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Published In:
Developmental Biology

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Transcription factor Ap-2α is necessary for development of embryonic melanophores, autonomic neurons and pharyngeal skeleton in zebrafish

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Received for publication 3 April 2003, revised 25 September 2003, accepted 25 September 2003

Abstract

The genes that control development of embryonic melanocytes are poorly defined. Although transcription factor Ap-2α is expressed in neural crest (NC) cells, its role in development of embryonic melanocytes and other neural crest derivatives is unclear because mouse Ap-2α mutants die before melanogenesis. We show that zebrafish embryos injected with morpholino antisense oligonucleotides complementary to ap-2α (ap-2α MO) complete early morphogenesis normally and have neural crest cells. Expression of c-kit, which encodes the receptor for the Steel ligand, is reduced in these embryos, and, similar to zebrafish c-kit mutant embryos, embryonic melanophores are reduced in number and migration. The effects of ap-2α MO injected into heterozygous and homozygous c-kit mutants support the notion that Ap-2α works through C-kit and additional target genes to mediate melanophore cell number and migration. In contrast to c-kit mutant embryos, in ap-2α MO-injected embryos, melanophores are small and under-pigmented, and unexpectedly, analysis of mosaic embryos suggests Ap-2α regulates melanophore differentiation through cell non-autonomous targets. In addition to melanophore phenotypes, we document reduction of other neural crest derivatives in ap-2α MO-injected embryos, including jaw cartilage, enteric neurons, and sympathetic neurons. These results reveal that Ap-2α regulates multiple steps of melanophore development, and is required for development of other neuronal and non-neuronal neural crest derivatives.

Keywords: Transcription factor ap-2; Zebrafish; Morpholino; Neural crest; Melanocyte; Branchial arches; Cranial nerves; c-kit; Enteric neurons; Sympathetic neurons; Hirschsprung’s disease

Introduction

Various growth factors are known to regulate the multiple steps of melanocyte development, but cell-autonomous requirements for melanocyte development are less well defined. For instance, misexpression and loss-of-function studies in zebrafish suggest Wnt signaling directs neural crest cells to adopt the melanophore fate (Dorsky et al., 1998). Analysis of mouse and zebrafish mutants suggest that steel factor (SCF), mediated by its receptor Kit, controls melanoblast migration and proliferation (Brannan et al., 1991; Nocka et al., 1989, 1990; Parichy et al., 1999; Reith et al., 1990; Tan et al., 1990). The ligand endothelin-3, working through its receptor Ednrb, also appears to promote melanoblast proliferation and perhaps differentiation (Bannash et al., 1994; Lahav et al., 1996; Reid et al., 1996; Shin et al., 1999). In mice, α-melanocyte-stimulating hormone...
Fig. 1. Gene structure of *ap-2α* and its expression in neural crest. (A) Multi-species comparison of predicted peptide sequence of exon 1 of AP-2 proteins. (Top) Exon 1, zebrafish *Ap-2α*, isoform 1 (GenBank accession number, AF457191) differs by just 1 of 15 amino acids from human AP-2α (NP_003211) (Williams et al., 1988) and mouse Ap-2α isoform 1 (P34056) (Moser et al., 1993), which are identical. Genbank accession number of each gene is included. The amino terminal sequence of zebrafish *Ap-2α* isoform 1 is also highly conserved in human AP-2α, AP-2γ, and AP-2δ, as well as in rat and chicken AP-2 proteins. (Bottom) Zebrafish Ap-2 isoform 2 (AF457192) differs from isoform 1 by an alternative exon 1, but utilizes the same exons 2–7. Exon 1 of isoform 2 shows no significant homology to known mouse or human isoforms, but shares 8 of 11 amino acids in sheep AP-2α, variant 6 (GenBank accession number, AAF70345) (Limesand and Anthony, 2001) and four of five amino acids with frog AP-2 (Winning et al., 1991). (B–H) Expression of *ap-2α* RNA revealed by whole mount in situ hybridization using a probe complementary to full-length *ap-2α*, isoform 1. Under the hybridization conditions employed here, this probe will hybridize to isoform 2 and any other first-exon variants of *ap-2α*. Anterior is to the left in all panels. (B) Dorsal view of a flat-mounted embryo at 12 hpf. High-level expression was evident in premigratory neural crest (NC) and pronephros (arrowhead), and low-level expression was present in non-neural ectoderm (NNE). Expression was absent from neural plate (NP). (C) Dorsal view of the anterior trunk lateral neural plate, of an 11 hpf, flat-mounted embryo, processed to reveal *ap-2α* RNA, in blue, and *foxd3* RNA, a marker of premigratory neural crest (Odenthal and Nusslein-Volhard, 1998), in red. All *foxd3*-expressing cells in lateral neural plate also expressed *ap-2α*, supporting the notion that these cells are premigratory neural crest (PNC). (D) The same embryo as shown in C, but focused in more caudal lateral neural plate. At this axial level, some lateral neural plate cells, presumed to be PNC, expressing *ap-2α* also expressed *foxd3* (asterisks) and others did not yet express *foxd3* (arrowhead), suggesting *ap-2α* is expressed in PNC before *foxd3*. (E) Dorsal head view and (F) lateral trunk view of embryos at 17 hpf. Expression was evident lateral to the midbrain and hindbrain, in presumed migrating cranial neural crest (arrowheads in E), and in dorsal neural tube, in trunk neural crest (arrowhead in F). (G) Lateral view trunk of 24-hpf embryo. Expression was detected in the migrating posterior lateral line primordium (arrowhead) and in interneurons of the hindbrain (asterisk) and trunk, but was absent from trunk neural crest by this stage. All scale bars, 100 μm.
regulates the type of melanin that is produced, although the role of this hormone in human pigmentation is unclear (Voisey and van Daal, 2002).

What are the early cell autonomous requirements for the melanocyte fate? The transcription factor Microphthalmia (Mif) is among the earliest known markers of the melanoblast fate, and is required for melanocyte development (reviewed in Tachibana, 2000). Other candidates for such genes include transcription factors that are expressed from early stages in neural crest, including activating protein 2 (AP-2). There are four member of the AP-2 family in humans and mice (α, β, γ, and more diverged δ) (reviewed in Hilger-Eversheim et al., 2000; see Zhao et al., 2001). To date, only one homologue each, most closely resembling mammalian AP-2α, has been characterized in zebrafish, frog, lamprey, amphioxus, and fruit fly (Bauer et al., 1998; Furthauer et al., 1997; Meulemans and Bronner-Fraser, 2002; Winning et al., 1991). In vertebrates, AP-2 homologues (except AP-2δ) are expressed in the epidermis and neural plate border and later in migrating neural crest (NC) cells at all axial levels (Chazaud et al., 1996; Furthauer et al., 1997; Luo et al., 2002; Meulemans and Bronner-Fraser, 2002; Mitchell et al., 1991; Moser et al., 1997; Zhao et al., 2001). In cephalochordates (e.g., Amphioxus), which are thought to have separated from the vertebrate lineage before the appearance of neural crest, AP-2 is expressed in non-neural ectoderm but not in the dorsal neural tube (Meulemans and Bronner-Fraser, 2002).

