A chimeric porcine circovirus (PCV) with the immunogenic capsid gene of the pathogenic PCV type 2 (PCV2) cloned into the genomic backbone of the nonpathogenic PCV1 induces protective immunity against PCV2 infection in pigs

Citation for published version:

Digital Object Identifier (DOI):
10.1128/JVI.78.12.6297-6303.2004

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Journal of Virology

Publisher Rights Statement:
Copyright © 2004, American Society for Microbiology

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
A Chimeric Porcine Circovirus (PCV) with the Immunogenic Capsid Gene of the Pathogenic PCV Type 2 (PCV2) Cloned into the Genomic Backbone of the Nonpathogenic PCV1 Induces Protective Immunity against PCV2 Infection in Pigs

M. Fenaux, T. Opriessnig, P. G. Halbur, F. Elvinger, and X. J. Meng

Center for Molecular Medicine and Infectious Diseases and Department of Large Animal Clinical Sciences, College of Veterinary Medicine, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061-0342, and Department of Veterinary Diagnostic and Production Animal Medicine, College of Veterinary Medicine, Iowa State University, Ames, Iowa 50011

Received 11 December 2003/Accepted 5 February 2004

Porcine circovirus type 2 (PCV2) is associated with postweaning multisystemic wasting syndrome in pigs, whereas PCV1 is nonpathogenic. We previously demonstrated that a chimeric PCV1-2 virus (with the immunogenic capsid gene of PCV2 cloned into the backbone of PCV1) induces an antibody response to the PCV2 capsid protein and is attenuated in pigs. Here, we report that the attenuated chimeric PCV1-2 induces protective immunity to wild-type PCV2 challenge in pigs. A total of 48 specific-pathogen-free piglets were randomly and equally assigned to four groups of 12 pigs each. Pigs in group 1 were vaccinated by intramuscular injection with 200 μg of the chimeric PCV1-2 infectious DNA clone. Pigs in group 2 were vaccinated by intralymphoid injection with 200 μg of a chimeric PCV1-2 infectious DNA clone. Pigs in group 3 were vaccinated by intramuscular injection with 10^4.5 TCID<sub>50</sub> of a wild-type pathogenic PCV2 virus. By 21 days postchallenge (DPC), 9 out of the 12 group 4 pigs were viremic for PCV2. Vaccinated animals in groups 1 to 3 had no detectable PCV2 viremia after challenge. At 21 DPC the lymph nodes in the nonvaccinated pigs were larger (P < 0.05) than those of vaccinated pigs. The PCV2 genomic copy loads in lymph nodes were reduced (P < 0.0001) in vaccinated pigs. Moderate amounts of PCV2 antigen were detected in most lymphoid tissues of nonvaccinated pigs but in only 1 of 36 vaccinated pigs. Mild-to-severe lymphoid depletion and histiocytic replacement were detected in lymphoid tissues in the majority of nonvaccinated group 4 pigs but in only a few vaccinated group 1 to 3 pigs. The data from this study indicated that when given intramuscularly in pigs, the attenuated chimeric PCV1-2 live virus, as well as the chimeric PCV1-2 infectious DNA clone, induces protective immunity against PCV2 infection and could potentially serve as an effective vaccine.

Postweaning multisystemic wasting syndrome (PMWS) was first observed in piglets of a high-health herd in Canada in 1991 (14). Since then PMWS has been reported in many pig-producing regions of North America, Europe, and Asia (2, 5, 6, 21, 28). The primary causative agent of PMWS is thought to be type 2 porcine circovirus (PCV2) (2, 7, 9, 10, 15, 20, 29).

The type 1 porcine circovirus (PCV1) was initially discovered as a persistent contaminant of a porcine kidney cell culture (PK-15) (36). PCV1 is a small nonenveloped icosahedral virus, with a single-stranded circular DNA genome of about 1.76 kb. PCV1 has not been found to cause disease and is generally considered to be nonpathogenic (1, 35). Pathogenic PCV2 and nonpathogenic PCV1 share only about 76% nucleotide sequence identity but have similar genomic organizations (10). Two open reading frames (ORFs) have been characterized: ORF1 encodes rep proteins required for viral replication (4) and ORF2 encodes the immunogenic capsid protein (25).

Both PCV1 and PCV2 are members of the Circoviridae family, along with psittacine beak and feather disease virus (3) and the tentative members columbid circovirus, goose circovirus, and canary circovirus (23, 31, 37). The human circoviruses TT virus, TT virus-like minivirus, and the SEN virus have genomic organization similar to that of PCV (27, 34, 38).

