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Camk2n1 Is a Negative Regulator of Blood Pressure, Left Ventricular Mass, Insulin Sensitivity, and Promotes Adiposity

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See Editorial Commentary, pp xxx–xxx

Abstract—Metabolic syndrome is a cause of coronary artery disease and type 2 diabetes mellitus. Camk2n1 resides in genomic loci for blood pressure, left ventricle mass, and type 2 diabetes mellitus, and in the spontaneously hypertensive rat model of metabolic syndrome, Camk2n1 expression is cis-regulated in left ventricle and fat and positively correlates with adiposity. Therefore, we knocked out Camk2n1 in spontaneously hypertensive rat to investigate its role in metabolic syndrome. Compared with spontaneously hypertensive rat, Camk2n1−/− rats had reduced cardiorenal CaMKII (Ca2+/calmodulin-dependent kinase II) activity, lower blood pressure, enhanced nitric oxide bioavailability, and reduced left ventricle mass associated with altered hypertrophic networks. Camk2n1 deficiency reduced insulin resistance, visceral fat, and adipogenic capacity through the altered cell cycle and complement pathways, independent of CaMKII. In human visceral fat, CAMK2N1 expression correlated with adiposity and genomic variants that increase CAMK2N1 expression associated with increased risk of coronary artery disease and type 2 diabetes mellitus. Camk2n1 regulates multiple networks that control metabolic syndrome traits and merits further investigation as a therapeutic target in humans. (Hypertension. 2019;74:00-00. DOI: 10.1161/HYPERTENSIONAHA.118.12409.)

Key Words: adiposity ■ blood pressure ■ hypertrophy ■ metabolic syndrome ■ rats

Metabolic syndrome (MetS) affects one in 4 people and is a major cause of coronary artery disease and type 2 diabetes mellitus (T2DM)1 MetS genetic determinants have been successfully elucidated in the spontaneously hypertensive rat (SHR), an established model of human polygenic MetS.2 Our previous studies identified Camk2n1 as a cis-regulated expression quantitative trait locus (cis-eQTL) in left ventricle (LV) and epididymal adipose tissue (EAT),3 and as a quantitative trait transcript that significantly positively correlates with relative fat pad weight ($r^2=0.67$, $P_{adj}=0.0002$) and adipocyte volume ($r^2=0.69$, $P_{adj}=0.0002$).4 Furthermore, Camk2n1 is close to the peak logarithm of the odds (to the base 10) score in QTLs for systolic blood pressure (BP; Bp292, Bp180, 433, and 441 Kb from peak) and relative LV weight (Cm24 and 441 Kb from peak) and resides in a QTL for T2DM (Niddm30).5 Camk2n1 has been reported through in vitro and in vivo peptide inhibition studies to be a specific inhibitor of CaMKII (Ca2+/calmodulin-dependent kinase II),6 an enzyme activated by Ca2+/calmodulin binding, which regulates multiple signaling pathways that control vascular tone,7 in vitro adipogenesis,8 and insulin-stimulated glucose uptake.9 However, the in vivo functions of endogenous Camk2n1 in cardiometabolic disease have not been studied directly, whereas experimental inhibition of CaMKII has been extensively studied because of the strong association between CaMKII hyperactivity, LV hypertrophy, and heart failure in humans.10 CaMKII inhibition has been induced experimentally using synthetic and transgenic CaMKII inhibitors. These studies have shown that such inhibitors can protect from Ang II (angiotensin II)–induced hypertension7 and LV hypertrophy.11 However, these CaMKII inhibitors have known off-target effects independent

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of CaMKII and may not mimic fully endogenous Camk2n1 functions.\textsuperscript{12} In other studies, knockout of CaMKII has been shown to protect from pressure-overload LV dysfunction but not prevent LV hypertrophy.\textsuperscript{13} CaMKII knockout improved hepatic insulin signaling in obese mice, while enhanced CaMKII activation has been shown to induce hyperinsulinemia and glucose intolerance.\textsuperscript{3,4} Taken together, these data suggest a potential causal role for Camk2n1 in cardiometabolic disease.

Here, we test the hypothesis that Camk2n1 regulates cardiometabolic traits by generating a Camk2n1 knockout in the SHR model of MetS.

