Transcriptome-Based Network Analysis Reveals a Spectrum Model of Human Macrophage Activation

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SUMMARY

Macrophage activation is associated with profound transcriptional reprogramming. Although much progress has been made in the understanding of macrophage activation, polarization, and function, the transcriptional programs regulating these processes remain poorly characterized. We stimulated human macrophages with diverse activation signals, acquiring a data set of 299 macrophage transcriptomes. Analysis of this data set revealed a spectrum of macrophage activation states extending the current M1 versus M2-polarization model. Network analyses identified central transcriptional regulators associated with all macrophage activation complemented by regulators related to stimulus-specific programs. Applying these transcriptional programs to human alveolar macrophages from smokers and patients with chronic obstructive pulmonary disease (COPD) revealed an unexpected loss of inflammatory signatures in COPD patients. Finally, by integrating murine data from the ImmGen project we propose a refined, activation-independent core signature for human and murine macrophages. This resource serves as a framework for future research into regulation of macrophage activation in health and disease.

INTRODUCTION

During the last two decades, a conceptual framework for the description of macrophage activation has been developed. According to this framework, macrophages can be polarized into classically (M1) or alternatively (M2) activated cells representing two polar extremes of signals computed by macrophages (Biswas and Mantovani, 2010). The M1 versus M2 model has been very helpful in describing immune responses during acute infections, allergies, asthma, and obesity (Chinetti-Gbaguidi and Staels, 2011). However, observations obtained from macrophages involved in chronic inflammation, chronic infection, or cancer strongly suggest that the myeloid compartment has a much broader transcriptional repertoire depending on different environmental signals received (Boorsma et al., 2013; Chow et al., 2011; Edin et al., 2012; Hodge et al., 2011; Lawrence and Natoli, 2011; Martinez et al., 2009; Mosser and Edwards, 2008; Murray and Wynn, 2011; Reinartz et al., 2013). Despite a number of genomic studies analyzing macrophage activation in response to bacteria, TLR ligands, and M1 or M2 stimuli, to date there have been no attempts to reconcile these observations by building new and integrative models of macrophage activation (Martinez et al., 2006; McDermott et al., 2011; Nau et al., 2002; Ramsey et al., 2008).

Transcriptomics has considerably contributed to a better understanding of immune cell function and regulation. Large consortia such as the ImmGen consortium (Best et al., 2013; Bezman et al., 2012; Cohen et al., 2013; Gautier et al., 2012; Miller et al., 2012) or the Human Immunology Project Consortium (Poland et al., 2013) compiled extensive data sets and defined a core transcriptional program for murine tissue macrophages and dendritic cells (DCs) under steady-state conditions (Gautier et al., 2012; Miller et al., 2012). A complementary approach has been introduced by InnateDB (Breuer et al., 2013). Data on molecular interactions between proteins of the innate immune system derived from smaller data sets have been compiled and can be used to reveal mechanistic insights into immune cell function (Hume et al., 2010; Mabbott et al., 2010). Unfortunately, meta-analysis of small data sets has been hampered by several challenges, including differences in the genetic background of mice and in stimulation conditions and the combination of in vitro and in vivo data limit or even bias model generation of incongruous data sets (Mabbott et al., 2010). Moreover, comparative studies have identified substantial differences in immune-cell gene expression between mice and humans (Schröder et al., 2012; Shay et al., 2013). Therefore, it remains to be fully elucidated, how immune cell activation—particularly in human macrophages—is transcriptionally controlled and to which degree these pathways are conserved across species...
Immunity
Spectrum Model of Human Macrophage Activation

A
CD14+ monocytes (n=299)
72 h: GM-CSF (n=275)
M-CSF (n=24)
Macrophages (baseline, M*)
28 stimuli con_1 con_2 con_28
Gene expression profiling
299 macrophage transcriptomes
Quality control
SOM clustering (H)
Co-regulation analysis
Probe filtering
M*+28 conditions, end point arrays, 9488 present genes
1000 variable genes
Coordinates of samples
10 defined clusters
Hierarchical clustering on Pearson correlation matrix (I)
Spectrum model (J)

B
IL4
IL13
IFNy
sLPS

C

D

E

F

G

H

I

J

All time points

End of activation time point
SOM

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standardization of data acquisition and assembling larger data sets, such as by the ImmGen consortium (Heng and Painter, 2008), is necessary to answer such questions.

