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The Human Cathelicidin LL-37 Has Antiviral Activity against Respiratory Syncytial Virus

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Abstract
Respiratory syncytial virus is a leading cause of lower respiratory tract illness among infants, the elderly and immunocompromised individuals. Currently, there is no effective vaccine or disease modifying treatment available and novel interventions are urgently required. Cathelicidins are cationic host defence peptides expressed in the inflamed lung, with key roles in innate host defence against infection. We demonstrate that the human cathelicidin LL-37 has effective antiviral activity against RSV in vitro, retained by a truncated central peptide fragment. LL-37 prevented virus-induced cell death in epithelial cultures, significantly inhibited the production of new infectious particles and diminished the spread of infection, with antiviral effects directed both against the viral particles and the epithelial cells. LL-37 may represent an important targetable component of innate host defence against RSV infection. Prophylactic modulation of LL-37 expression and/or use of synthetic analogues post-infection may represent future novel strategies against RSV infection.

Introduction
Respiratory syncytial virus (RSV) is the major cause of viral bronchiolitis in young children worldwide, and a leading cause of morbidity and mortality in infants, elderly and immunocompromised individuals [1,2]. In addition to major morbidity (33.8 million episodes worldwide per year), RSV bronchiolitis also leads to significant mortality in the developing world with up to 200,000 deaths annually in young children worldwide [1]. RSV infection results in inflammatory lower respiratory tract disease in about 30% of infants who become infected, and, in ~2% of all infants, RSV bronchiolitis is potentially life-threatening and requires admission to hospital (reviewed in [3]). Although most infants in developed countries recover, these children have an increased risk of recurrent wheeze and RSV infection has been implicated in the later development of asthma [4]. Despite the prevalence of RSV bronchiolitis, susceptibility remains poorly understood, there is no vaccine available and, apart from supportive measures, there is no specific effective treatment (reviewed in [5]). A clearer understanding of the host defence factors that contribute to effective protection against RSV in the majority of those exposed to this virus should inform the development of novel prophylactic and/or therapeutic strategies.

Cationic host defence peptides (CHDP; also known as antimicrobial peptides) are key components of innate defences against infection, with both microbicidal and host defence modulatory functions (reviewed in [6]). In addition to their well described bactericidal potential, CHDP have more recently been shown to have antiviral properties (reviewed in [7]). Cathelicidins are multifunctional immunomodulatory CHDP expressed in a broad range of tissues in infection and inflammation (reviewed in [8,9]). The sole human cathelicidin, Human Cationic Antimicrobial Peptide of 18KDa (hCAP-18), from which the active form LL-37 is generated proteolytically [10], is expressed primarily by neutrophils and epithelial cells. Airway epithelial cell expression of LL-37/hCAP18 is induced in vitro by RSV infection [11], and is significantly enhanced by the 1,25OH metabolite of vitamin D [12]. In addition, a recent study of children with RSV bronchiolitis demonstrated that median serum levels of hCAP-18 were significantly lower in children with RSV bronchiolitis than in those with human rhinovirus induced bronchiolitis [13]. Furthermore, RSV-infected children whose hCAP-18 levels were lower than the median were found to be more likely to be hospitalised for prolonged periods than those with hCAP-18 levels above the median. These findings suggest an antiviral role for human cathelicidin in host defence against RSV.

In order to determine the antiviral potential of LL-37 against RSV, we examined the impact of LL-37 on RSV infection in vitro, establishing the effect of this CHDP on viral particles and its protective impact upon human epithelial cells.
Materials and Methods