**Methods**

The authors declare that all supporting data are available within the article and detailed methods and supplementary results in the online-only Data Supplement.

**Rats**

SHR-Camk2n1\textsuperscript{−/−} knockout rats (referred to hereafter as Camk2n1\textsuperscript{−/−} rats) were generated on an SHR/NCrl background (Charles River, Margate, United Kingdom), containing a 38bp deletion in exon 1 of Camk2n1 confirmed by whole genome sequencing, polymerase chain reaction, and Immunoblot (Figure S1A through S1C in the online-only Data Supplement). All procedures were performed in accordance with UK Home Office regulations.

**Human Participants**

Visceral adipose tissue was obtained intraoperatively following ethical approval from 28 lean, obese, and obese type 2 diabetic subjects. All statistics were performed using Minitab Express (v1.5.1).

**Results**

**Camk2n1 Knockout Rat**

We generated a Camk2n1\textsuperscript{−/−} rat using zinc finger nuclease that created a 38 bp deletion in exon 1 of Camk2n1 confirmed by a polymerase chain reaction, Sanger sequencing, and whole genome sequencing (Figure S1A). Truncation of the transcript and absence of Camk2n1 protein were confirmed by a polymerase chain reaction and immunoblot, respectively (Figure S1B and S1C).

**Blood Pressure**

To determine the cardiovascular consequences of Camk2n1 deletion, we measured BP in SHR and Camk2n1\textsuperscript{−/−} rats. Mean systolic BP (Δ12 mmHg, \(P<0.001\)) and diastolic BP (Δ10 mmHg, \(P<0.005\)) BPs were significantly lower in Camk2n1\textsuperscript{−/−} than SHR, and although heart rate was similar (SHR=299±3 Camk2n1\textsuperscript{−/−}=305±6, \(P>0.05\)), rate pressure product was reduced significantly by 5% (Figure 1A through 1C).

To test whether lower BP in Camk2n1\textsuperscript{−/−} rats was associated with altered vasodilatory mechanisms, we tested in vivo vasoreactivity to acetylcholine in the presence and absence of N ω-Nitro-L-arginine methyl ester hydrochloride or Ang(1–7) antagonist A-779 and analyzed the ACE2 (angiotensin II–converting enzyme)-Ang-(1–7)-Mas pathway. Area under the blood pressure curve responses to the lowest level of acetylcholine infusion were similar in SHR and Camk2n1\textsuperscript{−/−} (Figure 1D). At higher doses of acetylcholine, Camk2n1\textsuperscript{−/−} had a similar response in the presence or absence of Nω-Nitro-L-arginine methyl ester hydrochloride, whereas responses in SHR treated with Nω-Nitro-L-arginine methyl ester hydrochloride were reduced (Figure 1D). These changes were associated with increased renal ACE2 activity and Ang (1–7) concentrations in Camk2n1\textsuperscript{−/−} kidney and serum (Figure 1D; Table). In addition, renal and serum eNOS (endothelial nitric oxide synthase) and serum nitrate levels were elevated in Camk2n1\textsuperscript{−/−} compared with SHR (Figure 1E; Table). Conversely, CaMKII activity was reduced by 50% (Figure 1F). A-779 did not affect BP modulation (\(P>0.05\), Figure S2E).

In Camk2n1\textsuperscript{−/−} rats, kidney wet mass was reduced by 4%, but no differences in hypertension-related vascular damage or kidney function markers were observed (Figure S2A and S2B; Table).

**LV Mass**

To determine the effects of Camk2n1 knockout on LVH, we investigated LV structure and function at baseline and with isoproterenol-induced (CaMKII-associated) hypertrophy in SHR and Camk2n1\textsuperscript{−/−} rats.

LV mass at baseline was reduced by 9% in Camk2n1\textsuperscript{−/−} compared with SHR, whereas baseline heart mass was similar (Figure 2A and 2B). Morphological signs of inflammation and fibrosis, blood vessel and cardiomyocyte density and morphology in LV were similar for both genotypes (Figure 2C and S2D).

To test whether Camk2n1 deficiency would protect against isoproterenol-stimulated LVH, we performed a 72 hours isoproterenol hypertrophic challenge. Isoproterenol treatment caused similar increases in heart mass and rate and reductions in BP in SHR and Camk2n1\textsuperscript{−/−} (Figure 2A and 2B; Figure S3A through S3C). However, rate pressure product increased in SHR only (Figure 2C).

