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Citation for published version:

Digital Object Identifier (DOI):
10.1093/nar/gkt840

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Nucleic Acids Research

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Human telomeres that carry an integrated copy of human herpesvirus 6 are often short and unstable, facilitating release of the viral genome from the chromosome


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Received July 23, 2013; Revised and Accepted August 28, 2013

ABSTRACT

Linear chromosomes are stabilized by telomeres, but the presence of short dysfunctional telomeres triggers cellular senescence in human somatic tissues, thus contributing to ageing. Approximately 1% of the population inherits a chromosomally integrated copy of human herpesvirus 6 (CI-HHV-6), but the consequences of integration for the virus and for the telomere with the insertion are unknown. Here we show that the telomere on the distal end of the integrated virus is frequently the shortest measured in somatic cells but not the germline. The telomere carrying the CI-HHV-6 is also prone to truncations that result in the formation of a short telomere at a novel location within the viral genome. We detected extra-chromosomal circular HHV-6 molecules, some surprisingly comprising the entire viral genome with a single fully reconstituted direct repeat region (DR) with both terminal cleavage and packaging elements (PAC1 and PAC2). Truncated CI-HHV-6 and extra-chromosomal circular molecules are likely reciprocal products that arise through excision of a telomere-loop (t-loop) formed within the CI-HHV-6 genome. In summary, we show that the CI-HHV-6 genome disrupts stability of the associated telomere and this facilitates the release of viral sequences as circular molecules, some of which have the potential to become fully functioning viruses.

INTRODUCTION

The ends of linear chromosomes are distinguished from double-strand breaks within the genome by specialized nucleoprotein structures known as telomeres. Human telomeres comprise a variable-length double-stranded DNA molecule that is predominantly 5'- (TTAGGG)n-3' (1) but includes, at the start of the repeat array, sequence-variant or degenerate repeats interspersed with TTAGG G repeats (2–4). Each telomere is terminated by an essential single-strand overhang of 50–300 nt (5). The telomeric DNA binds to the six-protein complex, known as Shelterin, via the double-strand binding TRF1 and TRF2 proteins and the single-strand binding protein POT1 (6). Telomeric DNA bound to the Shelterin complex forms looped structures (t-loops) in which the single-strand overhang invades the upstream duplex telomeric DNA forming a D-loop (7). The Shelterin complex on the capped telomere prevents inappropriate repair by non-homologous end-joining or by homologous recombination (HR) (8) and, in conjunction with other protein complexes, it regulates telomerase activity by controlling access to the single-strand overhang. However, telomerase is inactive in differentiated human cells and shows limited activity in stem cells; consequently in most cells, telomeric
DNA is eroded as a result of incomplete lagging-strand synthesis and end-processing that restores single-strand overhangs (6). Disturbance of the telomere capping function or telomere length regulation can result in genome instability leading to tumourigenesis, and the presence of a few short telomeres (9,10) induces senescence, which has known roles in ageing.

HHV-6A and B belong to the Ro深情ovirus genus of the β-herpesvirus subfamily, and though closely related, they present diverse biological, epidemiological, pathological and molecular properties. HHV-6B is prevalent in most populations and primary infection usually occurs in early childhood (<2 years) causing an intense fever and rash, whereas infections by HHV-6A tend to be less common (11–13). As for most herpesviruses, HHV-6A and B can remain latent lifelong, with the potential to reactivate that can have severe consequences. For example, HHV-6B reactivation in immunocompromised transplant patients can cause encephalitis and has been linked to bone marrow suppression and to graft failure (14). The HHV-6B genome is double-stranded DNA molecules comprising ∼145 kb of unique sequence encoding U1 to U100 ORFs, flanked by identical left and right direct repeats (DRL and DRR), each ∼8 kb (15–17). Each DR is terminated by the packaging and cleavage sequences, PAC1 and 2 (18), and includes two arrays of telomere-like repeats (T1 and T2) and a variety of ORFs (DR1–8). T2 is known to be a short array of pure (TTAGGG)n that varies in length between viral isolates, whereas T1 comprises an array of degenerate telomere-like repeats (19,20).