Ap-2α influences the fate of melanoma cells; however, a role for Ap-2α in development of melanocytes has not been adequately tested. Huang et al. (1998) showed that levels of AP-2α and KIT are reduced in aggressive but not non-aggressive melanoma cells. Misexpression of AP-2α in aggressive melanoma led to increased levels of KIT and decreased tumorigenicity and metastatic potential (Huang et al., 1998). These results suggest that AP-2α would regulate KIT expression in embryonic melanocytes as well. Mouse embryos homozygous for a targeted deletion of Ap-2α have defects in some neural crest derivatives, including jaw elements and cranial ganglia (Schorle et al., 1996; Zhang et al., 1996), and in vitro studies implicate Ap-2α in control of Schwann cell differentiation (Stewart et al., 2001). However, Ap-2α mutant embryos die at birth from severe morphological defects, thus melanocyte development could not be evaluated (Schorle et al., 1996; Zhang et al., 1996). To circumvent the requirement for Ap-2α in early morphogenesis, Nottoli et al. (1998) created mosaic embryos from wild-type and Ap-2α mutant cells. Of note, they reported that melanocytes derived from mutant cells with some frequency in these mosaics, but this experiment did not reveal population-level effects of Ap-2α on melanocytes. X. laevis embryos injected with antisense oligonucleotides targeting AP-2 have reduced expression of neural crest markers, however, they stall in development shortly after gastrulation, thus they were not useful for evaluating a role for Ap-2α in neural crest patterning (Luo et al., 2002).
To establish a model system in which to further explore the requirement for Ap-2α in melanocytes and other neural crest derivatives, we injected morpholino antisense oligonucleotides (morpholinos) to knockdown the function of Ap-2α in zebrafish. Early development was normal in embryos injected with ap-2α morpholinos and neural crest was present, although expression of crestin in neural crest was reduced. Injected embryos developed with a reduced number of melanophores, and melanophores migrated and differentiated abnormally. Concomitant with these visible defects was a reduction of c-kit expression, which is known to be required for melanophore cell number and migration, and of dopachrome tautomerase ( dct) expression, presumed to be necessary for melanophore differentiation. In addition to melanophore defects, we detected a reduction of jaw cartilage, sympathetic neurons, and enteric neurons. Analysis of zebrafish embryos deficient in Ap-2α has thus uncovered embryonic functions for this protein that were obscured in the more severely defective mouse and frog embryos lacking AP-2α.

Methods

Fish and embryo rearing

Zebrafish adults and embryos were reared as previously described (Westerfield, 1993) in the University of Iowa Zebrafish Facility and staged by hours or days post fertilization at 28.5°C (hpf or dpf) (Kimmel et al., 1995). Homozygous or heterozygous sparse mutant embryos, harboring a null allele of c-kit (sparsej) (Parichy et al., 1999), were generated by crossing homozygous adults to each other, or to wild-type adults, respectively.

Morpholino composition and injection

Morpholino antisense oligonucleotides (MO) (Gene Tools, Corvallis, OR) were designed to complement zebrafish ap-2α. The genomic structure of ap-2α was inferred from genomic traces and contigs found at , www.ensembl.org. Two isoforms of zebrafish AP-2 have been identified, which differ only by their first exon (Genbank accession numbers: , AF457191 and , AF457192, isoforms 1 and 2, respectively) (Furthauer et al., 1997). The first six exons, including the alternate first exon, are contained in contig ctg25479.

To target both isoforms simultaneously, we used ap-2α E2I2 MO (5’-AGCTTTTCTTCTTTACCGCAACTTCT-3’), which overlaps the exon2—intron2 splice site, or ap-2α E3I3 MO (5’-GAAATTGCTTACCTTTTTTGATTAC-3’), which overlaps the exon3—intron3 splice site. Negative controls included the standard random 25-mer from the supplier (Gene Tools), and a variant of ap-2α E2I2 MO, ap-2α E2I2 mismatch MO (5’-AGGTATCTTGCTACCTCAAGATCT-3’) which has polymorphisms at six bases. Morpholinos were reconstituted in Danieux buffer (Nasevicius and Ekker, 2000), then diluted in 0.2 M KCl to 1 mg/ml (E2I2) or 4 mg/ml (E3I3 and negative controls morpholinos). Embryos were injected with 2–4 nl of diluted morpholino at the 2–8 cell stage into the yolk immediately below the blastomeres. For all phenotypes reported here, at least 100 injected embryos were analyzed in three or more injection experiments. The reported phenotype was seen in at least 50% of embryos injected with either ap-2α E2I2 or ap-2α E3I3 MO at the indicated doses, except as noted in figure legends. When referred to in the text, the “strongly affected” phenotype was observed in 5–10% of embryos injected with E2I2 or E3I3 but never in embryos injected with control morpholinos.

RT-PCR

Fifteen embryos, either uninjected or injected with 2–4 nl of E2I2 MO at 1.0 mg/ml or E3I3 MO at 4.0 mg/ml, were quick-frozen to −80°C at 16 hpf, then RNA was harvested from them with RNAwiz (Ambion, Austin, TX). First-strand cDNA was synthesized with 200 ng of total RNA, 100 ng of random hexamer primers, 200 units of M-MLV Reverse Transcriptase (Invitrogen, Carlsbad, CA), 0.5 mM dNTPs, 50 mM Tris–HCl, pH 8.3, 75 mM KCl, 3 mM MgCl2, 0.01 mM DTT, 40 units of Rnase Inhibitor, (Invitrogen), incubated at 37°C for 1 h. Nonquantitative polymerase chain reaction (PCR) was performed with 2.5 units of JumpStart Taq polymerase (Sigma, St. Louis, MO), 10 mM Tris–HCl, pH 8.3, 15 mM KCl, 0.2 mM dNTP, 0.12 μM each primer, a forward primer in exon 2 (5’-CGCTCCTCCGTGTCTCAT-3’), a reverse primer in exon 4 (5’-TCTAGATCTGGTTTACGACCC-3’) (Xba1 restriction site added to the 5’ end), and 1/20 of the cDNA synthesis reaction. PCR was run on a thermal cycler (MJ Research, Waltham, MA), with the following program: 2 min at 94°C, followed by 35 rounds of 20 s at 92°C, 10 s at 94°C, 30 s at 60°C, and 30 s at 72°C, followed by a final round of 5 min at 72°C.

