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SLXL1, a Novel Acrosomal Protein, Interacts with DKKL1 and Is Involved in Fertilization in Mice

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Abstract

Background: Spermatogenesis is a complex cellular developmental process which involves diverse families of genes. The Xlr (X-linked, lymphocyte regulated) family includes multiple members, only a few of which have reported functions in meiosis, post-meiotic maturation, and fertilization of germ cells. Slx-like1 (Slxl1) is a member of the Xlr family, whose expression and function in spermatogenesis need to be elucidated.

Methodology/Principal Findings: The mRNA and protein expression and localization of Slxl1 were investigated by RT-PCR, Western blotting and immunohistochemistry in different tissues and at different stages of spermatogenesis. The interacting partner of SLXL1 was examined by co-immunoprecipitation and co-localization. Assessment of the role of SLXL1 in capacitation, acrosome reaction, zona pellucida binding/penetration, and fertilization was carried out in vitro using blocking antisera. The results showed that Slxl1 mRNA and protein were specifically expressed in the testis. SLXL1 was exclusively located in the acrosome of post-meiotic germ cells and interacts with DKKL1 (Dickkopf-like1), which is an acrosome-associated protein and plays an important role in fertilization. The rates of zona pellucida binding/penetration and fertilization were significantly reduced by the anti-SLXL1 polyclonal antiserum.

Conclusions/Significance: SLXL1 is the first identified member of the XLR family that is associated with acrosome and is involved in zona pellucida binding/penetration and subsequent fertilization. These results, together with previous studies, suggest that Xlr family members participate in diverse processes from meiosis to fertilization during spermatogenesis.

Introduction

Spermatogenesis is a complex process that can be divided into three stages: mitosis in spermatogonia, meiosis in spermatocytes, and spermiogenesis in round spermatids and later cell types. The end products of spermatogenesis are the spermatoozoa that contain specialized structures such as flagellum and acrosome. The acrosome plays an essential role during fertilization and is formed from a secretory vesicle by a Golgi-derived apparatus in the initial steps of spermiogenesis [1]. Acrosome biogenesis is a multi-step process including vesicular trafficking and organelle migration [1,2,3]. The acrosome contains a variety of proteins including protease zymogens, zona pellucida-binding proteins and DKKL1 protein [4,5,6,7]. Lack of acrosome reduces the rate of fertilization in vitro [8,9]. Acrosome reaction is an exocytic event in fertilization that releases enzymes to facilitate the spermatozoa’s binding to and penetration of the zona pellucida [10]. Clinical studies have identified patients whose infertility is associated with an abnormal acrosome reaction [11].

The expression of Xlr (X-linked lymphocyte-regulated complex) genes was initially identified in terminally differentiated B lymphoid cells [12,13]. The proteins of this family, sharing the COR1 domain, include XLR, SYCP3 (Synaptonemal complex protein 3), SLY (Syce3-like, Y-linked), and SLX/XMR (Syce3-like, X-linked). XLR is the prototype member, and is a nuclear protein expressed in the spermatocyte and oocyte nucleus during prophase of meiosis [14]. SYCP3 is expressed in meiotic cells and is a basic component of the lateral elements of the synaptonemal complex [15]. Syce3 is required for male fertility and the Syce3−/− male germ cells die around the zygote stage of meiosis [16,17]. Syce3-deficient females are fertile but the embryos are not viable due to massive aneuploidy [18,19]. Sly, a Y chromosome-linked gene of the family is only expressed in spermatids during spermatogenesis [20]. Interestingly, SLY interacts with DKKL1 which is involved in fertilization [20] probably by facilitating spermatozoa’s penetration of the zona pellucida [8,9]. In Sly-deficient mice, spermatozoa differentiation is severely impaired, and the spermatozoan heads are abnormal [21]. Slx/Xmr is an X-linked gene and its protein shares 94% and 43% homologies with Xlr and Sly, respectively [22]. Slx/Xmr was initially reported to be a nuclear protein expressed during meiosis [23,24]. Another study indicated that Slx encodes a spermatid cytoplasmic protein [25].
Moreover, the spermatozoa of Slx-deficient mice displayed abnormal heads and the males were sterile [26]. Six-like1 (Sxl1) is a multi-copy gene (~16 copies) located on the murine X chromosome [27]. SLXL1 and SLX expression were up-regulated in Syl-deficient mice [26]. However, the expression and function of SLXL1 in spermatogenesis have not been well-addressed. In the present study, we conducted an in-depth investigation of the expression of Sxl1 at both the mRNA and the protein levels. We report that Sxl1 protein is specifically present in the acrosomes of all post-meiotic germ cells. In addition, SLXL1 interacts with DKKL1 and plays an important role in zona pellucida binding/penetration and fertilization. Our results together with previous ones by others indicate that the XLR family proteins may be involved in diverse processes from meiosis to fertilization during spermatogenesis.

