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Genetic Dysmyelination Alters the Molecular Architecture of the Nodal Region

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We have examined the molecular organization of axons in the spinal cords of myelin-deficient (md) rats, which have profound CNS dysmyelination associated with oligodendrocyte cell death. Although myelin sheaths are rare, most large axons are at least partially surrounded by oligodendrocyte processes. At postnatal day 7 (P7), almost all node-like clusters of voltage-gated Na+ channels and ankyrinG are adjacent to axonal segments ensheathed by oligodendrocytes, but at P21, many node-like clusters are found in axonal segments that lack oligodendrocyte ensheathment. In P21 wild-type (WT) rats, the voltage-gated Na+ channels Na1.2, Na1.6, and Na1.8, are found in different subpopulations of myelinated axons, and md rats have a similar distribution. The known molecular components of paranodes—contactin, Caspr, and neurofascin 155—are not clustered in md spinal cords, and no septate-like junctions between oligodendrocyte processes and axons are found by electron microscopy. Furthermore, Kv1.1 and Kv1.2 K+ channels are not spatially segregated from the node-like clusters of Na+ channels in md rats, in contrast to their WT littermates. These results suggest the following: node-like clusters of voltage-gated Na+ channels and ankyrinG form adjacent to ensheathed axonal segments even in the absence of a myelin sheath; these clusters persist after oligodendrocyte cell death; dysmyelination does not alter the expression of different nodal voltage-gated Na+ channels; the absence of paranodes results in the mislocalization of neurofascin155, contactin, and Caspr, and the aberrant localization of Kv1.1 and Kv1.2.

Key words: myelin; oligodendrocytes; mutant; septate junctions; axon-glia interactions; proteolipid protein

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The molecular organization of the axonal membrane is highly related to that of its myelin sheaths (Arroyo and Scherer, 2000; Peles and Salzer, 2000; Rasband and Shrager, 2000). In both the CNS and PNS, the nodal membrane contains high concentrations of voltage-gated Na+ channels, which are linked to the spectrin cytoskeleton by ankyrinG. The paranodal region is distinguished by septate-like junctions that link the axonal membrane to the spiral of glial endfeet. Contactin and contactin-associated protein (Caspr; also known as paranodin) are localized to the paranodal axonal membrane. An alternatively spliced isoform of neurofascin (Caspr; also known as paranodin) is found in different subpopulations of myelinated axons, and md rats have a similar distribution. The known molecular components of paranodes—contactin, Caspr, and neurofascin 155—are not clustered in md spinal cords, and no septate-like junctions between oligodendrocyte processes and axons are found by electron microscopy. Furthermore, Kv1.1 and Kv1.2 K+ channels are not spatially segregated from the node-like clusters of Na+ channels in md rats, in contrast to their WT littermates. These results suggest the following: node-like clusters of voltage-gated Na+ channels and ankyrinG form adjacent to ensheathed axonal segments even in the absence of a myelin sheath; these clusters persist after oligodendrocyte cell death; dysmyelination does not alter the expression of different nodal voltage-gated Na+ channels; the absence of paranodes results in the mislocalization of neurofascin155, contactin, and Caspr, and the aberrant localization of Kv1.1 and Kv1.2.

A number of inherited dysmyelinating or demyelinating diseases that affect the PNS and/or the CNS that have been linked to mutations in genes that are expressed in the myelinating cells themselves. In humans and mice, different mutations of the proteolipid protein gene (PLP/Plp) cause a range of phenotypes (Nave and Boespflug-Tanguy, 1996). Although these inherited dysmyelination–demyelinating diseases are caused by cell autonomous defects in the myelinating glial cells, nonautonomous damage to axons has been increasingly implicated as a crucial aspect of these diseases (Griffiths et al., 1998). How demyelination leads to axonal loss is not known, but the reorganization of the axonal membrane is the earliest known alteration. This was first observed for voltage-gated Na+ channels: nodal clusters are lost after demyelination, but reappear after remyelination (Dugandzija-Novakovic et al., 1995; Novakovic et al., 1996). Similarly, juxtaparanodal Kv1.1 and Kv1.2 channels disperse after demyelination and reorganize with remyelination (Rasband et al., 1998). In inherited dysmyelinating–demyelinating diseases, the sequelae of demyelination and remyelination probably coexist even on the same myelinated fiber, resulting in a complex pathological picture. In this paper, we have investigated the organization of the axonal membrane in myelin-
deficient (md) rats, which have a severe dysmyelinating disease associated with oligodendrocyte cell death (Gow et al., 1998; Grinspan et al., 1998; Lipsitz et al., 1998). Although few large CNS axons are myelinated, they are ensheathed by oligodendrocyte processes. Node-like clusters of Na⁺ channels and ankyrinG develop at the edges of oligodendrocyte processes, but are subsequently found in regions devoid of oligodendrocytes. Ensheathed and even myelinated axons do not have molecular or structural specializations at paranodes, and Kv1.1 and Kv1.2 channels abut the node-like clusters of Na⁺ channels and ankyrinG.

