the post-hepatocytic wave of infection would epigenetically mark these var genes for expression in their descendent population, whilst leaving their unexpressed and thus unmarked var genes to become silenced. Such an ‘early loose-tight late’ model for var gene transcription in blood stage infection is compatible with experiments demonstrating that transcription of a particular var gene promoter leads to the silencing of the other var genes in the repertoire [73].

Our var gene transcript level, PiEMP1 surface expression, and cytoadhesion data may be indicating that the primary role of the P. falciparum var genes is the sequestration reaction and that at least in the earliest phases of blood stage infection, escape of IE from the circulation to a sequestration site which ensures successful replication takes precedence over shielding the antigenic variation repertoire from immune surveillance. The tiny amounts of PiEMP1 present at very low parasitaemia are unlikely to be sufficient to promote protective seroconversion. As infections develop and variant-specific parasitaemia rises, the repertoire become protected by epigenetic silencing until finally a novel cytoadhesion phenotype expressing a new PiEMP1 variant.
outgrows the increasingly immunocompromised founder population and the process starts over.

In summary, we have shown protein expression of two different var genes on the membrane of single erythrocytes infected with *P. falciparum* 3D7 to facilitate cytoadhesion of the infected cells to two different human receptors. These observations contradict the hypothesis of mutually exclusive PfEMP1 expression and at the same time offers an additional molecular explanation for how individual IE can mediate adhesion to multiple host receptors.

**Materials and Methods**

**Malaria parasites and in vitro selection procedure**

The *P. falciparum* isolate NF54 and the NF54 derived clone 3D7 were cultured in blood group 0 erythrocytes as previously described [74]. Cultures were routinely genotyped by PCR using primers targeting the polymorphic loci MSP2 and Glurp as described [75] and mycoplasma tested using the MycoAlert Mycoplasma Detection Kit (Lonza) following the manufacturer’s instructions.

IgG from rabbits immunized with DBL4γ or CIDR1z of 3D7 PFD1235w and CIDR2β of 3D7 PF11_0008 were used to obtain three different sub-lines of 3D7 parasites. 3D7PF1235w, originating from a previously selected line referred to as 3D7Dodowa1 [31] was obtained using sera targeting a PFD1235w-DBL4γ recombinant antigen. 3D7PF11_0008 originating from 3D7 was selected using sera targeting the CIDR2β of PF11_0008 and 3D7PF1235w/PF11_0008 similarly originating from 3D7 was obtained using sera targeting a CIDR1z domain recombinant protein of PFD1235w. NF54VAR2CSA kindly provided by Morten A. Nielsen was selected using rabbit antisera raised against a DBL5ε-6ε recombinant protein based on the var2csa gene. Briefly, the different rabbit sera were depleted on human uninfected erythrocytes type 0, incubated with gelatine purified trophozoite-stage 3D7/NF54 parasites for 30 min at 37°C, and unbound antibodies were removed by washing. Subsequently, IE were incubated with Protein A-coupled Dynabeads (Invitrogen) for 30 min at 37°C and bound IE were trapped using a magnet. Trapped IE were transferred to new culture flasks for continued in vitro culturing and the procedure was repeated until cultures stained positive by the selecting antisera in flow cytometry.

**Dynabeads sorting of parasites**

Gelatine purified 3D7PF1235w/PF11_0008 IE was incubated for 30 min at 37°C with rabbit sera targeting PFD1235w-DBL4γ, PFD1235w–CIDR1z, and PF11_0008–CIDR2β and incubated with Protein A-coupled Dynabeads. Bound IE were trapped as described above, grown for less than one cycle to rings, following which RNA was purified, reverse transcribed and used for real-time quantitative PCR.

**Flow cytometry sorting of parasites**

MACS purified 3D7PF1235w/PF11_0008 IEs were double stained with rat PFD1235w-DBL4γ and rabbit PF11_0008-CIDR2β antisera as described below and analysed on a FACSARia (Becton Dickinson). A total of 1×10^6 double positive IE were collected and grown for less than one cycle to rings, following which RNA was purified, reverse transcribed and used for real-time quantitative PCR.