Results

Sxl1 was a novel testis-specific member of the mammalian Xlr superfamily

Sxl1 was initially identified as one of the large number of testis-specific genes by mining microarray expression data (unpublished data). The protein (155 aa) was annotated by the NCBI Conserved Domain Database to belong to the XLR superfamily, whose members share the COR1 domain. SLXL1 does not contain additional domain(s)/motif(s) such as signal peptide, transmembrane region(s) indicating it is most likely a cytoplasmic protein. Its closest paralogs in the mouse genome are SLX and SLY with identity of 69% and 42% to SLXL1, respectively (Fig. 1).

The expression of Sxl1 mRNA was detected by RT-PCR using three pairs of primers (Fig. 2A) in various tissues of mice. Sxl1 mRNAs was only detected in the testis (Fig. 2B). Moreover, Sxl1 mRNA expression in testis was weakly detected at 14 dpp (day post partum) when spermatogenesis progressed to the pachytene spermatocyte stage, and was greatly increased at 21 dpp and thereafter when spermatids are produced in the first wave of spermatogenesis [20]. To examine the expression in a more precise manner, additional time points, one every two days from 10.5 dpp to 22.5 dpp, were included for the RT-PCR detection. As shown by Fig. 2C, the Sxl1 mRNA was first detected at 14 dpp and increased it level gradually until 22.5 dpp.

In order to study the protein expression of SLXL1 by immunostaining, two rabbit polyclonal antisera (anti-SLXL11-155aa and anti-SLXL165-155aa) were developed using the GST-fusion proteins of full-length SLXL1 and the C-terminal fragment, respectively. A similar antiserum against the C-terminal part of SLX (anti-SLX95-212aa) was also produced. The predicted molecular mass of SLX and SLXL1 are 25 kDa and 18 kDa respectively. As shown in Fig. 2D, the anti-SLXL195-212aa in Western blotting detected a protein of about 25 kDa in adult testis but not in mature spermatozoa, indicating that this antiserum specifically recognizes SLX. While anti-SLXL11-155aa in Western blotting detected two proteins of about 25 kDa and 18 kDa in the testis, it only detected the 18 kDa one in the spermatozoa, indicating that this antiserum recognizes both SLX and SLXL1 when both are present. In contrast, the anti-SLXL155-155aa only detected SLXL1 in both testis and spermatozoa. The SLXL1 was expressed only in the testis of the several mouse tissues examined using both anti-SLXL11-155aa and anti-SLXL165-155aa (Fig. 2E and 2F). Consistent with the results of a previous study [25] and of the Sxl1 mRNA expression in the present study, SLXL1 was dramatically up-regulated post-meiotically starting from a time point between 21 and 28 dpp.