MATERIALS AND METHODS

Animals. Male md rats and their wild-type (WT) littermates were obtained from a colony at the University of Pennsylvania. Postnatal day 14 (P14) and P21 rats have an obvious tremor and gait difficulties; P7 rats were genotyped by PCR as described previously (Grinspan et al., 1998).

Immunostaining. P21 md rats and their WT male littermates were overdosed with pentobarbital, then perfused with freshly prepared 4% formaldehyde containing 0.5% Triton X-100 in PBS, and incubated at 4 °C overnight. Sections were post-fixed and permeabilized by immersion in 20% sucrose PB overnight before embedding. Ten-micrometer-thick cryostat sections were thaw-mounted on SuperFrost Plus glass slides (Fisher Scientific, Pittsburgh, PA) and stored at −20 °C. Sections were post-fixed and permeabilized by immersion in −20 °C acetone for 10 min, blocked at room temperature for at least 1 hr in 5% fish skin gelatin containing 0.5% Triton X-100 in PBS, and incubated 24–48 hr at 4°C with various combinations of primary antibodies (Table 1) diluted in blocking solution. After incubating with the primary antibodies, the slides were washed, incubated with the appropriate fluorescein-, tetramethylrhodamine-, and cyanine 5-conjugated donkey cross-avidity-purified secondary antibodies (diluted 1:200; Jackson ImmunoResearch Laboratories, West Grove, PA), and mounted with Vectashield (Vector Laboratories, Burlingame, CA). The slides were examined by epifluorescence with tetramethylrhodamine isothiocyanate (TRITC) and fluorescein isothiocyanate (FITC) optics on a Leica (Nussloch, Germany) DMR light microscope and photographed with a Hamamatsu (Tokyo, Japan) digital camera or with a Leica TCS laser-scanning confocal microscope followed by image manipulation with Adobe Photoshop and Canvas.

To demonstrate the specificity of the Na⁺ channels against which it was raised and incubated overnight at 4°C. An equal amount of each antiserum was treated in a similar manner, substituting PBS for the blocking peptide solution. The next morning, a double labeled with the rabbit antiserum against Nav1.6 and the pan Na⁺ monoclonal antibody was added to each tube, and the contents of the "blocked" and "unblocked" tubes were used to label slides of P21 WT and md spinal cord as described above. Quantitative analysis. To determine whether the number of node-like clusters was higher in md spinal cords, we embedded the cervical spinal cords from three P21 md rats and three age-matched male WT littermates in the same block. Ten-micrometer-thick frozen sections were double labeled with the rabbit antiserum against Na⁺, and the pan Na⁺ channel mouse monoclonal antibody. A 130 μm² area of the ventral funiculus nearest to the midline was selected because this region contains many of the largest myelinated fibers in the spinal cord. We reasoned that examining large myelinated fibers would more easily reveal an increase in

Table 1. The source and dilutions of antibodies used in this study

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<th>Antibody</th>
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<th>Source/reference</th>
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<td>FITC-cholera toxin</td>
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Rb, Rabbit; M, mouse.
Digital images. By simultaneously comparing the Nav1.6 (TRITC) and with P21 WT littermates, we enlarged nonoverlapping parts of the above spinal cords were removed, cut into 2 with 0.9% NaCl followed by 3% glutaraldehyde in PB. The cervical night at 4 °C and incubated for 24 hr at 4°C in blocking buffer with rabbit antisera against NaV1.2 (1:1000), NaV1.6 (1:1000), NaV1.8 (1:1000), contactin (1:5000), or Caspr (1:5000). After washing in blocking solution, the blots were incubated in peroxidase-coupled secondary antibodies against rabbit (Jackson ImmunoResearch; diluted 1:10,000) for 1 hr at room temperature (RT). After washing in blocking solution and Tris-buffered saline containing 0.5% Tween 20, blots were visualized by enhanced chemiluminescence (Amersham, Arlington Heights, IL) according to the manufacturer's protocols. To visualize GAPDH, the blots were first washed in blocking buffer with 0.01% sodium azide and subsequently probed with a mouse monoclonal antibody against GAPDH (1:10,000) followed by washing in blocking solution. The blots were incubated in peroxidase-coupled secondary antibodies against mouse (Jackson ImmunoResearch; diluted 1:10,000) for 1 hr at RT and visualized by enhanced chemiluminescence.

RESULTS

Node-like clusters of Na+ channels and ankyrinG in md spinal cord
To determine the localization of voltage-gated Na+ channels in md rats, we immunostained sections with either a mouse monoclonal antibody or a rabbit antiserum that both recognize the same conserved peptide sequence common to all type 1 channels (Goldin, 1999). We embedded spinal cords of P21 md rats and their WT littermates to obtain longitudinal sections of the ventral funiculus, which contains the largest axons. As shown in Figures 1, 2, 3, 7, and 8, there were node-like clusters of Na+ channels in ventral funiculus of both WT and md rats: the clusters were thin (~1 μm in width) and perpendicular to the axons. In transverse sections of md spinal cord, these node-like clusters were often crescent-shaped rather than complete circles as in WT spinal cords (Fig. 3A–F), indicating that many do not surround the entire circumference of the axonal membrane.