HHV-6A and B are distinct among herpesviruses, as they can integrate into the human telomeric DNA, probably via HR with the terminal T2 region of DR (21–25). Once integrated in the germline, the chromosomally integrated virus (CI-HHV-6) can be inherited. Approximately 0.8% of the UK population are CI-HHV-6 carriers (26), but it is not known whether integration is a natural biological form of HHV-6 latency or whether it affects normal telomere function and length regulation. We show that the telomere carrying the integrated virus is often the shortest measured and that the presence of the full-length viral genome disrupts stability of the telomere, resulting in frequent truncations. Moreover, we detected extra-chromosomal circular DNA comprising viral sequences and showed that the viral genome can be released from the telomere, possibly via a t-loop excision mechanism. Altogether, our data are consistent with the proposal that telomeric integration is a natural biological form of HHV-6 latency.

MATERIALS AND METHODS
Oligonucleotides

Oligonucleotides for amplification of HHV6 sequences were primarily designed based on the GenBank consensus sequences for HHV-6A (U1102 accession no. X83413.1) or HHV-6B (HST accession no. AB021506.1; Z29 accession no. AF157706.1).

DNA samples and lymphoblastoid cell lines

To identify CI-HHV-6 carriers, we screened the HapMap Phase I (27), the CEPH-HGDP (28) and parental DNA samples from the CEPH family panel (29). Therefore, we screened 1178 samples from unrelated donors in these three panels (duplicated samples between the panels were removed). We also screened 528 samples in the People of the British Isles panel (30) and 2153 samples from the Orkney Complex Disease Study (31). The panel of 92 sperm donors, a gift from Alec J. Jeffreys (32), was also screened for the presence of CI-HHV-6.

Lymphoblastoid cell lines (LCLs) from one CI-HHV-6A (designated 3-10q26.3) and five CI-HHV-6B carriers (designated 1-9q34.3, 2-9q34.3, 4-11p15.5, 5-17p13.3 and 6-17p13.3) reported previously (24) were donated by Duncan Clark. Other LCLs were from Bruce Winney; NWA090 (CI-HHV-6A), BAN519, COR264, CUM082, DER512, NWA008, YOR546 (all CI-HHV-6B); from David Gurwitz CEPH-HGDP00628 (CI-HHV6A); from the cell bank at the Coriell Institute for Medical Research: GM18999 (CI-HHV-6A), GM07022 (CI-HHV-6B), CEPH 1375.02 [also known as GM10863 (CI-HHV-6B)] and from other members of the CEPH 1375 family. All the LCLs were grown in RPMI1640 medium with 10–15% foetal calf serum at 37°C in a humidified incubator with 5% CO2 and subcultured using standard methods. To measure the telomere shortening rate in the GM18999 (CI-HHV-6A) and CEPH1375.02 (CI-HHV-6B) cell lines, the cells were cultured over a longer period. The number of population doublings was calculated from cell counts obtained using a haemocytometer, and the percentage of dead cells was determined using Trypan blue staining at each subculturing. Cell pellets were obtained throughout the time course.

Identification of HHV-6-positive DNA samples

Panels of DNA samples in 96-well format were screened by polymerase chain reaction (PCR) for the presence of U11 or U18 sequences. Standard 10 μl PCRs were prepared containing 5 ng genomic DNA, 0.3 μM of each primer, 0.05 U/μl of Taq polymerase (Kapa Biosystems) and a buffer supplied with the enzyme. Thermal cycling conditions were 96°C for 1 min then 35 cycles of 96°C for 15 s; 62–64°C for 30 s; and 68°C for 1 min). Forward and reverse primers: U11F 5'-TTTTTACATCAGGCAACGATC; U11R 5'-GGGACGCGGAATCGGAGGAAC; U18F 5'-CATATCTGATCAACCTTGCGATG; and U18R 5'-AGGCG; DR7F(B): 5'-AACAGCATCGATAATGCAGGCCC. Identification of the virus type was achieved by amplification with primers DR5F(A): 5'-CGTGACCTTTCTGTTTTATGC; DR5R(A): 5'-CACATACATTGAAACGGACACAC for HHV-6A and DR7F(B): 5'-AGGCG; DR7R(B): 5'-CCGAAATACGTCATGGTCGCTTCT for HHV-6B. Amplicons of the expected sizes were detected by agarose gel electrophoresis.

