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A Stem Cell Reporter for Investigating Pluripotency and Self-Renewal in the Rat

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SUMMARY

Embryonic stem cells (ESCs) are pluripotent, immortal, embryo-derived cell lines that can differentiate into all cells of a developing embryo, including the germ cells, and can be used to introduce genetic modifications through the germline (Doetschman et al., 1987; Evans and Kaufman, 1981; Hooper et al., 1987; Martin, 1981; Thomas and Capecchi, 1987). Although mouse ESCs have been available for more than 30 years, the application of this technology to other experimental animals has proved challenging. Nonetheless, recent advances in understanding the control of self-renewal in ESCs and the development of rationally designed culture systems enabled the first derivation of authentic rat ESCs (Buehr et al., 2008; Li et al., 2008; Ying et al., 2008). Rat ESCs are derived efficiently de novo from blastocysts using this 2i+LIF culture medium, but rESC lines are typically less stable than their mouse counterparts under conditions of clonal expansion and continuous culture (Blair et al., 2012; Meek et al., 2010). This instability can be mitigated to some extent by titrating the level of GSK3 inhibition, to limit the prodifferentiative actions of β-catenin in association with the transcription factor LEF1, which is highly expressed in rESCs (Chen et al., 2013; Meek et al., 2013). Understanding the molecular basis of the different responses of these two demonstrably pluripotent ESCs (that efficiently colonize embryos to generate chimeric animals) affords valuable insights into how signaling and intrinsic mechanisms combine to control pluripotency and differentiation in early embryonic development.

Standard mouse ESC culture conditions containing serum and leukemia inhibitory factor (LIF) do not support rESC culture (Buehr et al., 2003, 2008). However, a two-inhibitor (2i) culture system developed for mouse ESCs, which uses small molecule inhibitors to suppress mitogen-activated protein kinase kinase (MEK1/2) and glycogen synthase kinase 3 (GSK3) activity in conjunction with the cytokine LIF, allows the continuous proliferation of rESC culture (Buehr et al., 2008; Li et al., 2008; Ying et al., 2008). Rat ESCs are derived efficiently de novo from blastocysts using this 2i+LIF culture medium, but rESC lines are typically less stable than their mouse counterparts under conditions of clonal expansion and continuous culture (Blair et al., 2012; Meek et al., 2010). This instability can be mitigated to some extent by titrating the level of GSK3 inhibition, to limit the prodifferentiative actions of β-catenin in association with the transcription factor LEF1, which is highly expressed in rESCs (Chen et al., 2013; Meek et al., 2013). Understanding the molecular basis of the different responses of these two demonstrably pluripotent ESCs (that efficiently colonize embryos to generate chimeric animals) affords valuable insights into how signaling and intrinsic mechanisms combine to control pluripotency and differentiation in early embryonic development.

Fluorescent stem cell reporter genes provide accurate and sensitive feedback on the state of the cells in live cultures, and are useful and important tools for studying the behavior of stem cells and their derivatives. A valuable ESC reporter gene in this regard is the ESC-associated transcription factor REX1/ZFP42, which is highly expressed in the naïve ESCs, the cell type captured in vivo in 2i+LIF cultures that most closely represents pluripotent stem cells in the preimplantation blastocyst embryo (Boroviak et al.,...
The REX1 zinc finger protein arose through duplication of the YY1 transcription factor gene during radiation of eutherian mammals and is most highly expressed in the preimplantation embryo, within a specific region of the placenta, and in the testis (Kim et al., 2007; Rogers et al., 1991). It is reported to regulate X chromosome activity through induction of the antisense RNA Tsix that represses Xist expression (Navarro et al., 2010). REX1 may also function as an epigenetic regulator through association with Polycomb, and as a repressor of endogenous retroviruses or visceral endoderm-associated genes (Garciá-Tunión et al., 2011; Guallar et al., 2012; Kim et al., 2011; Masui et al., 2008). Although there are indications that loss of REX1 may affect embryonic development and reduce fertility in aged mice, REX1-deficient mice are generally viable and healthy (Kalkan et al., 2017; Masui et al., 2008; Rezende et al., 2011). Indeed, in mouse ESCs the protein is dispensable for pluripotency and the Rex1-EGFP knockin transgene is used as a sensitive reporter of pluripotency in vitro and as a tool to assess stem cell potential in vivo (Bhatia et al., 2013; Boroviak et al., 2014; Kalkan et al., 2017; Toyooka et al., 2008; Wray et al., 2011).

In this study we report the generation of a Rex1-EGFP knockin allele in the rat and examine expression of the Rex1-EGFP reporter during preimplantation embryo development and ESC derivation. We evaluate the general requirement for REX1 in vitro and in vivo, and characterize the utility of the Rex1-EGFP reporter for studying self-renewal and differentiation status of rESC cultures.

RESULTS

Generation of a Rex1-EGFP Reporter rESC and Rat

To assess the pluripotent state of rat ESCs accurately in live cultures we used conventional homologous recombination to insert a promoterless EGFP-IRESngeo selection cassette immediately downstream of the Rex1 (Zfp42) promoter, replacing the entire coding region of the Rex1 gene (Figure 1). Germline competent Dark Agouti

![Figure 1. Generating a Rex1-EGFP Reporter in the Rat](image-url)
(DAK31) male rESCs (Blair et al., 2012) were electroporated with the linearized targeting vector, allowed to recover for 48 h, and then subjected to selection with the antibiotic G418 for a further 7 days. Ten G418-resistant ESC clones were expanded and all were shown by Southern blot analysis to carry the EGFP-IREs-neomycin cassette inserted within the Rex1 gene (Figure 1B). Targeted clones displayed the typical rESC colony morphology and exhibited EGFP fluorescence as identified by fluorescence microscopy and flow cytometry (Figures 1C and 1D). qRT-PCR confirmed that Rex1 mRNA levels were reduced by approximately 50% in the targeted Rex1 heterozygous cells relative to wild-type parental cells (Figure S1).