To determine whether these node-like clusters colocalized with other molecular components of nodes, we double-labeled sections with pan-Na+ channel antibodies (with either the mouse monoclonal antibody or the rabbit antiserum) and ankyrinG (either a rabbit antiserum or a mouse monoclonal antibody; the rabbit

The number of node-like clusters were compared by Student's t test using Microsoft Excel (mean ± SEM). To determine whether the proportion of NaV1.6-positive clusters was different in the ventral funiculi of md spinal cords compared with P21 WT littersmates, we enlarged nonoverlapping parts of the above digital images. By simultaneously comparing the NaV1.6 (TRITC) and the pan-Na+ channel (FITC) images on the computer screen, we determined whether each pan-Na+ channel cluster was NaV1.6-positive. These data were compared by ANOVA test using Microsoft Excel.

Electron microscopy. P21 md and WT male littersmates were perfused with 0.9% NaCl followed by 3% glutaraldehyde in PB. The cervical spinal cords were removed, cut into 2-3 mm wide sections, fixed overnight at 4°C in the same fixative, washed in PB, osmicated in 1% OsO4 for 1 hr at room temperature, then dehydrated in graded ethanols, infiltrated with propylene oxide followed by Epon, and polymerized at 60 °C. Sections were examined by epiluminescence as described above, and 40 μm digital images of the ventral funiculus were made for each animal. All the nodes in each image were counted and classified into one of three different categories: (1) naked clusters: node-like clusters of Na+ channels that were not flanked by MAG-positive ensheathed axonal segments; (2) heminodes: node-like clusters that were flanked on only one side by MAG-positive axonal segments; or (3) nodes: Na+ channels that are flanked on both sides by MAG-positive axonal segments. The percentage of nodes was calculated at each age; ANOVA statistical analyses were used to compare the samples.

Immunoblot analysis. To determine whether the levels of axonal proteins were altered in md rats, we made protein homogenates from spinal cords (stripped of dura and rootlets) and sciatic nerves dissected from P21 md rats and their male WT littersmates. Samples were immersed in liquid nitrogen, pulverized were a mortar and pestle on dry ice, and resuspended in ice-cold 50 mM Tris, pH 7.0, 1% SDS, and 0.017 mg/ml phenylmethylsulfonyl fluoride (Sigma, St. Louis, MO), followed by a brief sonication on ice with a desmembrator (Fisher Scientific). Protein concentrations were determined using the Bio-Rad kit (Bio-Rad, Hercules, CA) according to manufacturer's instructions. For each sample, ~100 μg of protein lysate was loaded onto a 5% SDS-polyacrylamide gel, electrophoresed, and transferred to nitrocellulose (for NaV1.2, NaV1.6, and NaV1.8) or Immobilon-polyvinylidene fluoride (Millipore, Bedford, MA; for contactin and Caspr) membrane over 1 hr, using a semidy transfer unit (Fisher Scientific). The blots were blocked (5% powdered skim milk and 0.5% Tween 20 in Tris-buffered saline) overnight at 4°C and incubated for 24 hr at 4°C in blocking buffer with rabbit antisera against NaV1.2 (1:1000), NaV1.6 (1:1000), NaV1.8, contactin (1:5000), or Caspr (1:5000). After washing in blocking solution, the blots were incubated in peroxidase-coupled secondary antibodies against rabbit (Jackson ImmunoResearch; diluted 1:10,000) followed by washing in blocking solution. The blots were incubated in peroxidase-coupled secondary antibodies against mouse (Jackson ImmunoResearch; diluted 1:10,000) for 1 hr at RT and visualized by enhanced chemiluminescence.

Figure 1. Node-like clusters of voltage-gated Na+ channels in md spinal cord.
These images were taken from longitudinal sections of the ventral funiculus from P21 md (A–D) or WT (E, F) spinal cords, immunostained with a rabbit antiserum against ankyrinG (A, C, E; TRITC) and a monoclonal antibody against voltage-gated Na+ channels (B; FITC) or tenascin-R (D; F; FITC). Note that node-like clusters of colocalize with voltage-gated Na+ channels and tenascin-R. The pairs of arrowheads mark the some of the node-like clusters in C–F. Scale bars, 10 μm.
We also double-labeled sections for ankyrin G and tenascin-R. The above results indicate that Na\textsuperscript{+} channels are more reliably recognized in P21 md rat spinal cords (Fig. 2F) stained nodes of some small axons in all funiculi, most abundantly in the dorsal funiculus, as well as the somatic membrane of most neurons throughout the gray matter, and an occasional initial segment. The Na\textsubscript{v}1.2 antisera (Fig. 2D) stained the nodes of a few small myelinated fibers in all funiculi, a few initial segments in the intermediate and dorsal gray matter, and the unmyelinated axons of the corticospinal tract. The nodes labeled with the Na\textsubscript{v}1.2, Na\textsubscript{v}1.6, or Na\textsubscript{v}1.8 antisera, as well as the unmyelinated axons of the corticospinal tract, were also labeled with the pan-Na\textsuperscript{+} channel mouse monoclonal antibody (data not shown; Arroyo et al., 2001).