Single telomere length analysis

Single telomere length analysis (STELA) was conducted as described (33) using the specialized PCR buffer and other
modifications reported previously (34,35). The primer concentrations in the 10 μl STELA reactions were flanking primer 0.3 μM, Telorette2 0.225 μM and Teltail 0.05 μM. The Taq polymerase (Kapa Biosystems) was used at 0.04 U/μl and Pwo (Genaxxon Bioscience) at 0.025 U/μl. The primers located adjacent to the telomere repeat and used to amplify the double-stranded portion of the 12q, 17p and XpYp telomeres were 12q-STELA: 5′-CTTGCTCTCGGCAGGAGTCC-3′ and 17p6: 5′-GACGAGGATCGCGGTGTT-3′ and 17p6: 5′-TTGCCCTCAAGGTCTAGTG-3′. Primers used for STELA from the integrated HHV-6A or B were DR1R: 5′-GAAGAAGGATCGCGGTGTT-3′, DR392R: 5′-GAACGTGGCCGCTACAGTTC-3′, DR8F: 5′-CAATAGATCGG-3′, DR421R: 5′-GGATTACGGAGGTGAAT-3′ and DR8R(HHV-6B): 5′-CGCCCGCGGACTGC-3′. The isolated DNA fragment was sequenced. The primer that did not yield STELA amplicons were: DR1F: 5′-ACCCTGGGCCAGCAGAAATGC-3′, DR3F: 5′-TCCGTTCCCTATCGGCATCT-3′ and DR8F: 5′-CATAGATCGG GACGCTCGTGA-3′. The DNA in each STELA reaction ranged between 100 and 1000 pg. Agarose gels (0.8%) were used for telomere length analysis with size markers (GeneRuler 1 kb and GeneRuler High Range DNA ladder, Fermentas) run in every gel. Amplicon length analysis was conducted on the phosphor-images (minimum two gels per sample) using the Imagequant software (Typhoon 9400, GE Healthcare). The length of the flanking DNA was subtracted from each amplicon, and the median telomere length and interquartile range of the telomere molecules were determined. For each donor, data from four telomeres was selected for independent reanalysis. Within each donor, the median length of the four measured telomeres was ranked to identify the shortest. To determine which telomere was most frequently the shortest across the donors, the shortest telomere in each donor was assigned a value of 1 and the other three telomeres given a value of 0. In donors where the difference between the two shortest telomeres was <100 bp, a value of 0.5 was assigned to each. The shortest telomere frequencies were compared using the non-parametric Kruskal–Wallis analysis (GraphPad Prism), and if \( P < 0.05 \), the Dunn’s multiple comparison test was used to compare specific telomeres.

**Sequence analysis**

Single STELA products were reamplified, gel purified and sequenced by the Sanger dyeoxy termination method using DR1R, DR392R 5′-CCAGATCGCGGATKAGTG; DR421R 5′-GAAGGTTGAAAGAGGGGT AG or Teltail. Other PCR products were either sequenced directly or, if necessary, reamplified, gel purified and then sequenced.

**Isolation and characterization of chromosomal-HHV-6 junctions**

The 10q-CI-HHV-6A junction fragment was isolated by inverse PCR using the primers IPCR-1 and IPCR-2 (25). The isolated DNA fragment was sequenced. The primer 10qF: 5′-ATCCTTCTCTTTGCGAGCCG designed from the sequence was used with the DR8F(A): 5′-GCAAGAGAAAATGATGCGGAAAG primer to verify amplification of the 10q-CI-HHV-6A junction from genomic DNA in this CI-HHV-6A carrier. The other four chromosome-CI-HHV-6-junction fragments were amplified with a primer designed from the 17p subtelomeric sequences (36). The primer SubT17-539: 5′-CCCATTTTACTGGT AATGGACT anneals to subtelomeric DNA sequences on several chromosomes, and it was used with the DR8F primer to isolate junction fragments from one CI-HHV6A and three CI-HHV-6B carriers. The isolated amplicons were sequenced.

**Sperm DNA analysis**

The panel of sperm donors (32) was screened for the presence of HHV-6 as described. There was sufficient DNA to conduct telomere length analysis by STELA on one CI-HHV-6A and three CI-HHV-6B carriers. Telomere length measurements in the sperm DNA samples were made from phosphor-images of STELA blots and scatter plots produced (as above). Frequency histograms were generated by sizing individual molecules and allocating them to 1 kb bins. Distributions were then fitted using a model with either one or two Gaussian distributions (GraphPad Prism). For single Gaussian distributions, outliers were then defined as those telomere lengths of greater or less than three standard deviations from the mean (which includes >99.7% of values from that population). Where the data showed two Gaussian distributions, the distribution of the major population was used to determine the number of outliers. The frequency of short outliers was then determined as a percentage of the total number of telomere molecules analysed and plotted against the mean telomere length. The data were tested for linear correlation using Pearson’s correlation coefficient (GraphPad Prism).