CaMKII activity and hypertrophy-related transcripts Acta, Actc, and Nppb were assessed in LV. CaMKII activity was reduced by 23% in Camk2n1\textsuperscript{−/−} compared with SHR LV (Figure 2D). Acta1 transcripts were 1.5-fold greater in saline-treated Camk2n1\textsuperscript{−/−} than in SHR, and after treatment with isoproterenol, Acta and Actc increased in SHR LV only, thereby increasing the ratio of Acta1:Actc1 in SHR compared with Camk2n1\textsuperscript{−/−} LV (Figure 2E). Camk2n1 deletion did not affect Nppb expression (Figure 2F).

**Insulin Sensitivity**

To establish a function for Camk2n1 in glucose metabolism, we measured the effectiveness of endogenous insulin to stimulate peripheral tissues glucose uptake, after an oral glucose bolus. Fasting plasma glucose concentrations in Camk2n1\textsuperscript{−/−} were significantly lower than SHR (\(P<0.01\); Figure 3A), although, area under the glucose curve was similar (\(P>0.05\)). Plasma insulin concentrations at \(t_0\) and \(t_{10}−t_0\) minutes and area
under the insulin curve following glucose bolus were significantly reduced together with homeostatic model assessment of insulin resistance (SHR, 1.22±0.06, Camk2n1−/−, 0.51±0.06, P=0.00001) in Camk2n1−/− compared with SHR (Figure 3A).

Adiposity
To assess a causal role for Camk2n1 in adiposity, we assessed adipose tissue mass, morphology, and adipocyte function of SHR and Camk2n1−/− fat pads. Relative masses of visceral (EAT, mesenteric, and retroperitoneal adipose tissue) and brown adipose tissue were significantly reduced in Camk2n1−/− compared with SHR, despite similar growth rate and total body mass (P>0.05; FigureS4A) with an overall 23% reduction (P=0.004) in total relative fat mass (Figure 3B). Subcutaneous fat mass was unaltered by Camk2n1 deletion (Figure 3B). Morphometric assessment of EAT showed that the reduced fat mass in Camk2n1−/− rats was associated with a reduction in adipocyte number rather than adipocyte volume (Figure 3B).

To determine whether Camk2n1 knockout altered CaMKII activity, visceral fat respiration, and whole-body adipokine production, we analyzed CaMKII activity, cellular energetics in epididymal adipocytes, and quantified serum concentrations of high molecular weight adiponectin and leptin in SHR and Camk2n1−/− rats (Figure 3D). Mitochondrial oxygen consumption: basal, ATP-linked, maximal, and leak respiration were similar in SHR and Camk2n1−/− adipocytes (Table S3). However, we found a 20% decrease (P=0.038) in circulating HMW adiponectin and a significant 70% decrease (P=0.01) in circulating leptin (Table). Given the reduction in fat mass and circulating leptin, we assessed food intake, body temperature, and locomotor activity as indicators of energy homeostasis in SHR and Camk2n1−/− rats. Food intake was reduced in Camk2n1−/− from 6 to 8 weeks age, whereas from 9 to 10 weeks of age, food intake was greater compared with SHR (Figure 3D); across the 6 to 10 week period, average food intake was not significantly different (P>0.05; Figure 3D, inset). There were no significant differences in body temperature and locomotor activity (P>0.05; Figure S4B and S4C).

In Vitro Camk2n1 Knockdown in 3T3-L1 Adipocytes
To establish whether Camk2n1 deficiency reduced adipogenic capacity, we knocked down Camk2n1 expression in mouse 3T3-L1 fibroblasts. Camk2n1 knocked down of 85% to 96% was confirmed from day 1 to 9 following lipofectamine treatment (Figure S5A) and was associated with a significant reduction in lipid formation assessed by oil red O staining intensity and reduction in the proportion of cells...
Table. Serum and Urinary Biochemical Analyses