Gene expression analysis

DIG-labeled antisense RNA probes (Roche Diagnostics, Mannheim, Germany) for in situ hybridization were generated from plasmids as follows: ap-2α, (Furthauer et al., 1997), NotI/T7; crestin, (Rubinstein et al., 2000), NotI/T7; c-kit (Parichy et al., 1999), XhoI/T3; dct, (Kelsh et al., 1996), EcoR1/T7; dlx2 (Akimenko et al., 1994), BamHI/T7; ednrb1, (Parichy et al., 2000a), XhoI/T3; foxd3 plasmid (fd6—Zebrafish Information Network) (Odenthal and Nusslein-Volhard, 1998), BamHI/T7; mifit, (Lister et al., 1999), EcoR1/T7; sox10, (Dutton et al., 2001b), SalI/T7. Two-probe in situ analysis was conducted with foxd3 probe synthesized using FITC-UTP and ap-2α synthesized using DIG-UTP (Roche Diagnostics). Embryos were hybridized
with ap-2α and foxd3 probes simultaneously, washed, blocked, incubated in 1:15,000 dilution of anti-FITC conjugated to alkaline phosphatase, washed, and developed in Fast Red (Sigma product F-4648). Embryos were subsequently stripped of antibody by washing in 1% glycine, pH 2.0, 3 × 10 min at RT, then reblocked, incubated in anti-DIG alkaline phosphatase, and developed as usual in NBT/BCIP (Roche).

Markers of trigeminal ganglia, used in whole mount preparations, included Zn12 (Metcalfe et al., 1990) and anti-Hu (Marusich et al., 1994) immunoreactivity at 28 hpf and 4 dpf respectively, and ngn1 RNA expression at 18 hpf (Andermann et al., 2002; Korzh et al., 1998). Monoclonal antibodies were used at the following dilutions: zn12, 1:4000 (Metcalfe et al., 1990); and anti-HU (monoclonal antibody 16a11, Molecular Probes, Eugene, Oregon) (Marusich et al., 1994), 1:100. Whole mount samples were developed with horse radish peroxidase as described elsewhere (Cornell and Eisen, 2002).

Alcian green was used to label pharyngeal cartilage as described (Kimmel et al., 1998). Anti-tyrosine hydroxylase immunoreactivity just ventral to the dorsal aorta
reveals sympathetic neurons (An et al., 2002). Frozen sections (15 μm) of embryos were collected on gelatin-coated slides, blocked in 0.5 M NaCl, 0.01 M phosphate buffer (PBS) (pH 7.0) with 5% normal goat serum and 0.1% Triton X-100 (block solution), incubated with polyclonal anti-tyrosine hydroxylase (Pel-Freez Biologicals, Brown Deer, Wisconsin) at 1:100 dilution in block solution, rinsed extensively in PBS with 0.1% Triton X-100, incubated in commercially available rhodamine-labeled goat anti-rabbit IgG secondary antibody (Jackson ImmunoResearch, West Grove, PA) diluted in block solution, and rinsed again. Embryos were examined on a Leica DMRA2 compound microscope and photographed with a Q-imaging Retiga 1300 digital camera and Open...

Fig. 4. Melanophore cell number and migration regulated by Ap-2α and C-kit. Lateral views of fixed embryos at 42 hpf. (A) Uninjected wild-type embryo and (B) ap-2α morpholino-injected wild-type embryo. ap-2α MO embryos had a significant reduction of the total number of melanophores (uninjected: 317.9 ± 23.5 total melanophores; ap-2α MO embryos: 120.6 ± 27.3 total melanophores; n = 10, P < 0.0001), and the fraction of them found in the ventral stripes (arrowheads) (uninjected: 45.6 ± 2.8%; ap-2α MO embryos: 27.3 ± 6.0%; n = 10, P < 0.0001). (C) Uninjected c-kit homozygous mutant embryos similarly have fewer total melanophores, with fewer migrated to the ventral stripe, than wild-type embryos (Parichy et al., 1999). (D) c-kit homozygous mutant embryos injected with ap-2α morpholino have still further melanophores than uninjected c-kit homozygous mutant embryos (c-kit mutants: 196 ± 23.3 melanophores, ap-2α MO injected c-kit mutants: 69.4 ± 24.1 melanophores, n = 10 embryos, P < 0.0001), and a smaller fraction in ventral stripes (c-kit mutants: 13.2 ± 3.2%, ap-2α MO c-kit mutants: 8.3 ± 4.4%, n = 10 embryos, P < 0.01). Scale bar, 100 μm.

Fig. 5. Analysis of cell autonomy of melanophore differentiation phenotype. Close-up views of melanophores in 30-hpf embryos. (A and D) bright field, (B and E) fluorescent view, (C and F) merged view. (A–C) A labeled melanophore derived from a cell transplanted from a control donor into a control host (arrowhead). The labeled melanophore was similar in size, number of dendrites, and density of pigmentation as the adjacent host-derived melanophore (60 control-to-control transplants scored, three or more labeled melanophores observed in 14 embryos). (D–F) Labeled melanophores, derived from a control donor, in an ap-2α MO host (arrowheads). The melanophore was similar in shape and color to host-derived melanophores, but smaller, less dendritic, and under-pigmented in comparison to labeled melanophores in equivalently staged control-to-control transplants (50 control-to-ap-2α MO transplants scored, three or more labeled melanophores observed in 12 embryos). In converse experiment, 200 ap-2α MO-to-control transplants were scored, 95 embryos with labeled cells in neural tube were closely examined, but labeled melanophores were detected in none.
lab 3.1.2 software, and figures were assembled, and brightness and contrast manipulated, in Photoshop 7.0.

