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Recombinant Plants Provide a New Approach to the Production of Bacterial Polysaccharide for Vaccines

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Abstract

Bacterial polysaccharides have numerous clinical or industrial uses. Recombinant plants could offer the possibility of producing bacterial polysaccharides on a large scale and free of contaminating bacterial toxins and antigens. We investigated the feasibility of this proposal by cloning and expressing the gene for the type 3 synthase (cps335) of Streptococcus pneumoniae in Nicotiniana tabacum, using the pCambia2301 vector and Agrobacterium tumefaciens-mediated gene transfer. In planta the recombinant synthase polymerised plant-derived UDP-glucose and UDP-glucuronic acid to form type 3 polysaccharide. Expression of the cps35 gene was detected by RT-PCR and production of the pneumococcal polysaccharide was detected in tobacco leaf extracts by double immunodiffusion, Western blotting and high-voltage paper electrophoresis. Because it is used a component of anti-pneumococcal vaccines, the immunogenicity of the plant-derived type 3 polysaccharide was tested. Mice immunised with extracts from recombinant plants were protected from challenge with a lethal dose of pneumococci in a model of pneumonia and the immunised mice had significantly elevated levels of serum anti-pneumococcal polysaccharide antibodies. This study provides the proof of the principle that bacterial polysaccharide can be successfully synthesised in plants and that these recombinant polysaccharides could be used as vaccines to protect against life-threatening infections.

Introduction

Polysaccharide encapsulated bacteria are major causes of disease and death in humans and animals. For example, diseases caused by Streptococcus pneumoniae (the pneumococcus), Neisseria meningitidis and Haemophilus influenzae are responsible for more than two million deaths every year, the majority children under the age of five [1,2][1,2]. Streptococcus pneumoniae alone is responsible for more than 50 percent of invasive disease worldwide. Despite the extensive use of pneumococcal vaccines, incidences of disease caused by S. pneumoniae remain high, mainly due to serotypes not included in the vaccine [3]. Current anti-pneumococcal vaccines are composed of capsular polysaccharide alone or conjugated to protein. Whatever the formulation, pneumococcal vaccine design has to deal with the facts that there are over 90 different capsular and the serotype distribution varies with time and geography. However, for reasons of economics and biology the current vaccines are limited in coverage (23 in the polysaccharide-only vaccine and 13 in the new version of the conjugate) to the most dominant serotypes in Europe and North America. Ideally multiple versions of these vaccines are required and they would be regularly reformulated to offer maximum protection. Cost of polysaccharide production then becomes a concern. One of the challenges for pneumococcal vaccine production is to manufacture bacterial polysaccharide on a large-scale, without need for purification procedures to remove contaminating toxins and pyrogens. Currently the preparation of polysaccharides requires expensive fermentation equipment, microbiological containment and high levels of quality control to prevent contamination. Plants offer a solution because they synthesise a large number of high molecular weight polysaccharides, they have many of the sugar precursors of bacterial capsular polysaccharide readily available and plants have compartmentalised metabolic pathways and transport processes that could facilitate polysaccharide extraction [4]. However, until now heterologous antigen production in plants has been limited to the production of proteins [5,6,7]. Here we report that plants can be engineered to synthesise bacterial polysaccharides and these polysaccharides provide protective immunity. We demonstrated this principle using the serotype 3 capsular polysaccharide of S. pneumoniae, a serotype that is
The pneumococcal cps3S gene {Dillard, 1995 #888} was amplified from genomic DNA of the pneumococcal type 3 strain WU2 using primers CPSFOR and CPSREV and cloned with PR1b signal sequence (which was used to direct secretion of the transgene to the apoplast) into the Agrobacterium binary vector pCambia 2301, to give pCMS4. This placed cps3S under the control of duplicated constitutive cauliflower mosaic virus promoters, CaMV35S and also enabled selection of transformed plants with kanamycin. Nucleotide sequence analysis of the cloned cps3S in pCMS4 showed 100 % identity with the published sequence [8].

Nicotiana tabacum was transformed with pCMS4 by A. tumefaciens-mediated gene transfer. A T1 generation was grown from the seeds of six plants and PCR showed that four plants contained the 1.3kb cps3S gene (Figure 1A). PCR also confirmed the absence of contaminating Agrobacterium DNA (Figure 1B). RT-PCR, with cps3S-specific primers, showed that the transgene was expressed in the transgenic plants (Figure 2). No amplicon was generated by direct PCR amplification of RNA extracts, confirming the absence of contaminating cps3S DNA (Figure 2A). No amplicon was generated by RT-PCR of untransformed plants (Figure 2A lane 3).