<table>
<thead>
<tr>
<th>Fluid</th>
<th>Parameter</th>
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<th>Camk2n1−/−</th>
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</thead>
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<td></td>
<td>Calcium, mmol/L</td>
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<td>2.41±0.02†</td>
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<td></td>
<td>Chloride, mmol/L</td>
<td>100.0±0.4</td>
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<td>Potassium, mmol/L</td>
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<td>Sodium, mmol/L</td>
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<td>Creatinine</td>
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<td>54.9±0.7</td>
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<td>Urea</td>
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<td></td>
<td>Adiponectin HMW, ng/mL</td>
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<td>2.39±0.09†</td>
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<td>Leptin, ng/mL</td>
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<td>Urine</td>
<td>eNOS, pg/mL</td>
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<tr>
<td></td>
<td>Ang-(1–7), pg/mL</td>
<td>78±6</td>
<td>101±6†</td>
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<td>Potassium, mmol/L</td>
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<td>Creatinine, mmol/L</td>
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<tr>
<td></td>
<td>Protein, g/L</td>
<td>1.76±0.39</td>
<td>1.17±0.16</td>
</tr>
</tbody>
</table>

Mean±SEM, n=10/group for analyses; n=5–7/group for adiponectin, leptin, eNOS, nitrate, and angiotensin-(1–7) (Ang-(1–7)). Ang indicates angiotensin; eNOS, endothelial nitric oxide synthase; HMW, high molecular weight; and SHR, spontaneously hypertensive rat.

*P<0.01.
†P<0.05.

containing lipids by day 9 (P<0.05; Figure 3E). Markers of adipogenesis, Cebpa and Pparg were significantly altered by Camk2n1 deficiency. Cebpa and Pparg expression increase from day 1 to 9 in scramble treated cells, whereas expression of these genes was significantly perturbed by Camk2n1 siRNA by 63% (P<0.01) and 80% (P<0.001) on day 9, respectively (Figure 3F). By day 9, the mature adipocyte markers Lep and Adipoq in siRNA-treated cells were similar to control (Figure SSB).

Transcriptomics

The molecular networks regulated by Camk2n1 were defined by weighted gene coexpression analysis (WGCNA) of the SHR and Camk2n1−/− LV and EAT transcriptomes. Differential expression was validated in LV and EAT by comparing expression of 12 genes (Table S4).

In LV, there were 192 differentially expressed genes (DEGs), 118 DEGs were ≥2-fold different between genotypes (P adj<0.05; Figure 4A; Table S5). Of the most significant DEG, Kirrel3, Fabp4, Atf3, Capn3, Hdac4, and Pcb4 control cardiomyocyte size and function, whereas Cj1, Irak3 Rnf144b, and Hspa8 regulate inflammatory processes. WGCNA of the LV transcriptome defined 8 modules with significant correlation to Camk2n1 (Figure 4B). Kyoto Encyclopedia of Genes and Genomes functional enrichment analysis of these modules showed that Camk2n1−/− LV are significantly enriched for cell function and maintenance, intracellular signaling, metabolism, and antigen presentation networks (Table S6). GO analysis showed that across modules, there were consistent themes relating to transcript and protein regulation (Table S7). Levels of FABP4 (fatty acid binding protein 4) in LV, a protein that regulates several enriched pathways, was reduced in Camk2n1−/− compared with SHR (Figure 4C).

In EAT, there were 129 DEGs after adjustment for multiple testing; 90% of the most DEGs (≥2-fold) were downregulated with 20% associated with metabolic pathways (Pla2g2a, P4ha3, Pla2g5, Slc39a8, Pcdh7, and Gcn2) and cell proliferation (Cldn15, Fg9, Diaph3, and Mki67; Figure 4D; Table S8). WGCNA of the EAT transcriptome defined 8 modules correlated significantly with Camk2n1 (Figure 4E; Table S9). Five modules were significantly enriched in Camk2n1−/− EAT for KEGG pathways regulating cell maintenance and survival, and innate immunity and antigen presentation (Table S9), with transcript regulation and immune defense GO terms significantly enhanced (Table S10).

Soluble phospholipase A2 activity, which regulates adipogenesis and apoptosis, was reduced significantly in Camk2n1−/− compared with SHR (Figure 4F).