The Rex1-EGFP clone E3 retained a normal karyotype and normal in vitro differentiation capacity. We also tested the developmental capacity of the E3 clone by assessing its ability to contribute to rat chimaeras following blastocyst injection. Clone E3 generated coat color chimaeras at a frequency of 41%, which was comparable with the 34% frequency obtained previously with the unmodified parental cell line, DAK31 (Table S1) (Meek et al., 2013). Seven male chimaeras were bred to test for ESC germline contribution, and two chimaeras fathered pups that demonstrated transmission of both coat-color and the Rex1-EGFP allele (Table S1 and Figure 1E).

To characterize the expression pattern of the Rex1-EGFP reporter in vivo, timed matings were used to generate Rex1-EGFP transgenic embryos. EGFP fluorescence was not detected in 1-, 2- and 4-cell embryos, but was evident in 8-cell (day E3.5) and blastocyst (day E4.5) stage embryos (Figure 2A). Interestingly, strong EGFP fluorescence was observed in both the inner cell mass (ICM) and the surrounding trophectoderm cells of the blastocyst, and persisted in the trophoblast cells after overnight culture, suggesting that Rex1-EGFP expression was probably not due to perdurance of EGFP protein from the earlier embryonic stages (Figure 2A). To assess the activity of the Rex1-EGFP reporter during ESC derivation, we plated heterozygous Rex1-EGFP E4.5 blastocysts into individual wells and monitored EGFP fluorescence during explant outgrowth for 7 days (Figure 2B). Rex1-dependent EGFP fluorescence was maintained throughout the expanding epiblast outgrowth. By contrast, EGFP fluorescence was gradually lost from the trophoblast cells and the surrounding flattened differentiated cells. After 7 days, the ICM outgrowths were dissociated and replated to produce heterozygous Rex1-EGFP ESC lines that were generated at a frequency comparable with wild-type ESC lines (Figure S2).

To assess the functional requirement for Rex1 in rats we genotyped the offspring from multiple crosses between Rex1-EGFP heterozygous rats. Litters displayed a near-normal Mendelian ratio of wild-type (20%), Rex1-EGFP heterozygous (60%), and Rex1-EGFP homozygous (20%) pups (Table S2). Moreover, the behavior and general health of the Rex1-EGFP homozygous rats was indistinguishable from wild-type or heterozygous littermates. Previous reports have suggested that REX1-deficient male mice are sub-fertile (Rezende et al., 2011). We therefore compared the litter sizes of Rex1-EGFP homozygous males with age-matched wild-type control males (9–12 months old) but could not detect any obvious difference in average litter numbers at E4.5 or mid-gestation between homozygous and wild-type rats, suggesting that the fertility of Rex1 mutant male rats at the ages we tested was normal (Table S3).

We next investigated how disruption of Rex1 affected the derivation and maintenance of rESCs in vitro. The culture of E4.5 embryos generated by cross-breeding heterozygous Rex1-EGFP rats showed that the derivation efficiency of Rex1-EGFP homozygous ESCs under 2i+LIF conditions (83%) was close to that obtained for wild-type (100%) and heterozygous (91%) lines (Figure S2). Furthermore, Rex1-EGFP homozygous ESC colonies were morphologically indistinguishable from those of Rex1-EGFP heterozygous or wild-type cells (Figure 2C). Notably, gene expression analysis by qRT-PCR confirmed the absence of Rex1 expression in homozygous lines, but most significantly also demonstrated the absence of Tsix expression, a downstream target of REX1 (Navarro et al., 2010), thus confirming the loss of Rex1 function (Figure 2D). Importantly, we did not detect consistent differences between clones in the expression of other key ESC genes (Figure 2E). In summary, we were unable to identify any consistent identifiable phenotype associated with the disruption of Rex1 in the rat or ESCs. These findings indicate that the rat Rex1 gene function is largely redundant in the rat and ESCs under standard conditions, and as a result provides a convenient neutral locus suitable for harboring a fluorescent reporter for monitoring stem cell pluripotency and self-renewal.

**Rex1-EGFP Reporter Expression Discriminates between Pluripotent and Differentiated ESC States**

To evaluate the specificity of the rat Rex1-EGFP reporter, we monitored the correspondence between expression of Rex1 mRNA and Rex1-EGFP mRNA under culture conditions that promote ESC differentiation. Withdrawal of LIF and 2i inhibitors from rESCs in suspension cultures to promote embryoid body differentiation reduced the level of EGFP expression as assessed by fluorescence microscopy (Figure 3A). qRT-PCR confirmed coincident downregulation of EGFP and endogenous Rex1 mRNAs, as well as reduced expression of the pluripotency-associated gene Nanog in embryoid bodies (Figure 3B). Similarly, a reduction in the concentration of MEK inhibitor applied to the monolayer
cultures produced a dose-dependent decline in EGFP-positive cells within the culture that was accompanied by a coordinate reduction in \(\text{Rex1}\) and \(\text{Rex1-EGFP}\) mRNA expression, and increased expression of an early differentiation marker Gata4 (Figures 3C and 3D).