Peptide synthesis

LL-37, TAMRA-labelled LL-37, scrambled LL-37 and truncated LL-37 partial peptides (all sequences in Figure 1a) were either purchased from Activec (Cambridge, UK) or synthesised as described below. Peptide purity was >95% by RP-HPLC and net peptide content determined by amino acid analysis. NovaSYN TGT resin preloaded with Fmoc-Ser(Bu)-OH (0.05 mM) was transferred to an automated peptide synthesiser (ABI 433A) reaction vessel for peptide chain elongation employing 10 equivalents of each standard Fmoc-protected amino acid employing the Fastmoc® protocol. Following chain elongation the fully protected resin-bound peptide was treated with trifluoroacetic acid: ethanedithiol: water (95: 2.5:2.5, 4.0 ml) for 5 hours. The resin was filtered off and the filtrate was added to cold diethyl ether (40 ml), which induced precipitation. The precipitate was collected by centrifugation at 3000 rpm, 4°C for 15 min. The ether layer was then decanted and the precipitated peptide was washed once more with cold diethyl ether (40 ml). The white precipitate was then dissolved in water and purified by semi-preparative reverse phase (RP)-HPLC (gradient: 5–60% acetonitrile/45 min). Fractions containing product were identified by LC-MS, pooled and lyophilised. Lyophilised peptides were reconstituted in endotoxin free water at 5 mg/ml stock concentration and determined to be >95% by RP-HPLC. Peptides were added before, after or concomitantly with RSV infection as described for individual experiments. Monolayers were fixed in absolute methanol/2% hydrogen peroxide solution for 30 minutes and washed twice with PBS. Cells were exposed to monoclonal biotinylated anti-RSV antibody in 5% BSA/PBS for 1 hour (1:200, AbD Serotec, Kidlington, UK); washed twice with PBS and were incubated with extravidin peroxidase in 5% BSA/ PBS for 30 minutes (1:500, Sigma Aldrich). Cells were stained using 3-aminocarbonylcarbazole substrate (Invitrogen/Life technologies) according to manufacturer’s instructions for 15 minutes, washed 3 times with PBS and incubated RSV positive foci were quantified by light microscopy, counting 3–4 wells per condition. For later timepoint analysis (48 and 72 hours post infection with RSV MOI = 0.0005), fixed and stained monolayers were examined by light microscopy and images were captured using an EVOS fl imaging system (PEQ-LAB Ltd, UK). Quantification was performed using the ImageJ plugin Treshold colour to measure the percentage of cells positive for RSV infection. 3 wells were assessed per condition, using 4 random fields of view captured per well. To assess the production of infectious viral particles, supernatants were serially diluted and transferred to fresh HEp-2 monolayers, incubated for 2 hours at 37°C, washed, incubated for a further 24 hours and then assessed by immunoplaque assay.

Direct viral inactivation assay

A modified viral inactivation assay [17] was performed. 1 ml RSV (PFU = 2 × 10^5, equivalent to MOI = 0.05) was mixed with 25 μg/ml LL-37, followed by immediate 10-fold dilution in serum-free medium to reach sub-therapeutic concentrations of LL-37 (2.5 μg/ml) and RSV MOI = 0.005. 25 μg/ml scrambled LL-37 or PBS were used as negative controls. HEp-2 monolayers were infected with this virus/peptide mixture for 2 hours then washed, before cells were cultured in serum-free medium for 24 hours. Fixed monolayers were subjected to immunoplaque assay.

Visualisation of TAMRA-labelled LL-37

HEp-2 cells were treated with unlabelled- or red-fluorescent tetramethylrhodamine (TAMRA)-labelled LL-37 at 25 μg/ml for 1 hour, followed by a PBS wash. Hoechst stain was added at 1 μg/ml for 30 minutes to visualise nuclei, and live cells were assessed by confocal microscopy, using a Leica SP5 confocal microscope with a 63× objective and Leica Application Suite Advanced Fluorescence software. A minimum of 4 fields of view were assessed for each condition.