The above results indicate that Na\textsubscript{v}1.6 is the predominant voltage-gated Na\textsuperscript{+} channel at nodes and initial segments in the spinal cord (Caldwell et al., 2000), with a minority of nodes expressing Na\textsubscript{v}1.8 followed by Na\textsubscript{v}1.2. To determine whether the axonal expression of these voltage-gated Na\textsuperscript{+} channels was affected by severe dysmyelination, we double-labeled sections of P21 md spinal cords with the Na\textsubscript{v}1.2, Na\textsubscript{v}1.6, and Na\textsubscript{v}1.8 antisera and the pan-Na\textsuperscript{+} channel monoclonal antibody. Compared with WT P21 rats, there was no apparent alteration in the spatial distribution of Na\textsubscript{v}1.2, Na\textsubscript{v}1.6, or Na\textsubscript{v}1.8 or pan-Na\textsuperscript{+} channel staining (Fig. 2A–C): most node-like clusters and initial segments were Na\textsubscript{v}1.6-positive; most neuronal cell membranes were Na\textsubscript{v}1.8-positive; the corticospinal tract (CST) was Na\textsubscript{v}1.2-positive; there were node-like clusters of Na\textsubscript{v}1.2, Na\textsubscript{v}1.6, and Na\textsubscript{v}1.8 in the same places as in WT P21 rats. The number of Na\textsubscript{v}1.6-positive node-like clusters, however, appeared to be increased in md spinal cords, in keeping with our results of following staining with the pan-Na\textsuperscript{+} channel monoclonal antibody (see above). To determine whether the proportion of Na\textsubscript{v}1.6-positive node-like clusters was affected, we analyzed the same 130 \( \mu \text{m}^2 \) square of the ventromedial funiculus used to determine the number of node-like clusters (see above). Although the number of node-like clusters tended to be higher in md spinal cords (see above), the proportion Na\textsubscript{v}1.6-positive clusters was the same (77%) as in P21 rats.
WT spinal cords (p = 1; ANOVA). Examples of double-labeled nodes are shown in Figure 3A–F. To support these findings, we performed “blocking” experiments with the peptides that were used to generate the Na\(^+\)v1.2, Na\(^+\)v1.6, and Na\(^+\)v1.8 antisera and immunoblot analysis. Preincubation of these antisera with their cognate peptides greatly attenuated all aspects of the staining described above (data not shown). Finally, Na\(^+\)v1.7 and Na\(^+\)v1.9 antisera did not label nodes in the CNS in either WT or md spinal cords, although there was Na\(^+\)v1.9 staining of what appeared to be unmyelinated afferents in the dorsal horn (data not shown; Fjell et al., 2000). Immunoblot analysis for Na\(^+\)v1.2, Na\(^+\)v1.6, and Na\(^+\)v1.8 revealed comparable levels of these voltage-gated Na\(^+\) channels in md and WT spinal cords (Fig. 4).

Juxtaparanodal and internodal specializations in md rats

In normal myelinated fibers, Kv1.1, Kv1.2, Kv\(\beta\)2, and Caspr2 are found at highest levels in juxtaparanodes and at lower levels in the internodes and paranodes, but they are not found at nodes (Arroyo and Scherer, 2000; Peles and Salzer, 2000; Rasband and Shrager, 2000). To investigate the localization of these proteins in md spinal cords, we stained longitudinal sections with mouse monoclonal antibodies and rabbit antisera against Kv1.1 and Kv1.2 (Table 1). In contrast to WT P21 spinal cords (Fig. 3H), Kv1.1 and Kv1.2 were more diffusely localized in md rats (Fig. 3G). Furthermore, Kv1.1 and Kv1.2 staining frequently abutted or even overlapped that of voltage-gated Na\(^+\) channels; this was never seen in WT spinal cords. Double-labeling for Kv1.1 and Kv1.2 demonstrated that these two channels were co-localized even in their abnormal distributions in md rats, as they are in WT rats and mice (data not shown). We could not perform comparable analyses for Kv\(\beta\)2 and Caspr2, because these antibodies stained too weakly in the fixed material (data not shown).

Contactin, Caspr, and NF155 are not localized to paranodes in md rats

The lack of separation between Kv1.1 and Kv1.2 K\(^+\) channels and voltage-gated Na\(^+\) channels in md rats led us to examine the expression of the paranodal proteins, contactin, Caspr, and NF155. In sections of P21 WT spinal cord, these proteins were colocalized to paranodes with multiple antibodies (Table 1). In md spinal cords, however, none of these proteins were localized to the paranodal region. Rather, there were diffuse staining in the white matter for contactin, Caspr, and NF155, and staining of oligodendrocyte cell bodies for NF155 (Tait et al., 2000). In the
ventral and dorsal rootlets, however, in which the axons are myelinated by Schwann cells, Caspr, contactin, and NF155 completely overlap at paranodes (Fig. 5), presumably because PNS myelination is normal in md rats (Dentinger et al., 1982).

To determine whether the altered distribution of contactin, Caspr, Kv1.1, and Kv1.2 was associated with altered amounts of these proteins, we performed immunoblot analysis. As shown in Figure 4, the amount of contactin and Caspr were similar in P21 md rat spinal cords and in their WT littermates. Immunoblotting for Kv1.1 and Kv1.2 was unsuccessful. These results indicate that the aberrant myelination in md rats causes the redistribution of contactin and Caspr without altering their overall levels.