High levels of β-catenin (CTNNB1) activity in association with the transcription factor LEF1 drives rat ESC differentiation, causing a loss of ESC colony morphology, induction of mesendoderm-associated differentiation markers Brachyury and Cdx2, and increased instability and collapse of rESC cultures (Chen et al., 2013; Meek et al., 2013). We therefore examined the response of the \(\text{Rex1-EGFP}\) reporter to manipulation of the β-catenin/LEF1 signaling pathway in rSCs. Fluorescence microscopy and flow cytometry confirmed that induction of β-catenin activity using concentrations of the GSK inhibitor CHIR99021 (CH) >3 μM efficiently suppressed \(\text{Rex1-EGFP}\)-dependent fluorescence (Figures 4A and 4B) and induced ESC differentiation (Figure 4C). Depletion of β-catenin (Ctnnb1) or LEF1 in these culture conditions, by either small interfering RNA (siRNA)-mediated knockdown (Figures S3A and S3B) or TALEN-mediated gene editing, maintained \(\text{Rex1-EGFP}\)

Figure 2. In Vivo and In Vitro Rex1-EGFP expression pattern

(A and B) Compound brightfield and fluorescent images of \(\text{Rex1-EGFP}\) expression (A) in 2-cell, 4-cell, 8-cell, and blastocyst-stage rat embryos (in sequence from left to right), and (B) during ESC derivation in a day 0 (d0), d2, d4, and d7 outgrowth from an E4.5-derived rat blastocyst (arrows highlight \(\text{Rex1-EGFP}\)-negative extraembryonic cells).

(C) Compound brightfield and fluorescent image of \(\text{Rex1-EGFP KO}\) rat ESCs (magnification ×100).

(D) qRT-PCR analysis for \(\text{Rex1}\) and \(\text{Tsix}\) expression in wild-type (WT) and knockout (KO) rat ESC (mean ± sd of three biological replicates).

(E) qRT-PCR analysis of the core pluripotency transcription factors \(\text{Oct4}\), \(\text{Nanog}\) and \(\text{Sox2}\) in wild-type (WT) and knockout (KO) rat ESC (mean ± sd of three biological replicates).
expression and ESC self-renewal (Figures 4C and 4D). Sequence analysis of β-catenin and LEF1 TALEN-transfected clones confirmed the presence of inactivating mutations in the Ctnnb1 and Lef1 alleles, respectively (Figure S3C). Transfection with control siRNAs or control TALENs, in contrast, did not prevent CH-induced rESC differentiation or downregulation of Rex1-EGFP reporter expression.

To determine if the Rex1-reporter could also be reactivated upon dedifferentiation into a pluripotent stem cell state through induced pluripotent stem cell (iPSC) reprogramming, we transfected rat embryonic fibroblasts (REFs) derived from a Rex1-EGFP heterozygous embryo with a piggyBac vector co-expressing human cDNAs encoding OCT4, SOX2, KLF4, and cMYC (Gao et al., 2019). The starting REF cultures did not express EGFP but 7–10 days after transfection with the piggyBac construct, compact colonies typical of undifferentiated rESCs emerged within the cultures that had reactivated the Rex1-EGFP reporter and expression of the endogenous rESC transcription factor NANOG (Figures 4E and S3D). This confirmed that the Rex1-EGFP reporter can be used to monitor iPSC reprogramming and together with rESC differentiation data indicates that the rat Rex1-EGFP allele provides a useful marker of undifferentiated pluripotent rat stem cells and experimentally manipulated derivatives.

Figure 3. Downregulation of Rex1-EGFP Reporter Expression during In Vitro Differentiation

(A) Compound brightfield and fluorescent images of Rex1-EGFP expression during embryoid body formation at d0 (left panel), d2 (middle panel), and d4 (right panel).

(B) qRT-PCR analysis of EGFP, Rex1, and Nanog expression in Rex1-EGFP rat ESCs harvested at d0, d2, and d4 during embryoid body differentiation (mean ± SD of three technical replicates).

(C) Flow cytometry analysis of Rex1-EGFP rat ESCs cultured for four days in 3 μM CHIR99021+LIF containing 1 μM PD0325901 (top panel), 0.25 μM PD (middle panel), and without PD (bottom panel).

(D) qRT-PCR analysis of EGFP, Rex1, and Gata4 in Rex1-EGFP rat ESCs cultured for four days in 3 μM CHIR99021+LIF containing 1 μM PD, 0.25 μM PD, and without PD (mean ± SD of three biological replicates).

Rex1-EGFP Reporter Expression Reveals Functional Heterogeneity in Rat ESC Cultures Maintained in 2i+LIF Conditions

Rat ESCs propagated in 2i+LIF culture medium exhibit background levels of spontaneous differentiation (Blair et al., 2012). To assess how the Rex1-EGFP reporter could be used to monitor the pluripotent status of cells within rESC cultures, we analyzed expression of the reporter in rat ESCs grown under standard 2i+LIF growth conditions with feeder support. Under these circumstances the basal level of spontaneous differentiation is most evident as morphologically differentiated cells located at the periphery of some ESC colonies. Immunocytochemistry revealed that many of these cells expressed the primitive endoderm-associated transcription factor GATA4 (Figure 5A). Expression of the Rex1-EGFP reporter showed the opposite pattern, being restricted to undifferentiated cells and excluded from the peripheral differentiated cells (Figure 5A). GATA4+/Rex1-EGFP+ cells were also evident within...
Figure 4. Response of Rex1-EGFP Reporter to CHIR-Induced Differentiation and Rescue by Targeting β-Catenin and LEF1

(A) Flow cytometry analysis of Rex1-EGFP rat ESCs cultured for five days in 2i medium containing 3 μM, 4 μM, or 6 μM CHIR99021.

