



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Rodent genetic models of neurodevelopmental disorders and epilepsy

Citation for published version:

Gonzalez-Sulser, A 2020, 'Rodent genetic models of neurodevelopmental disorders and epilepsy', *European Journal of Paediatric Neurology*, vol. 24, pp. 66-69. <https://doi.org/10.1016/j.ejpn.2019.12.012>

Digital Object Identifier (DOI):

[10.1016/j.ejpn.2019.12.012](https://doi.org/10.1016/j.ejpn.2019.12.012)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

European Journal of Paediatric Neurology

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Title: Rodent Genetic Models of Neurodevelopmental Disorders and Epilepsy

Authors: Alfredo Gonzalez-Sulser¹

Affiliations: Centre for Discovery Brain Sciences and Simons Initiative for the Developing Brain, University of Edinburgh¹.

Corresponding author at all stages: Alfredo Gonzalez-Sulser¹ – agonzal2@ed.ac.uk

Permanent Address: Centre for Discovery Brain Sciences, University of Edinburgh

1 George Square, EH8 9JZ, Edinburgh, United Kingdom.

Declaration of Interest: none.

Funding: This work was supported by Epilepsy Research UK and the Simons Initiative for the Developing Brain.

Abstract: Neurodevelopmental disorders (NDDs) are characterized by cognitive, social and motor deficits and are highly comorbid with intractable epilepsies. Through advances in genetic sequencing technologies a vast number of genes have been implicated in NDDs. State-of-the-art gene-editing techniques have led to the generation of hundreds of mouse models of NDDs. As an example, rodent models of Rett and Dravet syndromes as well as the syndromes caused by mutations in *CDKL5* and *Syngap1* display cognitive deficits in conjunction with seizure phenotypes. These models allow researchers to understand the underlying mechanisms as well as develop novel treatment strategies that can potentially be translated to the clinic. Furthermore, it may be possible to gain insights into the contribution of epilepsy to the progression of cognitive, social and motor phenotypes in NDDs.

Keywords (6): Neurodevelopmental disorders, epilepsy, MECP2, SCN1A, CDKL5, SYNGAP1.

1.0 Introduction

Neurodevelopmental disorders (NDDs) including autism spectrum disorders (ASDs) and intellectual disability (ID) can affect cognitive, social and motor abilities¹. NDDs are commonly associated with severe and intractable epilepsies with approximately 26% of patients having seizure comorbidities¹. Thus far, even if anti-epileptic drugs (AEDs) are effective in treating seizures, it is unclear whether neurodevelopmental symptoms are improved upon seizure control.

Recent advances in genetic sequencing have identified over 100 genes²⁻⁴ implicated in syndromic ASD and a further 700 x-linked, autosomal-dominant and autosomal-recessive genes that can be utilized to diagnose NDDs⁵⁻⁷. Coinciding with an increase in clinical data on the association of these genes with NDDs and epilepsy, gene-editing technologies such as CRISPR-Cas9 have made it increasingly faster, accurate and inexpensive to generate rodent models of these disorders with over 250 mouse models generated to study ASD alone^{8,9},

Animal models allow researchers to investigate the underlying mechanisms of these disorders, identify biomarkers for the clinic and develop potential novel treatment strategies. Furthermore, as many of the genes identified converge across biological pathways, it is important to define whether this results in symptomatic convergence and therefore possible common treatments.

For animal models to be effective it is necessary that they display face validity that resembles symptoms in patients. Although genes in brain disorders are frequently conserved in rodents, their function in other species may not be as critical to the complex behavioural disabilities associated with NDDs. Furthermore, it is possible that compensatory mechanisms come into effect upon gene-editing which are not present or have more of an impact than compensation in patients. Nonetheless, mouse models have shown the highest level of construct validity across species so far tested¹⁰⁻¹²

As recently reviewed by Silverman and Ellegood¹², deficits in many behavioural assays in rodent models of NDDs pertinent to ASD and ID include social communication experiments such as the three-chambered approach, reciprocal dyad interactions, social recognition, social place preference, and ultrasonic vocalizations. Cognitive inflexibility and insistence on sameness has also been modelled in NDD models^{13,14}. Other relevant behavioural deficits found in mouse models include quantification of spontaneous behaviours such as self-grooming, circling, jumping, back flipping and overall hyperactivity, which may be linked to abnormal or epileptic brain network activity.

