Inhibition of the NFAT pathway alleviates amyloid neurotoxicity in a mouse model of Alzheimer's disease

Citation for published version:

Digital Object Identifier (DOI):

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Journal of Neuroscience

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Introduction

Amyloid β (Aβ) soluble oligomers are thought to be an important source of neurotoxicity in Alzheimer’s disease (AD) (Walsh et al., 2002; Lesné et al., 2006; Shankar et al., 2007; Koffie et al., 2009). While synaptic loss correlates best with cognitive decline (Terry et al., 1991; DeKosky et al., 1996), the molecular mechanisms underlying Aβ synaptotoxicity and memory impairment remain largely unexplained, and new therapeutic approaches acting to protect neurons toward Aβ-related injuries need to be investigated.

Calcium dyshomeostasis and the consequent activation of calcineurin (CaN, or protein phosphatase 2B) may trigger Aβ-related pathological effects (Liu et al., 2005; Shankar et al., 2007; Kuchibhotla et al., 2008; Wu et al., 2010). An increased CaN activity has been described in AD brains (Liu et al., 2005; Wu et al., 2010), and CaN activation was reported in astrocytes surrounding amyloid deposits, thus exacerbating neuroinflammation (Norris et al., 2005). In neurons, CaN activation leads to pathological morphological changes, which could be blocked by abolition of CaN enzymatic activity using FK506 (Wu et al., 2010; Rozkalne et al., 2011). FK506 also improves cognitive deficits in the Tg2576 AD mouse model (Taglialatela et al., 2009), thus making calcineurin a potential therapeutic target. However, because FK506 prevents CaN phosphatase activity in all the cell types and toward all its substrates, the mechanisms underlying FK506 neuroprotection are unclear.

Calcineurin is a unique neuronal Ca2+/calmodulin-dependent serine/threonine phosphatase that plays fundamental physiological roles during development and regulates processes such as neurotransmitter release, synaptic plasticity, and learning (Klee et al., 1979; Groth et al., 2003). Upon activation, CaN leads to post-translational modification of postsynaptic proteins such as cofilin (Zhou et al., 2004) and AKAP79, which is associated with long-term depression (Bhattacharyya et al., 2009; Jurado et al., 2010). In addition, CaN dephosphorylates the nuclear factor of activated T cells (NFAT), which induces its nuclear translocation and the expression of target genes implicated in neuronal survival, axonal outgrowth, and dendritic complexity (Benedito et al., 2005; Nguyen and Di Giovanni, 2008; Schwartz et al., 2009). CaN activation therefore triggers short-term effects directly at the synapse as well as long-term modifications of synaptic plasticity by modulating gene expression through NFAT.

To decipher more precisely the role of CaN activation in Alzheimer disease, we tested the hypothesis that neuronal morphological changes caused by Aβ may be improved by inhibiting the CaN-induced NFAT pathway. We used a genetically encoded
VIVIT peptide initially developed as a specific competitive inhibitor of the interaction between CaN and NFAT, without affecting CaN phosphatase activity (Aramburu et al., 1999). We established that (1) VIVIT efficiently inhibits the neurotoxic effects associated with a constitutively activated CaN (CACA-N); (2) expression of a constitutively active form of NFAT produces a phenocopy of the effects induced by CACA-N or by Aβ exposure; (3) Aβ-induced decrease of spine density and dendritic complexity is inhibited by VIVIT; (4) restricted expression of VIVIT in the nucleus retains the beneficial effects of VIVIT, whereas a membrane-bound version of this peptide has no effect; (5) changes associated with amyloid plaques are alleviated when VIVIT or NLS-VIVIT were introduced by AAV-mediated gene delivery in the cortex of an AD mouse model in vivo.

Together, our findings offer new therapeutic opportunities by targeting Aβ downstream events through inhibition of NFAT transcriptional pathways.

Materials and Methods

Culture of primary cortical neurons and collection of neuronal conditioned media. Primary neurons were cultured from cerebral cortices of embryonic day 16 mice (Charles River Laboratories). Embryos resulted from the mating between a Tg2576 male that heterozygously overexpresses a human mutated APP gene and a wild-type female, thus giving rise to both transgenic (Tg) and littermate (WT) cultures. The genotype of each embryo was determined by PCR. Neurons were prepared as previously described (Wu et al., 2010) and plated at a density of 6.7 × 10^5 viable cells/35 mm culture dishes coated with poly-L-lysine (100 μg/mL, Sigma-Aldrich). The medium of the cells was not renewed to allow Aβ peptides to accumulate and conditioned media were collected from 14 DIV cultures (Wu et al., 2010). The concentration of Aβ peptides was quantified by a mouse/human ELISA kit (Wako) and reached ~7000 pMol. Transfection experiments were done at DIV 5 using Lipofectamine 2000 (Invitrogen).

Spine density and Sholl analyses. At 5 DIV, primary neurons were transfected with either pEGFP-N1 or pDsRed-Express-N1 plasmids (Clontech Laboratories), altogether with pVIVIT-EGFP and pVIVIT-NLS-EGFP constructs generously provided by Dr. Norris (Sanders-Brown Center On Aging, University of Kentucky, Lexington, KY) and Dr. Ruthazer (McGill University, Montreal, QC, Canada), respectively. For a subset of experiments, cells were also transfected with a wild-type or a constitutively active form of calcineurin described previously (Wu et al., 2010). Live imaging of GFP- or DsRed-expressing primary neurons (18 DIV) was used to analyze the morphological parameters of the cells. Images of the whole cell and of dendritic segments were captured using a LSM 510 Zeiss microscope to determine the morphological parameters of the cells. Images of the whole cell and of dendritic segments were captured using a LSM 510 Zeiss microscope to determine the morphological parameters of the cells. GFP and DsRed were respectively excited at 488 and 543 nm and emitted light was collected between 500–550 nm and 565–615 nm. Dendritic spines were analyzed using the NeuroStudio software (CNIC tools) that automatically detects three different spine types (thin, stubby, and mushroom) according to their morphological measures (i.e., length of the “neck” and size of the “head” of each spine) (Rodriguez et al., 2008). The complexity of the neuronal dendritic tree was determined by Sholl analysis (Spires-Jones et al., 2011), reporting the number of branch points with respect to the distance from the cell body. All these parameters were evaluated on transfected cells that did not show any obvious morphological alterations such as dendritic dystrophies.