Several elegant studies have demonstrated the value of analyzing networks based on expression profiling in macrophages (Martinez et al., 2006; Nau et al., 2002; Ramsey et al., 2008) or T helper 17 (Th17) cells (Ciofani et al., 2012; Yosef et al., 2013). These studies show how technological and analytical advances can reveal network structures in immune cells, e.g., by using algorithms that integrate transcriptome data with database-stored information. Other approaches that require large data sets, such as reverse network engineering (RNE), have previously been used to characterize B cell activation (Basso et al., 2005) and have been further refined during the last few years (Marbach et al., 2012). However, so far, RNE has not been applied to other immune cells most likely due to the lack of large enough data sets.

In this study, we generated a resource data set to assess transcriptional regulation during human macrophage activation by comparing a diverse set of stimuli on a single microarray platform under highly standardized conditions. Network modeling of this data set led us to extend the current M1 versus M2 polarization model to a “spectrum model” with at least nine distinct macrophage activation programs. Further characterization of individual programs identified transcription factors associated with particular phenotypes, such as STAT4, which was associated with stimuli linked to chronic inflammation (TNF, prostaglandin E2, TLR2-ligand, “TPP”) as seen in granulomatous diseases (Ciofani et al., 2012; Marino et al., 1997; Reiling et al., 2002; Shay and Celeste Simon, 2012). Furthermore, we applied this resource data set to define activation states of human alveolar tissue macrophages in vivo. Finally, we used our resource data set to refine the previously suggested core macrophage signature to encompass species differences and account for the spectrum of macrophage activation.

RESULTS

Extending the Current Model of Macrophage Polarization

We generated macrophages from human monocytes by differentiation with GM-CSF or M-CSF (Figure 1A) and compared their transcriptomes with DCs and T, B, and natural killer (NK) cells (see Figure S1A, Table S1, and Supplemental Information available online). With coregulation analysis (CRA) to assess overall sample-to-sample relationships, macrophages were clearly distinguishable from other cell types including DCs (Figures S1B–S1E), which was confirmed on protein level by flow cytometry (Figures S1F and S1G). To better understand the complexity of transcriptional regulation after macrophage activation, we next analyzed the transcriptional programs of macrophages activated with 28 different stimuli including pattern recognition receptor ligands, cytokines, and metabolic cues (Figure 1A). To determine the overall relationship of these activation states within the data set, we first applied CRA (Figures 1B–1G). In agreement with the existing model, a virtual axis was formed, where macrophages at baseline (M0) were placed in between macrophages stimulated with interferon-γ (IFN-γ) (M1) and interleukin-4 (IL-4) (M2) (Figure 1B; Movie S1). Adding other conditions linked to M1 (sLPS, TNF) or M2 (IL-13) polarization (Biswas and Mantovani, 2010) (Figure 1C) did not change the overall M1 and M2 axis (Figure 1B). Including further M1- and M2-associated stimuli (IFN-γ+TNF, IL-10) increased the variance in the correlation matrix but the overall bipolar structure was maintained (Figure 1D). However, when adding stimuli not linked to either M1 or M2 polarization, such as free fatty acids, high-density lipoprotein (HDL), or combinations of stimuli associated with chronic inflammation (Table S1), a spectrum of macrophage-activation signatures beyond the initial bipolar axis became apparent (Figures 1E and 1F). Furthermore, when samples generated at earlier time points of stimulation were added, the spectrum of macrophage activation was shown to consist of a dense network of individual signatures (Figure 1G). To bioinformatically validate these findings, we applied Self-Organizing-Map (SOM) clustering (Figure 1H) and correlation coefficient matrices (CCM) (Figure 1I). Performing SOM clustering on the conditions shown by CRA in Figure 1E revealed that every stimulus was characterized by a specific cluster structure (Figure 1H) further supporting the spectrum model. Similarly, we did not identify a bipolar structure within the CCM, but rather a condition-specific spectrum of correlation coefficients in 10 major clusters (Figure 1I). By using the coordinates of the samples defined by CRA within the 10 clusters defined by CCM to build sum vectors in three-dimensional space, we propose a model of macrophage activation best described by a spectrum of transcriptional programs (Figure 1J). Taken together, these data clearly extend the current model of M1 versus M2 polarization to a spectrum model of macrophage activation.