**Figure 4.** Immunoblot analysis of voltage-gated Na\(^+\) channels, contactin, and Caspr. Homogenates of spinal cords and sciatic nerves were prepared from P21 md and their WT littermates, and 100 \(\mu\)g of protein was analyzed for Na\(_{1.2}\), Na\(_{1.6}\), Na\(_{1.8}\), Caspr, and contactin. For Na\(_{1.2}\), Na\(_{1.6}\), Na\(_{1.8}\), the films were exposed for 20 min, then rehybridized with a mouse monoclonal antibody to GAPDH, and exposed to film for 5 min. For Caspr, the film was exposed for 30 sec, then rehybridized with a mouse monoclonal antibody to GAPDH, and exposed to film for 30 sec. For contactin, the film was exposed for 5 sec, then rehybridized with a mouse monoclonal antibody to GAPDH, and exposed to film for 2 min. The Na\(_{1.2}\), Na\(_{1.6}\), and Na\(_{1.8}\), bands were all \(250\) kDa; the Caspr doublet band \(190\) kDa; the contactin band \(135\) kDa. Note the similar amounts of Na\(_{1.2}\), Na\(_{1.6}\), Na\(_{1.8}\), Caspr, and contactin in md and WT samples.

**Figure 5.** Caspr and neurofascin are not localized to CNS paranodes in md rats. These images were made from longitudinal sections of P21 md (A–C) or WT (D–F) spinal cord, after double-labeling with a rabbit antiserum against NF155 (A, D, TRITC) and a mouse monoclonal antibody against Caspr (B, E, FITC); C and F show the merged images. In md rats, Caspr and neurofascin are colocalized in the paranodes in the ventral roots (arrows) but not in the spinal cord. The arrowhead marks an incisure, which is stained for NF155 but not for Caspr (Tait et al., 2000). In WT rats, Caspr and NF155 are colocalized at all CNS paranodes. Asterisks mark oligodendrocyte cell bodies, which are stained for NF155 but not Caspr (Tait et al., 2000); double arrowheads mark nodes. Scale bar, 10 \(\mu\)m.
Paranodal specializations in the absence of myelination?

The above data, taken together, indicate that the lack of paranodal specializations in md spinal cords results in the lack of separation between Kv1.1 and Kv1.2 K⁺ channels and voltage-gated Na⁺ channels. To investigate axon–oligodendrocyte interactions further, we examined P21 md spinal cords by electron microscopy, focusing on the ensheathment of the largest axons in the ventral funiculus, as these would normally be well myelinated. In transverse sections, the ventral funiculi had few myelinated fibers (<1% of axons larger than 2 μm in diameter); these had thin myelin sheaths whose characteristics have been previously described (Dentingier et al., 1982; Barron et al., 1987; Duncan et al., 1987; Rosenbluth, 1987). The few myelinated fibers that we found in longitudinal sections had disorganized paranodes; the glial endfeet were chaotically arranged, and even those apposed to the axolemma often did not terminate on it owing to the intrusion of astrocytic processes (Rosenbluth, 1987). Septate-like junctions–transverse bands were not seen even when the terminal loops directly apposed the axolemma (Rosenbluth, 1987).

We were particularly interested in one aspect that has not been previously emphasized; most large myelinated axons were individually ensheathed by glial processes (Fig. 6A,B). Many of these processes appeared to belong to oligodendrocytes because they contained relatively electron-dense cytoplasm and lacked intermediate filaments; some processes were focally devoid of cytoplasm, and thus appeared like a single wrap of compact myelin. In transverse sections, most large axons were at least partly surrounded by oligodendrocyte processes, often by more than one process, but some large axons also apposed astrocytes as well as other axons. In longitudinal sections (Fig. 6C–E), it was apparent that oligodendrocytic processes typically ensheathed large axons for short distances (<10 μm) and abutted other oligodendrocyte processes or even astrocytic processes. No paranodal specializations such as septate-like junctions were seen between oligodendrocyte processes and ensheathed axons.

Figure 6. Electron microscopy of P21 md spinal cord. These are electron micrographs of transverse (A, B) and longitudinal (C–E) sections of the ventromedial funiculus. In A and C, note several large axons (ax) that are not myelinated. A shows a portion of an oligodendrocyte (ol) with dilated cisternae; these are common in md rats. B shows one axon in higher magnification; note the multiple processes (asterisks) surrounding the axon. D and E show the rectangular regions; note the astrocytic processes (as) in D and the stack of five oligodendrocyte processes in E.
Oligodendrocytes ensheathe axons in *md* rats

The ultrastructure of *md* spinal cords indicated that many large axons are ensheathed by oligodendrocytes, yet are not myelinated. To visualize how oligodendrocytes ensheathe axons, we labeled longitudinal sections. We used antibodies that stained oligodendrocytes based on our previous study of *md* rats [Rip, MAG, myelin-oligodendrocyte glycoprotein (MOG), and MBP] (Grinspan et al., 1996), as well antisera against claudin-11 [also known as oligodendrocyte-specific protein (OSP)] (Table 1). Rip, MAG, MBP, MOG, and claudin-11/OSP antibodies labeled particularly well several myelin sheaths in most high-power fields. As shown in Figure 7, these well stained sheaths typically had one or more strands of claudin-11/OSP staining that extended from end to end, and bands of staining at the two ends, likely corresponding to paranodes (Gow et al., 1999; Morita et al., 1999). Although some of these highly stained sheaths probably correspond to the myelin sheaths seen by electron microscopy, they were much more numerous, demonstrating that many axons are ensheathed by oligodendrocyte processes but not myelinated. These studies also demonstrated that the Rip and MAG antibodies stain oligodendrocytes and their processes more completely than the other antibodies we examined (Fig. 6A–F).