Immunocytochemistry. Immunostaining was performed using a standard protocol (Wu et al., 2010). Briefly, primary neurons were fixed with 4% paraformaldehyde in PBS, pH 7.4, for 15 min, permeabilized with 0.5% Triton X-100 in PBS for 20 min, and blocked with 3% bovine serum albumin at room temperature for 1 h. Primary antibodies to detect NFATc4 (polyclonal s.c.-13036, 1:200 dilution, Santa Cruz Biotechnology), laminin-B1 (polyclonal ab16048, 1:1000, Abcam), HA tag (Clone 16B12, 1:200 dilution, Covance), and GFP (polyclonal ab6556, 1:1000, Abcam or chicken anti-GFP, Aves) were incubated overnight at 4°C. Secondary antibodies conjugated to either cyanine 3 (Cy3, 1:1000; Jackson ImmunoResearch), or Alexa 488 (1:1000; Invitrogen, Eugene, OR) were then applied. Fluorescent images were obtained using a LSM 510 Zeiss confocal microscope.

Evaluation of NFAT subcellular localization. The changes in the subcellular distribution of NFATc4 were evaluated after immunostaining of the endogenous protein in primary neurons or in mouse brain slices. GFP-positive neurons that were previously transfected or transduced with GFP, VIVIT-GFP, or NLS-VIVIT-GFP were specifically analyzed. Using Imaged (National Institutes of Health open software), the fluorescence intensity of NFATc4 in the nucleus was determined by overlap with the nuclear staining DAPI (Vector Laboratories), whereas the fluorescence intensity in the cytoplasm was quantified in the rest of the cell body, excluding the nuclear compartment (Fig. 1C). The intensity of nuclear NFATc4 was then divided by the intensity of cytoplasmic NFATc4.

Evaluation of NFAT transcriptional activity using a Luciferase reporter system. Cortical primary neurons (7 DIV) were transduced with an AAV-NFAT–TA–Luc, altogether with AAV-wtCaN or AAV-CACA-N and AAV-GFP, AAV-VIVIT-GFP, or AAV-NLS-VIVIT-GFP. Three days after transduction, cells were harvested and the luciferase activity was measured with a luminometer using a reagent kit (Luciferase Assay System with Reporter Lysis Buffer; Promega). The background luciferase activity, calculated when pNFAT-Luc alone was added, was subtracted from all experiments.

Animals. In vivo experiments were performed using APPswes/PS1d9 double transgenic mice (APP/PS1, obtained from Jackson Laboratory) that overexpress a human mutant amyloid precursor protein gene containing the Swedish mutation K594M/M597L and a variant of the Presenilin 1 gene deleted for the exon 9, both under the control of PrP promoter (Jankowsky et al., 2004). Substantial amyloid deposition is visible by 6 months of age and we used 7-month-old animals. Wild-type littersmates were used as controls. Experiments were performed in accordance with NIH and institutional guidelines.

Viral vectors construction and production. Plasmids containing VIVIT-GFP and NLS-VIVIT-GFP were digested using NotI and NcoI restriction enzymes and subcloned into an AAV backbone containing the chicken β-actin promoter and a Woodchuck Hepatitis Virus Post-Transcriptional Regulatory Element (WPRE). The Myr-tagged VIVIT-GFP was obtained after amplification by PCR and insertion into an AAV backbone using XhoI and BsrGI restriction sites. The AAV-NFAT–TA–Luc plasmid was purchased from Clontech Laboratories and the constructions of AAV-CBA-β-actin-GFP and pAAV-CBA-β-actin-CACA-N backbones were described previously (Wu et al., 2010). As a control, we used an AAV-GFP that was available at the Harvard Gene Therapy Core.

High titer of AAV serotype 2 vectors (AAV-2-GFP, AAV-2-VIVIT-GFP, and AAV-2-NLS-VIVIT-GFP) and AAV-2-WtCaN, AAV-2-CACA-N and AAV-2-NLS-VIVIT-GFP vectors (AAV-2-Luc) were produced using the vector production protocol established by the Harvard Gene Therapy Core, except for the AAV8-ttdTomato that was provided by the Gene Transfer Vector Core from University of Iowa, Iowa City, IA. Lentiviral vectors coding for WtNFAT and CANFAT were obtained after amplification of HA-mNFATc4 (WNFAT) and HA-CANFAT (CANFAT) by PCR. Both PCR products were then digested by Nhel and BsrGI restriction enzymes and cloned into a pCSCW2-IREs lentiviral backbone. Lentiviral vectors were produced by the Vector Development and Production Core at Massachusetts General Hospital. All the constructs were verified by sequencing.

Stereotactic intracortical injections. Stereotactic intracortical injections of AAV serotype 2 vectors were performed as described previously (Spires et al., 2005). Animals were anesthetized by intraperitoneal injection of ketamine/xylazine (100 and 50 mg/kg body weight, respectively) and positioned on a stereotactic frame (David Kopf Instruments). Injections of vectors were performed in the cerebral cortex of each hemisphere with 3 μl of viral preparation (titer 2 × 10^12 vg/ml) using a 33-gauge sharp micropipette attached to a 10 μl Hamilton syringe (Hamilton Medical) at a rate of 0.12 μl/min. Stereotactic coordinates of injection sites were calculated from bregma (antero–posterior −1 mm, mediolateral ± 1 mm, and dorsoventral −1.2 mm).

Cranial window implantation and multi photon imaging. Four weeks after AAV intracortical injection, mice were anesthetized with isoflurane (1.5%) and a cranial window was implanted by replacing a piece of skull
by a glass coverslip of 8 mm diameter [as described previously (Spikes et al., 2005)]. For imaging, a wax ring was built on the edge of the window to create a well of water for the objective (Olympus XLPlan N 25/0.11 objective with a numerical aperture of 1.05). To visualize the amyloid deposits, transgenic animals received an intraperitoneal injection of methoxy-XO4 (5 mg/kg) 24 h before surgery, a fluorescent compound that crosses the blood–brain barrier and binds to amyloid deposits (Bacskai et al., 2002; Klunk et al., 2002). When needed, Texas Red dextran (70,000 Da molecular weight; 12.5 mg/ml in sterile PBS; Invitrogen) was injected into a lateral tail vein to provide a fluorescent angiogram.