Spontaneous epileptic activity has been recorded both through behavioural monitoring and electrophysiology in multiple genetically modified mouse models of NDDs¹⁵⁻¹⁸. A range of spontaneously emerging seizure types have been reported including tonic-clonic, atypical and typical absence, myoclonic spasms and interictal spikes. Increases in susceptibility to audiogenic seizures, hyperthermia and chemically induced seizures is also prevalent across genetic rodent models of NDDs¹⁸⁻²³. These models offer the opportunity to test the effect of anti-epileptic drugs (AEDs) and to disentangle the contribution of seizures to the progression of NDD's cognitive, social and motor phenotypes. Below, we summarize findings about epileptic activity in four mouse models of NDDs.

2.1 methyl-CpG-binding protein 2 (*MECP2*)

In Rett syndrome, both ID and ASD develop due to gain and loss of function of the X-linked *MECP2* gene. Patients develop progressive dementia, infantile hypotonia, gait abnormalities and seizures following normal early development for the first 2 years of life²⁴. Approximately half of patients with *MECP2* gene duplication suffer from epilepsy during their lifetime with a high level of refractoriness of approximately 32 to 56%^{25,26}. Lennox-Gastaut syndrome, in which patients suffer from a variety of seizure types is the most common form of epilepsy in patients with *MECP2* duplication. Whereas patients with late truncating deletions have a lower prevalence of epilepsy, several mutations exist with varying risks for epilepsy.

Mouse and rat models of MECP2 overexpression and duplication reliably replicate the seizure phenotype^{15,16}, with animals developing tonic-clonic seizures and increasing the sensitivity to pentylentetrazole (PTZ)-induced and kainic acid-induced seizures. Loss of function models display cortical discharges consistent with absence epilepsy¹⁹.

Seizure and ND phenotypes are reversible in Rett syndrome rodent models utilizing gene modification strategies^{27,28} and implementation of gene therapy in the clinic may be soon attainable. Pharmacological strategies that target downstream targets of MECP2 are also possible but made difficult to implement as the exact function of the gene is still unknown. Nonetheless, the effectiveness of AEDs in treating seizures in Rett syndrome have not been extensively tested in rodent models and could inform clinicians on potential therapeutics for Rett syndrome.

2.2 SCN1A

Dravet syndrome is a rare form of epilepsy accompanied by neurodevelopmental deficits and is mediated by the SCN1A gene, which encodes for the alpha subunit of the voltage-gated sodium channel²⁹. The syndrome is characterised by multiple seizure types including febrile and febrile generalised tonic-clonic and focal-clonic, tonic, status epilepticus, myoclonic and atypical absences. The epileptic symptoms are highly refractory to AEDs and the onset of the disease occurs within the first year of life³⁰.

As recently reviewed by Griffen et al.³¹, multiple mouse models of Dravet with *SCN1A* loss of function exist. Rodent models display ictal and interictal activity, as well as myoclonic jerks and flexions¹⁷. Seizures in *SCN1A* knock-out mice are also inducible via hyperthermia²⁰.

First line AEDs are ineffective in treating seizures and sodium channel modulators such as carbamazepine are known to exacerbate seizures³². However, recent clinical trials show positive seizure reduction utilizing stiripentol, cannabidiol, and fenfluramine with 50 to 70% reduction in seizures and nearly a quarter of patients attaining seizure freedom³³. In mouse models, stiripentol is effective in reducing hyperthermia-induced seizures³⁴, while

cannabidiol similarly reduced spontaneous seizures³⁵. Fenfluramine was found to be effective in a zebrafish model of Dravet³⁶.