SLXL1 was an acrosome-associated protein

To examine the cellular and sub-cellular localization of SLXL1 protein in testis, immunostaining was performed on adult testis sections using anti-SLXL11-155aa and anti-SLXL165-155aa (anti-SLXL195-212aa failed to work immunohistochemically). Small bright SLXL1-positive particles adjacent to the nuclei of post-meiotic germ cells were detected, and

Figure 1. Amino acid sequence alignment of several representative members of the XLR family. doi:10.1371/journal.pone.0020866.g001
this type of signal was in strong contrast to that of SLX which was diffusely cytoplasmic in post-meiotic germ cells (Fig. 3A and 3B). The change in shape of the SLXL1 signal from round to oval/crescent particles as spermatids progress from round to elongated ones suggested that SLXL1 could be localized to acrosomes of different developmental stages (Fig. 3C–3F). Indeed, SLXL1 co-localized with an acrosomal protein DKKL1 at these stages (Fig. 4C). Moreover, the SLXL1 particles did not co-localize with γH2AX (phosphorylated H2A histone family, member X) (Fig. 3C), a marker for DNA double-strand break repair and the formation of sex body during meiosis of spermatocytes [29]. The acrosome localization of SLXL1 was most evident in released spermatozoa in the epididymis as the acrosomes can be easily recognized by its crescent shape on the apex of spermatozoa (Fig. 3G and 3H).

SLXL1 interacted with acrosomal protein DKKL1

Since both SLXL1 and DKKL1 are localized in the acrosome and SLY, a close paralog of SLXL1 interacts with DKKL1, we wondered whether SLXL1 and DKKL1 interact with each other. This turned out to be the case as indicated by co-immunoprecipitation (Co-IP) results. First, SLXL1-GFP and DKKL1-FLAG proteins expressed in K293T cells co-immunoprecipitated with each other (Fig. 4A). Second, the two native proteins in testis lysate also interacted with each other when the anti-SLXL1 and anti-DKKL1 antisera were used for IP and IB, respectively (Fig. 4B). Consistent with this, SLXL1 and DKKL1 co-localized in round and elongating spermatids (Fig. 4C).

In vitro fertilization was reduced by SLXL1 antiserum

We used in vitro fertilization assays to investigate a possible role for SLXL1 in fertilization. Successful in vitro fertilization was identified by the appearance of embryos at the 2-cell and 4-cell stages. While the fertilization rates of the untreated group, the pre-immune serum treated group, and the anti-SLX95–212aa treated group are all above 45%, that of the anti-SLXL11–155aa and anti-SLXL165–155aa treated groups dropped significantly to 8% and 17%, respectively (Fig. 5A). To exclude the possibility that the antiserum itself may contain any toxic factor to fertilization, we
SLXL1 Is Involved in Fertilization

Figure 3. Localization of SLXL1 in the testis and mature spermatozoa. (A) Co-staining of adult testis sections for the DAPI (blue) and SLX (green) using anti-SLX95–212aa. (B) Immunostaining of adult testis sections using anti-SLXL11–155aa. (C–D) A Stage VIII tubule immunostained by antisera for SLXL1 (red) and γH2AX (green); (E–F) A Stage VI–VII tubule showing the SLXL1 (red) localization in the round spermatid and elongating spermatozoa. (G) Immunostaining of SLXL1 in the spermatozoa of epididymis (red) showing its localization in the acrosome. (H) SLXL1 (red) was present in the acrosome of mature spermatozoa. D and F are the magnified views of the rectangular areas in C and E, respectively.

doi:10.1371/journal.pone.0020866.g003

added recombinant GST-SLXL1 protein to the anti-SLXL11–155aa treated group and the fertilization rate reverted back to above 40%. Therefore, SLXL1 plays a role in fertilization and its action can be blocked by its neutralizing antiserum.