Western blotting

HEp-2 cells were treated with LL-37 for 1 hour, or left untreated, followed by PBS washes or left unwashed. Cells were lysed on ice using lysis buffer (20 mM Hepes, 0.3 mM NaCl, 1.5 mM MgCl2, 0.2 mM EDTA, 1 mM DTT, 1 mM Orthova- nadate, 0.5% TritonX100) with Complete Protease Inhibitor Cocktail tablets (Roche Applied Science, UK; 1 tablet dissolved in 1 ml ddH2O and used at a 1 in 50 dilution). A BCA test was used to determine protein concentration, as per manufacturer’s instructions (Thermo Scientific, UK) and equal amounts of protein were incubated (5 min, 95°C) with sample buffer and reducing agent (Invitrogen). SDS polyacrylamide gel electrophoresis was performed, using 16% denaturing gels (100 V, 5 min, then 120 V, 85 min), before proteins were transferred to PVDF membranes by electro-blotting (100 V, 90 min). Membranes were probed with monoclonal antibodies against β-actin (1:2000; Sigma Aldrich) and the results were visualised using enhanced chemiluminescence.
blocked with 5% milk in PBS for 1 hour and incubated for 24 hours at 37°C, and then the number of infected cells was quantified. d) HEP-2 cells were exposed to LL-37 and RSV (MOI=0.005) simultaneously (“Simultaneous” condition), for 2 hours, or exposed for 2 hours to RSV that had been preincubated with LL-37 for 1 hour (“37°C pre-incubation” and “4°C pre-incubation” conditions), or exposed to RSV for 2 hours and washed before a 1 hour incubation with LL-37 (“Virus first” condition), or treated with LL-37 for 1 hour and washed before a 2 hour infection with RSV (“LL-37 first” condition). Every treatment was followed by a wash step. After 24 hours at 37°C, the number of infected cells was quantified. b, c & d) Infected cells were quantified using an immunoplaque assay and expressed as a percentage relative to untreated, RSV infected cells. Results are shown as mean ± SEM of n = 3-5. For statistical analysis, RSV-positive foci counts were log transformed and analysed by one way ANOVA with Dunnett Multiple Comparison Post-tests to compare treatments to untreated infected controls * p<0.05, ** p<0.01, *** p<0.001.

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Assessment of cytopathic effects

HEP-2 cells were treated with LL-37, a scrambled control LL-37 at 50 μg/ml or ribavirin (200 μM; Sigma Aldrich), in the presence or absence of simultaneous infection with RSV (MOI=0.005). Cells were incubated for 2 hours at 37°C, washed and placed in fresh media. In some wells LL-37 or ribavirin was added again at this point (for “continuous treatment”). Cells were cultured at 37°C for 24 hours or 5 days, after which the number of viable metabolically active cells was assessed using an MTT assay (Sigma Aldrich) according to manufacturer’s instructions (7.5 mg/ml; 37°C, 4 hours). Resulting formazan was dissolved in dimethylsulfoxide (Sigma Aldrich) and optical density was measured at 450 nm. Alternatively, cells simultaneously treated with or without LL-37 in presence or absence of RSV infection were allowed to dry for 10 minutes before they were fixed and permeabilised using ice-cold acetone/methanol (1:1) for 10 minutes at 4°C. After a wash with PBS, cells were subjected to TUNEL assay using an In situ cell death kit (Roche Applied Science, UK) according to manufacturer’s instructions. Cells were washed three times with PBS and 1 μg/ml DAPI was included in the second wash (3 minutes staining). Cells were examined by fluorescent microscopy, four representative images per well were captured using an EVOS fl imaging system (PEQLAB Ltd) and TUNEL positive and negative cells were quantified using ImageJ (http://rsweb.nih.gov/ij/index.html).

Statistical analysis

Statistical analyses were performed as described using Graphpad Prism version 5 for Windows (GraphPad Software, Inc., La Jolla, CA, USA). For statistical analysis, immunoplaque counts were log transformed. Statistical significance was assessed either

<table>
<thead>
<tr>
<th>Peptide</th>
<th>Sequence</th>
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<tbody>
<tr>
<td>LL-37 N-terminal</td>
<td>H2N-LLGDFFRKSKEGKEKRIVQIKDFDNLVPRTE5-SOH</td>
</tr>
<tr>
<td>LL-37 Core</td>
<td>H2N-KIGKEKRIVQIKDFDNLVPRTE5-SOH</td>
</tr>
<tr>
<td>LL-37 C-terminal</td>
<td>H2N-EFRRVQIKDFDNLVPRTE5-SOH</td>
</tr>
<tr>
<td>ScrLL-37</td>
<td>H2N-RSLEGTRPPVVLKSNRSKLEFKDGIKIRE5-FV5-SOH</td>
</tr>
</tbody>
</table>
using a one-way analysis of variance (ANOVA) with Dunnett Multiple Comparison post test, or a two-way ANOVA with Bonferroni’s post test where appropriate. p ≤ 0.05 was considered significant.