A second generation of plants were grown from the seeds of these plants and PCR confirmed stable transgene expression (Figure S1). All subsequent assays were done with second generation (T2) plant material.

Results

The Pneumococcal Type 3 Capsule Synthase Gene was Cloned into Nicotinia tabacum by Agrobacterium-mediated Gene Transfer

The pneumococcal cps3S gene was amplified from genomic DNA of the pneumococcal type 3 strain WU2 using primers CPSFOR and CPSREV and cloned with PR1b signal sequence (which was used to direct secretion of the transgene to the apoplast) into the Agrobacterium binary vector pCambia 2301, to give pCMS4. This placed cps3S under the control of duplicated constitutive cauliflower mosaic virus promoters, CaMV35S and also enabled selection of transformed plants with kanamycin. Nucleotide sequence analysis of the cloned cps3S in pCMS4 showed 100 % identity with the published sequence [8].

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A second generation of plants were grown from the seeds of these plants and PCR confirmed stable transgene expression (Figure S1). All subsequent assays were done with second generation (T2) plant material.

Pneumococcal Type 3 Polysaccharide was Detected in the Leaves of Transformed Plants

Double immunodiffusion showed that type 3 antibody-antigen complexes were seen (Figure 2B) between wells which contain purified type 3 pneumococcal polysaccharide, sonicated plant cell extract from transgenic plants (wells 1-4) and type 3 polysaccharide specific antiserum (well A). This was not seen in wells containing extract from a wildtype tobacco plants (wells 5 and 6).

Western blotting of transgenic and wildtype plant extracts using type 3 polysaccharide specific antiserum also showed the presence of type 3 polysaccharide in transgenic plant extracts only (Figure 2C). High-voltage paper electrophoresis of hot-acid-hydrolysates of cold-acid-extractable tobacco leaf polysaccharides confirmed these findings (Figure 2D). Acid hydrolysis of polysaccharides from transgenic leaves produced a relatively hot-acid-resistant, singly-ionised disaccharide with the same mobility as the β-d-GlcA-(1→4)-d-Glc seen following acid hydrolysis of pneumococcal type 3 polysaccharide. This disaccharide was barely detectable in wild type non-transformed plants (Figure 2D).

Immunisation with Transgenic Plant Extracts Protected Mice from Pneumococcal Disease

To test the immunogenicity of the plant-derived type 3 polysaccharide, mice were immunised with three doses of apoplast extracts from transgenic or wildtype plants. Sera were collected on the day before each immunisation and ten days after the final dose, and anti-type 3 polysaccharide IgG was determined by ELISA. Significantly more (P<0.05) specific anti-type 3 antibody was detected after a single dose of the transgenic leaf extract, with a further increase (P<0.05) after a second dose (Figure 3A), whereas antibody levels remained unchanged in those given wildtype extracts (P>0.05).

Mice were challenged intranasally with the serotype 3 S. pneumoniae strain HB565 230 days after the final immunisation. Mice immunised with transgenic plant extract survived significantly longer (P<0.001) than those given wildtype extracts (mean survival: 181±23h for transgenic and wildtype, respectively). Mice immunised with the wildtype extract did not survive longer (P>0.05) than sham-immunised mice (91±38h). None of the fifteen animals given wildtype extract were alive ten

Figure 1. Detection of the cps3S gene in transformed tobacco plants. A. DNA was used as a template for PCR (Lanes 2, 3: wild type plants; Lanes 4 – 7: transformed plants) using cps3S-specific primers. PCR products were analysed by agarose gel electrophoresis. The results show the presence of the cps3S gene in the transformed plants (Lanes 4 – 7) but not the wild type plants. The PCR reaction in Lane 9 contained purified plasmid DNA containing cps3S (pCMS4) as a positive control and Lane 8 contained no template DNA. Molecular sizes are indicated. B. PCR showing the absence of Agrobacterium DNA contaminating DNA preparations from wild type (Lanes 2, 3) and transformed (Lanes 4 - 7) tobacco plants. PCR was done with Agrobacterium-specific primers. The results show that there was no Agrobacterium DNA present in the transgenic plant samples. The PCR reaction in Lane 9 contained Agrobacterium DNA as a positive control and shows the expected 730bp band and Lane 8 contained no template. Molecular sizes are indicated. DNA was extracted from the same six N. tabacum plants for the PCRs shown in A and B.