Binding/penetration of spermatozoa to zona pellucida was inhibited by SLXL1 antiserum

As fertilization is a final readout of several earlier steps such as capacitation, acrosome reaction, and binding/penetration to zona pellucida of spermatozoa, it is necessary to check whether these steps could be affected by the SLXL1 antiserum. The capacitation of spermatozoa collected from the epididymis was induced by incubation in the HTF (Human Tubal Fluid) medium and were measured using the CASA system. As shown in Fig. 5B and 5C, neither the general motility nor the forward progression of spermatozoa was changed by the anti-SLXL11–155aa antiserum. In addition, acrosome reaction (AR) of spermatozoa is not changed significantly by the antiserum compared to the control preimmune serum group as shown by Fig. 5D.

The binding/penetration of spermatozoa to the zona pellucida of eggs was examined in the absence and presence of the anti-SLXL11–155aa antiserum. As shown by Fig. 6, while the control preimmune serum did not change the binding/penetration compared with the normal in vitro fertilization group (HTF), the antiserum reduced the binding/penetration significantly. The inclusion of SLXL1 recombinant protein in the antisera treatment reversed the binding/penetration back to the normal level, indicating the inhibitory effect of the antiserum was specific to the endogenous SLXL1.

Discussion

SLXL1 shares the highest amino acid identity with SLX, but its expression and potential role during spermatogenesis and/or fertilization has not been well addressed. The expression of Slxl1 mRNA and protein along with SLX in spermatogenesis have been reported in several studies [24,25], but some questions remain open. First, what is the cellular and sub-cellular localization of SLXL1? The answer to this question was not straightforward. The main problem was that anti-SLXL11–155aa developed against full-length SLXL1 detected both itself and its close paralog SLX on Western blot while anti-SLXL185–155aa, did not work on testis sections for immunostaining although it specifically recognized SLXL1 on Western blot. The cross-reaction of an anti-SLX antibody to SLXL1 on Western blot was also reported by others [26]. SLX was initially misidentified as being expressed in meiotic germ cells due to the use of an antibody that had not been well characterized [24], but has now been recognized as being expressed cytoplasmically in spermatids [25]. The signal detected by our anti-SLXL11–155aa was exclusively present in the acrosome of post-meiotic germ cells suggesting that on sections it detects SLXL1 but not SLX. On this basis, and given the interactions described below, it is evident that SLXL1 is an acrosomal protein, contrary to the expectation that it would be cytoplasmic based on its homology to SLX [26].

What does SLXL1 interact with in the acrosome? Based on the homology of SLXL1 and SLY, as well as the fact that SLY interacts with the acrosomal protein DKKL1, we hypothesized that SLXL1 also interacts with DKKL1. This turned out to be true as evidenced by IP assays using both over-expressed proteins in 293T cells and native proteins in testis lysates. Because SLY was reported to be a cytoplasmic protein and involved in regulating the expression of multiple genes [20,21], the implication of its interaction with the acrosomal protein DKKL1 is unclear at the moment. In contrast, the interaction of SLXL1 with DKKL1 is easier to understand because they are both present in the acrosome. It will be interesting to examine whether SLX interacts with DKKL1 given its cytoplasmic localization. It is possible that DKKL1 interacts with different members of the same family as the acrosome forms in a step-wise manner. Therefore, it is critical to define the subcellular localization of these XLR proteins as well as to identify more interacting partners.

Perhaps the most important question related to its function is whether SLXL1 plays a role in fertilization. Although SLXL1 was suggested to be important for male fertility by one study [26], the simultaneous knockdown of both Slx and Slxl1 gene expression by siRNA made it impossible to evaluate the contribution of SLXL1. In the present study, we used an in vitro fertilization assay to investigate the role of SLXL1 using neutralizing antisera. The
observation that the fertilization rate was significantly reduced by two antisera against SLXL1 (anti-SLXL1 1–155aa and anti-SLXL1 65–155aa) but not by the antiserum against SLX (anti-SLX 95–212aa) indicated convincingly that SLXL1 plays an important role in fertilization. The observations that neither the capacitation nor the acrosome reaction but the binding/penetration of spermatozoa was reduced by the antiserum further narrow down the time window during which SLXL1 executes its function. It is likely that SLXL1 is released from the acrosome and plays a role in the penetration of the spermatozoa into the oocytes, and during this time window the neutralizing antiserum has a chance to block its action. Alternatively, the antiserum may block the sperm-egg binding. If this is the case, it means that proteins released from the acrosome may stabilize the binding of spermatozoa to the eggs.