GFP-filled neuronal processes, amyloid deposits (in the case of APP/PS1 animals), and blood vessels were imaged using the Olympus FluoView FV1000MPE multiphoton laser-scanning system mounted on an Olympus BX61WI microscope (Olympus). A DeepSee Mai Tai Ti:sapphire mode-locked laser (Mai Tai; Spectra-Physics) generated two-photon fluorescence with 860 nm excitation. Emitted light was detected through three filters in the range of 420–460, 495–540, and 575–630 nm (Hamamatsu). Neurites were imaged at depths of 50–200 μm from the surface of the brain. High-resolution images were captured using the optical zoom feature in the Fluoview software (63×63×1 μm sections; 40–100 sections per stack).

**Image processing and analysis.** Two-dimensional projections of GFP-filled neurites were obtained using ImageJ. Only dendrites that were at least 20 μm long and had prominent dendritic spine protrusions were considered for analysis. Spine density and morphology were evaluated using the NeuronStudio software, as described previously (Rodriguez et al., 2005).
Genetically encoded VIVIT blocks NFAT accumulation in the nucleus and prevents spine loss and dendritic simplification induced by constitutively active calcineurin (CaN)

The use of pharmacological inhibitors of CaN phosphatase activity, such as FK506, is associated with a broad range of side-effects in transplanted patients (Ponticelli and Campise, 2005; Lee et al., 2008). In an effort to develop a safer drug, Aramburu et al. (1999) screened for an NFAT-specific reagent and developed a new inhibitor peptide called VIVIT (16 aa, MAGPHPVPHTGPHEE) that was able to interact with the CaN docking motif of NFAT with a high affinity, without interfering with its enzymatic activity.

In vitro, we first tested whether the overexpression of VIVIT-GFP in cortical primary neurons was able to counteract the nuclear translocation and the transcriptional activity of NFATc4 [the most prominent NFAT isoform in neurons (Ho et al., 1994)], as well as the neuronal morphological changes induced by a constitutively active form of calcineurin (CACA\text{N}). The constitutively active CaN was initially identified as a calpain-dependent truncated product of calcineurin (45 kDa), in which the regulatory autoinhibitory domain was removed, so that CaC\text{N} has an enhanced phosphatase activity (Wu et al., 2004). By contrast, overexpression of wild-type calcineurin (WtCaN) is tightly regulated and does not lead to an increase of calcineurin activity. At 5 DIV, cortical primary neurons were cotransfected with either CACA\text{N} or WtCaN and with either GFP or VIVIT-GFP. Cells were cultured for 16–18 DIV before analysis. Because of the presence of an HA-tag in the WtCaN and CACA\text{N} constructs, we verified that >90% of GFP-filled neurons also coexpressed each form of CaN (data not shown). As expected, the nuclear/cytoplasmic ratio of NFATc4 immunofluorescence intensity was significantly higher when CACA\text{N} was overexpressed (0.94 \pm 0.025) compared with WtCaN (0.62 \pm 0.028), whereas WtCaN by itself did not affect NFAT subcellular localization compared with GFP transfected cells (0.67 \pm 0.08) (Fig. 1A, B). This suggests that increased CaN activity leads to an accumulation of NFATc4 in the nucleus. By contrast, co-transfection of CACA\text{N} with VIVIT-GFP prevents the nuclear accumulation of NFATc4, as indicated by lower nuclear/cytoplasmic ratios (0.62 \pm 0.036) that were comparable to WtCaN transfected neurons (Fig. 1A, B). When VIVIT-GFP alone was overexpressed, no change could be detected in the subcellular localization of NFATc4 and the nuclear/cytoplasmic ratio remained comparable to GFP-transfected cells (data not shown). To further evaluate the effect of VIVIT on NFAT transcriptional activity, a reporter system containing several NFAT transcriptional response elements driving the expression of luciferase was used. Primary neurons were transduced with AAV-GFP, AAV-WtCaN+GFP, AAV-CACA\text{N}+AAV-GFP, AAV-CACA\text{N}+AAV-VIVIT-GFP at 10 DIV, and luciferase activity was quantified after 3 d. Overexpression of a constitutively active form of calcineurin induced a marked decrease in NFAT transcriptional activity compared with WtCaN. When VIVIT-GFP was co-transduced with CACA\text{N}, a significant decrease of the Luciferase activity was observed, suggesting that VIVIT efficiently inhibits NFAT-dependent transcriptional activity (Fig. 1C).

To determine whether VIVIT-GFP was able to prevent the pathological morphological changes induced by CACA\text{N}, we analyzed the spine density and dendritic complexity of GFP or VIVIT-GFP-expressing neurons that were also transfected with WtCaN or CACA\text{N} plasmids. The density of spines in CACA\text{N}/GFP overexpressing neurons (0.18 \pm 0.046 spines/\mu m) was significantly lower compared with cells overexpressing WtCaN/GFP (0.42 \pm 0.09 spines/\mu m; Fig. 1D, E). Sholl’s plot analyses also indicated that uncontrolled activation of calcineurin in neurons induced a marked dendritic simplification compared with wild-type calcineurin (Fig. 1D, G). However, when VIVIT-GFP was introduced together with CACA\text{N}, a significantly higher spine density (0.4 \pm 0.081 spines/\mu m) and a more complex neuritic arborization were observed, so that these morphological parameters reached those of WtCaN transfected neurons (Fig. 1D, E, G). Importantly, no difference was observed between cells that were transfected with GFP alone (0.43 \pm 0.13 spines/\mu m) or WtCaN+GFP (0.42 \pm 0.09 spines/\mu m), suggesting that WtCaN did not change the spine density and dendritic complexity of cortical neurons in culture (Fig. 1D, E, G). Similarly, overexpression of VIVIT-GFP alone did not affect the morphological parameters of the cells (data not shown). The beneficial effect of VIVIT-GFP was therefore specifically related to the inhibition of CACA\text{N}-dependent NFAT activation.

Interestingly, when the different spine types were compared, a decreased proportion of “mushroom” type spines was associated with CACA\text{N}+GFP (38\% \pm 1.8\%) compared with GFP (47\% \pm 2.8\%), WtCaN+GFP (44\% \pm 2.3\%), or CACA\text{N}+VIVIT-GFP (46\% \pm 1.4\%) (Fig. 1F), implicating calcineurin activation in the collapse of mature, mushroom-shaped spines that are known to be more stable than thin spines (Holmata et al., 2005).