Accumulated evidence indicates that PCV2 is the primary, but not the sole, causative agent of PMWS (2, 7, 9, 10, 15, 20, 29, 30). Clinical PMWS has been reproduced in conventional pigs coinfected with PCV2 and either porcine parvovirus or porcine reproductive and respiratory syndrome virus (PRRSV) (15, 29). Ladekjaer-Mikkelsen et al. (20) recently reproduced PMWS in 3-week-old specific-pathogen-free (SPF) piglets inoculated with PCV2 alone. PMWS was also reproduced in PCV2-inoculated piglets immunostimulated with keyhole limpet hemocyanin in incomplete Freund’s adjuvant (18). Opriessnig et al. (30) showed that pigs vaccinated with Mycoplasma hyopneumoniae and Actinobacillus pleuropneumoniae prior to PCV2 inoculation had increased length of PCV2 viremia and more-severe lymphoid lesions compared to unvaccinated pigs. It is generally believed that immunostimulation...
either by vaccination or secondary viral infection plays a role in the occurrence of PMWS (12, 17, 18, 19). However, the exact role of immunostimulation in the progression to clinical PMWS is not known.

There is a need for a vaccine to prevent PCV2 infections and its role in the progression to clinical PMWS. We previously showed that a chimeric PCV1-2 infectious DNA clone (with the immunogenic capsid gene of PCV2 cloned into the backbone of the nonpathogenic PCV1) is infectious when injected directly into the lymph nodes of SPF pigs and induces a strong antibody response to PCV2 capsid antigen while remaining attenuated in pigs (11). Therefore, chimeric PCV2-2 appears to be a good candidate vaccine. In this study, we evaluated the efficacy of this candidate vaccine by subjecting SPF pigs to intramuscular and intralymphoid immunization with chimeric PCV1-2 virus and an infectious chimeric PCV1-2 DNA clone followed by challenge with wild-type pathogenic PCV2. We showed that the PCV1-2 chimeric virus and the infectious DNA clone both induce protective immunity against wild-type PCV2 challenge in SPF pigs.

**MATERIALS AND METHODS**

**PCV2 and chimeric PCV1-2 infectious DNA clones.** The construction of PCV2 and the chimeric PCV1-2 infectious DNA clone was reported previously (9, 11). The original wild-type PCV2 was from a pig with naturally occurring PMWS on an Iowa farm (isolate 40895) (10). The PCV2 infectious DNA clone was constructed by cloning two tandem copies of the complete PCV2 genomes into pBluescript vector. The PCV2 infectious DNA clone and the PCV2 virus generated by transfection of PK-15 cells with the PCV2 infectious DNA clone have been shown to induce the hallmark pathological lesions of PMWS (9). The chimeric PCV1-2 infectious DNA clone was constructed by replacing the ORF2 capsid gene of nonpathogenic PCV1 with that of PCV2 in the genomic backbone of PCV1 (11). The PCV2 and PCV1-2 infectious clone plasmids used in this study were prepared essentially as previously described (9, 11). The concentration of the plasmid DNA used in vaccination of pigs was determined by spectrophotometry.

**Generation and infectivity titration of PCV1-2 and PCV2 virus stocks.** PCV2 and chimeric PCV1-2 live viruses were generated by transfection of PK-15 cells with the respective infectious DNA clone as previously described (9, 11). To determine the infectivity titers of the PCV2 and chimeric PCV1-2 virus stocks, PK-15 cells were cultivated on 8-well LabTek chamber slides (Nalge Nunc International). The TK-15 cells reached 70 to 80% confluency, the cells were infected with a 10-fold serial dilution of either PCV2 or PCV1-2 virus stock. After 3 days of incubation, the infected cells were stained in an immunofluorescence assay to determine the infectivity titers as previously described (9, 11). Briefly, the infected cells were fixed to the LabTek chamber slides through the use of an 80% acetone and 20% methanol fixing solution. Both PCV2- and PCV1-2-infected cells were then incubated with PCV2 polyclonal rabbit antibody. After washing three times with phosphate-buffered saline buffer, the cells were incubated with a secondary fluorescein isothiocyanate-labeled goat anti-rabbit IgG antibody (KPL Inc., Gaithersburg, Md.). Slides were mounted using Fluoromount-G and coverslipped. Viral infectivity titers were calculated using the Karber method.

**Vaccination of SPF pigs with a chimeric PCV1-2 infectious DNA clone as well as PCV1-2 chimeric live virus.** A total of 49-week-old SPF piglets were randomly assigned to four groups of 12 pigs each, and each group was housed separately. Pigs in group 1 were vaccinated with 200 μg of chimeric PCV1-2 infectious DNA clone by intramuscular injection. Pigs in group 2 were vaccinated with 200 μg of chimeric PCV1-2 infectious DNA clone by intrasplenic injection. Pigs in group 3 were vaccinated with 10^5 50% tissue culture infective doses of the chimeric PCV1-2 live virus by intramuscular injection. Pigs in group 4 were not vaccinated and served as controls. All animals were monitored daily for clinical signs, and serum samples were collected at 1, 7, 14, 21, 28, 35, and 42 days postvaccination (DPV) and weekly on the appropriate days postchallenge (DPC) until necropsies were performed at 21 DPC.