The development of node-like clusters in *md* rat spinal cords

We wished to determine the relationship between node-like clusters and oligodendrocyte ensheathe. We selected antibodies that reliably stain oligodendrocytes (against MAG, Rip, MOG, MBP), and antibodies that labeled paranodes (against claudin-11/OSP, but not against contactin, Caspr, and NF155) combined with antibodies that label node-like clusters (either the pan-Na⁺ channel monoclonal antibody or the rabbit antisera against Na⁺, or ankryin_G). Double-labeled longitudinal sections of P21 *md* rat ventral funiculi revealed that node-like clusters of Na⁺ channels (double arrowheads) in WT rats, but many node-like clusters are not associated with paranodal OSP staining in *md* rats. Scale bars: A–C, G, H, 10 μm; D–F, 20 μm.

**Figure 7.** MAG-, MBP-, and OSP-positive oligodendrocytes ensheathe axons in *md* spinal cord. These images are taken from longitudinal sections of a P21 *md* (A–G) or a WT (H) spinal cord, double labeled with a rabbit antiserum against OSP (TRITC) and a monoclonal antibodies (FITC) against MAG (A–C), MBP (C–F), or pan-Na⁺ channels (G, H). As in WT rats (G), the paranodes in *md* rats contain a spiral of OSP staining (arrows), and (MBP- and MAG-positive) internodes—oligodendrocyte processes often contain OSP-positive strands. An asterisk marks an oligodendrocyte nucleus in A–C. In G and H, paranodal OSP staining (arrows) flanks most node-like clusters of Na⁺ channels (double arrowheads) in WT rats, but many node-like clusters are not associated with paranodal OSP staining in *md* rats. Scale bars: A–C, G, H, 10 μm; D–F, 20 μm.
of staining with these other antibodies made it difficult to find the paranodes (data not shown).

Most node-like clusters, however, were not adjacent to ensheathed axonal segments; many were in areas that lacked oligodendrocytes altogether (Figs. 7G, 8A, B). These results indicate that oligodendrocyte ensheathment may not be necessary for the formation node-like clusters, or that node-like clusters form adjacent to ensheathed axonal segments but persist after becoming isolated following oligodendrocyte cell death (Gow et al., 1998; Grinspan et al., 1998; Lipsitz et al., 1998). To evaluate the latter possibility, we labeled longitudinal sections of the ventral funiculus from the cervical cord from P7 (four md rats and four WT), P14 (two md and two WT), and P21 rats (two md and two WT) with the pan Na⁺ channel monoclonal antibody (to label nodes) and a rabbit antiserum against MAG (to label ensheathed axonal segments). We counted and classified the node-like clusters in relation to the MAG staining, as either clusters (not associated with MAG staining), heminodes (MAG staining present on one side of the cluster), or nodes (MAG staining present on both sides of the cluster); these data are summarized in Figure 9 and Table 2. Note that the percentage of nodes increases in WT rats from P7 to P21, whereas in md rats, the proportion declines; at P7, the percentage of nodes is similar between md and WT rats, but it is significantly different at P14 and P21. Thus, these data indicate that isolated node-like clusters in P21 md rats result from oligodendrocyte cell death during development.

In the developing optic nerve, node-like clusters of voltage-gated Na⁺ channels form adjacent to paranodes, as marked by Caspr staining (Rasband and Trimmer, 2001a). Our results also demonstrate that node-like clusters of voltage-gated Na⁺ channels develop adjacent to ensheathed axons, as marked by MAG staining. However, because contactin, Caspr, and NF155 are not localized to paranodes in P21 md rats, septate-like junctions do not appear to be necessary for the clustering of voltage-gated Na⁺ channels. To exclude the possibility that these components of septate-like junctions might be initially present, but lost by P21, we examined longitudinal sections of P7 and P14 rat spinal cord that were double-labeled with antisera against contactin, Caspr, or NF155 and the pan-Na⁺ channels monoclonal antibody. In contrast to WT rats, we did not find any evidence for paranodal clustering of contactin, Caspr, or NF155 in md rats (data not shown). Thus, although voltage-gated Na⁺ channels form adjacent to paranodes in WT and md rats, septate-like junctions do not appear to be necessary for this to occur.