Successful face validity of Dravet animal models suggests that novel treatments to achieve a higher level of seizure control and reversal of neurodevelopmental deficits may be possible. Studies to modify gene expression in rodent models of Dravet Syndrome have shown to effectively reduce seizures³⁷. The utility of animal models in Dravet is therefore likely to yield fruitful new treatment paths to test in the clinic.

2.3 cyclin-dependent kinase-like-5 (CDKL5)

CDKL5 is likely involved in correct dendritic spine structure and synapse activity in excitatory neurons by binding and phosphorylating the NGL-1 cell adhesion molecule which allows it to form a stable association with PSD95³⁸. *CDKL5* is amongst the most common single-gene predictors of epilepsy diagnosis³⁹. Mutations in *CDKL5* lead to a NDD with severe developmental delay and early-onset epilepsy. Other features include impaired hand function with stereotypies, impaired social interaction with autistic features and severe motor impairment with only exceptional patients who are independently ambulant⁴⁰. 98% of patients present with a history of epilepsy including infantile spasms, myoclonus and prolonged generalized tonic-clonic seizures⁴¹. Most of these types of epilepsy appear to be refractory to AEDs.

CDKL5 models have been generated and display certain phenotypes including hyperexcitability²¹, visual impairments⁴², social interaction deficits, motor control decreases and loss of fear memory⁴³. Nonetheless, despite the prevalence of seizures in patients, mouse models have not been reported to display spontaneous ictal or interictal activity. It is likely that either compensatory mechanisms exist to compensate *CDKL5* loss of function or the gene does not play as critical a role in rodents. *CDKL5* mice however are susceptible to NMDA-induced seizures²¹. More research is necessary to elucidate whether animal models of this disorder have translational value.

2.4 SYNGAP1

SYNGAP1 is associated with NMDAR-activated Ras signalling dynamics in dendritic spines as well as AMPA receptor membrane insertion^{23,44}. Although mutations are relatively rare, heterozygous loss-of-function in *SYNGAP1* results in a genetically defined NDD with ID, which also predisposes patients to ASD. Symptoms of this syndrome include cognitive impairment, loss of language abilities and seizures^{45,46}. In a recent clinical report, 56 of 57 patients had epilepsy with a median age of onset of 2 years⁴⁷. Most seizures are generalized, with absence seizures reported in 53 of the patients, although focal seizures also occur. 65% of patients had intractable epilepsy with valproate and lamotrigine commonly prescribed as well as cannabidiol.

As found in patients⁴⁸, mouse models display increased risk-taking behaviours as assessed by the elevated plus maze as well as cognitive and learning impairments⁴⁵. *SYNGAP1* haploinsufficient mice have spontaneous interictal activity and have a reduced fluorothyl-induced seizure threshold as well as being prone to audiogenic seizures^{18,22,23}. Restoration of the gene in adult mice is able to improve behavioural and electrophysiological measures of memory and seizures¹⁸. More work is needed to test potential treatment strategies in these animals and determine whether these are translatable to the clinic.

3.0 Conclusion

Rodent models display phenotypes reminiscent of symptoms in genetically defined NDDs such as those affecting cognition, motor abilities and sociability. Genetically defined NDDs very frequently are comorbid with severe intractable epilepsy. Brain network dysfunction resulting in seizure activity is common in rodent models. Furthermore, if spontaneous epilepsy is absent, such as in the case of *CDKL5* models, the mutant animals can still be challenged to determine whether they have compromised seizure thresholds.

Pharmacology treatments as well as other strategies such as gene therapy can be tested on rodent models and findings can be translated to the clinic as with the positive results of

stiripentol³⁴, cannabidiol³⁵, and fenfluramine³⁶ being effective in seizure control in animal models. Rodents give researchers the opportunity to develop new treatments by probing deeper into the mechanisms resulting in NDDs. An enticing avenue of research with rodent genetic models of NDDs is to test the hypothesis that if seizures are controlled early enough in development, they are able to arrest the neurodevelopmental deficits.