**Transplants**

Two to five nanoliters of 1% rhodamine-labeled dextran (10000 mw, Molecular Probes) was injected into the yolk cell of donor embryos and 2–5 nl of 0.9 mg/ml unlabeled ap-2 E2I2 morpholino was injected into the yolk cell of host embryos at the 2–4 cell stage. Alternatively, 2–5 nl of 0.9 mg/ml rhodamine-labeled ap-2 E2I2 morpholino was injected into donor embryos, and hosts were uninjected (while the labeled morpholino was sufficiently bright to serve as lineage tracer for donor-derived cells, in some experiments, rhodamine-labeled dextran was added to the morpholino to increase brightness further). At the 1000 cell stage, 20–50 cells were removed from donor embryos by gentle suction and deposited near the margin of hosts of the same stage. Embryos were raised to 30 hpf. Using a stereomicroscope mounted with fluorescence optics, embryos with labeled cells in the neural tube were identified (approximately 50% of surviving transplants, regardless of whether donor or host was injected with morpholino). Such embryos were examined for labeled melanophores on a compound fluorescence microscope. At 30 hpf, morpholino-injected embryos were approximately 2 h delayed in development with respect to control embryos. Comparison of melanophore morphology was performed between equivalently staged embryos.

**Results**

**Zebrafish ap-2α is expressed in premigratory neural crest**

The two zebrafish variants of ap-2 cDNA isolated by Furthauer et al. (1997) most closely resemble mammalian Ap-2α (84.7% identical to human AP-2α, Williams et al., 1988; 68.1% identical to human AP-2β, Moser et al., 1995; and 59.9% identical to human AP-2γ; McPherson et al., 1997), so we refer to them as ap-2α isoforms 1 and 2. The isoforms differ from one another only at their 5′ ends, and analysis of genomic sequence revealed the alternate leader sequences are present as single exons upstream of the shared exons (see Methods). The leader sequences are homologous to those of two mouse isoforms of Ap-2α (Fig. 1), suggesting there may be functional significance to them. In mouse embryos, several of first-exon isoforms of AP-2α are expressed in overlapping but distinct domains (Meier et al., 1995; Mitchell et al., 1991). The zebrafish isoforms differ by less than 100 bp, and our attempts to resolve their

<table>
<thead>
<tr>
<th>Dose of E2I2 MO</th>
<th>Trunk melanophores</th>
<th>c-kit&lt;sup&gt;−/−&lt;/sup&gt;</th>
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<tr>
<td></td>
<td>Total&lt;sup&gt;a&lt;/sup&gt;</td>
<td>Ventral (%)&lt;sup&gt;b&lt;/sup&gt;</td>
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<tr>
<td>uninjected</td>
<td>69.1 ± 8.1</td>
<td>43.5 ± 3.4</td>
</tr>
<tr>
<td>0.2 mg/ml</td>
<td>60.8 ± 5.5</td>
<td>41.3 ± 5.3</td>
</tr>
<tr>
<td>0.4 mg/ml</td>
<td>52.2 ± 6.1</td>
<td>35.1 ± 6.6</td>
</tr>
<tr>
<td>0.8 mg/ml</td>
<td>47.4 ± 8.4</td>
<td>30.1 ± 6.4</td>
</tr>
</tbody>
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<sup>a</sup> Total number of melanophores in dorsal, lateral and ventral stripes, counted in the eight somite region of trunk rostral to the anus. n = 10 for all treatments.

<sup>b</sup> Percentage of total melanophores in ventral stripe.

<sup>*</sup> Significantly different from wild-type at same dose, P < 0.001 (Student’s t test).

<sup>**</sup> Significantly different from wild-type at same dose, P < 0.005 (Student’s t test).
individual expression patterns by whole mount in situ hybridization were unsuccessful.

Whole mount in situ hybridization of neurula stage embryos, using a probe that hybridizes to both isoforms of ap-2α, revealed that at 12 hpf, ap-2α was expressed in presumptive neural plate (PNC) of the lateral neural plate and in presumptive epidermis but excluded from rest of the neural plate, as previously reported (Furthauer et al., 1997). Double in situ analysis with an early marker of premigratory neural crest, foxd3, confirmed that these genes are expressed in the same cells, and showed that ap-2α expression slightly precedes foxd3 expression in this region (Fig. 1). At 17 hpf, expression was detected in migrating cranial neural crest (Fig. 1E) and trunk dorsal neural tube (Fig. 1F). By 24 hpf, expression was detected in lateral line primordium (Fig. 1G), perhaps in Schwann cells associated with it, but not near melanoblasts or elsewhere on trunk neural crest migratory pathways. These results suggest ap-2α is expressed in premigratory trunk neural but extinguished shortly after these cells delaminate from the dorsal neural tube. It is not clear when cells of vagal neural crest, whose migratory pathway has not been well characterized, lose expression of ap-2α.

Antisense morpholinos disrupt splicing of ap-2α transcripts

In zebrafish embryos, injection of morpholinos that are complementary to exon boundaries can direct splice machinery to choose alternate splice donor sites (Draper et al., 2001). To target all first-exon variants of ap-2α, morpholinos were designed to complement the exon 2–intron 2 (E2I2 MO) or the exon 3–intron 3 (E3I3 MO) splice donor sites. The effect of these MO on splicing of the ap-2α transcripts was tested by isolating RNA from uninjected control or morpholino-injected embryos, synthesizing first-strand cDNA, and amplifying ap-2α cDNAs (RT-PCR analysis) (Fig. 2). Both morpholinos caused mis-splicing of a fraction of ap-2α RNA, resulting in internal deletions of these transcripts that would be expected to prevent function of the encoded proteins (Fig. 1; Wankhade et al., 2000). Embryos injected with E2I2 MO, referred to below as ap-2α MO embryos, were analyzed for defects in neural crest derivatives. Specificity of each phenotype was subsequently confirmed by analysis of embryos injected with E3I3 MO, or with negative-control morpholinos (see Methods).

Neural-crest-derived melanophores are reduced in ap-2α MO embryos

In ap-2α MO embryos, there were no gross abnormalities of gastrulation and early morphogenesis, in contrast to mice homozygous for targeted mutation of Ap-2α (Schorle et al., 1996; Zhang et al., 1996), and frog embryos injected with ap-2-function-blocking oligonucleotides (Luo et al., 2002). The first visible phenotypes in zebrafish ap-2α MO embryos were within neural crest-derived melanophores, which had multiple defects. At 28 hpf, when black, dendritic melanophores were visible in the heads of uninjected embryos (Fig. 3A), in ap-2α MO embryos, melanophores were pale and small (Fig. 3B). By contrast, eye pigmentation was normal at this and later stages. At 42 hpf, only about 40% of the normal number of melanophores were present in ap-2α MO embryos (Figs. 4A and B). Moreover, the fraction of trunk melanophores present in ventral stripes (contacting the hindyolk) was also reduced in ap-2α MO embryos (Figs. 4A and B, Table 1), implying Ap-2α function is required for normal melanophore migration. At 48 hpf, the morphology and color (but not number) of melanophores had largely recovered in ap-2α MO embryos. It is unclear whether melanophores recover because they become independent of Ap-2α function at this stage, or because of diminishing efficacy of morpholinos. No defects on xanthophores and on iridophores were detected in ap-2α MO embryos.