(B) Flow cytometry analysis of Rex1-EGFP rat ESCs cultured for 3, 4, or 5 days in 2i medium containing 6 μM CHIR99021, and 5 days in 2iL.

(C) Brightfield and fluorescent images of Rex1-EGFP rat ESCs cultured for four days in 2iL self-renewal (3 μM CHIR99021) and differentiating (6 μM CHIR99021) rat ESC culture conditions, following transfection with control (-ve), Ctnnb1, or Lef1-specific TALENs (magnification ×100).

(D) Flow cytometry analysis of Rex1-EGFP rat ESCs cultured for four days in conditions described in (C).

(E) Brightfield image of Rex1-EGFP REFs, and brightfield, fluorescent (EGFP), and immunostained (Nanog) images of a Rex1-EGFP rat iPSC colony (magnification ×100).
some of the otherwise morphologically undifferentiated ESC colonies, implying that differentiation could also occur within the interior of the colonies.

We performed flow cytometry on the bulk cultures to quantify Rex1-EGFP expression in standard rESC 2i+LIF cultures and found that the ESCs could be assigned to three populations: cells expressing high levels of EGFP (Rex1-EGFP<sup>high</sup>), which typically represented the majority (>90%) of cells in the culture; cells expressing an intermediate or low level of EGFP (Rex1-EGFP<sup>low</sup>), which constituted a variable and less distinct population (1–10%); and cells that did not express EGFP (Rex1-EGFP<sup>neg</sup>) and contributed to 1–3% of the overall population (Figure 5B). qRT-PCR of the flow-sorted populations showed that Rex1-EGFP<sup>neg</sup> mRNA expression was downregulated in the Rex1-EGFP<sup>low</sup> cells, and almost absent from the Rex1-EGFP<sup>neg</sup> population (Figure 5C). By contrast, expression of the differentiation marker Gata4 was absent from the Rex1-EGFP<sup>high</sup> population, but was upregulated stepwise in Rex1-EGFP<sup>low</sup> and Rex1-EGFP<sup>neg</sup> cells, confirming the progressively increasingly differentiated status of cells within these populations. Expression of endogenous Rex1 mRNA was downregulated in Rex1-EGFP<sup>neg</sup> cells but unexpectedly was expressed at high levels in the Rex1-EGFP<sup>low</sup> population. This suggested that Rex1-EGFP<sup>low</sup> cells represented an intermediate state between the undifferentiated Rex1-EGFP<sup>high</sup> and differentiated Rex1-EGFP<sup>neg</sup> cells, characterized by asynchronous downregulation of the Rex1 alleles. This asynchronous pattern of downregulation of targeted and non-targeted Rex1 alleles was confirmed in three additional, independently targeted, Rex1-EGFP ESC clones (Figure S4).

We next examined the stem cell self-renewal potential of cells within the Rex1-EGFP sub-populations, by purifying Rex1-EGFP<sup>low</sup> and Rex1-EGFP<sup>high</sup> cells by fluorescence-activated cell sorting (FACS), replating the cells in 2i+LIF ESC self-renewal conditions, and monitoring the growth of the resulting cultures for 4 days (Figure 5D). Most Rex1-EGFP<sup>high</sup> cells formed EGFP-positive, compact, spherical colonies typical of undifferentiated rESC colonies. By
contrast, \(Rex1\)-EGFP\textsuperscript{low} cells formed more irregularly shaped and flattened colonies containing cells expressing variable levels of EGFP. Many of these colonies were also surrounded by flat, morphologically differentiated cells (Figure 5D). To stringently assess the self-renewal potential of individual rat ESCs within the EGFP sub-populations, we sorted cells according to their levels of EGFP expression (high, low, and negative), and assessed the colony-forming potential of the cells in a single-cell cloning assay. Individual cells were deposited into 96-well plates and after 7 days of culture the resulting colonies were scored for EGFP fluorescence and morphology (Figures 5E and 5F). \(Rex1\)-EGFP\textsuperscript{high} cells exhibited the highest colony-forming potential and generated undifferentiated EGFP\textsuperscript{high} colonies at the highest frequency (42%). A significant proportion of the EGFP\textsuperscript{high} cells (55%) also produced colonies that expressed uniformly low levels of EGFP. \(Rex1\)-EGFP\textsuperscript{low} sorted cells gave rise to fully differentiated \(Rex1\)-EGFP\textsuperscript{neg} colonies (31%), but also produced morphologically undifferentiated colonies that maintained a low level of EGFP expression (65%), indicating that the \(Rex1\)-EGFP\textsuperscript{low} state could be maintained through multiple cell divisions. By comparison, \(Rex1\)-EGFP\textsuperscript{neg} cells exhibited reduced cloning efficiency, gave rise to some \(Rex1\)-EGFP\textsuperscript{low} colonies (22%) and produced the greatest proportion of morphologically differentiated \(Rex1\)-EGFP-negative colonies (67%). The clonal analysis demonstrated that expression of the \(Rex1\)-EGFP reporter could discriminate between different rat ESC states. It also identified a loss of pluripotent potential among the \(Rex1\)-EGFP\textsuperscript{low} population and indicated that this state could persist for a period of time under standard 2i+LIF culture conditions. To assess how this loss of potential affected developmental potency in vivo, we purified \(Rex1\)-EGFP\textsuperscript{low} cells by FACS, allowed them to recover in culture for 4 days, and tested their contribution to embryonic development by blastocyst injection. The cells contributed to live-born chimaeras with an efficiency close to the parental ESCs (6 chimaeras out of 26 pups: 23%), demonstrating that the \(Rex1\)-EGFP\textsuperscript{low} cells either fully retained the capacity to functionally reintegrate into the developing embryo or were in a state where this potential was readily reinstalled within the blastocyst environment.