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days after the challenge, whereas eight of the fourteen immunised with transgenic extract survived (Figure 3B).

**Discussion**

This study has shown that bacterial polysaccharide vaccine antigens can be synthesised in plants and that simple extracts of these plants are immunogenic and protect against an otherwise
gene into cps3S type 3 synthase (Cps3S). Therefore, we cloned the pneumococcal these substrates into type 3 polysaccharide requires the enzyme, for cell wall polysaccharide synthesis [4,10]. Polymerisation of which transport them into the endomembrane system as substrates glucuronic acid (UDP-GlcA) are naturally synthesised by plants, precursors, UDP-glucose (UDP-Glc) and UDP-glucuronic acid (UDP-GlcA) are naturally synthesised by plants, which transport them into the endomembrane system as substrates for cell wall polysaccharide synthesis [4,10]. Polymerisation of these substrates into type 3 polysaccharide requires the enzyme, type 3 synthase (Cps3S). Therefore, we cloned the pneumococcal cps3S gene into N. tabacum using pCambia 2301. This strategy not only placed cps3S under the control of duplicated constitutive cauliflower mosaic virus promoters, CaMV35S, but it also enabled selection of transformed plants with kanamycin. Growth of a second generation of kanamycin-resistant plants confirmed stable transgene expression. Although not the primary purpose of the study, we did a limited investigation of how to extract the pneumococcal polysaccharide from plant tissue. The method that yielded the highest concentration of pneumococcal polysaccharide was to grind the plant tissue under liquid nitrogen, suspend the tissue in water and lyse the cells by sonication. Despite cloning the signal sequence PR1b we detected no type 3 polysaccharide in its destination, the apoplastic fluid. This implied that PR1b was not functioning correctly. Previous studies replaced the start codon of the transgene with PR1b [3], however, we maintained the start codon and cloned an in-frame sequence of cps3S. This may have led to a reduction in PR1b activity and improving this may increase the yield of type 3 polysaccharide. Another method to increase yield is to use root tissues, since the continuously growing primary cell wall may contain higher concentrations of the UDP-precurors. In this study we focussed on leaf tissue as we were working with parent and F1 generations and removal of the roots may have restricted growth of the plant and seed development. Leaf tissue was also much easier to obtain. For these reasons, the levels of polysaccharide extracted from plant tissue may not have been optimal.

Having shown the principle of in planta synthesis using the linear type 3 polysaccharide, the next challenge is the production of more complex, branched, bacterial polysaccharides. All the genes involved in pneumococcal capsular polysaccharide synthesis are closely linked on the bacterial chromosome, arranged within a single locus (a “type specific” cassette). Therefore, it is possible that the introduction of whole cassettes could lead to the synthesis of sugar precursors not naturally occurring in the plant and their assembly into more complex polysaccharides. Furthermore, effective signal or transport peptides should allow easier extraction by compartmentalising different polysaccharides.

Because anti-polysaccharide antibodies are protective against several bacterial pathogens of humans and animals there is great interest in polysaccharides as vaccines. However, some of the problems with these vaccines are illustrated by vaccines against S. pneumoniae. The current pneumococcal vaccine contains twenty-three polysaccharide serotypes but protection is serotype-specific and some are not immunogenic in children. The vaccine was formulated on the prevalence of serotypes in North America and Europe, but elsewhere the coverage can be considerably less [11]. In addition to protection being serotype-specific, polysaccharide immunogenicity also varies with serotype and age [12]. Furthermore, temporal variation occurs in the serotypes isolated from adults and children [13]. Thus it has been suggested that different vaccine formulations should be manufactured for differing situations [14], but unless low-cost solutions are found it will not become a reality. Polysaccharide vaccines can be expensive, which restrains their use in developing countries. Production of polysaccharide vaccines in plants can introduce economies of scale that can drive down the production costs. The alternative, of using microorganisms as the vaccine production system, requires expensive fermentation equipment and high levels of quality control to prevent contamination. In contrast, the use of plants for vaccine production offers an achievable solution, opening the possibility of local production, which increases the likelihood of adoption of the vaccine [15].