Expression of pigment cell markers was examined in ap-2α MO embryos. mitf, a gene required for specification of melanoblasts (nacre mutant, Lister et al., 1999), was expressed in fewer cells than normal at 24 hpf (Figs. 3C and D), although the level of expression of this gene within single cells appeared normal, suggesting mitf is not directly regulated by Ap-2α. In contrast, dopachrome tautomerase (dct), which encodes a tyrosinase family member necessary for melanin synthesis (Pawelek and Chakraborty, 1998), was expressed at much lower levels within individual cells in 24 hpf ap-2α MO embryos (Figs. 3E and F). A reduction of Det may explain why melanophores are underpigmented at 28 hpf. Similarly, c-kit, which encodes a receptor tyrosine kinase implicated in regulation of proliferation and survival of melanophores (Parichy et al., 1999), was expressed in fewer neural crest cells and at lower levels in 24 hpf ap-2α MO embryos than in uninjected embryos (Figs. 3G and H). By contrast, expression of c-kit in cells near the anus was unchanged in ap-2α MO embryos, revealing distinct regulation of c-kit expression in that tissue (Figs. 3G and H). Finally, expression of ednrb1, which encodes a putative receptor for endothelin-3 and is essential for adult but not embryonic melanoblasts (Parichy et al., 2000a), was similar to mitf in that it was expressed at normal levels, but in fewer cells than normal (not shown). Expression of c-fms, a c-kit homologue expressed in xanthophore precursors (Parichy et al., 2000b), appeared normal in ap-2α MO embryos (not shown).

Genetic interaction between Ap-2α and C-kit

The reduction of c-kit expression in ap-2α MO embryos suggests that Ap-2α and C-kit function in the same pathway to regulate melanophore cell number and migration. Consistent with this notion, c-kit heterozygotes injected with sub-maximal doses of ap-2α morpholino had fewer melanophores than similarly injected wild-type embryos (Table
absent, but they are merely smaller in cerratobranchials, derived from p3 – p7, were variably reduced in strongly affected embryos. p1 derivatives, Meckel’s cartilage (m) and palatoquadrate (pq), reminiscent of the lockjaw mutant phenotype(Schilling et al., 1996), except less severe. For instance, in mouse Ap-2α mutants, ceratobranchials were less affected but were variably reduced in size in strongly affected ap – 2α MO embryos (about 10% of embryos injected with either E2I2 or E3I3 morpholino). In contrast, Meckel’s cartilage and the palatoquadrate, first-arch (p1) derivatives, were relatively normal in size; however, in strongly affected embryos they pointed ventrally (Fig. 6).

Some neuronal derivatives of neural crest were affected in ap-2α MO embryos. Ap-2α directly binds and activates the tyrosine hydroxylase and dopamine β-hydroxylase (DBH) promoters (Greco et al., 1995; Kim et al., 2001). In ap-2α MO embryos, the number of cells expressing tyrosine hydroxylase near the dorsal aorta at 4 dpf, indicative of neural crest-derived sympathetic neurons and adrenal chromaffin cells (An et al., 2002), was highly reduced (Fig. 6). Dorsal root ganglion neurons appeared normal in ap-2α MO embryos, similar to mouse Ap-2α mutants (Schorle et al., 1996; Zhang et al., 1996). Interestingly, enteric neurons, which are derived from vagal neural crest (Le Douarin and Kalcheim, 1999), were dramatically reduced in ap-2α MO embryos (Fig. 6). In contrast to mouse Ap-2α mutants, markers of trigeminal ganglia (Zn 12 immunoreactivity at 30 hpf, anti-Hu immunoreactivity at 4 dpf) (results not shown, see Methods) were not dramatically changed in ap-2α MO embryos.

Multiple neural crest derivatives were reduced in ap-2α MO embryos

Because Ap-2α is required for normal jaw patterning in mouse embryos (Schorle et al., 1996; Zhang et al., 1996), pharyngeal cartilage was examined in ap-2α MO embryos at 4 dpf. Characteristic cartilage defects were observed, most clearly in the bilateral ventral elements of pharyngeal arch two (p2), the serratohyals (Fig. 6). In ap-2α MO embryos, the serratohyals were reduced in size and pointed medially (instead of rostrally) (Fig. 6). More posterior arches (p3 – p7; cerratobranchials) were less affected but were variably reduced in size in strongly affected ap – 2α MO embryos (about 10% of embryos injected with either E2I2 or E3I3 morpholino).

Control of melanophore development by Ap-2α is both cell autonomous and non-autonomous

Because ap-2α is expressed in neural crest and encodes a transcription factor likely to bind the c-kit promoter, its function is presumably at least partially cell autonomous to melanophores. However, ap-2α is also expressed in the presumptive epidermis, so potentially, Ap-2α could regulate the expression of a gene in the skin that would influence melanophore development in a cell-non-autonomous fashion. To test these possibilities, mosaic embryos were created by transplanting cells at blastula stage between control embryos and ap-2α MO embryos. Melanophores that derived from control cells transplanted into control hosts appeared normal, and were indistinguishable in color and shape from host-derived melanophores (Figs. 5A – C). Unusually, we observed that melanophores derived from control cells transplanted into ap-2α MO host embryos were underpigmented and non-dendritic, just as were host-derived, morpholino-containing melanophores (Figs. 5D – F). If the activity of Ap-2α was entirely cell autonomous to melanophores, melanophores deriving from control cells should have been clearly distinct in size and color from host-derived melanophores in ap-2α MO embryos. This implies that Ap-2α has a cell-non-autonomous role in regulation of melanophore differentiation. In the converse experiment, melanophores that derived from ap-2α MO cells transplanted into control embryos were never detected (Fig. 5C), despite analysis of more than 100 such transplants. These results suggest that Ap-2α has both cell-autonomous and cell-non-autonomous roles in regulation of melanophore size and color.