To examine the gene expression profile of the \(Rex1\)-EGFP\textsuperscript{low} intermediate cell type, we assessed the transcriptional activity of cells within the three populations of \(Rex1\)-EGFP cells in 2i+LIF culture by single-cell RNA sequencing. We sorted cells into replicate 96-well plates according to their levels of EGFP expression and followed the Smart-seq2 protocol, with minor modifications as we have described previously (Kirschner et al., 2017; Picelli et al., 2014) (Figure 6A). Consistent with the sorting criteria, the average level of EGFP RNA decreased in the populations in the order EGFP\textsuperscript{high}>EGFP\textsuperscript{low}>EGFP\textsuperscript{neg} (Figure 6B).

Conversely, expression of the differentiation markers Gata4 and Sox17 was upregulated in the EGFP\textsuperscript{neg} population. Expression of the ESC pluripotency-associated factors Nanog and Klf2 was also downregulated in the \(Rex1\)-EGFP\textsuperscript{neg} cells (Figure 6B). Most ESC factors, however, were expressed at comparable levels in the \(Rex1\)-EGFP\textsuperscript{high} and \(Rex1\)-EGFP\textsuperscript{low} cells, indicating that \(Rex1\)-EGFP\textsuperscript{low} cells might represent a very early step in the process of exiting pluripotency. Indeed, \(Rex1\) mRNA expression from the non-targeted \(Rex1\) allele in the \(Rex1\)-EGFP\textsuperscript{low} cells was at a similar or higher level than in \(Rex1\)-EGFP\textsuperscript{high} cells, even though \(Rex1\) mRNA was clearly downregulated in the differentiating \(Rex1\)-EGFP\textsuperscript{low} population (Figure 6B). This differential downregulation of wild-type \(Rex1\) mRNA and \(Rex1\)-EGFP mRNA in the \(Rex1\)-EGFP\textsuperscript{low} cells is consistent with bulk analysis of the rESC population (Figures 5C and 5D) and supports the notion that asynchronous downregulation of the \(Rex1\) alleles identifies an intermediary rESC state. The direct comparison of EGFP mRNA and \(Rex1\) mRNA expression in individual cells from all three populations showed that within many \(Rex1\)-EGFP\textsuperscript{high} (\(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{+}}) and \(Rex1\)-EGFP\textsuperscript{neg} (\(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{−}}) cells, both \(Rex1\) alleles were active or inactive, respectively (Figure 6C). By contrast, the majority of the cells within the \(Rex1\)-EGFP\textsuperscript{low} population expressed high levels of \(Rex1\) mRNA (\(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{−}}). To assess the differentiation potential of \(Rex1\)-EGFP\textsuperscript{low} cells in vitro, we purified the cells by FACS, expanded the population for 4 days after plating, and then analyzed the cells by flow cytometry and single-cell sequencing. In line with previous results, a significant proportion of the cells retained a \(Rex1\)-EGFP\textsuperscript{low} state (Figure 6D). In addition, expression of \(Rex1\) mRNA was downregulated in many of the remaining cells and the overall transcriptional profile of the cells shifted away from the pre-plated \(Rex1\)-EGFP\textsuperscript{low} and \(Rex1\)-EGFP\textsuperscript{high} cells, consistent with many of the cells having differentiated (Figures 6E and 6F).

Collectively these results suggest that the \(Rex1\)-EGFP reporter reveals a background level of differentiation in rESC 2i+LIF cultures that proceeds in the following order: from \(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{+}} \(\rightarrow\) \(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{−}} \(\rightarrow\) \(Rex1\)\textsuperscript{mRNA−/EGFP\textsuperscript{−}}. To investigate this possibility further we specifically compared the transcription profiles of stem cell pluripotency regulator genes in cells within these three categories (Figures 6G and 6H). This analysis revealed a trend of downregulation of pluripotency-associated factors in the \(Rex1\)\textsuperscript{mRNA−/EGFP\textsuperscript{−}} cells compared with \(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{+}} cells, some of which, such as Tbx3, Klf2, Klf4, and Klf5, are commonly associated with naive pluripotency (Figures 6I and S5). Gene list enrichment analysis (ToppGene Suite) showed enrichment of genes involved in chromosome organization among those differentially regulated between \(Rex1\)\textsuperscript{mRNA+/EGFP\textsuperscript{+}} and \(Rex1\)\textsuperscript{mRNA−/EGFP\textsuperscript{−}} cells (Figure S6).
In summary, these results show that the Rex1-EGFP reporter can be used to distinguish between pluripotent rat ESC states and their differentiated derivatives, and provides a useful tool for exploring early stages during establishment and dissolution of ESC pluripotency.