In conclusion, we have identified SLXL1 as an acrosome-associated member of the XLR family that plays an important role in fertilization probably involved in the penetration of spermatozoon through the zona pellucida or in the sperm-egg interaction. Given the different expression pattern and function of the characterized members of the family such as SYCP3, SLY, SLX, and SLXL1, it seems that XLR proteins may execute diverse functions during spermatogenesis.

Materials and Methods

Animals and Reagents

Adult ICR mice were obtained from the Experiment Animal Center, Chinese Academy of Sciences. New Zealand rabbits were purchased from the Experiment Animal Center, School of Medicine, Peking University. The animals were fed with normal chow and water in accordance with the NIH Guide for the Care and Use of Laboratory Animals. Ethical clearance was granted by the Animal Research Committee guidelines of the Institute of Zoology, Chinese Academy of Sciences (protocol study number 2004–35). Mouse anti-γH2AX (Ser139) was purchased from Millipore’s Temecula, CA. Goat anti-DKKL1 was purchased from R&D systems, Inc. Mouse anti-GFP monoclonal antibody was purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Mouse anti-Flag monoclonal antibody was purchased from Sigma (Sigma-Aldrich St. Louis, MO). Unless specified otherwise, the general reagents were purchased from Sigma and Invitrogen (Carlsbad, CA). Primers used in PCRs were synthesized by Invitrogen and listed in Table 1.

RNA extraction and RT-PCR

Total RNAs from mouse tissues were extracted using Trizol solution following the standard protocol. cDNAs of Slx and Sxl1 were cloned by standard RT-PCR using specific primers and sequencing confirmed.

Recombinant protein expression and polyclonal antibody production

To produce glutathione S-transferase (GST) fusion protein, the mouse Sxl1 cDNA fragments (amino acids 1–155 aa and 65–155 aa) and Sx cDNA fragments (amino acids 95–212 aa) were subcloned into pGEX-FT-1 vector (Pharmacia Biosciences) using gene-specific primers (Table 1). Recombinant proteins were expressed in Escherichia coli strain BL21 and purified using GSTrap_5ml_FF columns (GE Healthcare) according to the manufacturer’s instructions. About 0.5 mg of each recombinant GST fusion protein emulsified in Freund’s complete adjuvant was

Figure 4. Co-immunoprecipitation and co-localization of SLXL1 and DKKL1. (A) Coimmunoprecipitation of DKKL1-FLAG with SLXL1-GFP expressed in 293T cells. (B) Coimmunoprecipitation of DKKL1 with SLXL1 from testis lysates. Protein lysates were analyzed by standard Western blotting with anti-FLAG, anti-GFP, anti-SLXL1 and anti-DKKL1 antisera, respectively. (C) Co-localization of SLXL1 (red) and DKKL1 (green) in seminiferous tubules. The nuclei were stained with DAPI (blue). doi:10.1371/journal.pone.0020866.g004
injected subcutaneously into two New Zealand rabbits. Subsequently the rabbits were injected with 0.25 mg of the recombinant protein Freund’s incomplete adjuvant every 2 weeks for 2 months. Finally, polyclonal antisera were recovered from the rabbit blood by standard methods.