**Limiting dilution of parasites**

Late stage 3D7PF1235w/PF11_0008 IE was obtained by MACS purification and cloned using a slightly modified version of the protocol described by Walliker and Beale [76]. In brief, 0.25 or 0.5 IE in a total volume of 100 µl RPMI 1640 (Lonza) with 1% hematocrit, 5 mg/ml Albumax II (Life Technologies), 0.18 mg/ml glutamine (Sigma-Aldrich), 0.05 mg/ml gentamicin (Gibco) and 10% Normal Human Serum were seeded into each well of flat bottomed 96-wells plates (NUNC). New media was added every second day and additional 1% hematocrit was added on day 5. A total of 64 wells were checked for the presence of IE by Giemsa staining on day 12 and 16 yielding nine different clones. RNA was purified from all nine clones for real-time quantitative PCR analysis and the PFD1235w and PF11_0008 surface expression was similarly analysed by flow cytometry. One clone, clone 3 was selected for FISH and confocal microscopy analysis.

**Protein expression**

The DNA sequence encoding amino acid # 73-739 (DBL1z–CIDR1z: CELDYRF…DTKTNTPC), amino acid #473-817 (CIDR1z*: DYCQICP…NGEPCTG), amino acid # 407-798 (CIDR1z: KDAKTDS…TNLDNIG), amino acid # 1293-1689 (DBL3β: CAETGGV…YATACD), amino acid #1719-2255 (DBL4γ: PRDKTGG…LKGDKSL), amino acid #2258-2764 (DBL5α: ACALKYG…SAKQKDC), and amino acid # 2242-3016 (DBL5α-CIDR2β: CATVAKA…VTQPNIC) of 3D7 PFD1235w, amino acid #1553-1924 (CIDR2β: KQKEKL…NYVANPC) and amino acid #1994-2378 (DBL4β: CNTKEH…HDDACAC) of 3D7 PF11_0008 and the DNA amplified by primers listed in Table S1 was cloned and expressed in a baculovirus system as described [39,77–79]. DBL1x VAR2CSA protein was kindly provided by Ali Salanti and Madeleine Dahlback.

**Generation of antisera**

All procedures complied with European or national regulations. Prior to immunization each animal was pre-bled and these sera were used as negative controls in the flow cytometry and confocal microscopy experiments. VAR2CSA DBL5ε-6ε rat antisera, rabbit zDBL5ε-DBL6ε and mouse zDBL5ε antisera [80] were kindly provided by Ali Salanti and Madeleine Dahlback. PFD1235w–CIDR1z, PFD1235w–DBL4γ, and PF11_0008–CIDR2β rabbit, and PFD1235w–DBL1z–CIDR1z, PFD1235w–CIDR1z*, PFD1235w–DBL3β, PFD1235w–DBL4γ, PFD1235w–DBL5α, PFD1235w–DBL5α–CIDR2β, and PF11_0008–DBL4β were raised by subcutaneous injection of 10–20 µg protein in complete Freund’s adjuvant followed by several boosters of protein in incomplete Freund’s adjuvant.

**Ethics statement**

All experiments including immunizations and bleeding of animals was approved by The Danish Animal Procedures Committee (”Dyreforsøgsstyrrelsyn”) as described in permit no. 2008/561-1498 and according to the guidelines described in act no. LBK 1306 (23/11/2007) and BEK 1273 (12/12/2005).

**Affinity purification of antibodies**

0.5 mg of PFD1235w-DBL4γ and PF11_0008-CIDR2β in each 1 ml were dialysed ON against coupling buffer and subsequently coupled to HiTrap NHS activated HP columns as described by the manufacturer (GE Healthcare). A pool of 3 ml PFD1235w–DBL4γ antisera from six rats and 6 ml PF11_0008-CIDR2β antisera from one rabbit were diluted 1:1 in PBS and affinity purified on the HiTrap columns. Following elution in Glycin buffer (0.1 M, pH 2.8) antibodies were neutralised in Hepes buffer (1M, pH 8.0).
Depletion and blocking of antibodies