Constitutively active NFAT induces morphological abnormalities in primary neurons that phenocopy the effects of constitutively active CaN

As both spine loss and dendritic simplification were rescued when a genetically encoded VIVIT-GFP was overexpressed in neurons, we postulated that the morphological effects of CACA\text{N} were related to NFAT activation. We therefore investigated the specific effects of NFATc4 on these morphological parameters. We transduced 14 DIV cortical neurons with ei-
ther a lentiviral vector encoding a HA-tagged wild-type (WtNFAT) or constitutively activated (CANFAT) form of NFATc4 that lacks its N terminus regulatory CaN binding domain. This truncated NFATc4 is therefore activated without calcineurin (Molkentin et al., 1998). To detect morphologival variables, neurons were previously transfected with GFP at 5 DIV and we verified that a high percentage (80%) of the GFP-filled cells were also positive for HA. As observed in Figure 2A, WtNFAT was mainly detected in the cytoplasm, outside the nucleus delimited by laminin B1. On the contrary, the constitutively activated form of NFATc4 was concentrated into the nucleus of the cells and fit the laminin B1 staining. The nuclear localization of CANFAT is not affected by the overexpression of VIVIT-GFP (right). Scale bar, 10 μm. B, Images of GFP-filled neurons (top) and dendrites (bottom) that were transduced with either WtNFAT or CANFAT. Scale bar, 100 μm. C, Overexpression of CANFAT is associated with a decreased number of spines per dendritic segment compared with WtNFAT transduced cells. The loss of dendritic spines observed in CANFAT-expressing neurons is not improved by VIVIT-GFP. D, The overall dendritic complexity is also decreased by CANFAT compared with WtNFAT. The lower dendritic complexity induced by CANFAT cannot be rescued by VIVIT-GFP (*p < 0.05 and **p < 0.01. (n > 30 cells per condition)).

Figure 2. Overexpression of a constitutively activated NFATc4 is sufficient to induce abnormal morphological changes in primary neurons. A, Representative images of the nuclear membrane marker laminin B1 and of HA-tagged NFATc4 in cortical neurons that were transduced with WtNFAT/CANFAT (lentiviral vectors). WtNFAT (left) is mainly present in the cytoplasm, outside the nucleus delimited by laminin B1. The constitutively activated form of NFATc4 was concentrated into the nucleus of the cells and fit the laminin B1 staining. The nuclear localization of CANFAT is not affected by the overexpression of VIVIT-GFP (right). Scale bar, 10 μm.

B, Images of GFP-filled neurons (top) and dendrites (bottom) that were transduced with either WtNFAT or CANFAT. Scale bar, 100 μm. C, Overexpression of CANFAT is associated with a decreased number of spines per dendritic segment compared with WtNFAT transduced cells. The loss of dendritic spines observed in CANFAT-expressing neurons is not improved by VIVIT-GFP. D, The overall dendritic complexity is also decreased by CANFAT compared with WtNFAT. The lower dendritic complexity induced by CANFAT cannot be rescued by VIVIT-GFP (*p < 0.05 and **p < 0.01. (n > 30 cells per condition)).

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In contrast to CACaN-induced NFATc4 nuclear recruitment, accumulation of CANFAT into the nucleus could not be prevented by VIVIT-GFP (Fig. 2A). Overexpression of CANFAT led to a significant decrease of spine density (0.22 ± 0.045 spines/μm) and dendritic simplification compared with WtNFAT (0.39 ± 0.07 spines/μm). None of these changes could be improved by co-transfection with VIVIT-GFP (0.24 ± 0.023 spines/μm) (Fig. 2B–D), thus demonstrating that VIVIT acts upstream of NFAT activation and does not have an effect nonrelated to calcineurin. We therefore concluded that ectopic expression of a constitutively active form of NFATc4 is sufficient to induce spine loss and dendritic simplification in primary neurons, suggesting that NFATc4-related transcriptional events are critical for CaN-induced neurotoxicity in vitro.
Overexpression of VIVIT in Tg2576 transgenic neurons or in wild-type neurons treated with Aβ-containing medium rescues their morphological abnormalities

We previously observed that both transgenic neurons cultured from Tg2576 embryos and wild-type neurons treated with Aβ-containing conditioned media develop AD-like pathological morphological changes that include dendritic spine loss and neurtic simplification. The involvement of CaN in these processes was suggested by the fact that both phenotypes could be rescued when FK506 was added to the medium or used in transgenic mice (Wu et al., 2010; Rozkalne et al., 2011). Here we showed that the overexpression of constitutively activated CaN or NFAT phenocopy the morphological changes induced by Aβ. Based on these observations, we asked whether the inhibition of NFAT activation by calcineurin can also alleviate Aβ synaptotoxicity.

We first tested the efficiency of VIVIT-GFP in transgenic neurons that were cultured from Tg2576 embryos. Because the mutated APP transgene was present as a heterozygous state, transgenic (Tg) or littermate (Wt) cultures could be analyzed in parallel. Cells were transfected with either GFP or VIVIT-GFP at 5 DIV and analyzed at 18 DIV. A decreased density of dendritic spines was observed in Tg neurons (0.21 ± 0.11 spines/μm) compared with their wild-type counterpart (0.45 ± 0.1 spines/μm) (Fig. 3 A, B). The spine loss detected in Tg neurons was also associated with a simplification of the dendritic tree (Fig. 3C). Importantly, when VIVIT-GFP was overexpressed in Tg neurons, both parameters were rescued to normal wild-type levels (spine density: 0.39 ± 0.12 spines/μm) and significantly improved compared with Tg neurons transfected with GFP alone.