Identification of Genes Specifically Associated with Distinct Stimuli

We next determined whether the different stimuli could be distinguished on the gene level within the complete spectrum of the model. From all macrophage samples included in the analysis, we identified 9,498 genes that were expressed in at least one

Figure 1. Extending Macrophage Polarization to a Spectrum Model

(A) Schema describing the workflow for Figure 1.

(B–G) Correlation networks of 299 macrophage transcriptomes representing 29 conditions from end of activation time points (B–F) also including intermediate time points (G).

(H) Self-Organizing Map (SOM) clustering using samples displayed in (E). Clusters with the top up- or downregulated genes for each stimulus are marked with a frame.

(I) Matrix of hierarchically clustered Pearson’s correlation coefficient matrix (CCM) standardized from −2.78 to 2.78 (blue to red) based on 1,000 most variable probes.

(J) Spectrum model (3D) based on the ten clusters defined in (I) and sample values (coordinates) defined by correlation network from (F). Baseline macrophages (M0) are set as origin, activation states are represented by colored arrows, x, y, z axes are in dashed lines with double arrows. See also Figure S1, Movie S1, and Table S2.
condition (Table S1D). We used these genes for SOM clustering, allowing us to select genes specifically regulated and enriched for individual stimuli (Figure 2A). In fact, by using this approach we could identify genes that were selectively elevated in only one of the stimulation conditions included in our data set. For example, IFN-β selectively induced ZNF77, while IFN-γ selectively induced FEM1C (Figure 2B; Figure S2). However, we also found stimuli where a single gene was insufficient to distinguish between closely related conditions, e.g., SERINC2 was induced by PGE2 but also by PGE2+P3C, suggesting that gene combinations are necessary to distinguish complex input signals at the transcriptional level. Overall, although some input signals might be associated with the induction of single genes, future studies will require the assessment of a substantial number of markers as surrogates for distinct activation programs of macrophages.

**Network Analysis Defines Stimulus-Associated Programs of Macrophage Activation**

To investigate stimulus-specific gene sets associated with the respective macrophage activation programs, we applied
Figure 3. Activation-Specific Genes Revealed by Weighted Correlation Network Analysis

(A) Schema describing the workflow for Figure 3.
(B) Heatmap showing the correlation of the module eigengene (first principal component; ME) to the traits (stimulation conditions). Blue means negative correlation; orange means positive correlation.

(legend continued on next page)
to these activation programs, e.g., IL-4-condition. We also identified additional TFs that are linked as a central hub in the IFN-γ-condition and STAT6 as a hub in the IL-4-condition. We also identified additional TFs that are linked to these activation programs, e.g., STAT2, IRF7, and IRF9s for IFN-γ activation and IRF4 and the forkhead box proteins FOXO1 and FOXO2, which were not previously associated with the IL-4 activation network. For macrophages stimulated with TTP, the TF network also included STAT4, as well as TFs associated with negative regulation of TLR signaling (HEY1; Hu et al., 2008), macrophage activation (TGIF1; Ramsey et al., 2008), or TFs at the interface between inflammation and metabolism (HIFTA; Shay and Celeste Simon, 2012). Other TFs identified in this network have not yet been linked to macrophage activation as determined by pubatlas.org-based literature mining (data not shown). WGCNA revealed activation-associated gene sets responsible for important biological functions of different macrophage populations. These gene sets harbor specific TF networks including well-established TFs associated with major activation programs but also TFs not previously associated with macrophage activation programs. Taken together, this large data set of macrophage activation forms the basis for the establishment of transcriptional networks that are linked to specific activation signals in human macrophages.