**DISCUSSION**

Rex1 (Zfp42) gene expression is a benchmark marker of pluripotency in cultures of ESCs and reprogrammed iPSCs (Bao et al., 2011; Bhatia et al., 2013; Kalkan et al., 2017; Rodríguez et al., 2012). Here we describe the characterization of a Rex1-EGFP knockin transgene in the rat, and show that it provides a sensitive reporter of the pluripotent state in rESCs. Rat ESCs differ somewhat from mouse ESC in their growth factor requirements and growth characteristics, thereby providing a unique alternative to mouse cells with which to investigate self-renewal signaling and pluripotency (Chen et al., 2013; Li et al., 2008; Meek et al., 2013). Based on the work presented here we propose that the Rex1-EGFP reporter provides a useful tool to interrogate...
the early stages of ESC differentiation in the rat, and enrich our understanding of the regulation of pluripotency in mammals.

Replacement of the *Rex1* open reading frame with the EGFP cassette eliminated *Rex1* expression and function. Nonetheless, disruption of *Rex1* expression did not adversely affect either the derivation or growth of rESCs, or apparently the health of *Rex1* knockout rats. This is in line with previous reports showing that *Rex1* function was dispensable for maintaining pluripotency in mouse ESCs, and not essential in mice (Masui et al., 2008). However, in contrast to reports in mice, we did not observe signs of selective loss of *Rex1* homozygous mutant embryos or a loss of fertility in male knockout rats (Kalkan et al., 2017; Rezende et al., 2011), nor observe a bias toward endoderm differentiation previously described for mouse *Rex1* knockout ESCs (Masui et al., 2008). Nonetheless, deletion of the *Rex1* coding region in rat ESCs did eliminate expression of *Tsix*, a non-coding RNA that suppresses X chromosome inactivation and a reported target of *Rex1* in mouse (Navarro et al., 2010). This result confirmed that the *Rex1*-EGFP knockin created a non-functional allele and pointed to a conserved role of *Rex1* in epigenetic regulation of X chromosome activity in rat and mouse. Taken together our results indicate that *Rex1* function is largely dispensable, and implies that the *Rex1*-EGFP knockin allele provides a neutral reporter that does not noticeably interfere with ESC self-renewal or normal embryonic development in the rat.

Embryonic expression of the *Rex1*-EGFP reporter was first detected at the 8-cell stage and thereafter persisted through compaction to the formation of the blastocyst. *Rex1*-EGFP was detected within the external trophoblast cells of the blastocyst as well as the ICM, implying that early expression *in vivo* in the rat embryo is associated with an early undifferentiated embryonic state, rather than exclusively as a marker of pluripotency. Indeed, *Rex1* expression has been detected in trophoblast cell types in the mouse, including cells within the ectoplacental cone, extraembryonic ectoderm, and the placenta of post-implantation embryos (Kalkan et al., 2017; Kim et al., 2011; Rogers et al., 1991). In the mouse blastocyst, however, *Rex1* or *Rex1*-EGFP mRNA expression was restricted to the ICM, implying that the level or onset of *Rex1* expression in the trophoblast lineage differs between the mouse and the rat embryos (Kalkan et al., 2017; Pelton et al., 2002). Nevertheless, when rat blastocyst were cultured in 2i+LIF ESC culture conditions, *Rex1*-EGFP expression became rapidly confined to the expanding pluripotent stem cell compartment, consistent with *Rex1*-EGFP expression marking the undifferentiated ESCs. This restricted pattern of *Rex1*-EGFP expression might be further sharpened by failure of trophoblast cells to thrive in the 2i culture conditions (Ying et al., 2008). Withdrawal of self-renewal conditions or active stimulation of differentiation, by contrast, resulted in downregulation of the *Rex1*-EGFP reporter along with the unmodified wild-type *Rex1* allele and other markers of ESC pluripotency.

Even under standard 2i+LIF culture conditions, the activity of the *Rex1*-EGFP reporter gene could discriminate between undifferentiated ESCs and cells spontaneously escaping pluripotency and embarking on differentiation. An early transition state was identified by a low-level, semi-stable expression of the *Rex1*-EGFP reporter and was functionally characterized by an increased tendency to differentiate under clonal growth conditions. Surprisingly, expression of the untargeted *Rex1* allele in these transitional cells was as high as in rat ESCs expressing high levels of *Rex1*-EGFP (*Rex1*-EGFP*high*). This pointed to selective loss of transcriptional activity from the targeted *Rex1*-EGFP allele and might arise from a loss of positive regulatory elements from the targeted *Rex1*-EGFP allele or interference induced by insertion of the targeting cassette, thus sensitizing expression of the EGFP knockin allele to a weakening of control of pluripotency. An alternative possibility, however, is that asynchronous expression of *Rex1* alleles might occur normally in this peri-naive ESC transition state. Indeed, the detection of *Rex1*-EGFP*low* cells expressing low levels of endogenous *Rex1* mRNA (*Rex1*mrNA*/EGFP*), thus mirroring the *Rex1*-EGFP*low* (*Rex1*mrNA*/EGFP*) population, supports the notion that the EGFP transgene might simply reflect asynchrony in the response of *Rex1* alleles to relaxation of the control of pluripotency in the earliest stages of differentiation. Indeed, the pluripotency factor *Nanog* has been reported to exhibit mono-allelic expression prior to attaining full biallelic expression in the ICM of the blastocyst (Miyanari and Torres-Padilla, 2012).