**Analysis of the HHV-6 DR regions**

The length of the DR\(_{1,-T2}\) was determined following PCR using primers UDL6R: 5′-TTTGCTCAGCTGGCAGCT-3′ and DR8F. The presence of PAC2 in these amplicons was determined by PCR with UDL6R or U100F2-DR1R amplicons (DRR) with SmaI as a control for digestion. The absence of the PAC1 sequence in STELA products (encompassing part of DR\(_{1,-T1}\) and the telomere) was determined by sequencing reamplified gel-purified products using primers DR1R, DR392R, DR421R or Teltail. The length of the DR\(_{R,-T1}\) region was determined by PCR amplification between U100F2: 5′-TATCTCCGGAACTGATGCTG-3′ and DR1R, and the presence of DR\(_{R,-PAC1}\) was determined by secondary PCR with U100F2 and PAC1RB 5′-CGGGGAGTTTTA AAGTAATTTTTT-3′. The presence or absence of PAC1 was confirmed by digestion of STELA products (DR\(_{1}\)) or U100F2-DR1R amplicons (DR\(_{R}\)) with SalI (cuts at PAC1) or Sall as a control for digestion.
Sequencing the CI-HHV-6B genome using ion semiconductor technology

Overlapping amplicons across the unique region of HHV6 and the DRs (without T1 and T2) were generated from CEPH1375.02 CI-HHV-6B carrier, using primers based on the HST sequence (AB021506.1) (37). The amplicons were pooled and sonicated using a Bioruptor (Diagenode Inc.). The subsequent preparation of the sonicated DNA for sequence analysis using an IonTorrent personal genome sequencer (Life Technologies, Irvine, CA, USA). The size-selected DNA was then prepared for Ion Torrent sequencing on a 314 chip (100 bp read length) according to Life Technologies’ protocol.

Detection of HHV-6 RNA transcripts

Total RNA was extracted using Tri-reagent (Sigma-Aldrich). The RNA was dissolved in water (100 μl) and treated with RNase-free DNase I (100 U/μl, 20 min, 37°C). The RNA was purified by phenol–chloroform extraction, precipitated with isopropanol and resuspended in RNase-free water (100 μl). cDNA was synthesized from 2 μg of total RNA using M-MuLV reverse transcriptase (New England Biolabs) and random primer hexamers (0.2 μM) in a total reaction volume of 20 μl at 42°C for 1 h, followed by enzyme inactivation at 90°C for 10 min. PCR products were then loaded in 10 μl, containing 40 ng of synthesized cDNA and 0.05 U/μl of Taq DNA polymerase. Oligonucleotides used were U38F: 5’-TTGT AATTCTGCGCCGTATTAC and U38R: 5’-TGCTGGTC CTTGCGATAGA; U73F: 5’-TTACGAGGCGGCGAG ATAAAC and U73R: 5’-TGATGAGGGCGATTTGAG TA; U90B2F: 5’-CCCTTACTATGCGACGGCTCTC and U90B2R: 5’-ACGACATCGTCTTTCCAAAGTG for HHV6B or U90A2R: 5’-CCTTCGATGACCTGTATTAT TCCA for HHV6A; U94-2F: 5’-AGACAGCCATTCCGA TGGTTC and U94-1R: 5’-TTGGTACGCTCAAAGCGGA GAA. Thermal cycling conditions were denaturation at 96°C for 1 min followed by 35 cycles at 96°C for 15 s, annealing at 58–64°C (depending on amplicons) for 20 s, extension at 70°C for 1 min and a final extension step at 70°C for 10 min. PCR products were analysed by agarose gel (1%) electrophoresis.

Detection of CI-HHV-6B truncations at DR1-T2

Genomic DNA (0.5 μg) was digested with ExoI (New England Biolabs) to remove 3’ single-strand overhangs from telomeres. Following ExoI digestion, the integrity of the double-stranded genomic DNA was verified by PCR amplification of ~6 kb fragment using primers U100FB 5’-CAGCTATGCTATTTGCGCG and DR1R. STELA was conducted as above using flanking primers DR1R or the UDL6R on genomic DNA with or without ExoI digestion.