Human CAMK2N1 Cis-eQTL and Cardiometabolic Trait Analysis

We analyzed human data from the GTEx and Type 2 Diabetes Knowledge Portals to investigate whether sequence variants that increase/decrease CAMK2N1 expression were associated with cardiometabolic traits. We identified 263 cis-eQTLs regulating CAMK2N1 (Table S11). The most significant single nucleotide polymorphism regulating CAMK2N1 were found in Adipose–Visceral (VAT), in which there were 51 in total for this tissue. To test whether cis-eQTLs for CAMK2N1 are enriched in VAT, we compared 1000 sets of 263 cis-eQTLs not associated with CAMK2N1. There were 21.7% of CAMK2N1 cis-eQTLs in VAT, compared with 20.7% to 21.0% non-CAMK2N1 cis-eQTLs in VAT in the 1000 sets of non-CAMK2N1 cis-eQTLs (Figure S6).

Of the CAMK2N1 cis-eQTLs in VAT, 44 were significantly associated with cardiometabolic traits in the knowledge portals (Table S12). T2DM was the most frequent disease trait, linked to 39 variants (effect size range: −0.43 to 0.40, P adj=0.048–0.0052; Figure 5A), followed coronary artery disease linked to 27 variants (effect size range: −0.43 to 0.35, P adj=0.047–0.016; Figure 5B). All variants that are associated with CAMK2N1 downregulation (negative effect size), are also associated with significant disease risk reduction, whereas variants that are associated with CAMK2N1 upregulation (positive effect size), are associated with increased disease risk (Figure 5A and 5B; Table S11). To test whether CAMK2N1 cis-eQTLs are enriched for cardiometabolic traits, we determined the association to traits of V AT cis-eQTLs in a randomly selected set of 263 cis-eQTLs not associated with CAMK2N1 (Table S13). There was a significantly greater number of CAMK2N1 cis-eQTLs associated with T2DM, coronary artery disease, and VAT volume than cis-eQTLs not associated with CAMK2N1 (Table S14).

To establish the effect size of CAMK2N1 expression on visceral fat mass (not reported in the knowledge portals, we quantified CAMK2N1 expression in human visceral fat...
samples and found that compared with lean subjects, obese nondiabetics, and obese diabetics had a significantly greater body mass index, weight, and fat mass compared with lean subjects; although, BP across groups was not significantly different (Table S15). CAMK2N1 expression in visceral fat was increased significantly by 1.82- to 2.76-fold in obese diabetic and nondiabetic subjects, respectively, compared to lean nondiabetics (effect size, obese =1.23, \( P=0.02 \), obese diabetic =1.56, \( P<0.005 \)), but between obese groups, expression was similar (\( P>0.05 \); (Figure 5A). Regression analysis showed that CAMK2N1 expression in visceral fat correlated significantly with body mass index (Figure 5D) and absolute fat mass (\( r^2=0.34, \ P=0.0069 \)).

Discussion

The major results in this study demonstrate that Camk2n1 knockout in SHR reduced CaMKII activity in the kidney and LV, but not in adipose tissue and that these changes lead to profound alterations in the cardiometabolic phenotype of the Camk2n1\(^{-/-}\) rat. Compared with SHR, Camk2n1\(^{-/-}\) rats had lower BP and increased vascular reactivity and lower LV mass and rate pressure product. Camk2n1 knockout increased insulin sensitivity, whereas visceral fat mass in vivo and adipogenic capacity in vitro were decreased. These data support the previous eQTL and linkage studies in rat RI strains that showed strong associations between Camk2n1 and these cardiometabolic traits.\(^{3,4}\)
We show that reduced BP and increased vasoreactivity are associated with enhanced ACE2-Ang-(1–7)-Mas signaling in Camk2n1−/− rats. In LV, CaMKII-associated prohypertrophic and upregulation antihypertrophic components of cell cycle were downregulated, specifying mechanisms for reduced LV mass, whereas in EAT, diminished pro-obesogenic cell cycle pathways and classical complement associated with insulin resistance provide mechanistic insights into reduced adiposity and ameliorated insulin sensitivity of Camk2n1−/− rats. In human visceral fat, we found that CAMK2N1 expression correlated with fat mass and body mass index, in keeping with previous reports of 34 cis-eQTLs (GTex Portal) that associated significantly with increased CAMK2N1 expression and elevated risk of T2DM.