Western blot analysis

Mouse tissues and testes were homogenized in the RIPA lysis buffer (50 mM Tris, pH 7.4, 10 mM MgCl2, 150 mM NaCl, 1% NP-40, 1 mM sodium orthovanadate, 1 mM NaF). The protein concentration of extracts was determined using Bradford Reagent (BioRad) and were diluted in 6×SDS-loading buffer (125 mM Tris-HCl pH 6.8, 2% SDS, 20% glycerol, 0.2% bromophenol blue). Protein extracts from each tissue were run on 12% SDS-PAGE gels. Transfer of proteins in gels to Hybond-C membrane (Amersham) was performed using the Bio-Rad membrane transfer apparatus at 80 V for 2 h, and the membrane was then blocked in PBSA (5% milk powder and 0.1% Tween-20 in PBS) for 1 h at room temperature. First antibodies (anti-SLX 95–212aa diluted 1000 times, anti-SLXL1 1-155aa diluted 1000 times, anti-SLXL1 165–155aa diluted 1000 times and anti-GAPDH (diluted 2000 times) were added and incubation performed at 4°C overnight. The peroxidase-conjugated secondary antibody was used with a 2000-time dilution and signals were detected by chemiluminescence.

Immunohistochemistry

Immunohistochemistry was performed following standard protocols. Briefly, 8 μm frozen sections of mouse testes were fixed immediately in 4% paraformaldehyde for 15 min at room temperature. The sections were washed with phosphate buffered saline (PBS) and blocked for 1 h in 5% bovine serum albumin in PBS. The sections were then incubated with SLXL1 1-155aa antibody (diluted at 1:100), SLXL1 95–212aa antibody (diluted at 1:100), mouse anti-pH2AX antibody (diluted at 1:200), and DAPI (0.5 μg/ml) for 10 min. After washing in PBS, the sections were covered with slide and photographed by confocal microscopy. Rabbit pre-immune serum was used as negative control.

Figure 5. Assessment of the effects of SLXL1 antisera on in vitro fertilization, motility and acrosome reaction of spermatozoa. (A) The inhibitory effect of SLXL1 antisera on in vitro fertilization (IVF) rate. Successful fertilization was indicated by zygote cleavage. Spermatozoa were treated with PBS, preimmune serum, anti-SLXL1 1-155aa, anti-SLXL1 95–212aa, anti-SLXL1 165–155aa and anti-SLX 95–212aa before in vitro fertilization assays were performed. (B) Assessment of effect of anti-SLXL1 1-155aa on the motility of spermatozoa. (C) Assessment of effect of anti-SLXL1 1-155aa on the progressive movement of spermatozoa. (D) Assessment of effect of anti-SLXL1 1-155aa on the acrosome reaction of spermatozoa. **Denotes group that is significantly different (p<0.01) from the control groups.

doi:10.1371/journal.pone.0020866.g005
Co-immunoprecipitation

The coding sequence of mouse Slxl1 cDNA and Dkk1 cDNA were cloned into EGFP-N1 (BD Clontech) and pFlag-CMV-4 (Sigma). The plasmid constructs of SLXL1-GFP and Flag-DKKL1 were co-transfected into HEK293T cells using Lipofectamine 2000 according to the manufacturer’s instructions (Invitrogen). pFlag-CMV-4 empty vector and Slxl1-GFP were transfected into HEK293T cells served as negative controls. Cells were solubilized in the RIPA lysis buffer 48 h after transfection and were incubated with Flag-conjugated sepharose beads overnight at 4°C. Beads and captured protein complexes (SLXL1-GFP and Flag-DKKL1) were washed 5 times. Beads were suspended in 6×SDS loading buffer and were analyzed by standard Western blotting with anti-GFP (1:1000 dilution), anti-FLAG (1:2000 dilution). Total protein lysates of the testis were incubated with anti-SLXL11–155aa antibody or rabbit preimmune serum for 2 h at 4°C, followed by incubation with protein A coated agarose beads overnight at 4°C. The agarose beads and captured protein complexes were washed 5 times in PBS and suspended in 6×SDS sample buffer for immunoblotting with anti-SLXL1 (1–155aa) (1:1000 dilution) and anti-DKKL1 (1:1000 dilution).

Table 1. Primer pairs used in the present study.