**Distinct Phenotype and Function of Macrophages Activated by TNF, PGE2, and TLR2 Ligand**

We used the stimulation condition TTP (MTPP), which is associated with chronic inflammation, to demonstrate phenotypic and functional differences to macrophages stimulated with IFN-γ or IL-4. As demonstrated by CCM (Figure 1) and WGCNA (Figure 3), MTPP differed considerably in their genomic signature from M1 or M2 macrophages. By using differentially expressed genes (FC > 2, FDR adjusted p value < 0.05) between MTPP (TTP), M1 (IFN-γ), M2 (IL-4), and M0 we identified cell surface markers expressed selectively on MTPP. A total of 51 cell surface markers were elevated in MTPP but not M1, M2, or M0. By using flow cytometry, we confirmed significantly elevated expression for CD14, CD23, CD25, CXCR7, and CD187 on MTPP (p value < 0.05), while CD86 was elevated on both M1 and MTPP (Figure 4A). We also identified a set of TFs induced in MTPP but not in M1 or M2, among them STAT4. Analysis of STAT4 protein expression clearly confirmed that STAT4 is only induced in MTPP (Figure 4B). Further differences between MTPP and macrophages stimulated with IFN-γ (M1) or IL-4 (M2) were observed for soluble effector molecules where, e.g., CXCL5 secretion was significantly induced by MTPP and to a lesser extent by M1 (p value < 0.05), but not M2, and IL-1α was only secreted by MTPP (Figure 4C).

No difference in CD3- and CD28-stimulated T cell proliferation was observed in the presence of M1- or M2-activated macrophages, whereas M0 reduced T cell proliferation although not statistically significantly (p value < 0.05) (Figure 4D). However, MTPP showed a strong inhibitory effect, clearly demonstrating that macrophage activation by TTP induced an effector program distinct from M1 and M2 macrophages. Because transcriptional programs are further modulated on posttranscriptional level we assessed the global spectrum of miRNA expression by mirRNA-Seq (Figure 4E). Again, MTPP clearly differed from M1.
Figure 4. Phenotypic and Functional Characterization of Macrophages Stimulated with TNF, PGE₂, and P3C

(A) Flow cytometry of CD14, CD23, CD25, CD86, CXCR7, CD197, in M⁰ (dark gray), IFN-γ (light blue), IL-4 (red), and TPP (light gray) activated macrophages. Mean fluorescence intensities (MFI) of at least three independent experiments are shown (mean and SEM; *p < 0.05 Student’s t test).

(B) Immunoblot analysis of STAT4 and β-actin. STAT4 expression was normalized to β-actin expression and set to 100% in MTPP (TPP).

(C) CXCL5 and IL1α in supernatants of macrophage cultures.

(D) T cell activation by CD3 and CD28 beads in presence or absence of macrophages assessed by CFSE dilution.

(E) Heatmap showing fold changes of highly abundant miRNAs up- or downregulated (FC > 2, FDR adjusted p value < 0.05) in M¹ (IFN-γ) or M2 (IL-4), or MTPP (TPP) compared to M⁰ (baseline). Fold changes colored from blue to red.

For Figures 4A–4D, mean ± SEM (p < 0.05, Student’s t test) are shown.
Figure 5. Enrichment of Activation Modules in Human Alveolar Macrophages

(A) Schema describing the workflow for Figure 5.

(B) Correlation network of human alveolar macrophages (n = 100) from two studies (Shaykhiev et al., 2009; Woodruff et al., 2005) using 374 genes differentially expressed between nonsmokers (n = 39) and smokers (n = 49) or COPD (n = 12) patients (FC > 2.0, FDR adjusted p value < 0.05).

(C) Volcano plots of normalized enrichment scores (NES) and enrichment p values based on GSEA using WGCNA modules defined in Figure 3. Shown are data for the stimuli IFN-γ, IL-4, TPP, and palmitic acid (PA). Red circles show gene sets positively significantly enriched (NES > 1, p value < 0.05); blue circles show gene sets significantly depleted (NES < −1, p value < 0.05).