Notwithstanding the asynchronous downregulation of the *Rex1* alleles, the reduction in EGFP expression in *Rex1*-EGFP*low* cells potentially identified an intermediary rESC state at a very early stage in differentiation. This notion was supported by the finding that cells expressing low levels of *Rex1*-EGFP mRNA while retaining high levels of *Rex1* mRNA (*Rex1*mrNA*/EGFP*−) also tended to express lower levels of naive pluripotency regulators (Nichols and Smith, 2009) compared with the *Rex1*-EGFP*high* (*Rex1*mrNA*/EGFP*+) cells. Pathway analysis of genes differentially regulated between these two populations also pointed to the involvement of chromosome reorganization during this transition. Interestingly, we did not observe significant levels of reversion of the *Rex1*-EGFP*low* cells to the *Rex1*-EGFP*high* state in single-cell clonal assays, in line with the results reported for mouse *Rex1*-EGFP*low* ESCs cultured in 2i+LIF conditions (Kalkan et al., 2017). This contrasts with the results reported for a *Rex1*-EGFP reporter in mouse ESCs propagated in serum+LIF medium.
(Toyooka et al., 2008) and suggests that the stringency associated with 2i+LIF clonal culture conditions provides an effective barrier for the reversion of Rex1-EGFP<sup>low</sup> cells to a Rex1-EGFP<sup>high</sup> state. In conjunction with the persistence of the intermediate state in rat ESCs, this might potentially provide a continuous source of instability in rat ESC cultures. Nonetheless, given that the progeny of Rex1-EGFP<sup>low</sup> cells did colonize blastocysts and contribute to chimeric rats, it remains possible that, under the appropriate modified culture conditions, Rex1-EGFP<sup>low</sup> cells might be induced to revert and be stably retained within a Rex1-EGFP<sup>high</sup> state.

In conclusion, we show here that the rat Rex1-EGFP fluorescent reporter allows the qualitative and quantitative analysis of ESC states in rat ESC cultures in vitro, and provides a useful and sensitive tool to scrutinize early stages of pluripotent stem cell differentiation in the rat, as well as contributing to more general insights into the regulation of stem cell potency and fate determination in mammals.

**EXPERIMENTAL PROCEDURES**

**ESC Culture**
ESC were derived from E4.5 rat blastocysts following removal of the zona pellucida with acidic Tyrode solution. The intact blastocyst was then cultured in the well of a 96-well plate, in 2iL (N2B27 medium, 1 μM PD0325901 [PD], 3 μM CHIR99021 [CH], 1000 U/mL mouse LIF) on γ-irradiated (5 Gy) OF1 mouse fibroblasts for 7 days prior to passaging. ESC were maintained on irradiated OF1 or DR4 mouse fibroblasts for 7 days prior to passaging. ESCs were derived from E4.5 rat blastocysts following removal of the zona pellucida with acidic Tyrode solution. The intact blastocyst was then cultured in the well of a 96-well plate, in 2iL (N2B27 medium, 1 μM PD0325901 [PD], 3 μM CHIR99021 [CH], 1000 U/mL mouse LIF) on γ-irradiated (5 Gy) OF1 mouse fibroblasts for 7 days prior to passaging. ESC were maintained on irradiated OF1 or DR4 mouse fibroblasts in 2iL. Colonies were passaged every 2–3 days using TVP (0.025% trypsin, 1% chicken serum, and 1 mM EDTA) and plated at a density of (0.5–1) × 10<sup>5</sup> cm<sup>-2</sup>. MEK (PD) and GSK3 (CH) inhibitors were supplied by Axon Medchem (http://www.axonmedchem.com).

**Gene Targeting by Homologous Recombination**
A targeting vector was constructed using BAC recombineering which consisted of approximately 3 kb and 2.7 kb, 5' and 3' homology arms respectively, in which the entire Rex1 coding sequence was replaced with a Kozak-EGFP-loxp-IRESneo-loxp-bGHpA reporter/selection cassette. Approximately 1 × 10<sup>7</sup> DAK31 rat ESCs (Blair et al., 2012) in 0.6 mL of PBS containing 100 μg of linearized Rex1-EGFP targeting vector were electroporated using a Bio-Rad Gene Pulser apparatus (0.8 kV, 3 μF). Electroporated cells were plated into 10 cm<sup>2</sup> wells containing 2iL medium. The aminoglycoside G418 (80 μg/mL) was added 48 h after electroporation and G418-resistant colonies picked 9 days later onto DR4 fibroblasts for 7 days prior to passaging. ESCs were maintained on irradiated G418-resistant colonies picked 9 days later onto DR4 fibroblasts for 7 days prior to passaging. ESCs were maintained on irradiated OF1 or DR4 mouse fibroblasts in 2iL. Colonies were passaged every 2–3 days using TVP (0.025% trypsin, 1% chicken serum, and 1 mM EDTA) and plated at a density of (0.5–1) × 10<sup>5</sup> cm<sup>-2</sup>. MEK (PD) and GSK3 (CH) inhibitors were supplied by Axon Medchem (http://www.axonmedchem.com).

**ESC Differentiation**
For embryoid body differentiation, a single-cell suspension of rat ESCs was plated into non-coated, low-adherence plastic wells of a 6-well plate at a density of 2 × 10<sup>5</sup>/cm<sup>2</sup> in 2iL. The cells were cultured for 2 days then transferred into fibroblast medium (Glasgow minimum essential medium [GMEM], 10% fetal calf serum [FCS]) for a further 2 days. Rat ESCs differentiated by culturing at a density of 5 × 10<sup>5</sup>/cm<sup>2</sup> in either 2iL with concentrations of PD lower than 1 μM for 3–4 days or in 2i (no LIF) with concentrations of CH >3 μM for 3–4 days.