Detection of extra-chromosomal circular HHV-6 DNA

Genomic DNA (90 ng/reaction) was amplified with primers UDL6R and U100F at 0.2 μM with Taq (0.04U/μl) and Pwo (0.025U/μl) DNA polymerases in 10 μl reaction prepared in the same PCR buffer used for STELA. Control PCRs were also performed with primers UDL6R and DR3F or U100F2 and DR3R. The PCR products were resolved in 0.8% agarose, and Southern blots were prepared and hybridized to a DR3 probe generated using DR3F and DR3R primers. The amplicons generated from extra-chromosomal circular DNA in five separate PCRs from CEPH1375.11 CI-HHV-6B were reamplified and the composition of the DR region was verified by secondary PCRs and sequencing.

RESULTS

Identification of CI-HHV-6 carriers and verification of viral integration using STELA

To determine whether HHV-6 integration is a natural biological form of latency and what effect it has on the associated telomere, we investigated six LCLs from unrelated CI-HHV-6 carriers (24) with integration sites as follows: one CI-HHV-6A at 10q, two CI-HHV-6B at 9q, one CI-HHV-6B at 11p and two CI-HHV-6B at 17p (24). We used STELA (33) to confirm integration and to orientate the viral genome within the telomere (Figure 1A and B).

To identify more CI-HHV-6 carriers, we screened 3859 DNA samples from unrelated donors (see Materials and Methods) using primers that amplify conserved segments from the viral U11 and U18 genes. Integration was verified using STELA. As a result, 58 additional CI-HHV-6 positive samples were identified among the various populations (4 CI-HHV-6A and 54 CI-HHV-6B). Where available we obtained LCLs or additional high
Figure 1. Characterization of CI-HHV-6 carriers and proof of telomeric integration using STELA. (A) The HHV-6A or 6B genome inserts in the reverse orientation such that DR₉ and the U100 gene are closest to the subtelomeric sequences. The DR₉ is expanded to show the reverse order of DR₈, 3 and 1 ORFs. Plain arrows represent primers. The telorette 2 oligonucleotide (grey line) and Teltail primer used in STELA are shown. Orientation of DRs is shown by dashed arrows. Cen, centromere. (B) Amplification of the CI-HHV-6-associated telomere using STELA in a CI-HHV-6B carrier [1-9p34.3 (24)]. Amplicons are generated with the DR1, 3 and 8 reverse (R) primers but not the forward (F) primers, confirming the orientation of the integrated virus. The increasing size of the STELA products generated by DR1R, 3R and 8R reflects the distance from the 3 single-strand overhang. The blot was hybridized to a telomere probe. (C–E) Identification of the CEPH1375 family that segregates CI-HHV-6B through three generations. (C) Family members were screened for the presence the HHV-6 U18 gene. An amplicon from the XpYp telomere-adjacent sequence was used as a PCR control. (D) CEPH1375 family members are positive for CI-HHV-6B and showed the presence of the HHV-6B DR7 and absence of the HHV-6A DR5. Φ, size marker; NC, negative control. (E) CEPH1375 family showing transmission of CI-HHV-6B through three generations. (F) Detection of STELA amplicons using the DR1R (R), but not the DR1F (F), primer confirmed the orientation and integration of HHV-6B in family members. All the STELA amplicons contain viral sequences (left) and (TTAGGG)ₙ repeats (right). The maternal grandfather (CEPH1375.11) shows particularly short telomeres on the end of the CI-HHV-6B.
molecular weight genomic DNA from the newly identified CI-HHV-6 carriers for further analysis. We also identified a family (CEPH 1375) that has transmitted CI-HHV-6B over three generations (Figure 1C–F).

Characterization of the DRs in CI-HHV6 carriers

Sequence analysis of single STELA products from seven individuals showed that the terminal PAC1 sequence (at the distal end of DR1) was absent in CI-HHV-6 A (*n* = 2; 3-10q26.3 and 1501) and CI-HHV-6B (*n* = 5; 1-9q34.3, 2-9q34.3, 5-17p13.3, 6-17p13.3 and HGDP0092) carriers. We isolated the 10q chromosome-HHV-6 junction from one CI-HHV-6 A carrier [3-10q26.3 (24)] using inverse PCR (25). The internal chromosome-HHV-6 junctions from four other carriers were isolated by amplification between a subterminal sequence (found at several chromosome ends, Sub17p-539) and the DR8F primer in DRR between a subterminal sequence (found at several chromosome ends, Sub17p-539) and the DR8F primer in DRR between a subterminal sequence (found at several chromosome ends, Sub17p-539) and the DR8F primer in DRR.