The affinity purified rabbit PFD1235w-DBL4γ and PF11_0008-CIDR2β antibodies were depleted for V5-His reactivity using a non-sense peptide (VLM-tag) VLMPFNEQHKR-GASTYWCPGKIPNPLLGLD STRTGHHHHHH (Schafer-N) containing the V5-His epitope (underlined) and recombinant V5-His tagged VAR2CSA-DBL1x. The VLM-tag peptide and VAR2CSA-DBL1x (both 1 mg/ml in PBS) were coupled to Epoxy M270 Dynabeads according to the manufacturer's instructions (Invitrogen), 200 μl of each of the two different species of affinity purified antibodies was incubated for 2 h at RT with 3 mg of coupled Dynabeads. Following this the supernatant containing the depleted antibody fraction was removed and re-incubated twice with freshly coupled Dynabeads. Antibody bound to the Dynabeads were eluted in Glycin/HCl (0.1 M, pH 2.75) and neutralised in Tris buffer (1 M, pH 9.0). The reactivity of depleted and non-depleted antibody fractions was tested by ELISA (as described below) using PFD1235w-DBL4γ, PF11_0008-CIDR2β, VAR2CSA-DBL1x, and the VLM-tag peptide as coating antigen. Additionally, the affinity purified and depleted PFD1235w-DBL4γ and PF11_0008-CIDR2β antibodies used for flow cytometry and confocal microscopy were tested by LumineX (as described below) on 49 different PfEMP1 recombinant proteins. Similarly rabbit antisera against PFD1235w-CIDR1α was depleted on recombinant V5-His-tagged VAR2CSA-DBL1x and tested by ELISA using PFD1235w-CIDR1α, PF08_0103-CIDR1α, PF11_0008-CIDR2β, VAR2CSA-DBL1x, and the VLM-tag peptide as coating antigen.

Rat PFD1235w-DBL4γ, rabbit PFD1235w-CIDR1α or PF11_0008-CIDR2β antisera binding was blocked by incubation for 1 hr at 4°C using excess purified recombinant PFD1235w-DBL4γ, PFD1235w-CIDR1α, PF11_0008-CIDR2β, or PF08_0103-CIDR1α protein (3 μg/ml containing 5 μl rat or 10 μl rabbit sera) and remaining cross-reactivity to parasite antigens expressed on the surface of infected erythrocytes was subsequently tested by flow cytometry as described below.

Flow cytometry

Parasite cultures were repeatedly antibody selected as described above and grown for 3-5 cycles prior to doing flow cytometry. Single colour flow cytometry surface staining was done with minor modifications as described [74]. In brief, IE were purified on a magnet-activated cell sorting column (MACS) and 2×10^5 ethidium bromide-labelled IE were incubated for 30 min at 4°C in 5 μl rat, 10 μl rabbit sera, or 15 μl affinity purified and tagged-depleted antibody depleted of anti-human erythrocyte antibodies and then incubated for 30 min at 4°C with FITC-conjugated goat-anti-rat IgG (1:150, Zymed) or FITC-conjugated goat-anti-rabbit IgG (1:200, Vector Laboratories). Prior to doing two colour flow cytometry surface staining the purity of the MACS-purified IE was verified by running a small sample of ethidium bromide-labelled IE on a flow cytometer. Sub-line MACS preparations with ≥80% IE were used for two colour flow cytometry. In the case of clone 3 MACS preparations with 75-80% IE were used. In brief, 2×10^5 MACS-purified unlabelled IE were incubated with 5 μl rat and 10 μl rabbit sera depleted of anti-human erythrocyte antibodies, followed by incubation with FITC-conjugated goat-anti-rat IgG (1:150, Zymed) and Alexa Fluor 610-R-PE-conjugated goat-anti-rabbit IgG (1:200, Molecular Probes). Samples were analysed on a Cytomics FC 500 MPL flow cytometer (Beckman Coulter) and data analysed using WinList version 6.0 (Verity Software House Inc.). IE stained with one surface colour was gated based on the ethidium bromide staining to exclude uninfected erythrocytes. Two colour stained non-ethidium bromide-labelled IE were gated based on forward and side scatter values since ethidium bromide could not be used due to overlapping spectra with the Alexa Fluor 610-R-PE-conjugated anti-rabbit used. For removal of surface PfEMP1 and analysis of antisera cross-reactivity with trypsin resistant surface IE antigens expressed on late-stage trophozoites and schizonts IE were treated with 1 g/l porcine trypsin and 0.2 g/l EDTA solution in Hanks balanced salt solution (Sigma-Aldrich) for 10 min at 37°C. The reaction was stopped by adding 10% foetal calf serum (FCS) and cells washed three times in PBS plus 2% FCS. Controls were incubated similarly in PBS plus 2% FCS or in a trypsin/EDTA solution containing 10% FCS.