Citations

1. Association AP., Force APAD-5 T. *Diagnostic and statistical manual of mental disorders : DSM-5*. Fifth edit. Arlington, VA: Arlington, VA : American Psychiatric Association; 2013.
2. Iossifov I., O’Roak BJ., Sanders SJ., Ronemus M., Krumm N., Levy D., et al. The contribution of de novo coding mutations to autism spectrum disorder. *Nature* 2014;**515**(7526):216–21. Doi: 10.1038/nature13908.
3. Huguet G, Benabou M BT. *The Genetics of Autism Spectrum Disorders*. Springer; n.d.
4. Krumm N., O’Roak BJ., Shendure J., Eichler EE. A de novo convergence of autism genetics and molecular neuroscience. *Trends Neurosci* 2014;**37**(2):95–105. Doi: 10.1016/j.tins.2013.11.005.
5. Kaufman L., Ayub M., Vincent JB. The genetic basis of non-syndromic intellectual disability: A review. *J Neurodev Disord* 2010;**2**(4):182–209. Doi: 10.1007/s11689-010-9055-2.
6. Gilissen C., Hehir-Kwa JY., Thung DT., Van De Vorst M., Van Bon BWM., Willemsen

- MH., et al. Genome sequencing identifies major causes of severe intellectual disability. *Nature* 2014;**511**(7509):344–7. Doi: 10.1038/nature13394.
7. Vissers LELM., Gilissen C., Veltman JA. Genetic studies in intellectual disability and related disorders. *Nat Rev Genet* 2016;**17**(1):9–18. Doi: 10.1038/nrg3999.
 8. Banerjee-Basu S PA. Sfari gene: An evolving database for the autism research community. *Dis Model Mech* 2010;**3**(3–3):133–5.
 9. Kumar A., Wadhawan R., Swanwick CC., Kollu R., Basu SN., Banerjee-Basu S. Animal model integration to AutDB, a genetic database for Autism. *BMC Med Genomics* 2011;**4**. Doi: 10.1186/1755-8794-4-15.
 10. Crawley JN. Behavioral phenotyping of transgenic and knockout mice: Experimental design and evaluation of general health, sensory functions, motor abilities, and specific behavioral tests. *Brain Res* 1999;**835**(1):18–26. Doi: 10.1016/S0006-8993(98)01258-X.
 11. Crawley JN. Behavioral Phenotyping Strategies for Mutant Mice. *Neuron* 2008;**57**(6):809–18. Doi: 10.1016/j.neuron.2008.03.001.
 12. Silverman JL., Ellegood J., Centre I. Behavioral and Neuroanatomical Approaches in Models of Neurodevelopmental Disorders: Opportunities for Translation Jill 2019;**31**(2):126–33. Doi: 10.1097/WCO.0000000000000537.Behavioral.
 13. Lewis MH., Tanimura Y., Lee LW., Bodfish JW. Animal models of restricted repetitive behavior in autism. *Behav Brain Res* 2007;**176**(1):66–74. Doi: 10.1016/j.bbr.2006.08.023.
 14. Bechard AR., Bliznyuk N., Lewis MH. The development of repetitive motor behaviors in deer mice: Effects of environmental enrichment, repeated testing, and differential mediation by indirect basal ganglia pathway activation. *Dev Psychobiol* 2017;**59**(3):390–9. Doi: 10.1002/dev.21503.