Efforts to improve the poor immunogenicity of polysaccharide vaccines in the young are focused on the development of polysaccharides covalently linked to protein, but these make difficulties with serotype coverage worse. When the US FDA licensed a 7-valent anti-pneumococcal conjugate vaccine (Prevnar) the serotypes covered by the vaccine caused 90% of disease in North America and Europe, but less than 70% in Asia [15]. This emphasises the desirability of a “tailor-made” vaccine. However, formulations of conjugates for a particular country or for
childhood and for adult vaccination programmes are even less likely than pure polysaccharide formulations, unless cheaper production methods can be found. Conjugate vaccines are very expensive and the pneumococcal conjugates will be more expensive than any other current vaccine. Even with tiered pricing the price of these vaccines is a real concern for their uptake [16]. The ability to produce polysaccharide vaccines in plants, on a large scale, could lead to a ready availability of polysaccharides for protein conjugate vaccine production. There is, however, a more exciting possibility, namely to exploit the plant’s glycosylation machinery to glycosylate heterologous proteins with heterologous polysaccharide to make conjugate vaccines in planta. Plants making heterologous immunogen represent an innovative technology for the development of childhood vaccines. The longer-term objective of this research is to synthesise polysaccharide-protein conjugates in plants.

In planta synthesis of bacterial polysaccharide and conjugates offers an innovative contribution to vaccinology. Synthesising heterologous polysaccharides in plants represents the first proof of concept step in this process. These experiments have yielded exciting data even though no attempt was made to optimise gene expression, polysaccharide purification or immunising protocol. These are developmental issues for the future now that the concept that bacterial polysaccharide can be synthesised in plants has been proven.

### Materials and Methods

#### Construction of the Plant Expression Vector

The sequence of the type 3 pneumococcal capsular polysaccharide biosynthesis cassette was obtained from GenBank (www.ncbi.nlm.nih.gov), accession number U15171. A 1.3 kb DNA fragment containing the cps3S gene was obtained by PCR from the genomic DNA of S. pneumoniae serotype 3 (WU2) using the oligonucleotide primers sense (CPSFOR) 5′-CTG GTA QCC ATG TAT ACA TTT ATT TTA ATG TTG TTG G-3′ corresponding to 2227bp - 2254bp with a KpnI restriction site inserted at the 5′ end; anti-sense (CPSREV) 5′-TCA TCA CTC TGT TAA ATT CCT AGT TCC -3′ corresponding to 3454bp - 3479bp of the cassette. The PCR amplification was performed in a total volume of 50 μl. using 2 μg genomic template under the following conditions: 94°C for 4 minutes and then a cycling procedure comprising denaturation at 94°C for 45 seconds, annealing at 58°C for 1 minute, and extension at 72°C for 1 minute 30 seconds, which was repeated either 10 or 30 times, and a final extension at 72°C for 10 minutes. Amplified DNA resulting from PCR were purified from the PCR reagents using the QIAquick PCR purification kit (Qiagen). The amplified fragment was inserted into the multiple cloning site of pCR4-TOPO (Invitrogen, Carlsbad, CA, USA). The Agrobacterium binary vector pPZP222 (GenBank U10463) [17] has been previously engineered to produce vector pCHF2 (Figure 4) containing the constitutively expressed cauliflower mosaic virus (CaMV) 35S promoter and a...
rbcS terminator (C. Fankhauser, personal communication). Here, synthetic primers homologous to the PRb1 signal peptide sequence [19] were annealed, phosphorylated and inserted into pCHF2 using SacI and KpnI cutting sites situated between the CaMV 35S promoter and the rbcS terminator. The cps3S gene was removed from the pCRI-TOPO vector using KpnI and PsI and inserted between the PRb1 signal sequence, and the rbcS terminator sequence, to form the clone pCMS4. The entire expression cassette was then excised from pCMS3 with EcoRI and HindIII and ligated into the binary plant vector pCAMBIA2301 (GenBank accession number AF234316) (containing a kanamycin-resistance gene and gus) to give the resulting plasmid, pCMS4.

Transformation of Plants

pCMS4 was introduced into Agrobacterium tumefaciens strain GV3101 directly by the heat shock method, using 0.5 μl (1 μg) pCMS4 and 0.1 ml of frozen CaCl2-competent A. tumefaciens cells. Cells were thawed at 37°C for 5 minutes, re-suspended in 1 ml of YEP broth (10 g/L Yeast Extract, 10 g/L Peptone, 5 g/L NaCl, pH 7.0) and incubated at 28°C for 2 hours with gentle shaking. Cells were harvested by centrifugation at 600 g for 5 minutes, the pellet re-suspended in 0.1 ml YEP and spread onto YEP agar containing 100 μg/ml kanamycin and 100 μg/ml rifampicin and incubated overnight at 28°C. Subsequently, A. tumefaciens carrying pCMS4 was used to transform tobacco (Nicotiana tabacum cv. SR1) leaf discs, as described previously [19]. Leaf discs were then transferred to Murashige-Skoog (MS) agar containing 3% (w/v) sucrose, the plate was sealed with parafilm and incubated in the humidity box for 1-2 weeks. Precipitin lines were observed and photographed in indirect light. Recombinant pneumococcal polysaccharides were estimated by comparison with the intensity of the precipitin lines of the positive controls.