Experimental inhibition of CaMKII has been investigated to develop new treatments for hypertension and cardiac hypertrophy.10–12,15 In addition, indirectly, these and other studies have investigated the function of Camk2n1 while also testing CaMKII function. However, because of a number of off-target effects, including regulation of calcium signaling, masking docking sites on CaMKII, and inhibiting CaM-associated and other kinases, associated with KN-92, KN-93, autacamide-derived inhibitory peptide (AC3-I), and CaMKIN-tides, used in these inhibitor studies, there is an incomplete understanding of Camk2n1 function and its regulation of CaMKII.12 Furthermore, these inhibitory peptides are based on the inhibitory domain of CAMKIIN/Camk2n2, which has a different expression pattern and is likely conformationally and functionally distinct from endogenous Camk2n1. In our study, we show that endogenous Camk2n1 is required for full activity of CaMKII in kidney and LV but is dispensable for CaMKII activity in adipose tissue. We propose that the cardiorenal physiological and molecular effects of Camk2n1 deletion are, in part, likely determined by reduced CaMKII activity, but may also be due, at least in adipose tissue, to CaMKII-independent functions of Camk2n1.

Hypertension, like other features of MetS, has a polygenic basis and is controlled by multiple genetic variants in both rats and humans. For example, BP QTLs have been found on all rat chromosomes,16 just as GWAS hits for hypertension reside across all human chromosomes.17 Moreover, there is good agreement between rat BP QTLs and genes, and their syntenic regions and orthologs in humans, with the likelihood that the networks regulating BP are conserved across species.16 Hypertension can be mitigated by NO-mediated vasodilation through the ACE2-Ang(1–7)-Mas axis; in humans, ACE2 and NOS3 variants modulate BP, whereas renal eNOS deficiency in SHR, or ACE2 deletion in mice, contribute to hypertension and hypertension-related renal
damage. CaMKII transfection in vitro into endothelial cells reduced Ca\(^{2+}\)/CaM binding to eNOS and decreased NO production following bradykinin stimulation, whereas in vivo transgenic overexpression of CaMKII or AC3-I did not alter baseline BP nor NO-dependent vasodilation, only partially protecting from Ang II–induced hypertension, without affecting CaMKII overactivity. The relationship between CaMKII and ACE2 has not been elucidated. However, in our study, Camk2n1 deletion reduced renal CaMKII activity, but increased renal ACE2 and eNOS, and their respective products, Ang-(1–7) and NO, which may, in part, be responsible for the lower BP in Camk2n1\(^{-/-}\). ACE2, eNOS, and CaMKII are regulated by Ca\(^{2+}\)/CaM binding; therefore, Camk2n1 may regulate the binding association of Ca\(^{2+}\)/CaM with these enzymes, thereby affecting their activation and vasodilatory capacity.
CaMKII overactivity in humans with hypertension or T2DM is considered a cause of pathological LVH and heart failure. We found that Camk2n1 deletion reduced cardiac CaMKII activity and LV mass in SHR and conferred partial protection from increased myocardial load. Thus, our data support previous studies showing reduced CaMKII activity ameliorates LV remodeling, but that this occurs through Camk2n1 deficiency. Our WGCNA defined altered hypertrophic pathways associate with Camk2n1 knockout. For example, the cell cycle pathway that included inhibitors Cdkn3 and E2f8 was upregulated in Camk2n1−/− LV. Furthermore, Fabp4 and FABP4 were downregulated in Camk2n1−/− LV and have been shown to promote cardiac hypertrophy in mice and regulate AMPK signaling, actin cytoskeleton, and oxidative phosphorylation, all of which were associated with Camk2n1 deletion in our study.

Together, these changed pathways indicate mechanisms by which Camk2n1 knockout has reduced LV mass and is protected from stress-related rate pressure product increases and remodeling through CaMKII modulation.

Insulin resistance, a key factor in MetS, was ameliorated in Camk2n1−/− rats and occurs independently of CaMKII in adipose tissue. Improved insulin sensitivity is associated in humans with increased circulating NO and Ang-(1−7), reduced leptin and visceral adiposity, all of which are features of Camk2n1−/− rats. In addition, improved insulin sensitivity in SHR by Camk2n1 deletion, mirrors the connection in humans, we found between cis variants that decrease CAMK2N1 expression and are associated with increased insulin sensitivity.