Figure 5 (legend continued on next page)
and M2 activation at the miRNA level; M\textsuperscript{TPP} had elevated hsa-miR-125a-5p expression and a lack of M1- (hsa-miR-23b-3p) or M2-associated microRNAs (miRNAs) (e.g., hsa-miR-125b-5p, hsa-miR-99a-5p). Similarly, a set of miRNAs was significantly reduced in M\textsuperscript{TPP} compared to M1 or M2 activation (FDR adjusted p value < 0.05). Therefore, macrophages differing in their global transcriptional program from M1 or M2, such as M\textsuperscript{TPP} macrophages, are also phenotypically and functionally distinct, further supporting the spectrum model of macrophage activation.

**Macrophage Activation Model Can Be Used to Predict Macrophage Programs In Vivo**

To address whether specific activation programs such as those described in Figure 3 can be found in human tissue macrophages, we compiled two data sets of human alveolar macrophages obtained by bronchoalveolar lavage (Shaykhiev et al., 2009; Woodruff et al., 2005) consisting of samples from nonsmokers, smokers, and COPD patients. Following filtering steps, data structure analysis, and data visualization, gene-set enrichment analysis (GSEA) was performed (Figure 5A). Three major clusters reflecting the three patient groups were revealed by CRA (Figure 5B) supporting distinct transcriptional programs in macrophages from the three groups. Next, stimulus-specific gene modules identified by WGCNA (Figure 3) were utilized as 49 gene sets from in vitro conditions for enrichment analysis. As positive controls, we applied GSEA to the comparison of IFN-\(\gamma\), IL-4-, TPP-, and palmitic acid (PA)-stimulated macrophages with unstimulated macrophages. We calculated normalized enrichment scores (NES), which were plotted against enrichment p values in a volcano plot (Figure 5C). As expected, the highest positive NES and lowest p values were observed for those gene sets (gene modules) that were most significantly enriched (NES > 1, p value < 0.05) in the WGCNA analysis for the respective stimuli (IFN-\(\gamma\), IL-4, TPP, and PA). We then applied GSEA to the patient sample groups (smoker, COPD) in comparison to nonsmokers (Figure 5D). Unexpectedly, in smokers a glucocorticoid (GC)-associated gene module (41, WGCNA) was most significantly enriched (p value < 0.05) followed by several genes modules associated with free fatty acid but also IL-4 and TPP stimulation, suggesting a complex network of stimuli acting on alveolar macrophages in smokers. In contrast to previous literature (Shaykhiev et al., 2009), we did not see an enrichment of IL-4-IL-13-associated signatures in COPD patients with our data-driven approach (Figure 5D). Rather we found a complete loss of the profound signature of enriched modules observed in smokers. Concurrently, the most significantly depleted (NES < -1, p value < 0.05) WGCNA module in COPD patients was module 8 (linked to IFN-\(\gamma\) stimulation), which was also significantly depleted in smokers (Figure 5D). Network visualization of GOEA further supported complex and profound transcriptional changes in alveolar macrophages from smokers while cells from COPD patients were rather characterized by loss of antigen processing, inflammatory response, and regulation of immune response, consistent with a depletion in the IFN-\(\gamma\) linked module (Figure 5E). Applying the WGCNA- and GSEA-defined macrophage differentiation programs to human ex vivo tissue macrophages, we have uncovered a hitherto unexplored biology in alveolar macrophages from smokers and COPD patients.