Results

LL-37 has antiviral activity against RSV

In order to assess the antiviral potential of LL-37 against RSV, HEp-2 human epithelial cells were infected with RSV and concomitantly treated with a concentration range of LL-37 peptide, then assayed by counting the number of RSV-infected cells by immunoplaque assay after 24 hours. This simultaneous exposure to virus and LL-37 resulted in a significant, concentration-dependent reduction in infectivity at ≥10 μg/ml of LL-37 (p < 0.05), compared to virus alone (Figure 1b). The same effect was observed when the human bronchial airway epithelial cell line 16HBE14o- was used (Figure 1c), demonstrating that the effect was not specific to the HEp-2 cell line. A scrambled LL-37 control peptide had no antiviral effect (Figure 1c), demonstrating that this was not a non-specific effect of a cationic peptide. In order to determine whether maximal antiviral effect required interaction between the peptide and virus in advance of infection, the sequence of peptide and virus addition to cell was then modified. As a mimic of the physiological encounter between RSV and LL-37 that might be expected to occur in the airway surface liquid in vivo before reaching epithelial cells, virus was pre-incubated with LL-37 (at 37°C or at 4°C, given the temperature sensitive nature of RSV[18]), before HEp-2 cell infection. These conditions, in which peptide may have effects on the cells, the virus or both, also resulted in significant antiviral activity (Figure 1d; LL-37 effect p < 0.05 by one-way ANOVA), resulting in approximately 1 log fewer infected cells at 25 μg/ml of LL-37 (p < 0.01 at 37°C and p < 0.001 at 4°C), but not substantially different to the simultaneous addition of peptide and virus without preincubation. In contrast, in these experiments, delaying LL-37 treatment until 2 hours after viral infection resulted in a loss of the antiviral effect (Figure 1d), indicating that LL-37 treatment after infection does not “rescue” infected cells. However, pre-incubation of HEp-2 cells with LL-37, followed by cell washing prior to infection, also resulted in significantly less RSV infection (p < 0.001; Figure 1d). These data demonstrate that LL-37 has significant antiviral activity against RSV infection of epithelial cells in vitro and suggest that this peptide may exert its effects by directly affecting the viral particle and/or by acting on epithelial cells to reduce their susceptibility to infection.

The antiviral effects of LL-37 are protective against cell death

We have previously demonstrated that LL-37 can induce apoptosis in Pseudomonas aeruginosa-infected epithelial cells [19]. In order to determine whether similar mechanisms could lead to reduced infection through epithelial cell death, or whether LL-37 was actively protecting cells from infection, we examined the number of viable, metabolically active HEp-2 cells by MTT assay and induction of cell death by TUNEL assay. The treatment of RSV-infected cells with LL-37 (≥50 μg/ml) did not have any substantial effect on the number of metabolically active HEp-2 cells at 24 hours (Figure 2a), nor any substantial effects on the low background levels of apoptosis in this cell line (0.5–1.5% of HEp-2 cells were found to be TUNEL positive in the presence or absence of RSV or LL-37 at 24 hours after infection; data not shown). In order to determine longer term effects, treated cells were examined for up to 5 days. The treatment of uninfected cells with LL-37

Figure 2. LL-37 is protective against RSV-induced cell death. HEp-2 cells were treated with LL-37 at 0–50 μg/ml, a scrambled control LL-37 at 50 μg/ml (Scr 50), or ribavirin (200 μM), in the presence (a & c) or absence (b) of simultaneous infection with RSV (MOI = 0.005). Cells were incubated for 2 hours at 37°C, washed and placed in fresh medium. In some wells LL-37 or ribavirin was added again at this point (c; “continuous treatment”). Cells were cultured for 24 hours (a) or 5 days (b & c), after which the number of viable metabolically active
LL-37 Possesses Anti-RSV Activity

LL-37 directly affects RSV viral particles

In order to determine whether LL-37 could have a direct effect upon RSV viral particles, a concentrated viral stock was exposed to LL-37 at 25 µg/ml, immediately followed by a 1 in 10 dilution, reducing the concentration of LL-37 to which cells were exposed to below the effective antiviral level (2.5 µg/ml) before incubation with HEp-2 cells. This was compared to pre-mixing of virus (at standard concentration) with LL-37 at 25 µg/ml before infection of HEp-2 cells (Figure 3a). The combination of virus exposure to 25 µg/ml LL-37, with cell exposure to 2.5 µg/ml LL-37, had significant antiviral effects (p<0.001), whereas the combination of virus and cell exposure to 2.5 µg/ml LL-37 had no significant effects and was significantly different from the former (p<0.01). No effect was observed using scrambled LL-37 under the same dilution conditions. These data demonstrate that the antiviral properties of LL-37 against RSV are at least partly due to a direct interaction with the viral particles, an effect that was separable, under these conditions, from any peptide-mediated effects on the cells.