15. Collins AL., Levenson JM., Vilaythong AP., Richman R., Armstrong DL., Noebels JL., et al. Mild overexpression of MeCP2 causes a progressive neurological disorder in mice. *Hum Mol Genet* 2004;**13**(21):2679–89. Doi: 10.1093/hmg/ddh282.
16. McLeod F., Ganley R., Williams L., Selfridge J., Bird A., Cobb SR. Reduced seizure threshold and altered network oscillatory properties in a mouse model of Rett syndrome. *Neuroscience* 2013;**231**:195–205. Doi: 10.1016/j.neuroscience.2012.11.058.
17. Yu FH., Mantegazza M., Westenbroek RE., Robbins CA., Kalume F., Burton KA., et al. Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy. *Nat Neurosci* 2006;**9**(9):1142–9. Doi: 10.1038/nn1754.
18. Creson TK., Rojas C., Hwaun E., Vaissiere T., Kilinc M., Jimenez-Gomez A., et al. Re-expression of SynGAP protein in adulthood improves translatable measures of brain function and behavior. *Elife* 2019;**8**. Doi: 10.7554/eLife.46752.
19. Wither RG., Colic S., Bardakjian BL., Snead OC., Zhang L., Eubanks JH. Electrographic and pharmacological characterization of a progressive epilepsy phenotype in female MeCP2-deficient mice. *Epilepsy Res* 2018;**140**(July 2017):177–83. Doi: 10.1016/j.eplepsyres.2018.01.015.
20. Ogiwara I., Iwasato T., Miyamoto H., Iwata R., Yamagata T., Mazaki E., et al. Nav1.1 haploinsufficiency in excitatory neurons ameliorates seizure-associated sudden death in a mouse model of dravet syndrome. *Hum Mol Genet* 2013;**22**(23):4784–804. Doi: 10.1093/hmg/ddt331.
21. Okuda K., Kobayashi S., Fukaya M., Watanabe A., Murakami T., Hagiwara M., et al. CDKL5 controls postsynaptic localization of GluN2B-containing NMDA receptors in the hippocampus and regulates seizure susceptibility. *Neurobiol Dis* 2017;**106**:158–70. Doi: 10.1016/j.nbd.2017.07.002.

22. Clement JP., Aceti M., Creson TK., Ozkan ED., Shi Y., Reish NJ., et al. Pathogenic SYNGAP1 mutations impair cognitive development by disrupting maturation of dendritic spine synapses. *Cell* 2012;**151**(4):709–23. Doi: 10.1016/j.cell.2012.08.045.
23. Ozkan ED., Creson TK., Kramár EA., Rojas C., Seese RR., Babyan AH., et al. Reduced cognition in Syngap1 mutants is caused by isolated damage within developing forebrain excitatory neurons. *Neuron* 2014;**82**(6):1317–33. Doi: 10.1016/j.neuron.2014.05.015.
24. A. R. [On a unusual brain atrophy syndrome in hyperammonemia in childhood]. *Wien Med Wochenschr* 1966;**116**(37):723-6.
25. Marafi D., Suter B., Schultz R., Glaze D., Pavlik VN., Goldman AM. Spectrum and time course of epilepsy and the associated cognitive decline in MECP2 duplication syndrome. *Neurology* 2019;**92**(2):E108–14. Doi: 10.1212/WNL.0000000000006742.
26. Nissenkorn A., Levy-Drummer RS., Bondi O., Renieri A., Villard L., Mari F., et al. Epilepsy in Rett syndrome - Lessons from the Rett networked database. *Epilepsia* 2015;**56**(4):569–76. Doi: 10.1111/epi.12941.
27. Sztainberg Y., Chen HM., Swann JW., Hao S., Tang B., Wu Z., et al. Reversal of phenotypes in MECP2 duplication mice using genetic rescue or antisense oligonucleotides. *Nature* 2015;**528**(7580):123–6. Doi: 10.1038/nature16159.
28. Tillotson R., Selfridge J., Koerner M V., Gadalla KKE., Guy J., De Sousa D., et al. Radically truncated MeCP2 rescues Rett syndrome-like neurological defects. *Nature* 2017;**550**(7676):398–401. Doi: 10.1038/nature24058.
29. Meisler MH., O'Brien JE., Sharkey LM. Sodium channel gene family: Epilepsy mutations, gene interactions and modifier effects. *J Physiol* 2010;**588**(11):1841–8. Doi: 10.1113/jphysiol.2010.188482.
30. Kearney JA., Wiste AK., Stephani U., Trudeau MM., Siegel A., Ramachandran R.,