**Common Denominators of Macrophage Activation**

While we clearly extended the concept of macrophage polarization (M1 versus M2) to a spectrum model, our large data set also allowed us to define common denominators of macrophage activation. To define these common macrophage activators, we used reverse network engineering (RNE) utilizing ARACNe (Algorithm for the Reconstruction of Accurate Cellular Networks, Figure 6A) (Margolin et al., 2006). We used the 9498 genes present in at least one stimulation condition (Table S1D) to generate a so-called all-versus-all network (Bonferroni corrected p value \(10^{-7}\)) by predicting interactions based on mutual information between each gene pair computed from the expression profiles (Figure S4A; Table S3A; for further technical details see Supplemental Information). We identified 66,744 interactions resulting in an average degree of connectivity of 14.7, meaning one gene is involved in 15 transcriptional interactions on average (Figure 6B; Table S3B; Figure S4B). We confirmed these findings with a second RNE approach (TINGe, Tool for Inferring Network of Genes) (Aluru et al., 2013), which demonstrated high similarity in the number of interactions, the average degree of connectivity and the rank of hubs based on degrees of connectivity as determined by ARACNe (Table S3C; Figure S4C). We summarized the network statistical properties (Figures S4D–S4F), since the entire network is too complex to be displayed. The top 10% of hub genes inferred in the network (n = 869 most interconnected genes) collectively participated in 30,431 interactions (Figure 6B). In the ten most highly interconnected genes, we identified FABP5, which has recently been implicated in lipid metabolism and inflammation crosstalk (Furuhashi et al., 2011), and TNFAIP6, a negative feedback regulator of myeloid cell activation (Choi et al., 2011). However, according to a pubatlas.org search (Table S3D), little is known about the role of the other most highly interconnected genes during macrophage activation, suggesting that RNE approaches reveal unknown aspects of macrophage activation. To further understand the biological processes of the top 10% of hub genes, we performed GOEA with visualization of GO enrichment networks by using BINGO (Figure 6C; Table S3E). This GO-term network subdivided into five major clusters, one of which was related to immune response processes (especially terms associated with “regulation of activation”). However, other clusters were associated with cell death, biosynthetic processes of small molecules, and metabolic and catabolic processes, which also constitute major but underappreciated aspects of macrophage activation.

By using the TFCat database (Fulton et al., 2009), we identified 27 TFs in the top 10% hub genes (TFs, Figure 6B). We reasoned...
Figure 6. Macrophage Activation Network Calculated by ARACNe

(A) Schema describing the workflow for Figure 6.

(B) Visualization of the 10% largest hub genes of the ARACNe predicted macrophage regulatory network (n = 299 transcriptomes). For the top ten genes (highest degree of connectivity, blue) and TFs, mean expression values (log2, derived from the ten clusters in Figure 1I) are highlighted in red colors. Node size reflects degree of connectivity.

(C) Network visualization of GOEA using BiNGO and EnrichmentMap on hubs shown in (B). Red nodes represent enriched GO-terms, and node size represent FDR-adjusted enrichment p value (q value). Edge thickness represents overlap of genes between neighbor nodes.
that the most highly expressed TFs are the most relevant for macrophage activation and therefore ranked them by average expression and generated a network of the top five TFs (JUNB, NFKB1, HIVEP1, CREB1, and HBPI) (Figure 6D; Table S3F). Roles in macrophage activation have been established for all of these TFs, except for the zinc finger protein HIVEP1: JUNB (part of the AP1 complex), NF-κB (global activator), and CREB1 (inducing survival signals) (Wen et al., 2010). HBPI has been linked to differentiation of malignant myeloid cells and to the regulation of other important TFs including PU.1, RUNX1, JUNB, or CEBP (Yao et al., 2005). By using position-weight matrices, we predicted binding of the 27 TFs to the gene loci of the top 10% hub genes (Table S3G). Twenty-six out of 27 of these TFs showed significantly enriched binding prediction (p value < 0.05).

As a complementary approach, we also applied two gene prioritization tools, ToppGene (Chen et al., 2009) and Endeavour (Tranchevent et al., 2008) to rank the potential association and closeness of the top 10% hub genes with macrophage cellular programs using the macrophage lineage TFs RUNX1 and SP11 (PU.1) as test genes. Of note, the top 11 ranked genes were TFs and in total, 20 of the 30 top ranked genes are associated with transcriptional regulation (Figure 6E). In addition to NFKB1, JUNB, and CREB1, we identified additional TFs already associated with macrophage activation (STAT3), as well as other TFs not yet associated with macrophage activation (HMGA1, NFE2, ZNF148, SMARCA2, DDX21, MNDa, TBLX1). Several macrophage-activation markers (e.g., MMP9 and CSF1 [M-CSF]) were also strongly linked to macrophage activation in this analysis. Furthermore, a strong enrichment of PU.1 binding and permissive histone marks H3K4me3 at the loci of the 869 major hub genes indicate that these genes are likely to be highly transcribed during macrophage activation (Figure S5; Supplemental Information). Together, this RNE analysis identified five distinct clusters of biological processes as part of the macrophage activation process, confirmed the involvement of known transcriptional regulators such as NFKB1, and identified unexpected yet unexplored candidate regulators.