ap-2α MO embryos

Fig. 6. Zebrafish ap-2α MO embryos have reduced neural crest derivatives. Lateral (A and B) and ventral (C and D) views of 4-dpf embryos processed to reveal craniofacial cartilage. (A and C) Uninjected embryo and (B and D) ap-2α MO embryo. In injected embryos, 41 of 110 exhibited characteristic abnormal craniofacial architecture. Hyosymplectic (hs) and ceratohyal (ch) are dorsal and ventral elements of the p2, respectively (see Schilling et al., 1996). In ap-2α MO embryos, the cerratohyals were reduced in size and abnormally shaped. The hyosymplectics were present but not as extended as in wild-type embryos. The ceratobranchials, derived from p3 – p7, were variably reduced in strongly affected embryos. p1 derivatives, Meckel’s cartilage (m) and palatoquadrate (pq), appeared somewhat reduced in size and in strongly affected embryos (10/110 embryos injected with E2I2 MO), pointed ventrally. These phenotypes are highly reminiscent of the lockjaw mutant phenotype (Schilling et al., 1996), except less severe. For instance, in lockjaw mutants embryos, ceratobranchial elements are absent, but they are merely smaller in ap-2α MO embryos.

The milder phenotype of ap-2α MO embryos suggests Ap-2α function is reduced but not eliminated in them. (E and F) Lateral view of embryos fixed at 4 dpf and processed to reveal anti-Hu-immunoreactivity. (E) In uninjected embryos, enteric neurons (e) dorsal root ganglion (drg) neurons are revealed by anti-Hu immunoreactivity. (F) In ap-2α MO embryos, enteric neurons (e) were highly reduced (anti-Hu-immunoreactive neurons in the section of gut overlying the hindyolk: uninjected embryos, 99.2 ± 13.2, ap-2α MO embryos, 22.2 ± 10.1, n = 6, P < 0.0001, Student’s t test). (G and H) Transverse sections, near axial level of pectoral fins, of 4-dpf embryos processed to reveal anti-tyrosine hydroxylase immunoreactivity (TH-IR). (G) In uninjected embryos, TH-IR ventral to notochord (n) is in sympathetic neurons, while TH-IR in more ventral and lateral position is in adrenal chromaffin cells (An et al., 2002) (arrowheads; average of 36 ± 5 sympathetic neurons, n = 5 embryos.) (H) In ap – 2α MO embryos, there was a significant reduction of TH-IR positive cells (average of 7 ± 1 TH-IR positive cells in ap-2α MO embryos that showed reduced numbers of melanophores at 30 hpf, n = 5, P < 0.005). All scale bars, 100 μm. Scale bar in B applies for both A and B, etc.
In mice, neural crest cells contribute to cardiac outflow tract development, and Ap-2α is required for outflow tract morphogenesis (Brewer et al., 2002). Recent studies in zebrafish have suggested that some cranial neural crest cells contribute to cardiac muscle and that ablation of neural crest cells leads to decreased contractility and aberrant cardiac morphology by 2 dpf (Li et al., 2003). However, in ap-2α MO embryos at 2 dpf, blood flow,
cardiac morphology, and heart rate were all normal (99 ± 7 bpm in controls, 97 ± 6 bpm in ap-2x MO embryos, n = 10, Student’s t test, P = 0.4). Starting at 3 dpf, ap-2x MO embryos exhibited pericardial edema (results not shown), as is typically found in zebrafish embryos with cardiovascular insufficiency. However, it is unclear whether edema arising at this stage results from a direct influence of Ap-2α on the heart or as a secondary consequence of abnormal aortic arch formation (Isogai et al., 2001) (e.g. p3 in Fig. 6B).

ap-2x MO embryos have reduced expression of cranial neural crest markers

Because multiple neural crest derivatives were abnormal in ap-2x MO embryos, a primary defect in neural crest was indicated. In addition, ap-2x is expressed in zebrafish neural crest (Furthauer et al., 1997; Nguyen et al., 1998), and inhibition of AP-2 in frog blocks expression of markers of premigratory neural crest (Luo et al., 2003). In contrast, in ap-2x MO embryos at 11 hpf, expression of two markers of

![Fig. 7. Zebrafish ap-2x MO embryos have reduced cranial neural crest cells. Lateral view of 24-hpf (A) uninjected embryo and (B) ap-2x MO embryo processed to reveal crestin expression, a marker of migrating neural crest at this stage (Luo et al., 2001). The intensity of crestin expression was much lower in individual cells of ap-2x MO embryos, and the number of crestin-expressing cells appeared to be reduced. (C and D) Lateral view of 20-hpf (C) uninjected embryo or (D) ap-2x MO embryo processed to reveal sox10 expression. Expression was normal, or very slightly reduced as in the embryo pictured here, in ap-2x MO embryos. (E and F) Lateral view of 24-hpf (E) uninjected embryo and (F) a strongly affected ap-2x MO embryo, processed to reveal dlx2 expression, a marker of neural crest migrating in pharyngeal arches at this stage (Akimenko et al., 1994). Relative to uninjected embryos, in a majority of ap-2x MO embryos, a clear reduction of dlx2-expressing cells was seen in p2 (arrowhead), and in strongly affected ap-2x MO embryos, a reduction of dlx2-expressing cells was also apparent in more caudal arches (8/90 embryos scored). The number of dlx2-expressing cells in p1 appeared normal in ap-2x MO embryos. All scale bars, 100 μm. Scale bar in B applies for both A and B, etc.](image-url)
premigratory neural crest, *foxd3* and *sox10*, was normal (results not shown; Dutton et al., 2001b; Odenthal and Nusslein-Volhard, 1998). In *ap-2* MO embryos at 20 and 24 hpf, expression of *sox10* in migrating neural crest marker appeared normal, although we note that this gene is not a marker of migrating neural crest in pharyngeal arches (Fig. 5 and not shown, Dutton et al., 2001a). Surprisingly, in contrast to *sox10*, expression of *crestin* was globally reduced at 24 hpf (Luo et al., 2001; Rubinstein et al., 2000).

At 24 hpf in *ap-2* MO embryos, abnormal expression of *dlx2*, a marker of cranial neural crest in pharyngeal arches (Akimenko et al., 1994), was detected. In majority of *ap-2* MO embryos, fewer cells of pharyngeal arch 2 (p2) expressed *dlx2*, and in strongly affected embryos, expression was also reduced in p2–p7 (Fig. 7). Expression of *dlx2* in p1 was relatively unaffected in all *ap-2* MO embryos (Fig. 7 and not shown). Thus, migrating neural crest is present in *ap-2* MO embryos, although its gene expression profile is abnormal and neural crest in p2–p7 may be reduced.