LL-37 modulates epithelial cell susceptibility to RSV infection

Having demonstrated that pre-treatment of epithelial cells with LL-37 could protect against subsequent infection by RSV (Fig. 1c), the longevity of that protective effect was determined by delaying RSV infection of cells for a variable period of time after LL-37 treatment and washing of cells (Figure 3b). A significant protection against RSV infection was observed for up to 3 hours after LL-37 treatment and cell washing (p<0.001). However, a time dependent loss of protection was observed, with no protection evident 24 hours after LL-37 treatment and cell washing.

The pre-treatment of cells with LL-37, followed by washing of the cells, was conducted in an effort to remove LL-37 from the cell culture system before addition of RSV. However, we have previously demonstrated that LL-37 can interact with epithelial...
LL-37 can protect epithelial cells against spread of virus infection

In order to examine the effects of LL-37 on the production of new infectious viral particles, HEp-2 cells were infected with RSV in the presence or absence of LL-37 for 2 hours, then cultured in fresh media for up to 3 days, with or without the continued presence of LL-37 or scrambled LL-37. At selected timepoints, infectivity of the supernatant from these cells was assessed for its ability to infect fresh HEp-2 cell cultures (Figure 5a). Treatment with LL-37 (but not scrambled LL-37) resulted in the production of significantly more (>0.001), and very substantially, fewer new infectious viral particles, even reducing infectivity up to day 3 (statistically significantly) (Figure 3a). The presence of LL-37 or scrambled LL-37 was assessed by western immunoblot. In the previous experiment (Figure 1d), addition of LL-37 at 2 hours after RSV infection had no effect on the number of infected cells at 24 hours. However, the effect on production of new infectious viral particles suggested that LL-37 might prevent the spread of infection from initially infected cells to uninfected cells in the culture over a longer time course. In order to examine this in vitro, HEp-2 cells were infected with a lower inoculum and evaluated by immunoplaque assays over 3 days (Figure 5b). The number of infected cells dramatically increased by day 2 in untreated cultures (Figure 5b, panel 1) and those exposed to scrambled LL-37 (Figure 5b, panel 4), rapidly reached uncountable levels of infected cells with death and substantial loss of cells by day 3 (Figure 5b, panels 5 & 8). In contrast, this spread of infection beyond the cells initially infected was minimised when LL-37 had been added simultaneously with RSV (Figure 5b, panels 2 & 6), or indeed even after RSV infection had been established (Figure 5b, panels 3 & 7). A quantitative image analysis approach to these data significantly underestimates the number of infected cells where cell death and loss has occurred, but nevertheless demonstrates significant protection against spread of
infection from initially infected cells over 2 to 3 days, regardless of whether LL-37 was added concomitantly with, or after viral infection (Figures 5c & d). These data demonstrate that LL-37 can significantly inhibit the spread of newly produced infectious particles.

The antiviral activity of LL-37 is retained by a truncated peptide fragment

Having established the anti-RSV activity of LL-37, partial LL-37 peptides were utilised to determine the antiviral properties of truncated forms. Three 22-mer peptides [24] were used, which correspond to the N-terminal (amino acids 1–22), central core (amino acids 12–33) and C-terminal (amino acids 16–37) sequence of the LL-37 peptide (Figure 1a). Simultaneous treatment of HEp-2 cells with RSV and peptides from this panel revealed significant antiviral activity for full length LL-37 and the central core fragment, but no effects for the N-terminal and C-terminal fragments (Figure 6). These data demonstrate that shorter peptide analogues, more amenable to the development of novel therapeutics, retain antiviral activity.

Discussion

RSV is a major cause of morbidity and mortality globally, particularly in infants, for which there is currently no effective vaccination or treatment available. In experimentally induced RSV infection in adult volunteers, early high viral titres have been found to correlate with severity of inflammatory disease manifestations [25]. In addition, more severe disease in infants is associated with high viral titres in early stages of naturally-occurring RSV bronchiolitis [26]. Thus, components of innate host defence with the capacity to reduce viral load would be expected to prevent or reduce severe disease. Here, we demonstrate for the first time that the cationic host defence peptide LL-37 has effective antiviral activity against RSV in vitro, implicating this
peptide as a targetable host defence factor with a potentially key role in susceptibility to RSV infection in vivo.