The CI-HHV6 telomere was often the shortest measured in carriers

To investigate the length of the telomere on the end of the virus, we used STELA with the DR1R primer to amplify telomere molecules from small aliquots of LCL-DNA from CI-HHV-6 carriers (Figure 3A). The median length of the virus-associated telomere was compared with the length of XpYp, 12q and 17p telomeres. In 50% (8/16) of the LCLs, the CI-HHV-6-associated telomere was the shortest measured (Figure 3B, Supplementary Figure S3A) (9,38), even compared with the 17p telomere that has been reported as often being the shortest (36,39). Corresponding analysis of blood DNA samples from 24 carriers (21 CI-HHV-6B from the Orkney Complex Disease Study study, and 3 CI-HHV-6 A siblings in a British family) again showed that the virus-associated telomere was most often the shortest [42% (10/24); Figure 3B and Supplementary Figure S3B], indicating that short virus-associated telomeres do not arise from the establishment or propagation of LCLs. The combined data show that the shortest-telomere frequency differs significantly between the chromosome-ends in somatic cells (Kruskal–Wallis test *P* < 0.0117) and is highest at the CI-HHV-6-associated telomere (shortest in 45% of the carriers).

CI-HHV-6-associated telomeres in the germline

To investigate virus-associated telomere lengths in the germline, we screened sperm DNA samples from 92 men from UK of North European descent (32) and identified five (5.4%) CI-HHV-6 carriers (4 HHV-6B and 1 HHV-6 A), a higher frequency than seen in the British Isles panel but not statistically significant. Telomere length analysis in four men showed that all were considerably longer in sperm DNA, as expected (40). Interestingly, the 17p telomere, not the CI-HHV-6-associated telomere, was the shortest we measured in the germline of all four sperm donors. Moreover, the CI-HHV-6-associated telomere was the longest measure in two of the sperm donors (Table 1 and Supplementary Figure S4).

Examination of the telomere length distributions in the sperm samples showed that a proportion of outlier molecules was considerably shorter than the mean length, as reported previously (40). The shortened telomere molecules likely arise through intra-telomere trimming via the t-loop excision mechanism (7,41-43). We showed that the frequency of short outlier molecules correlated with mean telomere length in the germline. Consequently, the longest CI-HHV-6-associated telomeres have the highest frequency of outliers (Table 1 and Figure 3C), indicating a high frequency of t-loop excisions within the telomere.

The HHV-6-associated 12q, 17p and XpYp telomeres shorten at similar rates in LCLs

The difference in the ranked length of the CI-HHV-6-associated telomere between somatic cells and the germline suggests that the presence of the viral genome affects length regulation. Analysis of telomere shortening rates in two LCLs (GM18999 CI-HHV-6 A and CEPH1375.02 CI-HHV-6B) showed similar attrition rates (average 79 bp/cell division) at the Xp/Yp, 12q, 17p and CI-HHV-6-associated telomere in both cell lines (Supplementary Figure S5). This suggests that the disparity between the length of the CI-HHV-6-associated telomere in somatic cells and the germline is not explained by a higher rate of telomere erosion at the virus-associated telomere. However, we detected low-level telomerase expression in the CEPH1375.02 CI-HHV-6B at all time points and in some time points for the GM18999 CI-HHV-6 A cell line. Clearly, the level of telomerase is insufficient to maintain telomere length in these cell lines but
we cannot exclude the possibility that it may target one telomere more than another.

**Somatic truncation of the integrated HHV-6 genome and viral excision**

The detection of short telomere molecules in the germline (Supplementary Figure S4 and Figure 3C) (40) prompted us to look for extra-chromosomal circular molecules. We detected extra-chromosomal circles containing (TTAGGG)_n repeats in the CEPH1375.02 LCL, which arise from low-frequency intra-telomere t-loop excision at all telomeres (41–43). We also detected extra-chromosomal circular molecules containing HHV-6 DR sequences (Supplementary Figure S6). These may arise from...
Figure 3. Telomere length analysis in CI-HHV-6 carriers. (A) STELA was used to measure telomere length at 12q, 17p, XpYp and on the end of the virus (DR1R) in CI-HHV-6 carriers. It shows a representative STELA-Southern blot on blood DNA from one CI-HHV-6A carrier (1501) and below the scatter plots of the data obtained from several blots. The median and inter-quartile ranges are shown as red lines. (B) The median values for each of the four telomeres measured were ranked by length in DNAs from 16 LCLs and 24 blood DNAs from CI-HHV-6 carriers. The proportion of the samples in which each telomere is the shortest is shown in pie charts for the LCLs and blood DNA samples. The histogram of the combined data shows that the shortest telomere frequency (+SEM) is different between the chromosome ends (Kruskal–Wallis test $P<0.0117$). (C) Relationship between telomere length and the frequency of short telomere molecules in the male germline. The short (outlier) telomere molecules, shorter than 3 standard deviations from the mean (<3SD), were counted as a proportion of the total molecules analysed for each telomere in four sperm donors and plotted against mean telomere length (Table1).
excision of a quasi t-loop formed by invasion of the telomeric 3' single-strand overhang into the DR1-T2 (Figure 4A). The reciprocal product of excisions should be a truncated CI-HHV-6 with a novel telomere at DR1-T2. Therefore, we conducted STELA using a flanking primer at the U1 gene and detected very short telomere molecules (140–200 bp) in five CI-HHV6B cell lines (two shown in Figure 4B). Amplification of the short telomere molecules is dependent on the presence of a telomeric 3' single-strand overhang, as ExoI digestion before STELA abolished the products. Sequence analysis showed these amplicons contain DR1-PAC2.