**Challenge of vaccinated pigs with wild-type pathogenic PCV2.** At 42 DPV, all pigs were challenged with 2 × 10^4 50% tissue culture infective doses of the wild-type pathogenic PCV2. To maximize the challenge conditions, each animal received one-third of the PCV2 challenge inoculum intramuscularly and two-thirds intranasally. All animals were subjected to necropsies at 21 DPC (63 DPV).

**Clinical evaluation.** Pigs were weighed at DPV 0 and at the time of necropsy. Body temperatures and clinical pathological scores (0, normal; 6, severe) were recorded every other day from −1 to 63 DPV (21 DPC). Clinical observations, including evidence of central nervous system disease, icterus, muscular skeletal disease, and changes in body condition, were recorded daily.

**Gross pathology and histopathology.** Necropsy tissues were prepared essentially as previously described (9, 11). The concentration of the plasmid DNA used in vaccination of pigs was determined by spectrophotometry.

**Evaluating the efficacy of this candidate vaccine by subjecting SPF pigs to intramuscular and intralymphoid immunization with chimeric PCV1-2 virus and an infectious chimeric PCV1-2 DNA clone followed by challenge with wild-type pathogenic PCV2. We showed that the PCV1-2 chimeric virus and the infectious DNA clone both induce protective immunity against wild-type PCV2 challenge in SPF pigs.**
amplify a fragment of 900 bp (11). The PCR products were sequenced to confirm the identity of the virus recovered from pigs.

Statistical analyses. All statistical analyses described below were performed using an SAS system (version 8.02; SAS Institute Inc., Cary, N.C.). Serum sample S/P ratios determined by ELISA were compared between vaccinated and nonvaccinated groups by analysis of variance using the MIXED procedure. The model included the effects of vaccine, DPV, and their interaction. Chimeric PCV1-2 vaccination effects for each DPV were evaluated by the slice option. S/P ratios were dichotomized to the presence or absence of seroconversion at S/P 0.20 and analyzed by logistic regression using the LOGISTIC procedure. Mean viral genomic copy loads in lymph nodes were compared by the Kruskal-Wallis test using the NPAR1WAY procedure and/or by analysis of variance of ranked data followed by a Bonferroni test of multiple mean ranks, using the GLM procedure. Gross pathological and histopathologic lesion scores were compared by the Kruskal-Wallis test with the NPAR1WAY procedure and/or by analysis of variance using the GLM procedure followed by a Bonferroni test of multiple means. The proportions of pigs with gross and histopathologic lesions in various tissues were compared between groups by Fisher’s exact test, using the FREQ procedure.

RESULTS

Chimeric PCV1-2 live virus and the chimeric PCV1-2 infectious DNA clone both replicate in pigs when injected intramuscularly or intralymphoid and induce specific antibody response against PCV2 capsid antigen. Prior to inoculation at −1 DPV, serum samples from all animals tested negative by real-time PCR for PCV1 or PCV2 nucleic acids.

Group 1 pigs vaccinated intramuscularly with the chimeric PCV1-2 DNA clone did not develop PCV1-2 viremia for the duration of the study, as no PCV1-2 DNA was detected in sera (Table 1). Three pigs in group 1 had detectable PCV2 maternal antibody titers at −1 DPV; the titer levels waned by DPV 14. Seroconversion to PCV2-specific antibodies occurred in 7 out of 12 pigs by DPV 42, the day of challenge with wild-type pathogenic PCV2 (Table 2).

Fenaux et al. have previously shown that pigs can be infected by intralymphoid injection of PCV2 or PCV1-2 infectious DNA clone (9, 11). Therefore, group 2 pigs were made positive controls by vaccination through intralymphoid injection of the chimeric PCV1-2 infectious DNA clone. Like the pigs in group 1, none of the vaccinated pigs in group 2 developed PCV1-2 viremia (Table 1). Of the 12 pigs, 3 had detectable PCV2 maternal antibody titers at DPV −1; the titer levels waned in all by DPV 14. Seroconversion to PCV2 capsid antibodies was first detected at DPV 21, and 7 of the 12 pigs had seroconverted by DPV 42 (Table 2).

Animals in group 3 were vaccinated with the chimeric PCV1-2 live virus by intramuscular injection. PCV1-2 viremia was not detected in any of the immunized pigs for the duration of the study. Four pigs in group 3 had detectable maternal PCV2 antibodies at −1 DPV; the titer levels waned in all by 7 DPV. Seroconversion to PCV2 capsid antibodies was first detected at 28 DPV in 4 of the 12 animals, and by 42 DPV all pigs in group 3 had seroconverted (Table 2).