Confocal microscopy

Laser scanning confocal microscopy was performed on all samples analysed by flow cytometry in order to observe the staining patterns on individual IE. MACS purified unlabelled infected cells from the same batches of parasites tested by flow cytometry were incubated with individual antibodies or in combination with the PFD1235w, PF11_0008, and VAR2CSA antibodies as previously described [81]. Briefly, 1 μl packed IE were washed in 1% BSA in PBS (BSA/PBS) and the pellet was incubated in 100 μl BSA/PBS and 3 μl of the respective antibodies for 30 minutes at 4°C. The IE were washed three times in BSA/PBS. The IE stained with primary antibody were then incubated with secondary antibodies, either Alexa 488 anti-rat IgG (Invitrogen), Alexa 568 anti-rabbit and/or 568 Alexa anti-mouse IgG (Invitrogen) and DAPI (3 μl of a 3 μg/ml solution) for 30 minutes at 4°C. The IE were washed three times and visualised as live, unfixed cells using a Nikon TE 2000-E confocal Nikon microscope with 60x oil immersion objective lens (DIC). The images were processed using Adobe Photoshop software and displayed with the 5 μm scale bar calculated by the EZ-C1 software.

ELISA and luminex

Affinity purified and depleted rat and rabbit antibodies were tested by ELISA. Wells of Maxisorp plates (Nunc, Roskilde, Denmark) were coated with 1 μg/ml of testing antigen in Glycin/HCl buffer (0.1 M, pH 2.75), antibodies were diluted 1:100 in blocking buffer (PBS, 0.5 M NaCl, 1% Triton-X-100, 1% BSA, pH 7.2), and plates were washed and developed as described previously [77]. In addition, the affinity purified and depleted antibodies were tested in the BioPlex100 System (BioRad) as previously described [81]. Briefly, 0.1 mg Baculovirus produced proteins and the VLM-tag peptide were individually coupled to 1.25×10^7 Luminesx xMAP technology microsphere beads (Ramcon). Plex 1 and 2 contained 6 and 45 different proteins, respectively. Three protein domains were coupled twice in plex 2 and two were identical to domains in plex 1 (Table S1). Prior to multiplexing, protein coupling was verified by incubating beads (1:333) with mouse anti-V5 antibody (1:10,000, Invitrogen) followed by biotinylated anti-mouse IgG (1:500, DakoCytoImation). The biotinylated antibody was detected using PE-streptavidin (1:500, Sigma) and dinitrobenzene was used PBS/TB (PBS, 0.05% (v/v) Tween-20, 0.1% (w/v) BSA, pH 7.4). Multiplexed beads (1:333) were incubated with rat or rabbit antibodies (1:1000) followed by incubation with biotinylated anti-rat IgG (Sigma) or anti-rabbit IgG antibody (The Binding Site) diluted 1:1000. Detection was done with PE-streptavidine 1:100 or 1:1000 for rat and rabbit antibodies, respectively. Beads were resuspended in 100 μl dinitrobenzene and a minimum of 100 beads from each set of multiplexed beads were analyzed to yield the mean fluorescence intensity (MFI). Each affinity purified and depleted antibody was analyzed in duplicates, the average MFIs and standard deviation.
were calculated based on three independently repeated experiments.

RNA extraction and cDNA synthesis

Antibody selected IE were grown for 3–5 cycles and erythrocytes infected by trophozoite/schizont-stage parasites (20–48 h post invasion) from in vitro cultures were MACS purified and used for flow cytometry, confocal microscopy and for re-invasion of uninfected erythrocytes in new cultures flasks. IE used for RNA extraction were harvested when re-invaded parasites were at the ring-stage as confirmed by microscopy of Giemsa stained thin smears.

Total RNA was prepared using Trizol (Invitrogen) as recommended by the manufacturers and treated with DNase1 (Invitrogen) for 15 min at 37°C. Absence of DNA in RNA samples was confirmed by stable base fluorescence after 40 cycles of real-time PCR with seryl-tRNA synthetase and fructose-bisphosphate aldolase primers as previously described [41]. Superscript II was used to reverse transcribe DNA-free RNA primed with random hexamer primers (Invitrogen) at 25°C for 10 min and 42°C for 50 min followed by 70°C for 15 min.