**Discussion**

*ap-2* MO embryos complete early development normally

Even at maximal doses, morpholinos targeting *ap-2* did not cause the gross defects in morphogenesis seen in mouse *Ap-2* mutants, which include failure of closure of cranial neural folds, or closure of the ventral body walls in mouse embryos (Schorle et al., 1996; Zhang et al., 1996). Moreover, in frog embryos injected with *AP-2* antisense oligonucleotides, there is an apparent absence of neural crest and a cessation of development shortly after gastrulation (Luo et al., 2003). In zebrafish, *ap-2* MO embryos migratory neural crest is clearly present, although its gene expression profile is altered, and cranial neural crest cells in P2–P7 may be reduced, as expression of *dlx2* is seen in fewer cells there. In zebrafish *ap-2* MO embryos, early roles for *AP-2* in morphogenesis and neural crest formation may be performed by residual *Ap-2* protein. Indeed, full-length *ap-2* transcripts that escaped the morpholino were always detected in RT-PCR experiments carried out on *ap-2* MO embryos. Alternatively, additional *Ap-2* homologues may function in early zebrafish development; a recent search of public sequence data revealed the presence of at least three additional *ap-2* homologues in the zebrafish genome (W.L. and R.A.C., unpublished observations, Sanger Consortium). Misexpression of dominant negative *Ap-2* proteins may be a more effective way to block all *Ap-2* function (see Buettner et al., 1993). Whatever the explanation for their normal early development, *ap-2* MO embryos are well suited for analysis of *Ap-2* function in neural crest derivatives, particularly in derivatives that arise after the time of death of mouse *Ap-2* mutant embryos.

*Ap-2α* regulates melanophore development via *C-kit* and other targets

Melanophore defects in *ap-2* MO embryos make sense in light of the suspected regulation of *C-KIT* by *Ap-2α* in human cells. *C-kit* encodes a receptor for the ligand Steel and is required for proliferation, survival, and perhaps differentiation of melanoblasts in mice (Ito et al., 1999; Kunisada et al., 1998; Steel et al., 1992). In zebrafish *c-kit* null mutants (i.e., *sparse*), melanophores are reduced in number and cluster near the dorsal neural tube (Parichy et al., 1999). In *ap-2* MO embryos, *c-kit* expression is reduced, trunk melanophores are reduced in number and in extent of migration. Importantly, our observation that a given dose of *ap-2* morpholino has a greater effect on melanophore cell number and migration in *c-kit* heterozygotes than in wild types is strong evidence that these two genes work in the same pathway in melanophores. The regulation of *c-kit* by *Ap-2α* is likely direct: the human *C-KIT* promoter contains *AP-2* consensus binding sites, and overexpression of *AP-2α* in melanoma cell lines increases *C-KIT* expression (Huang et al., 1998). However, *Ap-2α* appears to regulate melanophore number and migration via additional targets because injection of the *ap-2* morpholino further reduces melanophores in *c-kit* homozygous null mutant embryos. Moreover, in *ap-2* MO embryos at 28 hpf, melanophores are not as black as usual, indicating reduced melanin synthesis, and they appear smaller and less dendritic than normal. While activation of the *C-kit* pathway can stimulate differentiation of mouse melanoblasts (Kunisada et al., 1998), abnormal melanophore differentiation is not seen in zebrafish *c-kit* mutant embryos (Parichy et al., 1999), again suggesting other or additional effectors of melanophore differentiation by *Ap-2α*.

What other targets of *Ap-2α* regulate melanophores? Candidates include other *c-kit* orthologues, although *c-fms*, a *c-kit* homologue required by embryonic xanthophores, was normal in *ap-2* MO embryos, showing that not all *c-kit* homologues depend on *Ap-2α*. A reduction of melanin is consistent with reduced levels of *dct* expression in *ap-2α* MO embryos. The regulation of *dct* may be mediated by Melanocyte-Specific Gene 1 (MSG1), which contains potential binding sites for *Ap-2α* in its promoter (Fenner et al., 1998) and which upon overexpression in melanoma cells leads to expression of *tyrosinase* and *dopachrome-tautomerase* (*DCT*), and increased melanin (Nair et al., 2001). Other potential *Ap-2α* targets include components of the pathway stimulated by the Endothelin receptor type b (*Ednrb*). As in *ap-2α* MO embryos, mouse embryos homozygous for a targeted deletion of *Ednrb* have reduced numbers of melanocytes (Hosoda et al., 1994; Lee et al., 2003; Shin et al., 1999) and *Ednrb* signaling stimulates proliferation and perhaps differentiation of mammalian melanoblasts (Lahav et al., 1996). Furthermore, *Ednrb* function is also known to be required for correct migration of enteric neurons (Lee et al., 2003). Finally, both *Ap-2α*
and Ednrb appear to repress differentiation of Schwann cells (Brennan et al., 2000; Stewart et al., 2001). In zebrafish, only one ednrb orthologue has been described to date, and it is expressed in embryonic melanoblasts. Mutations in this gene lead to a reduction of adult melanophores, however, embryonic melanophores and enteric neurons appear normal (Parinchy et al., 2000a). Therefore, it is most likely that a separate Ednrb orthologue functions in these cell types. Nonetheless, the observation that melanoblast expression of ednrb1 is normal in residual melanoblasts in ap-2α MO embryos suggests that any interaction between Ednrb and Ap-2α pathways is not at the level of receptor expression. Because the role of Ap-2α is partially cell non-autonomous, one possibility is that Ap-2α regulates expression of the ligand endothelin-3.

**Cell autonomy of Ap-2α function in melanophores**

A cell-non-autonomous role for Ap-2 in melanophore development was unexpected, but is consistent with essential roles for Ap-2 homologues in epidermal development. Thus, melanophores derived from control cells transplanted into ap-2α MO embryos did not have a wild-type appearance, but instead were small and undertpigmented. This result could not be explained by a global delay of development caused by the morpholino, because the phenotype of small, pale melanophores was observed in closely stage-matched embryos, as judged by the development of the eye and progression of the lateral line primordium. This result implies that Ap-2α function is required in another tissue in addition to melanophores to support their normal differentiation. Melanophores are closely apposed to skin. In *X. laevis*, reduction of AP-2 function appears to cause epidermal precursors to adopt a neural fate, implying a very early role for AP-2 in specification or maintenance of the epidermal fate (Luo et al., 2002). In human skin, nuclear expression of Ap-2α is highest in the basal layer of keratinocytes, suggesting it regulates their proliferation, or perhaps limit their differentiation (Mazina et al., 2001; Oyama et al., 2002). Ap-2γ is expressed in all layers of epidermal cells, and may also be important for keratinocyte differentiation (Oyama et al., 2002). It will be important to assess epidermal development in ap-2α MO embryos to address this hypothesis.