PCV1-2 viremia was not detected in the nonvaccinated pigs

### TABLE 1. Detection of chimeric PCV1-2 and PCV2 viremia by real-time PCR in sera of vaccinated and nonvaccinated pigs

<table>
<thead>
<tr>
<th>Group</th>
<th>Vaccine</th>
<th>Route of vaccination</th>
<th>No. of pigs with viremia/total on DPVa</th>
<th>Total (PCV1-2)</th>
<th>No. of pigs with viremia/total on DPVb</th>
<th>Total (PCV2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PCV1-2 DNAa</td>
<td>Intramuscular</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
</tr>
<tr>
<td>2</td>
<td>PCV1-2 DNAa</td>
<td>Intralymphoid</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
</tr>
<tr>
<td>3</td>
<td>PCV1-2 virusd</td>
<td>Intramuscular</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
</tr>
<tr>
<td>4</td>
<td>None</td>
<td></td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
<td>0/12 0/12 0/12 0/12 0/12 0/12 0/12 0/12</td>
</tr>
</tbody>
</table>

a Results represent detection by real-time PCR of chimeric PCV1-2 and wild-type PCV2 nucleic acid.

b At 42 DPV, the animals in all four groups were challenged with the wild-type PCV2 virus.

c Cloned PCV1-2 genomic DNA in pSK plasmid.

d A PCV1-2 live vaccine virus stock generated by transfection of PK-15 cells with PCV1-2 infectious DNA clone.

### TABLE 2. Seroconversion to PCV2-specific antibodies in pigs vaccinated with PCV1-2 live virus or with PCV1-2 infectious DNA clone before and after PCV2 challenge

<table>
<thead>
<tr>
<th>Group</th>
<th>Vaccine</th>
<th>Route of vaccination</th>
<th>Maternal antibodya</th>
<th>No. of pigs testing positive/total no. of pigs on DPVb</th>
<th>No. of pigs testing positive/total no. of pigs on DPCc,d</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PCV1-2 DNAd</td>
<td>Intramuscular</td>
<td>No</td>
<td>0/9 0/9 1/9 1/9 1/9 1/9 1/9 6/9</td>
<td>8/9 9/9 9/9</td>
</tr>
<tr>
<td>2</td>
<td>PCV1-2 DNAd</td>
<td>Intralymphoid</td>
<td>Yes</td>
<td>3/3 1/3 0/3 0/3 0/2 1/3 1/3 1/3</td>
<td>2/3 2/3 2/3</td>
</tr>
<tr>
<td>3</td>
<td>PCV1-2 virusd</td>
<td>Intramuscular</td>
<td>Yes</td>
<td>3/3 1/3 0/3 1/3 1/3 2/3 2/3 2/3</td>
<td>3/3 3/3 3/3</td>
</tr>
<tr>
<td>4</td>
<td>None</td>
<td></td>
<td>Yes</td>
<td>3/3 1/3 1/3 1/3 1/3 1/3 1/3 1/3</td>
<td>1/3 1/3 3/3</td>
</tr>
</tbody>
</table>

a Some animals in each group had maternal antibodies at −1 DPV.

b PCV2 antibody was measured with an ELISA with the recombinant PCV2 capsid antigen: results represent the number of pigs testing positive/number tested.

c At 42 DPV, the animals in all four groups were challenged with the wild-type PCV2 virus.

d Cloned chimeric PCV1-2 genomic DNA in pSK plasmid.

e PCV1-2 candidate vaccine live virus stock generated by transfection of PK-15 cells with the chimeric PCV1-2 infectious DNA clone.
Three pigs in group 4 had detectable PCV2 maternal antibodies, and none of the animals seroconverted to PCV2. PCV2 DNA was extracted during necropsy at DPC 21 from vaccinated pigs in groups 1, 2, and 3. Wild-type PCV2 viral genomic copy loads in TBLN tissues collected from nonvaccinated pigs in group 4. PCV2 genomic DNA was extracted and subjected to real-time PCR amplification. Pigs positive for PCV2 genomic DNA in each group are indicated with an open circle. The numbers 9, 10, 7, and 3 within circles along the x axis indicate the numbers of animals in each group that were negative for PCV2 genomic DNA in the TBLN. The PCV2 genomic copy loads were determined by real-time PCR and are represented as a log value of PCV2 genomic copy loads per 10 μg of TBLN tissue (y axis).

in group 4, and none of the animals seroconverted to PCV2 antibodies prior to PCV2 challenge (Table 1 and Table 2). Three pigs in group 4 had detectable PCV2 maternal antibodies at DPV -1. By DPV 7, the maternal antibody had waned in all but one animal.