Quantitative real-time PCR

Quantitative real-time PCR was performed using a Rotorgene thermal cycler system (Corbett Research), Quanti-Tect SYBR Green PCR Master Mix (Qiagen), and real-time PCR-optimized and gene-specific primers (0.5 μM) for each of the full-length var genes in the P. falciparum 3D7 genome and to the endogenous control genes seryl-tRNA synthetase and fructose-bisphosphate aldolase. Quantification was done using Rotorgene software version 6.0. mRNA transcript copy numbers were calculated from real-time Q-PCR measurements of 10 fold genomic DNA dilutions as described [43]. The proportions of individual var genes transcripts relative to the total copy number of all var transcripts or to the total number of endogenous control genes was subsequently calculated for comparison of var transcript profiles within and between samples and depicted in pie-charts.

FISH analysis

RNA-FISH was done on ring stage parasites of 3D7PFD1235w, 3D7PF11_0008, and 3D7PF11_0008/PF11_0008 IE. PCR products were amplified using 3D7 genomic DNA and PFD1235w (fw: 5’-GGGATCCGACACGTCGAGACAGAGG-3’; rv: 5’-GGAGAAGCTTGCAGGCAAGGAG-3’) and PF11_0008PFD1235w (fw: 5’-GGGATCCCTAGTTATTTGAGCGACGACCAGC-3’ and 5’-CTTGAAATTTCCGTGCTACTTCCCTCCC-3’) specific primers containing restriction sites (underlined). The PFD1235w and PF11_0008 PCR products were subsequently cloned into the pSPT19 or pSPT18 vector (Roche Applied Science), respectively and sequenced on a 3130 Genetic Analyzer (Applied Biosystems). Digoxigenin (DIG)- and biotin-labelled antisense RNA probes were generated using a DIG RNA Labelling Kit and Biotin RNA Labelling Mix (Roche Applied Science). The var2CSA anti-sense probe described previously [50] was included as a negative control. The specificity of the RNA probes was confirmed by standard Northern blotting analysis according to the DIG Application Manual found at http://www.roche-applied-science.com/PROD_INF/MANUALS/DIG_MAN/dig_toc.htm and [39].

The RNA-FISH slides were prepared according to a standard FISH protocol with minor modifications [82,83]. Following hybridization over-night at 48°C, single stained anti-sense RNA were detected by α-DIG HRP conjugated Ab or α-biotin HRP conjugated Ab and the dual stained anti-sense RNA were detected with both antibodies. The signal was further detected using the TSA Plus Fluorescence Palette System (PerkinElmer) with FITC and/or Cy3 (Amersham) at a concentration of 100 μg/ml for 30 min at 37°C prior to hybridization with the probes. Slides were washed, mounted with anti-fade reagent containing DAPI (Invitrogen) and images captured using a Nikon TE 2000-E confocal microscope as described above. The positivity of a total number of 100 IE was scored in each experiment.

Intron real-time PCR and sequencing of intron regions

RNA was isolated from the 3D7PFD1235w, 3D7PF11_0008, and 3D7PF11_0008/PF11_0008 sub-lines and cDNA generated as described above. Real-time quantitative PCR was done using primers amplifying the intron spanning region and parts of exon 1 of the var genes listed in Table S2 as well as the seryl-tRNA synthetase and fructose-bisphosphate aldolase [41]. In addition, PCR products were amplified from cDNA using Takara LA Taq (Lonza) and the intron spanning primers of PFD1235w and PF11_0008 and the var genes listed in Table S2. The PCR products were sequenced on a 3130 Genetic Analyzer using the same set of primers. For comparison genomic DNA was purified from a batch culture of 3D7PFD1235w/PF11_0008 as well as from clone 3 and the intron spanning region of PFD1235w and PF11_0008 was sequenced.