To study the effect of exogenous Aβ peptides on neuronal morphological parameters, Aβ-containing conditioned medium (TgCM) was obtained by collecting the medium of Tg neurons after 2 weeks in culture and used to treat naive wild-type cells. The concentration of Aβ40, estimated as 7000 pm, was comparable to the levels observed in AD brains (Ingelsson et al., 2004). Western blot analysis showed that TgCM mainly contained soluble oligomeric species (data not shown) that have been described to have important neurotoxic effects (Walsh et al., 2002; Lesnè et al., 2006; Shankar et al., 2007, 2008). Wild-type primary neurons cultured in standard NB/B27-serum free medium for 16 DIV were exposed for 24 h to either wild-type (WtCM) or transgenic (TgCM) conditioned media diluted 1:1 in the initial medium. As observed in Figure 3, D and E, 24 h exposure to Aβ was sufficient to induce a significant loss of spines in wild-type primary neurons (0.28 ± 0.075 spines/μm) compared with WtCM treatment (0.48 ± 0.12 spines/μm). However, no change of the dendritic complexity was observed, suggesting that such a short exposure with oligomeric Aβ could not affect the whole dendritic arbor of the cells. Aβ immunodepletion (using the anti-amyloid-β monoclonal antibody 6E10) significantly inhibited the effect of TgCM on spines (0.39 ± 0.09 spines/μm), demonstrating that the spine loss was directly related to the presence of amyloid β species (immunodepletion led to a 70% decrease in TgCM Aβ content). Remarkably, when cortical primary neurons were transfected with VIVIT-GFP, before exposure to TgCM, the effect on spine density was greatly ameliorated (0.41 ± 0.108 spines/μm), despite the persistence of Aβ species in the medium. This suggests that blocking CaN-mediated activation of NFAT protects against neurotoxic effects of soluble exogenous Aβ oligomers.

A nuclear-targeted form of VIVIT, but not a membrane bound VIVIT, is a potent inhibitor of CaN-induced morphological changes in primary neurons

VIVIT was initially identified as a selective blocker of the CaN/NFAT interaction, mimicking the PxIxIT sequence of NFAT that is required to bind calcineurin. Although Aramburu et al. (1999) showed that calcineurin activity on critical substrates (such as CREB) was not affected by VIVIT, several recent studies suggested that it can also disrupt the interaction between CaN and other important cytosolic targets implicated in the regulation of synaptic activity such as AKAP or cabin (Dell’Acqua et al., 2002; Liu, 2003). In an attempt to precisely decipher the role of the transcription factor NFAT in Aβ-related neurotoxic events, we used a nuclear-targeted version of VIVIT that was shown to have a restricted localization within the nucleus and that was previously used to specifically counteract NFATc4-related transcriptional activity (Schwartz et al., 2009). As a negative control, we fused a myristoylation (Myr) tag to the VIVIT-GFP plasmid to direct the localization of this peptide toward the cell membrane. Thus, these constructs would disambiguate the transcriptional effects of VIVIT from other non-transcriptional functions at the spines.

We first tested whether these two constructs, NLS-VIVIT-GFP and Myr-VIVIT-GFP, showed the expected subcellular localization. At 5 DIV, primary cortical neurons were transfected with GFP, NLS-VIVIT-GFP, or Myr-VIVIT-GFP. As observed in Figure 4A, NLS-VIVIT-GFP was strictly restricted to the nuclear compartment and limited by the nuclear membrane marker laminin B1. By contrast, the fluorescent signal of Myr-VIVIT-GFP was completely excluded from the nucleus. When each plasmid was transfected altogether with DsRed (as a common denumerator to obtain the full morphology of each cell), both GFP and Myr-VIVIT-GFP could be detected in the processes of neurons and in the dendritic spines, which was not the case for NLS-VIVIT-GFP (Fig. 4B).

To determine the effect of NLS-VIVIT and Myr-VIVIT on NFAT transcriptional activity, primary neurons were transduced with an AAV-pNFAT-Luc reporter vector, altogether with AAV-WtCaN+GFP, AAV-CaCaN+AAV-GFP, AAV-CaCaN+AAV-NLS-VIVIT-GFP, or AAV-CaCaN+AAV-Myr-VIVIT-GFP. After 3 d, the luciferase activity was quantified. As previously observed, overexpression of a constitutively active form of calcineurin led to an increase of NFAT transcriptional activity compared with WtCaN. This increase was significantly diminished when NLS-VIVIT-GFP was expressed, but no change could be detected with Myr-VIVIT-GFP. This suggested that only a nuclear-targeted inhibitor was able to inhibit NFAT transcriptional activity in vitro (Fig. 5A). We next determined whether CaN-induced morphological changes could be rescued by the different VIVIT-GFP constructs by co-transfecting neuronal cultures with either WtCaN or CaCaN, altogether with GFP, NLS-VIVIT-GFP, or Myr-VIVIT-GFP. Because dendrites and spines were not detectable from the GFP signal when NLS-VIVIT-GFP was overexpressed, we used DsRed as a common denominator to evaluate spine density and dendritic complexity. At 18 DIV, both spine loss and dendritic simplification associated with CACaN overexpression was suggested by the fact that both phenotypes could be rescued by NLS-VIVIT-GFP (Fig. 5B–D). By contrast, no effect could be detected when Myr-VIVIT-GFP was used, consistent with the hypothesis that the neuroprotection of VIVIT against CaCaN-mediated neurodegeneration is related to transcriptional effects in the nucleus rather than non-transcriptional effects in the cytoplasm or dendrites.
Figure 3. The morphological abnormalities observed in Tg neurons and in TgCM-treated neurons are improved by VIVIT-GFP. A. Representative images of Wt and Tg GFP-labeled neurons (left) and dendrites (right) at 18 DIV. Scale bar, 100 μm. B, GFP-transfected Tg neurons present a lower spine density compared with Wt littermate cells, but the number of spines is (Figure legend continues.)
Nuclear-targeted VIVIT, but not membrane-targeted VIVIT, protects against Aβ neurotoxicity in primary neurons

In the previous experiment, we determined that a nuclear version of VIVIT was almost as potent as a nontargeted VIVIT at blocking the pathological changes induced by CACaN. Considering the fact that calcium dyshomeostasis and CaN activation have been shown to be downstream events to Aβ neurotoxicity (Busche et al., 2008; Kuchibhotla et al., 2008), we next asked whether a nuclear-targeted version of this peptide may be protective against Aβ toxicity as well.

Following the same procedures as described above, we tested the NLS-VIVIT-GFP or Myr-VIVIT-GFP constructs in Wt and Tg primary neurons that were co-transfected with DsRed (Fig. 6A). Both spine density and dendritic complexity were significantly improved when NLS-VIVIT-GFP was overexpressed in Tg neurons, but no improvement was detected with Myr-VIVIT-GFP (Fig. 6B, C).