PCV2 antibody S/P ratios differed between treatment groups (P < 0.0001) and over time (P < 0.0001) (data not shown). Following vaccination, up to DPV 42 (the day of PCV2 challenge) pigs in groups 1, 2, and 3 were 1.9 (95% confidence interval [CI] [0.7; 5.1]), 3.51 (95% CI [1.3; 9.1]), and 5.4 (95% CI [2.0; 14.1]) times more likely to have an S/P ratio greater than 0.20 than nonvaccinated pigs in group 4 (overall vaccine effect; P = 0.004).

The chimeric PCV1-2 candidate vaccine prevents PCV2 viremia and reduces virus load in lymph nodes after challenge with wild-type pathogenic PCV2. After the vaccinated pigs were challenged with wild-type pathogenic PCV2, PCV2 viremia was not detected by real-time PCR in any of the vaccinated pigs in groups 1, 2, and 3 (Table 1). In group 4 nonvaccinated pigs, PCV2 viremia was first detected at DPC 7 in 1 of 12 pigs and 9 of the 12 pigs had detectable PCV2 viremia by 14 DPC (Table 1). Seroconversion to PCV2 antibodies was first detected at DPC 14 in 6 of the 12 group 4 pigs, and by DPC 21 all pigs in group 4 had seroconverted to PCV2 (Table 2). The PCV2 viral genome loads in serum samples of group 4 pigs peaked at DPC 14; the loads ranged from 2,800 to 240,800 PCV2 genomic copies per ml of serum.

At necropsy (DPC 21), PCV2 genomic DNA was detected in the tracheobronchial lymph nodes (TBLN) in 3 of 12 group 1 pigs, 2 of 12 group 2 pigs, 5 of 12 group 3 pigs, and 9 of 12 group 4 pigs (P = 0.057) (Fig. 1). The range of PCV2 viral genomic copy loads per 10 μg of homogenized TBLN in positive-testing samples was 1,023 to 61,119 in group 1, 10,532 to 82,152 in group 2, 1,652 to 5,419,774 in group 3, and 363 to 621,285,534 in group 4 (Fig. 1). The median genomic copy loads differed between groups 1, 2, 3, and 4 (P = 0.012). Median PCV2 copy loads in TBLN did not differ between groups 1, 2, and 3 or between groups 3 and 4 (P > 0.05); however, loads did differ between groups 1 and 4 and between groups 2 and 4 (P < 0.05). PCR amplification using PCV2- and PCV1-2-specific primers followed by DNA sequencing confirmed that the genomic sequence detected by real-time PCR in the TBLN of animals in groups 1, 2, 3, and 4 originated from the pathogenic PCV2 and not from the chimeric PCV1-2 vaccine virus.

The chimeric PCV1-2 candidate vaccine reduces macroscopic and microscopic lesions as well as viral antigen load in tissues of vaccinated pigs after challenge with wild-type pathogenic PCV2. (i) Clinical evaluation. Clinical signs characteristic of PMWS were not observed in any animals of groups 1, 2, 3, and 4 for the duration of the study.

(ii) Gross lesions. All pigs were subjected to necropsy at DPC 21. The enlargement of lymph nodes of the vaccinated pigs in groups 1, 2, and 3 generally ranged from mild to moderate, with 1 or 2 pigs with severely enlarged (3 times normal size) lymph nodes in each group (Table 3). The lymph nodes of all nonvaccinated group 4 pigs were moderately to severely enlarged (Table 3). The mean gross lesion scores of the lymph nodes differed between groups 1, 2, 3, and 4 (P = 0.0007). The values for vaccinated groups 1, 2, and 3 did not differ (P > 0.05) among those groups but were less than the mean score of the nonvaccinated group 4 pigs (P < 0.05; Table 3).

(iii) Microscopic lesions. Microscopic lung lesions characterized as mild peribronchiolar lymphoplasmacytic and histiocytic bronchointerstitial pneumonia and liver lesions characterized as mild lymphoplasmacytic hepatitis were detected in pigs in all groups (Table 4). Mild LD of lymph node follicles was detected in 4 of 12 group 1 pigs, 1 of 12 group 2 pigs, and 1 of 12 group 3 pigs, and mild to moderate LD was detected in 11 of 12 pigs in group 4 (P < 0.001; Table 4). Mild HR of lymph node follicles was observed in 3 of 12 pigs in group 1 and 1 of 12 pigs in both groups 2 and 3. Mild to moderate HR was detected in 10 of 12 pigs in group 4 (P = 0.001; Table 4). Mild LD and HR of the tonsil follicles were found in 1 of 12 pigs in