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Figure 6. The inhibitory effect of SLXL1 antiserum on in vitro zona pellucida binding/penetration of spermatozoa. (A). Bright (left panel) and fluorescent (right panel) images of eggs and spermatozoa incubated with HTF, control, anti-SLXL11–155aa (antiserum) and anti-SLXL11–155aa plus SLXL1 (Antiserum+SLXL1). (B) Statistical analysis of binding/penetration of spermatozoa to oocytes. The number of bound spermatozoa on each egg was evaluated. *Denotes significantly different groups compared with control group (p<0.05). doi:10.1371/journal.pone.0020866.g006
In vitro fertilization

The female mice were superovulated and the stage MI oocytes were collected from mice oviducts as described [30]. Mouse spermatozoa from cauda epididymis were capacitated in HTF (Human Tubal Fluid) medium within 30 min, the spermatozoa (in drops of 50 µl with concentration of 5 x 10^7/ml) were incubated for 20 min with the following 6 sets of reagents: i) HTF; ii) rabbit preimmune serum; iii) anti-SLXL1 1–155aa; iv) anti-SLXL1 1–155aa+SLXL1; v) anti-SLXL1 65–155aa; vi) anti-SLXL1 65–212aa. The treated spermatozoa (50 µl) was deposited into each medium drop (50 µl) containing 30–35 mouse oocytes and incubated for 2 h. Unbound spermatozoa were washed away. To analyze the IVF rate, two pronuclei cells were examined 6 h after fertilization. The zygote cleavages were counted at 42 h.

Capacitation analysis

Spermatozoa were obtained from the mouse epididymis and washed in PBS. Washed spermatozoa (5 x 10^7/ml) were incubated to capacitate in an HTF medium or HTF medium with 0.3% SLXL1 antisera or 0.3% control preimmune serum by incubation for 2.5 h at 37°C under 5% CO2 as described [31]. To measure the motility of spermatozoa, a CASA system (Version 12 CEROS, Hamilton Thorne Research) was used with the following settings: minimal contrast, 50; minimal cell size, 4 pixels; and 60 frames per second. Samples were evaluated for general and progressive motility.

Acrosome reaction analysis

Capacitated spermatozoa in each sample were incubated in the HTF medium supplemented with progesterone (P) to induce the acrosome reaction [32]. The samples were transferred to microslides, air dried and stained with Coomassie brilliant blue G-250 dye (CBB) in 10 ml of 3.5% (v/v) perchloric acid for 3 min [33]. The slides were rinsed with distilled water, the spermatozoa samples were observed under a phase contrast microscope. Acrosome reaction of the spermatozoa was examined from randomly selected fields of the slides until 400 spermatozoa had been examined.

Zona pellucida binding/penetration assay

Oocytes were collected from ovaries and cumulus cells were removed with 0.01% (w/v) hyaluronidase [34]. About 8–10 oocytes with intact zona pellucida (ZP) were incubated in a medium drop with HTF medium containing 0.3% pre-immune serum or 0.3% anti-SLXL1 1–155aa. Fresh spermatozoa of cauda epididymis were capacitated in HTF medium for 30 min and deposited into each medium drop followed by incubation for another 30 min. Then oocyte-spermatozoon complex were washed thoroughly and fixed with 0.25% glutaraldehyde. In order to observe egg-spermatozoa binding/penetration, the complex were stained by 1 mg/ml Hoechst 33342 for 15 min and observed under a fluorescence microscope. The protocol followed that of a previous study with slight modifications [35].

Statistical analysis

Statistical analysis was carried out using SPSS software (SPSS Statistics 17.0.1). Each experiment was repeated at least three times and differences were considered significant with p<0.05.

Author Contributions

Conceived and designed the experiments: CH. Performed the experiments: X-JZ X-HL. Analyzed the data: CH X-JZ HJC MZ. Contributed reagents/materials/analysis tools: S-YL X-XW. Wrote the paper: CH X-JZ HJC.

References