Similarly, when Wt primary neurons were transfected with GFP, NLS-VIVIT-GFP, or Myr-VIVIT-GFP and then stained for laminin B1 and DAPI (left). On the right, higher magnification images show that GFP alone can be detected in the nuclear and cytoplasmic compartments, whereas NLS-VIVIT-GFP is surrounded by laminin B1 and colocalizes with DAPI, suggesting that it is restricted to the nucleus. Inversely, Myr-VIVIT-GFP is excluded from the nuclear compartment. Scale bar, 50 μm. B, Both GFP and Myr-VIVIT-GFP can be detected in the dendritic shaft and in spines, as suggesting by the colocalization with DsRed. NLS-VIVIT-GFP is, however, absent in the neuronal processes. Scale bar, 100 μm.

Together, these observations strongly suggest that NFAT-related transcriptional events play a role in Aβ-related neurotoxicity, thus predicting that inhibition of the CaN/NFAT pathway may be a protective strategy against Aβ-induced neuronal injuries in vivo.
Disruption of NFAT pathway with VIVIT and NLS-VIVIT increases spine density in the vicinity of amyloid plaques

In vitro, we observed that a selective disruption of the interaction between CaN and NFAT may alleviate Aβ-related neurotoxicity. We asked whether these neuroprotective effects could be reproduced in vivo. Neurites surrounding amyloid deposits have an increased calcium overload, decreased spine density, and an increased probability to develop neuritic dystrophies (Spires et al., 2005; Kuchibhotla et al., 2008). Interestingly, overexpression of a constitutively activated CaN (CaCaN) can be improved by co-transfection with a nuclear-targeted VIVIT (NLS-VIVIT), whereas no beneficial effects is observed with a membrane-bound VIVIT. Overexpression of NLS-VIVIT also leads to a more complex dendritic arborization of CaCaN transfected neurons, whereas overexpression of Myr-VIVIT cannot rescue the effects of CaCaN. *p < 0.05. (n > 30 cells per condition).

Figure 5. The restricted localization of VIVIT into the nucleus potently inhibits the morphological abnormalities induced by an overactivated form of CaN. A, NFAT transcriptional activity assessed using the pNFAT-luciferase reporter system shows that overexpression of NLS-VIVIT-GFP, altogether with CaCaN, is associated with a decreased luciferase activity. No effect is observed when Myr-VIVIT-GFP is overexpressed, suggesting that this inhibitor does not affect NFAT transcriptional activity. B, Representative images of cortical primary neurons (top) and dendrites (bottom) that were co-transfected with GFP, NLS-VIVIT-GFP, and Myr-VIVIT-GFP altogether with CaCaN and WTCaN. To be able to observe the neurites and quantify the dendritic spines in all the conditions, cells were co-transfected with a DsRed fluorescent reporter. Scale bar, 100 μm. C, The decreased spine density associated with the overexpression of a constitutively activated CaN (CaCaN) can be improved by co-transfection with a nuclear-targeted VIVIT (NLS-VIVIT-GFP), whereas no beneficial effects is observed with a membrane-bound VIVIT. D, Overexpression of NLS-VIVIT also leads to a more complex dendritic arborization of CaCaN transfected neurons, whereas overexpression of Myr-VIVIT cannot rescue the effects of CaCaN. *p < 0.05. (n > 30 cells per condition).
Figure 6. Aberrant neuronal morphology observed in Tg neurons and in TgCM-treated cells are prevented by NLS-VIVIT-GFP but not by Myr-VIVIT-GFP. A, Representative images of DsRed-filled Wt or Tg neurons (left) and dendrites (right) overexpressing GFP or the different VIVIT constructs show that Tg cells transfected by NLS-VIVIT-GFP present an increased spine density compared with GFP transfected Tg cells. Scale bar, 100 μm. B, Bar graph summarizing the averaged spine density of each group confirms that only the nuclear targeted peptide is able to improve the morphological abnormalities observed in Tg neurons, but this is not the case when Myr-VIVIT-GFP is overexpressed. C, Sholl’s plot analysis demonstrates that NLS-VIVIT-GFP also increases the dendritic (Figure legend continues.)
of APP/PS1 transgenic mice and examining its effects on neuritic degeneration.

To investigate the effect of VIVIT in vivo, AAV vectors encoding for VIVIT-GFP, NLS-VIVIT-GFP (both inhibitors having proven beneficial effects in vitro), and GFP (as a control) were stereotactically injected in the cortex of 7-month-old littermates or APP/PS1 mice, when amyloid deposits are already present. In the case of AAV-NLS-VIVIT-GFP injected mice, we first verified that the recombinant protein was properly addressed to the nuclear compartment and colocalized with Hoechst-stained nuclei (Hoechst solution was applied topically; Fig. 7A). AAV-NLS-VIVIT-GFP and AAV-GFP were then coinjected with a ratio of 3:1, so that a high proportion of GFP filled neurons were also transduced by the AAV-NLS-VIVIT-GFP vector. One month later, a cranial window was implanted. Neurites and spines were detected and quantified by multiphoton imaging in the living animal (Fig. 8A). High-magnification images were taken to visualize dendritic spines. Spine density was quantified for each neuritic segment using the NeuronStudio software (see Materials and Methods). Importantly, no significant difference was observed when comparing the spine density in wild-type littermate mice injected with either AAV-GFP (0.47 ± 0.11 spines/μm), AAV-VIVIT-GFP (0.49 ± 0.107 spines/μm), or AAV-NLS-VIVIT-GFP (0.53 ± 0.1 spines/μm) (data not shown), indicating that VIVIT-GFP does not have an effect on spine density by itself, i.e., independently of Aβ.

Compared with dendrites from GFP-injected wild-type littermates (0.47 ± 0.11 spines/μm), dendrites in the vicinity of amyloid deposits (<100 μm away from plaque) in APP/PS1 mice exhibited a decreased spine density (0.31 ± 0.12 spines/μm). This amyloid-associated spine loss was restored to essentially normal levels when either AAV-VIVIT-GFP (0.49 ± 0.13 spines/μm) or AAV-NLS-VIVIT-GFP + AAV-GFP (0.44 ± 0.1 spines/μm) were injected in APP/PS1 mice (Fig. 8B). In both APP/PS1 animals and human AD brains the spine densities are correlated with the distance from the edge of the amyloid deposits (correlation coefficient: 0.41) (Koffie et al., 2009) (Spires et al., 2005). However, this local effect of amyloid plaques was nearly abolished when AAV-NLS-VIVIT-GFP (correlation coefficient: 0.22) or AAV-VIVIT-GFP was expressed (correlation coefficient: 0.09; Fig. 8C).