**DISCUSSION**

The molecular organization of axons in md rats is severely altered, as summarized in Figure 10. Oligodendrocytes ensheath short segments of many large axons, but form few myelin sheaths. The axoglial junctions of these ensheathed axons, and even of the myelinated axons, lack septate-like junctions, and contactin, Caspr, and NF155 do not accumulate at paranodes. Nevertheless, node-like clusters form adjacent to ensheathed axonal segments and persist after oligodendrocyte cell death. The distribution of Na⁺, 1.2, Na⁺, 1.6, and Na⁺, 1.8 clusters, and the number Na⁺, 1.6 clusters in md rats are comparable to those in age-matched WT spinal cords. Kv1.1 and Kv1.2 abut and even overlap node-like clusters of voltage-gated Na⁺ channels and ankyrinG.

**Figure 8.** Clusters of Na⁺ channels and ankyrinG in regions devoid of oligodendrocytes. These images were taken from longitudinal sections of P21 md spinal cord, stained with a rabbit antiserum against MAG and a pan-Na⁺ channel monoclonal antibody (A; merged confocal images), or a rabbit antiserum against ankyrinG, and a mouse monoclonal antibody against MBP (B; merged epifluorescence images). Note the clusters of Na⁺ channels and ankyrinG staining in regions that are devoid of MAG–MBP staining (arrowheads), as well as adjacent to MAG–MBP-positive processes (arrows). An asterisk marks an oligodendrocyte nucleus. Scale bars: A, 10 μm; B, 20 μm.

**Figure 9.** Quantitative analysis of node-like clusters in the ventral funiculus. Longitudinal sections through the ventral funiculus of P7 (4 md and 4 WT), P14 (2 md and 2 WT), and P21 (2 md and 2 WT) were double-labeled with the pan Na⁺ channel monoclonal antibody (to label nodes) and the rabbit antiserum against MAG (to label ensheathed axonal segments). All node-like clusters of Na⁺ channels were classified as either naked clusters (not flanked by MAG-positive ensheathed axonal segments), heminodes (flanked on only side one by MAG-positive axonal segments), or nodes (flanked on both sides by MAG-positive axonal segments). The percentage of nodes was calculated at each age; ANOVA statistical analyses were used to compare the samples; the p values are shown for each comparison.
proteins. Nav1.2 is highly expressed by unmyelinated axons and in gray v

data show that different neurons express at least two other nodal

spinal cord of normal rats. The molecular organization of nodal

motic membrane staining of Na v1.8 have not been reported,

Although all nodes in adult optic nerve (a CNS myelinated tract) contain Nav1.6 (Caldwell et al., 2000; Boiko et al., 2001), our observations of WT spinal cord con

fi
differentiation of the axonal membrane in md rats. In this schematic image, the axon is depicted as intact, whereas the glial cells are depicted as being hemisected, to reveal the axoglial junctions. The localization of nodal (blue; voltage-gated Na” channels and ankyrinG), paranodal (red for contactin and Caspr; purple for NFI55), and juxtaparanodal proteins (green; Kv1.1, Kv1.2, Kvβ2, Caspr2) in WT rats are shown on the right. The left side of the figure depicts that in P21 md rats, nodal proteins can be localized with or without oligodendrocyte ensheathment, whereas contactin and Caspr are diffusely localized, and Kv1.1 and Kv1.2 abut the nodal membrane.

The localization of Na1,2, Na1,6, and Na1,8 in the spinal cord of normal rats

Our observations of WT spinal cord confirm and extend previous studies of voltage-gated Na” channels in the CNS. That the CST contains abundant Na1,2 is consistent with previous reports that Na1,2 is highly expressed by unmyelinated axons and in gray matter (Westenbroek et al., 1989), because the CST contains abundant unmymelinated axons (Langford and Coggshall, 1981). Although all nodes in adult optic nerve (a CNS myelinated tract) contained Na1,6 (Caldwell et al., 2000; Boiko et al., 2001), our data show that different neurons express at least two other voltage-gated Na” channels. Finally, to our knowledge, the somatic membrane staining of Na1,8 have not been reported, although Na1,8 mRNA has been detected (Schaller and Caldwell, 2000).

The organization axonal proteins in dysmyelinating mutants

There are two previous reports on the molecular organization of myelinated axons in md rats. The monoclonal antibody HNK-1 stains the optic nerve diffusely (Struckhoff et al., 1997), whereas it stains perinodal astrocytes in WT optic nerves (ffrench-Constatn and Raff, 1986). Thus, we expected to find diffuse tenascin-R staining, because tenascin-R is the only molecule localized to perinodal astrocytes that has an HNK-1 epitope (ffrench-Constatn et al., 1986). Kaplan et al. (1997) examined the optic nerves of P10 and P16 WT and md rats, and found fewer node-like clusters of voltage-gated Na” channels in md rats. Our findings demonstrate that the number of node-like clusters in the ventral funiculus of md rats is not reduced at P21. Whether this discrepancy is related to the later onset of myelination in the rat optic nerve (P10) (Hildebrand and Waxman, 1984; Trimmer and Wunderlich, 1990), as compared to the ventral funiculus (P1) (Baron et al., 1993), or another difference between the neuronal populations remains to be determined.