**Reprogramming of REPs**
Approximately 5 × 10<sup>5</sup> Rex1-EGFP REPs were suspended in Resuspension Buffer R (Invitrogen) and transfected with 1.5 μg of pB<sub>B</sub> TRE-hOSMK + 0.5 μg of PBbase + 0.5 μg of PB-RTTA (Gao et al., 2019), using the Neon transfection system (Invitrogen) set at 1350 V, 30 ms, 1 pulse. Transfected cells were plated in a 6-well plate at a density of 2.4 × 10<sup>5</sup>/cm<sup>2</sup> on γ-irradiated (100 Gy) STO mouse fibroblasts in M15G+S+ medium (GMEM base media + 15% fetal bovine serum + 1000 U/mL human LIF + 50 μg/mL Vc + 1 μg/mL doxycycline + 1 mM sodium butyrate) and kept at 37°C in 5% CO<sub>2</sub>. The medium was changed the day after transfection and every 2 days thereafter. Colonies emerged by day 10 and M15G+S+ media changed to t2i+Lif (N2B27 medium, 1 mM PD0325901, 1 mM CHIR99021, 1000 U/mL mouse LIF). The colonies were fixed for immunocytochemistry at day 14.

**Immunocytochemistry**
Cells were fixed in cold 100% methanol (5 min at −20°C), washed with PBS, then blocked in PBS + 10% FCS for 1 h at room temperature. GATA4 primary antibody (Santa Cruz, sc-25310, 1:50) diluted in PBS/2%FCS was applied at room temperature for 2 h, followed by washes with PBS. Secondary antibody (goat anti-mouse IgG2a, 1:1000) was diluted in PBS and applied for 1 h at room temperature in the dark. The cells were washed with PBS, then 10 μg/mL DAPI applied for 5 min at room temperature in the dark followed by washes with PBS. NANOG protein was detected in cells fixed with 4% paraformaldehyde/PBS (15 min at room temperature) and washed with PBS. The cells were permeabilized using ice-cold methanol (−20°C for 10 min), washed with PBS, and incubated with blocking solution (PBS/0.3% Triton X-100/10% goat serum) for 1 h at room temperature. NANOG primary antibody (Abcam, 80892, 1:100) was diluted in blocking solution and applied overnight at 4°C followed by washes with PBST (PBS + 0.3% TritonX100). Secondary antibody (goat anti-rabbit IgG, 1:1000) was diluted in blocking solution and applied for 1 h at room temperature in the dark. The cells were stained with DAPI as described above.

**Flow Cytometry and FACS**
The parameters for gating cell populations by flow cytometry were established using the EGFP-negative parental DAK31 cells. EGFP fluorescence was measured in single-cell suspensions of ESCs in N2B27 medium on a BD FACSCalibur or BD LSR Fortessa. Cell populations or single cells were sorted in 2iL using a BD FACSAria III cell sorter. Single cells were sorted into wells of a 96-well plate coated with mouse fibroblasts and containing 2iL medium.

**qRT-PCR**
RNA (1 μg), purified using an RNeasy Mini Kit (QIAGEN) was used to synthesize cDNA using SuperScript First-Strand Synthesis System (Invitrogen). Approximately 1/60 of the cDNA was amplified using a Platinum SYBR Green QPCR kit (Invitrogen) under the
following conditions: 50°C for 2 min, then 95°C for 2 min followed by 40 cycles of 95°C for 15 s, then 60°C for 30 s, with a final cycle consisting of 95°C for 1 min, 60°C for 30 s, and 95°C for 15 s. Primer sequences are in Table S4.

Single-Cell cDNA Library Preparation and Sequencing

Rex1-EGFP cells maintained in 2iL were sorted into single cells using a BD FACSAria cell sorter. Wild-type DAK31 parental cells (Blair et al., 2012) were used as a control to exclude EGFP-negative and auto-fluorescent cells. CF-1 fibroblast cells were used to exclude feeder cell contamination. Finally, E3 Rex1-EGFP cells were sorted into three populations, EGFP hi, EGFP lo, and EGFP neg, and single cells plated into a 96-well plate containing 2 μl of lysis buffer. To allow for batch effects, triplicate plates were generated in which 32 cells from each group were plated per plate. A population of 104 EGFPlow cells were also plated into a 2 cm2 well and maintained on CF-1 feeders, in 2iL for 4 days prior to flow cytometry analysis. cDNA from single cells was generated using the smart-seq2 protocol (Picelli et al., 2014). Illumina Nextera reagents were used for library construction and the library was sequenced on the HiSeq 4000 Sequencing System at 75PE to yield a minimum of 290M+290M reads. The single-cell sequencing data are available on Edinburgh Datashare (https://doi.org/10.7488/ds/2639).

Statistical Analysis

Unless specified, all experiments were performed on triplicate biological or experimental samples. The data presented represented means ± SD or SEM. Single-cell sequencing data were analyzed using R to generate principal-component analysis (PCA) and box plots.

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at https://doi.org/10.1016/j.stemcr.2019.12.001.

AUTHOR CONTRIBUTIONS

S.M., J.W., A.J., and T.B. conceived and designed the study. S.M., J.W., T.O., J.O., L.S., and T.W. performed the study. T.C. and A.S. planned and performed the single-cell RNA sequencing experiments. D.F.C. constructed the β-catenin TALENs. S.M., J.W., A.J., T.C., and T.B. analyzed the data. S.M. and T.B. wrote the manuscript. All authors reviewed the manuscript.

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