Overall, the injection of AAV-VIVIT-GFP was associated with a marked recovery of spine density around amyloid deposits in vivo. A significant beneficial effect was also observed in AAV-NLS-VIVIT injected mice, even though this improvement did not reach the levels of VIVIT-GFP. This difference might be due to the fact that some of the GFP-filled neurites observed in animals coinjected with AAV-NLS-VIVIT-GFP and AAV-GFP had only been transduced by the later vector. To examine this possibility, we coinjected the AAV-NLS-VIVIT-GFP and an AAV-TdTomato with the same 3:1 ratio as previously used, and we observed that 86% of the red fluorescent cells also expressed NLS-VIVIT (arrows indicate GFP/TdTomato double positive cells). Scale bars: 100 μm on the top and 50 μm on the bottom.

Neuritic abnormalities observed around amyloid deposits in APP/PS1 mice are partially restored by VIVIT-GFP and NLS-VIVIT-GFP

The presence of amyloid deposits not only affects the density of dendritic spines, but is also known to disturb the morphology of the neuritic shaft itself (Knowles et al., 1999). We therefore asked whether overexpression of VIVIT or NLS-VIVIT might have an additional rescue effect in regards to these morphological parameters. As previously shown, amyloid deposits are associated with the development of neuritic dystrophies, which can be visualized by AAV-GFP injection in APP/PS1 mice. Even though the size of the...
amyloid plaques that were analyzed was comparable between all the injected animals, plaques from AAV-VIVIT-treated mice were associated with fewer dystrophies (5 ± 4 dystrophies/plaque) compared with those from AAV-GFP (14 ± 9 dystrophies/plaque) and AAV-NLS-VIVIT (18 ± 10 dystrophies/plaque; Fig. 9A, B) injected mice. Because the density of GFP-filled neurons around plaques was similar among all the groups (data not shown), we conclude that the difference in the number of dystrophies was not due to a decreased amount of GFP-filled neurites between AAV-GFP and AAV-VIVIT-GFP treated animals. This finding suggests that dystrophies, unlike dendritic spine loss, might not be recovered by a restricted inhibition of NFAT transcriptional activity in the nucleus. Interestingly, like VIVIT, calcineurin inhibition with FK506 led to an improvement in neuritic dystrophies (Rozkalne et al., 2011). The remaining dystrophies in AAV-VIVIT-GFP or NLS-VIVIT-GFP treated groups are not significantly different from GFP injected littermates. A third morphological characteristic of neurites around amyloid deposits is a subtle change in their trajectories (Knowles et al., 1999). This abnormal neuritic curvature was assessed on paraffin-embedded sections after immunostaining for GFP and amyloid plaques. The curvature ratio was calculated by reporting the length of a neurite divided by the end-to-end length of the same segment. We observed that the average neuritic curvature of both AAV-VIVIT-GFP (1.054 ± 0.038) and AAV-NLS-VIVIT-GFP (1.059 ± 0.048) injected mice were improved around amy-
loid deposits compared with AAV-GFP-treated animals (1.081 ± 0.07) (Fig. 9C,D).

**The beneficial effects associated with VIVIT-GFP and NLS-VIVIT-GFP correlate with a decreased recruitment of NFATc4 into the nucleus**

Using an *in vivo* imaging approach, we observed that several neuronal morphological parameters (i.e., spine density, neuritic dystrophies and curvature) that are abnormal in the vicinity of amyloid deposits were significantly improved when either AAV-VIVIT-GFP or AAV-NLS-VIVIT-GFP was injected in the cortex of APP/PS1 mice. To verify that these beneficial effects were related to the efficient inhibition of NFATc4 recruitment into the nucleus, we performed a postmortem immunohistological analysis.

Brain sections of injected mice were stained to detect both GFP and endogenous NFATc4 to determine the ratio of NFATc4 in the nucleus versus cytoplasm in transduced neurons. We found that the distribution of NFATc4 nuclear/cytoplasmic ratios was shifted toward lower values in APP/PS1 mice injected with either VIVIT-GFP (0.89 ± 0.06) or NLS-VIVIT-GFP (0.83 ± 0.09) (Fig. 10A,B).

**Discussion**

The biological mechanisms that sculpt the fine structure of the adult brain and their alterations in neurodegenerative diseases remain largely unknown. In this report we demonstrated that Aβ neurotoxic damage can be prevented or even reversed by inhibiting calcineurin-mediated activation of NFAT. Importantly, the same profound morphological changes occur when a constitu-
NFAT replicate the decrease of spine density and dendritic simplification induced by the neurotoxic Aβ peptides. Along with recent data indicating that calcineurin is activated both in human AD brain and in transgenic models of AD (Liu et al., 2005; Dineley et al., 2007; Wu et al., 2010), we tested the hypothesis that inhibition of the excessive NFAT activity might improve the neuritic abnormalities observed in AD models. We demonstrate that VI-VIT, a genetically encoded inhibitor of the interaction between calcineurin and NFAT, increases both the spine density and the complexity of dendritic arbors in APPsws-transgenic neurons or in TgCM-treated wild-type neurons. In vivo, a gene transfer approach using adeno-associated vectors enabled the delivery of VIVIT to neurons and significantly increased the spine density in the vicinity of senile plaques. Transduction with an inhibitor that, due to a nuclear localization signal (NLS-VIVIT), specifically blocked the nuclear activation of NFAT, was also able to restore to nearly normal the morphological abnormalities associated with Aβ. The ability of NLS-VIVIT to reproduce the same beneficial effects as VIVIT is intriguing, as this peptide is thought to interact primarily with the docking site of NFAT upon CaN. However, several studies demonstrated that activated CaN is able to translocate to the nucleus in neurons (Pujol et al., 1993; Solàe et al., 1999; Yang et al., 2005; Schwartz et al., 2009), a phenomenon we also previously described in AD patients (Wu et al., 2010). We therefore propose that both CaN and NFAT translocate to the nuclear compartment, where VIVIT would compete with NFAT for CaN binding. Considering the higher affinity of VIVIT for CaN [~25-fold compared with NFAT (Aramburu et al., 1999)], the CaN/NFAT interaction would thus be interrupted and NFAT would rapidly be phosphorylated and shuttled back to the cytoplasm. This hypothesis is in agreement with the previous observation that NLS-VIVIT efficiently inhibited the expression of NFAT-target genes (Schwartz et al., 2009), and with our observation of a significant decrease in the NFAT nuclear/cytoplasmic ratio not only in AAV-VIVIT-GFP-injected animals but also in AAV-NLS-VIVIT-treated mice.