In humans, we showed that CAMK2N1 was reduced in visceral fat from lean compared with obese subjects, consistent with a previous study showing CAMK2N1 upregulation in obese compared with lean Pima Indians. This is analogous to the reduced visceral fat and CaMKII-dependent adipogenesis we have observed in Camk2n1−/− rats. This is distinct from nonspecific CaMKII inhibitor studies suggesting CaMKII regulation of adipogenesis in vitro. WGCNA of the adipose transcriptome defined CaMKII-dependent alterations in obesity- and MetS-related cell cycle, classical complement, and apoptosis pathways in Camk2n1−/− rats. For example, downregulation in Camk2n1−/− EAT of Pla2g5 and Pla2g2a and reduced soluble phospholipase A2 activity supports the amelioration in MetS phenotypes in Camk2n1−/− rats, as Pla2g5 is upregulated in obese adipose tissue and promotes leptin secretion, whereas Pla2g2a is causally related to obesity and MetS. Moreover, soluble phospholipase A2 has been shown to promote adipogenesis and apoptosis associated with obesity. Apoptosis is an unlikely cause of reduced adiposity in Camk2n1−/− rats as proapoptotic genes (Baiap2l1/Birc5, Dsg2, and Pcdh7) were downregulated. Alternatively, reduced proliferation is suggested by upregulation of Cdkn1a/p21 in Camk2n1−/− EAT, which has been shown elsewhere to prevent obesity and adipocyte hyperplasia, and downregulation of Rbl1/p107 and Cdk1 that have been shown by others to be pro-obesogenic and upregulated in obesity. Furthermore, upregulation of Id3 and downregulation of C1s and C4 provide additional mechanisms for reduced adiponectin production, reduced adiposity, and increased insulin sensitivity.
2. Aitman TJ, Critser JK, Cuppen E, et al. Progress and prospects in University of Edinburgh. Additionally, support from the BHF Centre for Research Excellence, Wellcome Trust New Investigator grant 100981/Z/13/Z awarded to FS/15/60/31510. S.M. de Procé is funded by a Medical Research Menzies is supported by a British Heart Foundation Fellowship.

Camk2n1 knockout in SHR, ameliorates multiple cardiomyopathological phenotypes including hypertrophy, LV mass, insulin sensitivity, and visceral adiposity, associated with reduced cardiorenal CaMKII activity and independent of adipose CaMKII activity. Together with our demonstration that visceral fat CAMK2N1 expression increases in obese subjects and correlates with adiposity and our analysis of cis-acting variants that regulate human CAMK2N1 and MetS traits, we conclude that Camk2n1 regulates multiple cardiovascular and metabolic processes, both dependently and independently of CaMKII, suggesting that endogenous Camk2n1/CAMK2N1 may not function exclusively as an inhibitor of CaMKII and requires a reappraisal of existing studies that have used nonspecific CaMKII inhibitors proposed to mimic Camk2n1 function. Furthermore, our data suggest that therapeutic targeting of CAMK2N1 may allow amelioration of MetS features in humans.

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Disclosures

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References

Novelty and Significance

What Is New?

- Camk2n1 negatively regulates blood pressure, left ventricle mass, and insulin sensitivity and is required for full adipogenic capacity in the spontaneously hypertensive rat.

What Is Relevant?

- Camk2n1 deletion has effects on multiple organ systems: lowering blood pressure and reducing renal CaMK (Ca2+/calmodulin-dependent kinase II) activity, which is associated with increased ACE2 (angiotensin II-converting enzyme)-Ang-(1–7)-Mas activity, enhancing nitric oxide-related vasoreactivity; in the left ventricle, Camk2n1 deletion reduced cardiac CaMKII activity, downregulating hypertrophy-related pathways, and preventing stress-induced increased myocardial load; and in visceral fat, Camk2n1, improved insulin sensitivity and reduced adipogenesis, by downregulating cell cycle pathways, independent of CaMKII.

Summary

Camk2n1 regulates multiple organ systems associated with metabolic syndrome, and its ablation improved MetS features in spontaneously hypertensive rat. In humans, visceral fat Camk2n1 expression correlated with adiposity, and SNPs that regulate Camk2n1 expression are associated with coronary artery disease and type 2 diabetes mellitus.