LL-37 is a pleiotropic CHDP, cleaved from the sole human cathelicidin hCAP-18. This peptide can be detected in a broad range of tissues and bodily fluids including plasma, bone marrow, airway surface fluid, skin, sweat, reproductive tract, semen, urine, breast milk and vernix (reviewed in [27]). Produced primarily by neutrophils and epithelial cells, hCAP-18/LL-37 is upregulated in response to infection, inflammation and wounding, and detected at higher concentrations in the bronchoalveolar lavage fluid of individuals in the context of lung infections, reaching average levels of approximately 20–25 µg/ml [28,29]. More specifically, the expression of CAMP (the gene encoding hCAP-18) can be regulated by the active vitamin D metabolite 1,25-dihydroxyvitamin D3 (1,25OH Vit D3), acting via a vitamin D response element in the CAMP gene promoter [12,30,31], an effect that can promote RSV-induced hCAP-18 production [11]. In addition to well described bactericidal properties, we, and others, have recently demonstrated that LL-37 has antiviral activity against a number of viruses (reviewed in [7]), including influenza in vitro [32,33] and in vivo [34]. Although the specific mechanisms involved remain largely undetermined.

We now demonstrate that LL-37 has concentration-dependent antiviral activity against RSV in vitro, capable of decreasing the number of initially infected cells by approximately 1 log at the physiologically relevant [28,29] concentration of 25 µg/ml. LL-37 prevented virus-induced cell death in epithelial cultures, significantly inhibited the production of new infectious particles and diminished the spread of infection. The effect was maximal when peptides were premixed with virus, or added simultaneously. However, significant depression of the extent of viral infection after three days was seen (Figure 5) even when the peptide was not added until two hours after infection of the cells, despite no effect on the number of entry events being observed at 24 hours under these conditions (Figure 1c). The specific events in the infection cycle that are affected by LL-37 remain to be determined.

Nevertheless, these data suggest that higher baseline endogenous levels of LL-37 and/or enhanced production of LL-37 in response to RSV infection would offer protection in the lung. This hypothesis was validated by the recent study demonstrating clinical associations between low serum cathelicidin levels in children and both the likelihood of RSV bronchiolitis and the severity of disease [13]. These studies raise the possibility that interventions capable of enhancing upregulation of LL-37 levels in vivo could be used in RSV infection, given prophylactically in high risk individuals, such as infants predisposed to repeated RSV bronchiolitis.

The development of strategies to induce LL-37 expression is currently focused on the use of drugs such as butyrate and/or the use of vitamin D supplementation (reviewed in [34]). Butyrate is a short-chain fatty acid produced by bacterial fermentation of dietary fibre in the colon. 4-phenylbutyrate (PBA), an analogue of butyrate that is already licenced for use in humans, can effectively upregulate hCAP-18/LL-37 expression in vitro, including in airway epithelial cells [35], and in vivo in a model of Shigella infection [36] and has potential where more acute, short term upregulation is required. Low serum 25(OH) VitD3 levels, and the decrease in levels observed during winter [37], are associated with the risk of respiratory infections, with a 7% reduction in adult respiratory infection reported for every 10 nM/l increase in serum 25(OH) VitD3 reported [38]. In addition, newborns with low cord blood 25(OH) VitD3 levels were found to have a significantly greater risk of developing RSV-associated lower respiratory tract infections [39]. Expression of hCAP-18/LL-37 can be regulated by vitamin D [12,30,31] and further upregulated by exposure to RSV in the presence of vitamin D [11]. An association between serum 25(OH) VitD3 and hCAP-18/LL-37 expression has not yet been definitely demonstrated in vivo. Nevertheless, these studies raise the possibility that vitamin D supplementation (particularly in winter in the northern hemisphere), should be considered as a novel preventative strategy against RSV infection. Furthermore, LL-37 expression may represent a candidate mechanism involved in understanding these observations.