<table>
<thead>
<tr>
<th>Group</th>
<th>Vaccinea</th>
<th>Route of vaccination</th>
<th>No. of pigs with gross lymph node lesions/total no. of pigsb</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PCV1-2 DNA</td>
<td>Intramuscular</td>
<td>12/12 (2.0)1</td>
</tr>
<tr>
<td>2</td>
<td>PCV1-2 DNA</td>
<td>Intralymphoid</td>
<td>12/12 (1.8)1</td>
</tr>
<tr>
<td>3</td>
<td>PCV1-2 virus</td>
<td>Intramuscular</td>
<td>12/12 (1.6)1</td>
</tr>
<tr>
<td>4</td>
<td>None</td>
<td></td>
<td>12/12 (2.7)1</td>
</tr>
</tbody>
</table>

a Animals in group 1 to 3 were vaccinated with chimeric PCV1-2 infectious DNA clone or chimeric PCV1-2 live virus. At 42 DPV, all animals in groups 1 to 4 were challenged with wild-type pathogenic PCV2.
b All 12 pigs in each group were subjected to necropsy at 21 DPC. Results represent the number of animals with enlarged lymph nodes/number of animals in each group (numbers in parentheses represent mean scores of estimated enlargement). Different superscripts Roman numerals (I and II) indicate different mean value scores between groups (P < 0.05).
group 1, 0 of 24 pigs in groups 2 and 3, and 7 of 12 pigs in group 4 \( (P = 0.0003; \text{Table 4}) \). Mild LD and HR of the spleen follicles were observed in 2 of 12 pigs in group 1 and in none of the pigs in groups 2 and 3, but 10 of 12 group 4 pigs exhibited mild to moderate LD and 9 of 12 group 4 pigs exhibited mild to moderate HR \( (P < 0.0001; \text{Table 4}) \). Results with respect to the presence of lesions in other tissues and organs are summarized in Table 4.

(iv) Detection of PCV2 antigen. At necropsy (DPC 21), PCV2 antigen was not detected in the lymph node tissues of pigs in vaccinated groups 1, 2, and 3 except for one pig in group 3. In the nonvaccinated group 4 pigs, low-to-high amounts of PCV2 antigen were detected in the lymph nodes \( (P < 0.0001; \text{Table 5}) \). No PCV2 antigen was detected in the tonsil of group 1, 2, and 3 pigs. However, low-to-high amounts of PCV2 antigen were detected in the tonsil of 8 of 12 nonvaccinated group 4 pigs \( (P < 0.0001; \text{Table 5}) \). PCV2 antigen was not detected in spleen tissues of vaccinated group 1, 2, or 3 pigs. Low-to-moderate amounts of PCV2 antigen were detected in the spleen tissue of 5 of 12 pigs in nonvaccinated group 4 \( (P = 0.0002; \text{Table 5}) \). No PCV2 antigen was detected in thymus tissues of any pigs (Table 5).

DISCUSSION

PMWS has become a serious global pig disease; hence, there is an urgent need for the development of a vaccine against PCV2-associated diseases, including PMWS. Fenoux et al. previously reported that a chimeric PCV1-2 virus is attenuated when inoculated into SPF pigs but is capable of inducing a humoral immune response against PCV2 capsid protein, suggesting that the chimeric PCV1-2 may serve as a candidate vaccine against PCV2 infection (11). In this study, we demonstrated that pigs vaccinated with the chimeric PCV1-2 candidate vaccine developed protective immunity against wild-type pathogenic PCV2 challenge. We also demonstrated that pigs can be effectively vaccinated using an intramuscular route by injections with both the infectious chimeric PCV1-2 DNA clone and chimeric PCV1-2 live virus.

The majority of the vaccinated pigs in all 3 groups seroconverted to PCV2 antibodies within 4 to 6 weeks postvaccination. The remaining seronegative pigs at the time of challenge had elevated PCV2 antibody titers with rising S/P ratios. Statistical analysis showed that there is a significant overall vaccine effect on S/P ratios \( (P = 0.004) \). No chimeric PCV1-2 viremia was detected throughout the study in the vaccinated pigs, which is in agreement with our earlier study (11). Most importantly, no PCV2 viremia was detected in vaccinated pigs after challenge with wild-type pathogenic PCV2. In contrast, PCV2 viremia was detected in 9 of 12 nonvaccinated pigs after challenge. After PCV2 challenge, PCV2 antigen was detected in low-to-high amounts in lymph node, tonsil, and spleen tissues of nonvaccinated pigs but not in the vaccinated pigs with the exception of one pig in group 3. Vaccinated pigs also had reduced PCV2 genomic copy viral loads in the lymph nodes. These data indicate that the chimeric PCV1-2 candidate vaccine can prevent PCV2 viremia and significantly reduce the amount of PCV2 virus in the lymphoid tissues, which are important factors in pathogenesis of PCV2-associated diseases.