**Refinement of Core Genes of Murine Tissue**

**Macrophages using Human Macrophage Activation Signatures**

Comparative transcriptomics of immune cells between human and mouse provides a framework for the use of model systems in the context of human biology and disease (Shay et al., 2013). We therefore propose a general strategy for how our resource of human macrophage transcriptional data can be linked to murine data (Figure 7A). First, we visualized the expression of the human orthologs of the ImmGen defined core macrophage (Figure 7B) and DC (Figure 7C) genes in human macrophages and monocyte-derived DCs under the different stimulation conditions. We visualized the expression values as heatmaps of genes ranked by overall differential expression between human macrophages and DCs (Figures 7A–7C; Table S4). Within the core signature genes defining murine macrophages, we identified three groups of genes (Figure 7B). The first group of genes (1) had high conservation of differential expression between human macrophages and DCs irrespective of macrophage activation, whereas the second group of genes (2) was characterized by loss of differential expression after activation with certain stimuli. The third group of genes (3) was either not differentially expressed between human macrophages and DCs or showed even opposite regulation (Figure 7B). A similar grouping was found for the core signature genes of DCs (Figure 7C). This approach identified cell surface markers (CD14, FCGR2A [CD32], MERTK, FCGR1A [CD64], CD13 [ANPEP]) that distinguish human macrophages from both DCs and CD14+ blood monocytes by flow cytometric analysis (Figure 7D). We propose that this set of cell surface markers should be used for the discrimination between macrophages and DCs in both species. Overall, this comparative transcriptomics approach has refined the core signatures for macrophages and DCs determined in mice to make them also applicable to human macrophages. This will further improve the interpretation of data obtained in species other than human and will guide animal model design to better reflect relevant human biology.

**DISCUSSION**

The generation of this large and unique transcriptomic data set of human macrophage activation represents an important step forward in understanding how macrophages integrate and compute signals from their local microenvironment under inflammatory conditions. The extension of macrophage activation from M1 versus M2 polarization to a spectrum model opens new avenues to study macrophage activation in the context of human diseases. For example, network-based description of global but also input signal-specific transcriptional programming could form the basis for further studies linking defined activation programs with in vivo human macrophage biology. In fact, the lack of any major inflammatory signals in human alveolar macrophages derived from COPD patients was an unexpected—but from a clinical perspective, highly important—result. This result might actually reflect clinical observations demonstrating inefficiency of anti-inflammatory treatment regimens in COPD patients making it necessary to search for alternative strategies (Barnes, 2013). A better understanding of the transcriptional regulation of human macrophages could help to selectively target specific macrophage subsets therapeutically and thereby could spare other cell types.

Integrating human and murine transcriptome data sets will aid to prioritize and focus future work in animal models. In this respect, overlaying our human data onto the ImmGen-derived core signatures of DCs and macrophages classified several genes to be conserved in expression regulation, whereas others clearly are not conserved. In the future, these studies have to be extended comparing macrophages from the same tissues in both species in homeostasis and pathophysiology. Alternatively, the
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A

Human

Mouse

166 macrophages, 33 DC and 22 monocyte
transcriptomes

216 samples 17981 present probes

Fold change calculation

Biobase

BioGPS

44 M core genes
43 DC core genes

40 M core genes
45 DC core genes

Z-transformation and heatmap (B-C)

Flow cytometry (D)

B

CD14
PLD3
CAMK1
TBXAS1
CSPG2
PLD1
CTSD
FCGR2A
COMT
FGD4
MERTK
PLAG1
FCGR1A
TLR7
PECAM1
TP53
DRAM2
Tlr4
TP1
PCY1X1
Ddx3
TOM1
BLVRB
MR1
GLUL
ARSG
MYO2A
PTPLAD2
CD164
CTLL2
FER
C10orf128
LAMP3
TCN2
NLR1
PLA2G4A
SQRDL1
SEPP1
SLC4A1
ABC1