We hypothesized that the entire viral genome could be released from the telomere through telomeric 3' single-strand overhang (eroded into the DRL-T1 region) invasion into the internal DRR-T1 region or between a viral genome already truncated at DRL-T2 into the internal DRR-T2 (Figure 4A). Resolution of the quasi t-loop, which would include the whole viral genome, could release the HHV-6 genome as a large circular t-loop, which would include the whole viral genome, and latency owing to alternative splicing and the use of different CI-HHV-6 LCLs consistently revealed transcripts from the U90 gene in all 10 LCLs, whereas detection of transcripts from other genes varied between the LCLs and between RNA samples. Seven LCLs showed two different length U90-reverse transcriptase PCR (RT-PCR) products, two LCLs showed only the shorter product and one LCL showed only the longer product. Sequence analysis showed that the shorter product had undergone a correct splicing to remove an intron (Figure 5). The HHV-6 U90 gene is an immediate early transactivator that produces different transcripts during productive infection and latency analysis of the base substitutions and indels indicates that none are predicted to cause disruptive frameshifts in the ORFs (Supplementary Figure S6).

### Spliced U90 transcripts detected in CI-HHV-6 carriers

Expression analysis of U38, U73, U90 and U94 in 10 different CI-HHV-6 LCLs consistently revealed transcripts from the U90 gene in all 10 LCLs, whereas detection of transcripts from other genes varied between the LCLs and between RNA samples. Seven LCLs showed two different length U90-reverse transcriptase PCR (RT-PCR) products, two LCLs showed only the shorter product and one LCL showed only the longer product. Sequence analysis showed that the shorter product had undergone a correct splicing to remove an intron (Figure 5). The HHV-6 U90 gene is an immediate early transactivator that produces different transcripts during productive infection and latency analysis of the base substitutions and indels indicates that none are predicted to cause disruptive frameshifts in the ORFs (Supplementary Figure S6).

### Discussion

Among the five internal CI-HHV-6 junction fragments we isolated (2 CI-HHV-6A and 3 CI-HHV-6B carriers), four were different, thus representing independent integration events, and all shared a similar sequence organization (25-47). The absence of DR2-PAC2 and the presence of telomere sequence-variant repeats (3) in each junction support the proposition that HHV-6 integrates by HR between DRr-T2 and the proximal end of a telomere. The absence of PAC1 at the terminal end of DR1 could also be a consequence of HR at the time of integration. However, the curious expansion of the T1 region,
identified through analysis of the internal DR<sub>R</sub>-T1, and the variable truncation of the terminal DR<sub>R</sub>-T1 in carriers suggest that the degenerate repeat region underwent erosion. Therefore, the expanded T1 region may serve a role during integration by acting as a buffer for replication-driven erosion that results in loss of PAC1 and part of the DR<sub>R</sub>-T1 region until (TTAGGG)<sub>n</sub> repeats are added by telomerase in the germline.

The CI-HHV-6-associated telomere was the shortest measured in somatic cells from 45% of the 40 carriers investigated but not the shortest in the four CI-HHV-6 sperm donors showing that the virus-associated telomere...
is readily lengthened by telomerase in the germline. The disparity between CI-HHV-6-associated telomere length in somatic cells and the germline suggests that the integrated virus perturbs an aspect of telomere function. We showed that the rate of telomere erosion through replication-driven processes was not higher at the CI-HHV-6-associated telomere in LCLs from two CI-HHV-6 carriers. However, as these cell lines showed a low level of telomerase activity, further analysis will be required to determine whether or not telomerase is targeted to the CI-HHV-6 telomere more frequently.