The mean scores of microscopic lesions in lymph node,
spleen, and tonsil tissues of the three vaccinated groups showed that the lesions were less severe (\( P < 0.05 \)) than those of the nonvaccinated group, indicating protection against PCV2 challenge by the candidate vaccine. The vaccinated pigs had significantly less enlargement of lymph nodes than the nonvaccinated pigs. The enlargement of the lymph nodes observed in vaccinated pigs may be attributed to normal vaccine activation of the immune system in response to the PCV2 challenge, since the enlarged lymph nodes in vaccinated pigs had no detectable microscopic lesions or viral DNA. In nonvaccinated pigs, LD and HR of follicles associated with PCV2 antigen were observed in lymph node, spleen, and tonsil consistent with the hallmark PCV2-associated pathological lesions observed in natural and experimental cases of PMWS. In contrast, only one of the vaccinated pigs had detectable PCV2 antigen in lymphoid tissues. These results strongly indicate that chimeric PCV1-2 candidate vaccine is effective in protecting pigs from PCV2-associated lymphoid lesions and thus in preventing the detrimental effects on the immune system.

The occurrence of LD during initial PCV2 infection may be linked to the eventual occurrence of leukopenia prior to the onset of clinical PMWS (26, 32). Therefore, the chimeric PCV1-2 candidate vaccine may have the ability to stop the eventual progression to clinical PMWS by preventing the initial LD of lymphoid tissues.

No significant differences were found among the three different routes of vaccination with the PCV1-2 candidate vaccine. Intramuscular vaccination with PCV1-2 DNA clone, intralymphoid vaccination with PCV1-2 DNA clone, and intramuscular vaccination with PCV1-2 live virus were all effective in inducing protective immunity against PCV2 infection. However, the intramuscular vaccination route is the only route likely to be acceptable to swine producers. The intralymphoid route of vaccination with chimeric PCV1-2 infectious DNA clone was included as a positive control, since Fenaux et al. had previously shown that this route has the ability to induce an infection (9, 11).

Low levels of maternal antibody found in a few animals in groups 1, 2, and 3 had no apparent effect on the induction of protective immunity by the chimeric PCV1-2 candidate vaccine. Since there were only a few animals with low levels of maternal antibodies in this study, a definitive answer cannot be given to the question of whether or not the low level of maternal antibodies has any effect on vaccination with a live vaccine. Since many newborns in commercial swine farms have PCV2 maternal antibodies following colostrum uptake, future studies with larger numbers of animals with different levels of maternal antibody are warranted to confirm our preliminary results.

Although not all the vaccinated animals seroconverted to PCV2 by the time of challenge, they were all protected against the pathogenic PCV2 challenge, suggesting that high S/P ratios of PCV2 antibody responses are not absolutely required for protection. It is possible that the chimeric PCV1-2 candidate vaccine induces a cell-mediated immune response (8), which may be equally or more important for induction of protection against PCV2. Further research is warranted to determine the exact role and the extent of cell-mediated immunity induced by the candidate vaccines in protection against PCV2 infections.

**ACKNOWLEDGMENTS**

We thank Mike Gill and Jay Srinivas of Fort Dodge Animal Health, Inc., and S. M. Boyle, L. A. Eng, R. B. Duncan, J. C. Sible, and T. E. Toth for their support and Jillian Fenaux for editorial assistance. This study was supported in part by a grant from Fort Dodge Animal Health, Inc., Fort Dodge, Iowa, and by a grant from the U.S. Department of Agriculture National Research Initiative Competitive Grant Program (NRI 2004-35204-14213).

**REFERENCES**


---

**TABLE 5. Immunohistochemical detection of PCV2 antigen in lymph nodes, tonsils, spleen, and thymus of vaccinated and nonvaccinated pigs at 21 DPC with wild-type PCV2**

<table>
<thead>
<tr>
<th>Group</th>
<th>Vaccine</th>
<th>Route of vaccination</th>
<th>Lymph node</th>
<th>Tonsils</th>
<th>Spleen</th>
<th>Thymus</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PCV1-2 DNA</td>
<td>Intramuscular</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)</td>
</tr>
<tr>
<td>2</td>
<td>PCV1-2 DNA</td>
<td>Intralymphoid</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)</td>
</tr>
<tr>
<td>3</td>
<td>PCV1-2 virus</td>
<td>Intramuscular</td>
<td>1/12 (0.08)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)(^1)</td>
<td>0/12 (0.0)</td>
</tr>
<tr>
<td>4</td>
<td>None</td>
<td>Intramuscular</td>
<td>9/12 (0.75)(^1)</td>
<td>8/12 (0.67)(^1)</td>
<td>5/12 (0.42)(^1)</td>
<td>0/12 (0.0)</td>
</tr>
</tbody>
</table>

\(^{a}\) The candidate vaccine was either chimeric PCV1-2 infectious DNA clone or chimeric PCV1-2 live virus.