C

ADAM19
CCRT
TRA15
ZBTB46
GPR152
ANPEP
SLAMF7
RBP30
RNUK3
SPINT2
PTX1
KMO
HLA-DRB2
CITA
GPR88
FLT3
DP4
CSF2R3
BCL11A
ASS1
JAK2
HLA-B
K1T
CD123
CNN2
BH3BP
GPR62
GPR13
SEPT6
NAPSA
PDSTP1
HLA-DMB
AMICA1
FOL2

D

CD14 (MFI) x 10^5

CD32 (FCGR2A)

MERTK

CD64 (FCGR1A)

CD13 (ANPEP)

(legend on next page)
Identification of human-specific regulation of gene expression will require new methodologies to study gene regulation in an entirely human context, and without the respective animal models. Beyond aspects comparing murine and human macrophages, this resource data set can help to answer open questions concerning differential activation of human macrophages with closely related stimuli, e.g., ultrapure and standard LPS, which have been often used synonymously in prior studies. Similarly, while free fatty acids seem to induce closely related transcriptional programs when compared to the remaining stimulation conditions, we have clear evidence that saturated and unsaturated fatty acids induce distinct transcriptional responses in human macrophages (S.V.S., data not shown).

We anticipate this data set of human macrophage activation to serve as a starting point for future studies into human macrophage biology. In addition to expanding our understanding of human macrophage biology, this resource will contribute to a better understanding of general mechanisms of transcriptional control, as well as the development of new mathematical models for signal integration and new therapeutic strategies in human disease.

**EXPERIMENTAL PROCEDURES**

Detailed description of all experimental procedures and links to analytical tools and databases used are provided in Supplemental Information. Abbreviations and description of bioinformatics tools are summarized in Table S5.

**Isolation, In Vitro Culture, and Functional Assessment of Cells under Study**

 Buffy coats from healthy donors were obtained according to protocols accepted by the institutional review board at the University of Bonn (local ethics votes no. 289/13). Human monocytes, B cells, T cells, and NK cells (Table S1) were purified from peripheral blood mononuclear cells by MACS in accordance with the manufacturer’s instructions. Macrophages (M0, baseline) were generated from monocytes by stimulation with GM-CSF or M-CSF for 72 hr and further activated with 28 stimulation conditions (Table S1). DCs were generated by GM-CSF in presence of IL-4 for 72 hr followed by further stimulation with uLPS, TNF+PGE2, or sCD40 mAbs+TNF. Cells were phenotypically assessed by flow cytometry using cell lineage and activation markers. Expression of STAT4 was measured by immunoblotting. CXCL5 and IL-1 were typically assessed by ELISA following the manufacturer’s instructions. Expression of murine macrophage and dendritic cell signature genes in humans (A) Spectrum Model of Human Macrophage Activation

methods, ARACNe and TiNGe. Networks were visualized in a force-directed layout in Cytoscape, followed with statistical analysis utilizing the plug-in Network Analysis (Cline et al., 2007). With the cytospot plug-in MultiColoredNodes (Warsow et al., 2010) mean expression values of the most highly interconnected genes and TFs were visualized.

**Linking Human Macrophage Activation to ImmGen Core Signatures**

Human macrophage samples (n = 166), DCs (n = 33), and monocytes (n = 22) were compiled and expression values of the human orthologs of the previously described murine macrophage (Gautier et al., 2012), and DC (Miller et al., 2012) core signatures were plotted as a heatmap of standardized and scaled log2 values (Figure 7; Table S4).

**ACCESSION NUMBERS**

Macrophage microarray data, ChiP-Seq data, miRNA-Seq data, and alveolar macrophage microarray data are accessible via super series GSE47189. Preprocessed data are also available at an online resource portal (www.macrophages.uni-bonn.de).

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes five figures, five tables, Supplemental Experimental Procedures, and one movie and can be found with this article online at http://dx.doi.org/10.1016/j.immuni.2014.01.006.

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