Detection of cps3S Gene Expression in Transgenic Plants

Total RNA was isolated from 100 mg of leaf tissue using a RNA isolation kit (RNasy Plant Mini Kit; Qiagen, Surrey, UK) and used in the production of first strand cDNA using a cDNA synthesis kit (RNase H− reverse transcriptase kit; Invitrogen) and a random hexa-nucleotide primer. PCR was then performed using the CPSFOR and CPSREV primers as described above, RNA extracts without prior reverse transcriptase treatment were used as a control to indicate the presence of cps3S specific DNA. RNA extracted from wild-type tobacco leaves also was used as negative control. To confirm the absence of contaminating Agrobacterium DNA PCR was done with the primers VCF (5′-ATG ATT TGT AGC GAC T-3′) and VCR (5′-AGC TCA AAC CTG CTT C-3′), designed to amplify a 730bp region of the vcr gene of Agrobacterium Ti and Ri plasmids [20].

Preparation of Leaf Extracts

Leaves were collected from tobacco plants, tissue ground under liquid nitrogen and nanopure water added to give 0.5 g/ml plant tissue. The cells were lysed by sonication: 6 x 30 second sonication at an amplitude of 50 microns with 30 second rests in between sonication. The cells lysate then was centrifuged for 5 minutes at 10,000 g and the supernatant divided into 1 ml volumes, lyophilised and stored at 4°C.

Extraction of type 3 Polysaccharide from Apoplastic Fluid

A modification of the method described by Fry and co-workers was used [21]. Leaf material (1 g) was added to 50 ml of 50 mM CaCl2 and vacuum-infiltrated for a period of 30 minutes. The leaves were removed and dried gently on a paper towel before being transferred to the barrel of a 25 ml syringe with the plunger removed. This was placed in a 50 ml Falcon tube and the assembly centrifuged at 900 g at 10°C for 10 minutes. The aqueous extract was stored at 4°C until required.

Double Immunodiffusion

A modification Ouchterlony’s method was used [22]. 0.2% (w/v) Ouchterlony agarose in barbitone buffer (1.84 g/l diethylbarbituric acid, 10.3 g/l sodium diethylbarbiturate, pH 8.6) was used to coat microscope slides. These were left to dry for 1 hour, and then overlaid with 4.5 ml 1% (w/v) agarose in barbitone buffer. Once set, 4 mm holes were cut and 20 μl of sample was placed in the outer holes. Type 3 polysaccharide from S. pneumoniae (ATCC), diluted in 20 μl of untransformed plant extract, was used as a positive control. The central hole contained 20 μl neat rabbit anti-type 3 polysaccharide antiserum (Statens Serum Institute, Copenhagen, Denmark). The slides were incubated at 4°C in a humidity box for 1-2 weeks. Precipitin lines were observed and photographed in indirect light. Recombinant pneumococcal polysaccharides were estimated by comparison with the intensity of the precipitin lines of the positive controls.

High-Voltage Paper Electrophoresis (HVPE)