\(^{b}\) All pigs were subjected to necropsy at 21 DPC. Values in parentheses are the mean estimated amounts of PCV2 antigen in lymphoid tissue (range: 0, no antigen detected; 3, high amounts of antigen). Different superscripts Roman numerals (I and II) indicate mean value score differences between groups (\( P < 0.05 \)).
characterization of type 2 porcine circovirus (PCV-2) from pigs with post-
weaning multisystemic wasting syndrome in different geographic regions of
North America and development of a differential PCR-restriction fragment
length polymorphism assay to detect and differentiate between infections

genicity and pathogenicity of the chimeric infectious DNA clones between
pathogenic type 2 porcine circovirus (PCV2) and non-pathogenic PCV1 in

Stevenson, J. C. Foster, J. A. Ellis, S. Krakowka, B. M. Adair, and G. M.
Allan. 2003. In vitro studies on the infection and replication of porcine
circovirus type 2 in cells of porcine immune system. Vet. Immunol. Immu-
nopathol. 94:149–161.

pathogenicity of two U. S. porcine reproductive and respiratory syndrome

weaning multisystemic wasting syndrome (PMWS). Swine Health Prod.
8:201–203.

Morozov, and P. S. Paul. 2001. Experimental reproduction of severe disease
in CD/CD pigs concurrently infected with type 2 porcine circovirus and
porcine reproductive and respiratory syndrome virus. Vet. Pathol. 38:528–
539.


17. Krakowka, S., J. A. Ellis, F. McNeilly, D. Gilpin, B. Meehan, K. McCul-
lough, and G. Allan. 2002. Immunologic features of porcine circovirus type
2 infection. Viral Immunol. 15:567–582.

2001. Activation of the immune system is the pivotal event in the production
Pathol. 38:31–42.

19. Kyriakidis, S. C., K. Saoulidis, S. Lekkas, Ch. C. Miliotis, P. A. Papoutsis,
and S. Kennedy. 2002. The effects of immune-modulation on the clinical and
pathological expression of postweaning multisystemic wasting syndrome.
J. Comp. Pathol. 126:38–46.

kowka, J. Ellis, F. McNeilly, G. Allan, and A. Botner. 2002. Reproduction of
postweaning multisystemic wasting syndrome (PMWS) in immunostimulated
and non-immunostimulated 3-week-old piglets experimentally infected with

21. Larochelle, R., M. Morin, M. Antaya, and R. Magar. 1999. Identification and
incidence of porcine circovirus in routine field cases in Quebec as deter-

petitive PCR analysis of porcine circovirus DNA in serum from pigs with
3477.

sequencing of columbid circoviruses (CoCV), a new circovirus from pigeons.

Lekchaoeunskul, and P. S. Paul. 2002. Modified indirect porcine circovirus
(PCM) type 2-based and recombinant capsid protein (ORF2)-based ELISA

Paul. 2000. Open reading frame 2 of porcine circovirus type 2 encodes a

porcine circovirus type 2 induced postweaning multisystemic wasting syn-

27. Nishizawa, T., H. Okamoto, K. Konishi, H. Yoshizawa, Y. Miyakawa, and M.
Mayumi. 1997. A novel DNA virus (TTV) associated with elevated transam-
inate levels in posttransfusion hepatitis of unknown etiology. Biochim. Bio-

28. Onuki, A., K. Abe, K. Togashi, K. Kawashima, A. Taneichi, and H. Tsu-
emitsu. 1999. Detection of porcine circovirus from lesions of a pig with

29. Opreissig, T., M. Fenaux, S. Yu, R. B. Evans, D. Cavanaugh, J. M. Gallup,
2004. Effect of porcine parvovirus vaccination on the development of PMWS
in segregated early weaned pigs coinfected with type 2 porcine circovirus and

30. Phenic, K. V., J. H. Weston, I. Ypelaar, A. Lavazza, J. A. Smyth, D. Todd,
circovirus of canaries and its relationship to other members of the genus

31. Segales, J., C. Alfonso, C. Rosell, J. Pastor, F. Chianini, E. Campos, L.
Lopez-Fuertes, J. Quinina, G. Rodriguez-Arrozio, M. Calsamiglia, J. Pujols,
J. Dominguez, and M. Domingo. 2001. Changes in peripheral blood leuko-
cyte populations in pigs with natural postweaning multisystemic wasting

1999. Development of a polyclonal-antibody-based immunohistochemical
method for the detection of type 2 porcine circovirus in formalin-fixed,

a new human DNA virus (TTV-like mini virus, TLMV) intermittently re-

276.


determinations and analyses of novel circoviruses from goose and pigeon.

Strathdee, D. Vlahov, and D. L. Thomas. 2001. Dynamics of SEN virus