We surmised that the presence of the HHV-6 genome might interfere with telomere capping by perturbing of t-loop formation and excision giving rise to telomere rapid deletion events that were first described in yeast (48,49). We showed that LCLs from CI-HHV-6B carriers are mixed populations of cells carrying the full-length CI-HHV-6B and a subset with CI-HHV-6B chromosomes truncated at DRL-T2 by the presence of a novel very short telomere with a single-strand overhang (Figure 4). The truncations at DR1-T2 could arise through a double-strand break at DR1-T2, followed by end processing to generate a 3' single-strand overhang. However, as we also detected extra-chromosomal circular molecules containing HHV-6 DR sequences, we favour a model in which the truncated chromosomes arise through processing of a t-loop (41–43) formed by strand invasion of the telomeric 3' single-strand overhang within DR1-T2. Excision of the quasi t-loop would result in a sudden truncation and formation of a telomere at DR1-T2. The presence of the novel short telomere may contribute to cell cycle arrest and the onset of senescence.

Sequence analysis of the CI-HHV-6B genome in the CEPH1375.02 showed that the viral genome is intact in this family; moreover, we detected molecules that contain a reconstituted DR region with both PAC1 and PAC2 in the grandfather CEPH1375.11. The reconstituted DR region could only arise from a recombination event between the terminal DRL (that contains a full-length T2 and PAC2) and the internal DRR (that contains a full-length T1 and PAC1) resulting in the release of a circular molecule containing the viral genome with a single DR (Figure 4). Therefore, consistent with our model invoking t-loop formation including the viral genome, we propose that occasional single-strand invasion of the telomeric 3' overhang into the internal HHV-6 DR-T1 facilitates release of the entire viral genome from the chromosome with a single reconstituted DR (Figure 4A). These large extra-chromosomal circular molecules arise from an intact viral genome (as shown by the next-generation sequence analysis), and so they have the potential to undergo rolling circle replication that could regenerate full-length HHV-6B genomes with two identical terminal DRs. This and the evidence that spliced
transcripts from the U90 gene, which is involved in latency, were detected in the cell lines investigated support the hypothesis that CI-HHV-6 is an alternative form of viral latency.

The reciprocal product of a large excision event would be retention of a DR, lacking both PAC sequences, within the telomere. Interestingly, we have identified two unrelated individuals (2/3859; population frequency 0.05%) that lack the unique portion of the HHV-6B genome but carry a single HHV-6B-DR (without PAC1 and 2) integrated into a telomere. These individuals may have inherited chromosomes that had already undergone a viral excision event, although partial insertion during integration cannot be excluded. The released viral genome with a single reconstituted DR in CEPH1375.11 could also arise by strand invasion of a short telomere on a CI-HHV-6 molecule already truncated at DR1-T2. Excision would release a circular HHV-6B genome with a reconstituted DR and remove the viral genome from the telomere (Figure 4A [3]).

In summary, we have shown that the CI-HHV-6-associated telomere is often one of the shortest in somatic cells and prone to sudden deletions that can critically shorten telomere at a new location in the viral genome. The presence of a short telomere associated with the CI-HHV-6 will increase the chance that the cell will become senescent, thus affecting tissue homeostasis (9,10). Moreover, cells carrying critically short telomeres associated with CI-HHV-6 may be prone to telomere fusion events that can drive instability. We also show that integrated copies of HHV-6B can be excised from the chromosome and we propose that this is achieved through use of the t-loop excision mechanism. This may facilitate spreading to another telomere within the cell or represent the first step towards viral reactivation.

ACKNOWLEDGEMENTS

The authors thank Sir Walter Bodmer for generously sharing the Population of the British Isles DNA panel; Mark Jobling, Howard Cann and David Gurwitz for sharing the HapMap, CEPH-HGDP DNA panels and for providing some cell lines. They thank Jenny Foxon and Dipti Vashi for help with cell culture and Farzaneh Atrain-Afaiy for some technical assistance. They thank Dzanan Osmanovic and Pierpaolo Maisano-Delser for their assistance with IonTorrent sequence analysis.

FUNDING

The Medical Research Council [G9001657 to N.J.R.]; the Wellcome Trust Institutional Strategic Support Fund [WT097828MF] and by Conacyt, Mexico [to A.H.-B.]. Funding for open access: University of Leicester RCUK.

Conflict of interest statement. None declared.

REFERENCES