Leaf material was harvested, washed, cut into pieces and ground into a fine powder under liquid nitrogen. Samples were stored at –20°C until required. To 10 g fresh weight was added 50 ml 5% (v/v) formic acid and the suspension was incubated with gentle shaking for 2 days at room temperature. This procedure is expected to extract the capsular polysaccharide, but only a small proportion of the leaf cell-wall polysaccharides and starch. The homogenate was filtered through Miracloth and rinsed with 25 ml water and then the combined filtrate was adjusted to pH 4.0 with pyridine. Co-extracted proteins were denatured at 100°C for 60 min; the supernatant was freeze-dried. The dried material was washed exhaustively at room temperature in several changes of 82.6% (v/v) ethanol, which dissolves low-MW sugars. The remaining insoluble, polysaccharide-rich material was then air-dried, incubated at 90°C for 30 min in 22.6 ml water and cooled; trifluoroacetic acid (TFA) was then added to a final concentration of 0.36 M, which solubilised the polysaccharide. For partial hydrolysis of the polysaccharide to yield the relatively acid-resistant, diagnostic dimer (aldobioseuronic acid; GlcA–Glc) a portion of the solution was hydrolysed in 2 M TFA at 120°C for 30 min [conditions optimised in preliminary runs with authentic type 3 polysaccharide; data not shown], then dried in vacuo. The hydrolys products were redissolved in water containing a trace of Orange G (internal anionic marker), and a volume (equivalent to 32 mg fresh weight of leaf) was spotted on to Whatman 3 MM paper. The samples were subjected to HVPE at pH 6.5, at 3.0 kV for 60 min [23] and then stained with silver nitrate [21] to reveal sugars. External markers, run on the same sheet, included hydrolysates of (i) purified Type 3 polysaccharide (yielding glucose plus GlcA–Glc), and (ii) the trimer α-1,3-mannosyl-(1→4)-α-2-glucuronosyl-(1→2)-myo-inositol [24] (which yields a...
comparable dimer, α-α-glucuronyl-[1→2]-myo-inositol, plus mannose. Other markers were commercial glucose and galacturonic acid. After staining with silver nitrate [21], electrophoretograms were scanned and relevant spots were quantified for grey density in PhotoShop, as described in Supplementary Figure 1 of Parsons et al [25].

Immunisation and Challenge
Nine-week-old MF1 female mice (HarlanOlac, Bicester, UK) were given three doses of control plant extract or plant extract containing 2 μg plant-derived pneumococcal polysaccharide per mouse (as estimated by the Ouchterlony method) in 67 μl PBS and 33 μl Imject alum adjuvant (Pierce, Rockford, IL, USA). Mice were immunised intraperitoneally on days 0, 10, 20 and 30. Sham-immunised mice received alum adjuvant containing an irrelevant immunogen (KLH) using the same schedule. Serum samples were obtained by tail bleeding the day before each immunisation. Mice were challenged intraperitoneally with 2.8×10⁶ cfu serotype 3 pneumococci on Day 260. The health status of animals was monitored, according to the scheme of Morton et al [26]. These experiments were done under a project licence from the UK Home Office.

ELISA
Maxisorb ELISA wells (Gibco BRL, Nunc products) were coated with 2 μg/ml purified type 3 pneumococcal polysaccharide (ATCC) in coating buffer (50 nM NaHCO₃ pH 9.6, 0.02% w/v NaNO₃) for 16 h at 22°C. After rinsing with PBS, the wells were blocked with PBS + 5% w/v dried milk at 37°C for 1 h and washed three times with washing buffer (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.05% w/v Tween20). Mouse sera were diluted 1:100 in blocking buffer and 100 μl added to the wells and incubated, shaking, for 2 h at 37°C. The plates were washed three times as before and bound antibodies were detected using alkaline phosphatase-conjugated goat anti-mouse IgG secondary antibody (Fc specific, Sigma; diluted 1:5000) and 1 mg/ml p-nitrophenyl phosphate (Sigma) dissolved in 1 M diethanolamine pH 9.8, 0.5 M MgCl₂. Absorbance was read at 405 nm after 1 hour at 37°C and IgG concentration determined by reference to a standard curve prepared with murine IgG (Statens Serum Institute).

Western blot analysis
Western blotting was performed on leaf extracts as described previously [27].

Statistical analysis
Statistical analysis was performed using GraphPad Prism 5 (GraphPad, San Diego, CA, USA). The differences in antibody titres from mice immunised with transgenic or wildtype plant extracts were analysed using a student’s T-test. Survival data were analysed by the Kaplan-Meier survival curve analysis.

Supporting Information

Figure S1 Confirmation of the stable transformation of tobacco plants with the cps3S gene. Plant RNA was used as a template for reverse transcriptase PCR (RT-PCR) (Lane 1: wild type plants; Lane 2: transformed plants) using cps3S-specific primers. RT-PCR products were analysed by agarose gel electrophoresis. The results show the presence of the cps3S gene in the transformed plants (Lane 2) but not the wild type plants.

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Author Contributions
Conceived and designed the experiments: CMS SCF KCG ISR PWA. Performed the experiments: CMS SCF AJFP SG ISR PWA. Analyzed the data: CMS SCF SG ISR. Contributed reagents/materials/analysis tools: SCF KCG ISR. Wrote the paper: CMS SCF ISR PWA.

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