Adhesion assays

Adhesion assays were done as described previously [84]. In brief, ring stage IE were cultured over night in hypoxanthine-free RPMI 1640 (Lonza) with 10% hypoxanthine-free Albumax (Invitrogen) and 8.75 MBq 3H-hypoxanthine (Amersham) per ml packed erythrocytes. 96-well flat bottomed plates (Nunc) were coated with 1% gelatine and HUVEC cells (PromoCell) grown in endothelial cell growth medium (Promocell) to maximum 6th passage were seeded at 500,000 cells per well and grown to become confluent. CHO cells (CHO-wild type, CHO-CD36, CHO-CD54/ICAM1 from ATCC) were grown in RPMI 1640 with 1% added glutamine and 10% foetal calf serum (Lonza) and similarly seeded at 550,000 cells per well. On the day of the assay, 3H-labelled parasite cultures were enriched for late-stages by gelatine floatation and 50 μl of labelled trophozoites (12,000 counts per minute) were added in duplicates or triplicates to the 96-well plates containing HUVEC or CHO cells. The plates were left to incubate on a rocking table for 2 hrs at 37°C. Monoclonal mouse anti-CD54/ICAM1 My13 (10 μg/ml, InVitrogen), anti-CD54/ICAM1 15.2 (20 μg/ml, AbD Serotec), anti-CD36 (10 μg/ml, R&D) and anti-CD31/PECAM-1 9G11 (20 μg/ml, R&D Systems) was added in anti-adhesion assays. Unbound IE were removed using a washing robot (Biomek 2000, Beckman Coulter) and cells binding IE were harvested onto filter paper (Unifilter-96, GF/C, PerkinElmer) using a Filtermate Harvester (PerkinElmer). Following addition of 50 μl/well of scintillation liquid (Microscint-20, PerkinElmer) counting was done on a Topcount NXT (PerkinElmer). The H max-values were determined as the counts per minute of 50 μl of parasite added to one cell-free well and harvested directly onto a filter paper. To standardise, all values obtained was given as a ratio compared to the max value obtained in the particular assay. The percentage binding of the different 3D7 sub-lines were calculated relative to the binding of the 3D7PFD1235w parasite line adhering to CD34/ICAM1 (Figure 6A–C) or the 3D7PF11_0008 parasite line adhering to HUVEC (Figure 6D) (e.g. binding of these equalling a 100%).
Supporting Information

Figure S1 PIEMP1 surface expression on 3D7 infected erythrocytes and antisera specificity. (A) Erythrocytes infected with the 3D7PF1235w sub-line. Surface staining of PIEMP1 by flow cytometry was done using rat anti-PIEMP1 antisera, (B) rabbit anti-PIEMP1 antisera, and (C) double-color surface staining of PfEMP1 by flow cytometry using (A) rabbit anti-PIEMP1 antisera, and (B) rat pre-bleed. Grey bars: exon I primers. Black bars: intron primers. White bars: house keeping gene primers. Numbers on the Y-axis are primer number (see Table S2). Found at: doi:10.1371/journal.ppat.1000183.s011 (0.11 MB DOC)

Figure S2 Trypsination of 3D7 infected erythrocytes and blocking of antisera show no cross-reactivity. (A) and (D) 3D7PF1235w IE, (B) and (E) 3D7PF1235w IE, (C) and (F) 3D7PF1235w IE. (A-C) Untreated (light gray bars), trypsin (black bars), and trypsin plus fotal calf serum (FCS) treated IE (dark gray bars) were surface stained for flow cytometry using rabbit sera against PFD1235w-DBL5, PFD1235w-DBL5, and PFD1235w-CIDR2B of PFD1235w and (9) DBL5 of PF11_0008 as well as using rabbit antisera against (9) CIDR1 of PFD1235w, (8) DBL4 of PFD1235w, and (10) CIDR2 of PFD1235w. Flow cytometry settings were identical for all panels and reactivity with pre-bleeds is shown as grey histograms. Found at: doi:10.1371/journal.ppat.1000183.s001 (0.11 MB DOC)

Figure S3 PIEMP1 surface expression on erythrocytes infected with 3D7PF1235w/PF11_0008 clone 3 and 4. Single colour surface staining of PIEMP1 by flow cytometry was done using (A) rabbit pre-bleed (1), (B) rabbit pre-bleed (2), (B) rabbit pre-bleed (3), (B) rabbit pre-bleed (4), (B) rabbit pre-bleed (5), (B) rabbit pre-bleed (6). Found at: doi:10.1371/journal.ppat.1000183.s002 (0.28 MB DOC)