The molecular organization of myelinated axons has been examined in other genetic models of CNS dysmyelination. In Plp”isozy mouse, which have a similarly severe phenotype to md rats, Baba et al. (1999) described diffuse staining of Kvβ2 except where myelinated axons are formed, and one can presume that Kv1.1, Kv1.2, and Caspr2 would be similarly distributed. Homozygous shiverer mice have a recessive mutation that results in a complete absence of MBP, but they are much longer lived, and oligodendrocyte cell death is not a prominent feature. Like md rats, shiverer oligodendrocytes ensheath but do not myelinate axons; unlike md rats, shiverer mice have elaborate paranode-like specializations (Rosenbluth, 1981) that likely correspond to the pattern of Caspr and NFI55 staining described by Tait et al. (2000). The lack of normal paranodes in shiverer mice may explain why Kv1.1 and Kv1.2 staining in their CNS tracts (that would be myelinated in WT mice) was described as “diffuse” (Wang et al., 1995). How nodal clusters of Na” channels are affected in shiverer mice is less clear. The number of Na” channels are increased in shiverer optic nerve (Noebels et al., 1991), and that there appears to be a diffuse increase in Na1,2 channels (Westenbroek et al., 1992; Boiko et al., 2001). In contrast, using pan-Na” channel antibodies, Rasband et al. (1999) found node-like clusters of Na” channels in shiverer optic nerve, although fewer in number than in age-matched WT mice. These clusters were typically irregular in appearance, did not colocalize with ankyrinG, and about one-quarter of them were not adjacent clusters of Caspr staining. Recently, Boiko et al. (2001) reported a developmental delay in the appearance of Na1,6-positive node-like clusters in shiverer optic nerve. At P40, only a few nodes were Na1,6-positive in shiverer optic nerves (and these were the ones flanked by Caspr-positive paranodes), whereas all nodes were Na1,6-positive in WT optic nerves.

Table 2. Development changes in the proportion of nodes, heminodes, and clusters in md and WT rats

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<td>Nodes</td>
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The percentage of nodes increases in WT rats from P7 to P21, whereas in md rats, the proportion declines. Thus, at P7, the percentage of nodes is similar between md and WT, but it is significantly different at P14 and P21.
In contrast to some of the above findings, in the md rat spinal cord, the number of Na⁺ channel clusters was not decreased compared with WT male littermates, and all of these clusters were colocalized with ankyrinG. We did not observe diffusely increased levels of Na⁺ channel staining, with either the pan-Na⁺ channel antibodies, or the antisera specific for Na⁺,L, Na⁺,C, or Na⁺,V8, by immunostaining or immunoblot analysis. Moreover, the topology of Na⁺,L, Na⁺,C, or Na⁺,V8 staining was not altered in md spinal cord, calling into question the generality of the finding that Na⁺,L is “maintained” by the dysmyelination in shiverer optic nerve (Westenbroek et al., 1989; Caldwell et al., 2000; Boiko et al., 2001). The failure of Rasband et al. (1999) to find colocalization with ankyrinG could reflect a real difference between these mutant animals, or a technical issue, as they used a different antiserum against ankyrinG. The most striking difference, however, is the lack of clusters of contactin, Caspr, or NF155 in md rats as opposed to shiverer mice. The lack of these clusters nicely correlates with the lack of septate-like junctions—terminal bands between oligodendrocyte processes and axons in md rats, as compared with shiverer mice, in which there are abundant, but disorganized septate-like junctions at axo-glial junctions (Rosenbluth, 1981, 1987, 1995).

**Septate-like junctions are not required for the formation of node-like clusters**

Our data demonstrate that node-like clusters of voltage-gated Na⁺ channels and ankyrinG form adjacent to ensheathed axon segments and become isolated after oligodendrocyte cell death. Furthermore, these node-like clusters form in the absence of septate-like junctions or the paranodal accumulation of contactin, Caspr, or NF155. These data confirm and extend similar findings from mice that lack contactin (Boyle et al., 2001) or Caspr (Bhat et al., 2001), or in cgr-null mice, in which NF155 does not cluster at the axoglial junctions (Dupree et al., 1999; Poliak et al., 2001). In these mice, however, myelin sheaths are well formed, whereas in md rats, axons are ensheathed by oligodendrocytes but seldom myelinated. Thus, there appears to be a mechanism that causes nodes to form adjacent to ensheathed axons independent of septate-like junctions; whether this is related to the oligodendrocyte-derived clustering factor (Kaplan et al., 1997, 2001) remains to be determined.

**Paranodal specializations exclude Kv1.1 and Kv1.2 channels**

Our finding that Kv1.1 and Kv1.2 abut nodes has also been reported in contactin-, Caspr-, and cgr-null mice (Dupree et al., 1999; Bhat et al., 2001; Boyle et al., 2001; Poliak et al., 2001). The common denominator in all these mutant mice is that contactin, Caspr, and NF155 are mislocalized, and that septate-like junctions do not form. The absence of a stable complex of contactin, Caspr, and NF155 may allow the complex of Caspr2 (and its associated molecules, Kv1.1, and Kv1.2) to be maintained in the paranodal region by its interactions with band 4.1B (Poliak et al., 2001). These juxtaposed Kv1.1 and Kv1.2 channels likely interfere with the propagation of action potentials (Popko, 2000; Boyle et al., 2001) and may play the key role in the pathogenesis of demyelinating diseases.

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