In addition to prophylactic modulation of LL-37 expression, our data raise the possibility that LL-37 peptide administration may also have potential in post-exposure prophylaxis, given in the early stages after infection to minimise the spread of RSV in the epithelium, viral load and thus the severity of disease. In vivo modelling is required to evaluate the efficacy of such an approach, but the observation that antiviral activity is retained by a truncated peptide demonstrates the potential to develop shorter synthetic analogues for future testing. Interestingly, our evaluation of these previously characterised [24] partial peptides representing the N-terminus (amino acids 1–22), a central portion (amino acids 12–33) and the C-terminus (amino acids 16–37) of LL-37 suggests that the antiviral activity may relate to the same core peptide region previously identified for antibacterial activity [24,40] and for the capacity to induce secondary necrosis of apoptotic cells [41].

The specific mechanisms by which LL-37 has antiviral activity against RSV are now under evaluation, but remain to be determined. Our data demonstrate that LL-37 is capable of having a direct effect on the viral particles. This was recently demonstrated to occur in LL-37 treatment of influenza virus [33]. However, damage to the influenza viral membrane did not alter the binding or initial uptake of virus by cells, but was proposed to affect viral propagation or survival downstream of this. Interestingly, similar to our observations with RSV, LL-37 was also found to have antiviral activities against influenza when cells where exposed to peptide in advance of infection [33]. However, in contrast, LL-37-dependent inhibition of HIV-1 infectivity is
mediated by inhibition of HIV-1 reverse transcriptase and protease and does not require direct peptide-virus interaction [42,43]. Furthermore, LL-37-mediated antiviral activity against the non-enveloped adenovirus has also been reported [44], suggesting important activities other than direct damage to viral envelopes.

Our data demonstrate that LL-37 treatment in advance of viral infection results in peptide retention and internalisation by the epithelial cells, providing significant protection for at least 3 hours after cell washing. This is in keeping with our previous observations of active cellular uptake of LL-37 [20], and suggests that either RSV may interact with cell-associated LL-37 or that LL-37 may induce an antiviral state in the epithelial cells. Interestingly, human β-defensin (hBD)2 has been shown to destabilise the RSV viral envelope upon contact with soluble hBD2 in solution, or following exposure to plasma membrane-associated hBD2 during cell entry [45]. Although LL-37 may function in a similar manner, the well described capacity of LL-37 to modify cellular responses to inflammatory stimuli raises the possibility that host defence modulatory capacity is important.

LL-37 has been demonstrated to modulate signalling downstream of pattern recognition receptors and differentially induce and inhibit certain cytokine and chemokines responses, with the potential to modify the nature of host responses to bacterial and viral infections [14,46,47]. Interestingly these effects are observed at peptide concentrations (typically 1–5 μg/ml) that are lower than those required for microbialicidal activity. Although our data indicate that LL-37-mediated modulation of cellular type I interferon responses is not required for the antiviral activity against RSV observed in this study, the capacity of cathelicidins to modulate cellular responses to TLR3, −7, −8 and −9 agonists [22,47,48] may prove to be significant. Indeed, the dramatic effects of LL-37 on cytokine production and survival in influenza infected mice, in comparison to the relatively modest effect on viral loads in vivo [32], also suggests that peptide-mediated modulation of inflammation and immunity may be an important component of their antiviral function. In addition to evaluating the impact of cathelicidins in vivo and the capacity for direct and indirect antiviral activities of these peptides to act simultaneously, future studies should also determine the extent to which synergies exist between cathelicidins and other CHDP (such as defensins) which have been shown to have activity against RSV.

In conclusion, this study demonstrates a novel antiviral activity for the sole human cathelicidin LL-37 against RSV, an important human pathogen for which no effective disease-modifying therapies exist. It implicates hCAP-18/LL-37 as an important, targetable component of innate host defence against RSV and suggests future potential in strategies aimed at prophylactic modulation of cathelicidin expression in vulnerable individuals and/or the development of synthetic peptide analogues for use in post-exposure prophylaxis.

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Author Contributions

Conceived and designed the experiments: SMC EGF HW PF JS DJD. Performed the experiments: SMC BM AM TM HW. Analyzed the data: SMC EGF HW PF JS DJD. Contributed reagents/materials/analysis tools: DM. Wrote the paper: SMC EGF DM PF JS DJD.

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