Figure S4 The specificity of antisera and antibodies used for surface labelling and selection of parasites. The specificity was tested by ELISA (A-C) and Luminex (D-F) on 49 different PIEMP1 domains (Table S1). (A) rat PFD1235w-DBL4 antisera, (B) rabbit PFI1_0008-CIDR2B antisera, (C) rabbit FDI235w-CIDR1x antisera, (D) antibodies purified on a PFD1235w-DBL4 column and depleted of tag reactivity, (E) antibodies purified on a PFD1235w-CIDR2B column and depleted of tag reactivity, and (F) PFD1235w-CIDR2x antisera depleted of tag reactivity. The coating antigens in ELISA were DBL4 γ and CIDR2x of PFD1235w, CIDR1x of PFI1_0008, CIDR2x of PF11_0008, DBL1x of VAR2CSA, and the VLIM-tag peptide as indicated by the vertical lines in (A-C). Found at: doi:10.1371/journal.ppat.1000183.s004 (0.02 MB PDF)

Figure S5 Simultaneous surface expression of PIEMP1 on single erythrocytes infected with 3D7 detected using purified antibodies. Parasite cultures (A-D) were identical to those used in Figure 1, Figure 2, and Figure S1. The affinity purified and depleted antibodies used were (A-D1) rat PFD1235w-DBL4 γ; (A-D2) rabbit PFI1_0008-CIDR2B antibodies; and (A-D3) rat PFD1235w-DBL4 γ combined with rabbit PFI1_0008-CIDR2B. For confocal microscopy rat antibody staining of PFD1235w expressed by the 3D7PF1235w (A1 & A3) and 3D7PF1235w/PF11_0008 (C1 & C3) sub-lines were detected using Alexa 488-labeled anti-rabbit antibodies (green) and rabbit antibodies staining PF11_0008 expressed by the 3D7PF11_0008 (B2) and 3D7PF1235w/PF11_0008 (C2 & C3) were detected using Alexa 568-labeled anti-rabbit antibodies (red). Double staining using the two Alexa fluorophores (A-D3) showed simultaneous expression of PFD1235w and PF11_0008 by RBC infected with the 3D7PF1235w/PF11_0008 with no co-localisation of the staining (C3). Flow cytometry histograms show single staining of the three different IE sub-lines using the purified and depleted antibodies. Double staining was not done due to limited amounts of depleted antibody. (D1-3) A N554VAR2CSA control sub-line did not stain positive with any of the antibody preparations. DAPI staining of DNA in the nuclei is blue. Scale bar 5 µm. Found at: doi:10.1371/journal.ppat.1000183.s005 (0.06 MB PDF)

Figure S6 Real-time Q-PCR Ct-values for exon I and intron crossing amplicons amplified from cDNA of 3D7. (A) 3D7PF1235w sub-line, (B) 3D7PF11_0008 sub-line, (C) 3D7PF1235w/PF11_0008 sub-line. Numbers on the Y-axis are primer numbers (See Table S2). Grey bars: exon I primers. Black bars: intron primers. White bars: house keeping gene primers. Found at: doi:10.1371/journal.ppat.1000183.s006 (0.01 MB PDF)

Table S1 Primers used for amplification of DNA encoding recombinant proteins coupled to Luminex beads. Found at: doi:10.1371/journal.ppat.1000183.s007 (0.09 MB DOC)

Table S2 Specific var gene primers used for var intron Q-RT PCR. Found at: doi:10.1371/journal.ppat.1000183.s008 (0.05 MB DOC)

Video S1 Simultaneous surface expression of PIEMP1 on single erythrocytes infected with Plasmodium falciparum 3D7PF1235w/PF11_0008-
Staining of PFD1235w and PF11_0008 is green and red, respectively. DAPI staining of DNA in the nuclei is blue. Scale bar 5 μm. Found at: doi:10.1371/journal.ppat.1001083.s009 (2.30 MB MOV)

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Author Contributions

Conceived and designed the experiments: LJ DCB LT TGT DEA ATRJ. Performed the experiments: LJ DCB AB ER SSB MBD MEV ATRJ. Analyzed the data: LJ DCB AB ER TGT DEA ATRJ. Contributed reagents/materials/analysis tools: LT GKKC TL. Wrote the paper: LJ DEA ATRJ.

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