Ap-2α is also expected to be required within melanoblasts, for instance to activate the c-kit promoter. However, it is unclear why we were unable to detect labeled melanophores in ap-2α MO-to-control transplants, despite scoring enough such transplants that we predicted seeing them in about 30 embryos. Perhaps the melanophores containing the ap-2α MO were delayed or hidden by the large, black melanoblasts of the host. An intriguing alternative explanation is that neural crest cells or melanoblasts inheriting ap-2α MO succumb to death signals, or signals that induce them to adopt a different fate, which are present in control hosts but are reduced in an ap-2α MO hosts.

**Other neural crest derivatives depend on Ap-2α**

Defects in several other neural crest derivatives were detected in ap-2α MO embryos, suggesting zebrafish may be a useful model for analysis of neurocraniopathies. ap-2α MO embryos like mouse *Ap-2α* mutants have an abnormal pharyngeal skeleton. Elements of p2 were most sensitive to the ap-2α morpholino, with more caudal arches reduced in strongly affected embryos, implying a quantitative difference in dependence on Ap-2α function in these structures. Derivatives of p1, Meckel’s cartilage and the palatoquadrate, were relatively normal in ap-2α MO embryos (Fig. 6). These elements were also normally patterned, although slightly reduced in size, in homozygous *lockjaw* mutant embryos, which harbor null alleles of *ap-2α* (Schilling et al., 1996; T. Schilling, personal communication). Thus, Ap-2α function may be limited to neural crest contributing to p2 and more posterior arches. Consistent with this model, in mouse embryos, p2 is the anterior limit of *hoxa2* expression, a gene that is activated by Ap-2α (Hunter and Prince, 2002; MacCormochie et al., 1999). Indeed, the role of Ap-2α in p2 elements may be largely or entirely mediated by its effect on hox paraglogue group 2 genes (*hoxa2* and *hoxb2*), because p2 elements are abnormal in embryos in which these genes have been inhibited with morpholinos (*hox pg2* morphants, see Hunter and Prince, 2002). Ap-2α may also interact with more posteriorly expressed Hox genes, because p3–p7 derivatives, which are relatively normal in *hox* pg2 MO embryos, are clearly abnormal in severely affected *ap-2α* MO embryos and *lockjaw* mutant embryos. Because the hyoid arch was the most affected in *ap-2α* MO embryos, it is notable in the mouse *Ap-2α* mutant, the hyoid, which derives from pharyngeal arches 2 and 3, was reported to be normal in mouse mutants (Zhang et al., 1996). The mouse phenotype was reported to include an absence of the stapes bone (p2 derived) but the presence of the mandible (p1 derived) (Schorle et al., 1996; Zhang et al., 1996). It is unclear in mouse *Ap-2α* mutants whether defects in pharyngeal skeleton reflect a direct dependence of cranial neural crest on Ap-2α, or whether they are secondary to defects in closure of cranial neural folds (Schorle et al., 1996; Zhang et al., 1996). In zebrafish *ap-2α* MO embryos, changes in *dlx2* expression predicted the changes in pharyngeal skeleton, implying a direct role for Ap-2α in neural crest patterning. These results are interesting because human *Ap-2α* is located on chromosome 6p24, a region that has been linked to nonsyndromic cleft lip and palate in many studies (Carinci et al., 1995; Prescott et al., 2000; Scapoli et al., 1997).

Analysis of *ap-2α* MO embryos revealed a requirement for Ap-2α function in autonomic neurons of sympathetic and enteric ganglia. Most of the sympathetic ganglion chain derives from trunk neural crest, but the first detectable sympathetic neurons in zebrafish are rostral ones that likely originate in vagal (hindbrain) neural crest (An et al., 2002). These cells express Tyrosine Hydroxylase (TH), an enzyme required for synthesis of catecholamines.
noreactivity was reduced in the normal vicinity of sympathetic neurons in ap-2α MO embryos. Ap-2α has been shown to directly bind the regulatory regions of genes encoding TH and dopamine β-hydroxylase, another enzyme involved in the biosynthesis of catecholamines (Greco et al., 1995; Kim et al., 2001, 1998; Yang et al., 1998). In addition, ErbB2, which encodes a receptor for the growth factor Neuregulin-1, is activated by Ap-2α (Bosher et al., 1995), and ErbB2 mutants suffer severe hypoplasia of the sympathetic ganglion chain, apparently from a failure of neural crest migration (Britsch et al., 1998). These observations suggest Ap-2α regulates the multiple steps of sympathetic neuron development. In addition, enteric neurons, which also derive from vagal neural crest, were virtually eliminated from the hindgut of strongly affected ap-2α MO embryos. It is unclear what step in the development of enteric neurons (i.e., specification, differentiation, survival, or migration) depends on Ap-2α. This result was not anticipated by previous work, but is of interest in light of congenital defects of the enteric nervous system, such as Hirschsprung’s disease, a multifactorial disorder characterized by aganglionic colon (OMIM 142623, reviewed in Passarge, 2002) or Waardenburg–Shah syndrome, characterized by pigment and enteric ganglia abnormalities (OMIM 277580, Shim et al., 1999). Indeed, it is possible that mutations in AP-2α confer susceptibility to these diseases.

In conclusion, because they complete early development relatively normally, analysis of zebrafish ap-2α MO embryos has revealed previously unrecognized roles for Ap-2α in neural crest patterning. The zebrafish model should be useful in the exploration of the roles the Ap-2α plays in normal development of neural crest derivatives, and possible roles for this protein in diseases that afflict neural crest derivatives, including malignant melanoma, Hirschsprung’s disease, and cleft lip and palate.

Acknowledgments

We are grateful to Tom Schilling for sharing data before publication. We appreciate excellent technical assistance from Saxon McCloud and Jennifer Paulson, and the gift of sparse homozygotes from Stephen Johnson. This work was supported by NIH grant HD22486 to J.S.E. and a Carver Foundation seed grant to R.A.C. C. d., and M.A. were supported by grants ICM P99-137-f and Fondecyt 1031003. E.K.O. was supported by Grant T32 DC00040 (Bruce Gantz, PI).

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Electronic resources