Our results indicate that (1) NFAT activation is a likely consequence of Aβ accumulation in Alzheimer’s disease, as suggested by its prominent nuclear localization in the brain of AD patients (Abdul et al., 2009; Wu et al., 2010), and by the ability of VIVIT to restore Aβ-associated morphological neurodegenerative changes; (2) once activated and translocated to the nucleus, NFAT presumably induces the transcription of target genes in mature neurons that lead to a pathological remodeling of dendrites and dendritic spines. The beneficial effect of a nucleus-directed NFAT inhibitor supports the role of transcriptional events in the regulation of dendritic spine stability in vitro and in vivo.

Figure 10. Overexpression of VIVIT-GFP and NLS-VIVIT-GFP prevent NFATc4 accumulation in the nucleus of transduced neurons. A, Representative images of GFP immunostaining and endogenous NFATc4 in injected animals allows detection of endogenous NFATc4 in transduced neurons. Scale bar, 50 μm; for higher magnification images, on the right, 20 μm. Higher magnification images of NFATc4 and DAPI (third column) show that an increased recruitment of NFATc4 is detected in AAV-GFP-injected APP/PS1 mice. The nuclear/cytoplasmic ratio is significantly increased in AAV-GFP treated transgenic mice compared with AAV-VIVIT and AAV-NLS-VIVIT treated animals. On the graph, the blue line represents the mean of NFATc4 nuclear/cytoplasmic ratio in all three groups tested. However, there is a shift toward higher values in AAV-GFP-injected APP/PS1 mice. The nuclear/cytoplasmic ratio is significantly increased in AAV-GFP treated transgenic mice compared with AAV-VIVIT-GFP and AAV-NLS-VIVIT-GFP (p < 0.05). (n = 4 animals per group and 3 slices per animal were analyzed).
NFAT to interact with other transcriptional factors (Crabtree and 2003). These varied results could be explained by the ability of NFAT to interact with other transcriptional factors (Crabtree and Olson, 2002; Ogata et al., 2003; Kao et al., 2009), a mechanism by which the pattern of NFAT-induced target genes can be modulated depending on the developmental stage, on the cell type, and on the physiological or pathological context. It is therefore conceivable that a particular set of NFAT coactivators are present in cortical neurons exposed to Aβ, so that its activation leads to the upregulation of genes implicated in dendritic spine shrinkage.

Although the transcriptional effects of NFAT (and its inhibition) await further investigation, the present study provides a novel neuroprotective therapeutic target against the downstream neurotoxic effects of amyloid-β and confirms the feasibility to pharmacologically arrest the “amyloid cascade” of neurodegeneration at a step after amyloid deposition has already occurred. The latter has critical therapeutic implications, since most amyloid deposition is thought to occur years or even decades before the onset of cognitive decline, and therefore, before AD diagnosis, whereas synaptic and neuron loss largely occurs in a second stage and correlates with the severity of cognitive impairment (Sperling et al., 2009). In this scenario, amyloid-directed therapies would not be sufficiently effective once cognitive symptoms (and the underlying neurodegenerative processes) have begun. Consequently, combined synergistic therapies aimed at reducing Aβ content (either by inhibiting the production of Aβ peptides or by increasing their clearance) and at restoring Aβ-induced neural damage would be needed.

The literature addressing the question of NFAT target genes specifically regulated in neurons is quite limited, but few interesting findings might be relevant in the context of Alzheimer’s disease. For example, NFAT was shown to contribute to the induction of apoptosis in neuroblastoma cells through upregulation of Fas Ligand (Fas-L) (Luoma and Zirpel, 2008; Alvarez et al., 2011), a gene that was also reported to increase in AD brains, especially in the hippocampal formation and entorhinal cortex that are primarily involved in memory encoding. In addition, Aβ peptides induce a pathological increase in Fas-L in cortical primary neurons and its accumulation was demonstrated in neuritic dystrophies surrounding the amyloid deposits (Su et al., 2003). Even though no direct connection between Fas-L and spine morphology has been reported so far, one of the down-stream target of Fas-L, capase-3, was recently shown to trigger early synaptic dysfunction and spine loss in AD (D’Amelio et al., 2011). This would suggest that activation of capase-3 may not only be involved in cell death but is also closely associated with the regulation of synaptic plasticity. It is therefore possible that the activation of Fas-L expression by NFAT indirectly causes a synaptic failure via the induction of a non-apoptotic capase-3 pathway. The expression of the potassium channel Kv2.1 can also be driven by NFAT and was shown to be upregulated by 72% in the hippocampus of rat injected with Aβ25–35 (Pan et al., 2004). Interestingly, the use of several potassium channel openers (minoxidil, pinacidil, cromakalim) can induce amnesia in mice (Ghelardini et al., 1998), thus suggesting that the regulation of the activity of the potassium channels might impair memory encoding, one of the best hallmark of AD. As learning and memory deficits mainly reflect a default in spine-mediated plasticity, we can hypothesis that dysregulation of the potassium channel Kv2.1 might lead to the alteration of dendritic spine morphology. Last, the upregulation of the inositol 1,4,5-trisphosphate type 1 receptor (InsP3R) gene by NFAT is of particular interest in the context of Alzheimer’s disease (Genazzani et al., 1999; Amberg et al., 2004). Indeed, InsP3R was shown to directly interact with presenilins 1 and 2, increasing its activity in response to Inositol-3-phosphate, thus leading to an excessive release of Ca²⁺ (Müller et al., 2011). The upregulation of InsP3R might therefore participate to exacerbate the increase of the resting calcium levels. The few examples mentioned above might be of interest in the context of AD, but other NFAT target genes previously identified does not seem relevant in a pathological context. Indeed, NFAT transcriptional activity was associated with an increased expression of the neurotrophic factor BDNF in Purkinje cells, an important mediator of axonal outgrowth during development (Graef et al., 2003). These varied results could be explained by the ability of NFAT to interact with other transcriptional factors (Crabtree and References