- et al. Recurrent de novo mutations of SCN1A in severe myoclonic epilepsy of infancy. *Pediatr Neurol* 2006;**34**(2):116–20. Doi: 10.1016/j.pediatrneurol.2005.07.009.
31. Griffin A., Hamling KR., Hong SG., Anvar M., Lee LP., Baraban SC. Preclinical animal models for Dravet syndrome: Seizure phenotypes, comorbidities and drug screening. *Front Pharmacol* 2018;**9**(JUN):1–15. Doi: 10.3389/fphar.2018.00573.
 32. Knupp KG., Wirrell EC. Treatment Strategies for Dravet Syndrome. *CNS Drugs* 2018;**32**(8):783. Doi: 10.1007/s40263-018-0546-0.
 33. Wirrell, EC NR. Recent Advances in the Drug Treatment of Dravet Syndrome. *CNS Drugs* 2019;**33**:867–81. Doi: 10.1136/pgmj.27.310.390.
 34. Cao D., Ohtani H., Ogiwara I., Ohtani S., Takahashi Y., Yamakawa K., et al. Efficacy of stiripentol in hyperthermia-induced seizures in a mouse model of Dravet syndrome. *Epilepsia* 2012;**53**(7):1140–5. Doi: 10.1111/j.1528-1167.2012.03497.x.
 35. Kaplan JS., Stella N., Catterall WA., Westenbroek RE. Cannabidiol attenuates seizures and social deficits in a mouse model of Dravet syndrome. *Proc Natl Acad Sci U S A* 2017;**114**(42):11229–34. Doi: 10.1073/pnas.1711351114.
 36. Dinday MT., Baraban SC. Large-scale phenotype-based antiepileptic drug screening in a zebrafish model of Dravet syndrome. *ENeuro* 2015;**2**(4):1–19. Doi: 10.1523/ENEURO.0068-15.2015.
 37. Colasante G., Lignani G., Brusco S., Di Bernardino C., Carpenter J., Giannelli S., et al. dCas9-Based *Scn1a* Gene Activation Restores Inhibitory Interneuron Excitability and Attenuates Seizures in Dravet Syndrome Mice. *Mol Ther* 2019. Doi: 10.1016/j.ymthe.2019.08.018.
 38. Ricciardi S., Ungaro F., Hambrock M., Rademacher N., Stefanelli G., Brambilla D., et al. CDKL5 ensures excitatory synapse stability by reinforcing NGL-1–PSD95 interaction in the postsynaptic compartment and is impaired in patient iPSC-derived

- neurons. *Nat Cell Biol* 2012;**14**:911.
39. Symonds JD., Zuberi SM., Stewart K., McLellan A., O'Regan M., MacLeod S., et al. Incidence and phenotypes of childhood-onset genetic epilepsies: a prospective population-based national cohort. *Brain* 2019. Doi: 10.1093/brain/awz195.
 40. Guerrini R., Parrini E. Epilepsy in Rett syndrome, and CDKL5- and FOXP1-gene-related encephalopathies. *Epilepsia* 2012;**53**(12):2067–78. Doi: 10.1111/j.1528-1167.2012.03656.x.
 41. Fehr S., Wong K., Chin R., Williams S., De Klerk N., Forbes D., et al. Seizure variables and their relationship to genotype and functional abilities in the CDKL5 disorder. *Neurology* 2016;**87**(21):2206–13. Doi: 10.1212/WNL.0000000000003352.
 42. Mazziotti R., Lupori L., Sagona G., Gennaro M., Della Sala G., Putignano E., et al. Searching for biomarkers of CDKL5 disorder: early-onset visual impairment in CDKL5 mutant mice. *Hum Mol Genet* 2017;**26**(12):2290–8. Doi: 10.1093/hmg/ddx119.
 43. Wang ITJ., Allen M., Goffin D., Zhu X., Fairless AH., Brodtkin ES., et al. Loss of CDKL5 disrupts kinome profile and event-related potentials leading to autistic-like phenotypes in mice. *Proc Natl Acad Sci U S A* 2012;**109**(52):21516–21. Doi: 10.1073/pnas.1216988110.
 44. Zhu JJ., Qin Y., Zhao M., Van Aelst L., Malinow R. Ras and Rap control AMPA receptor trafficking during synaptic plasticity. *Cell* 2002;**110**(4):443–55. Doi: 10.1016/S0092-8674(02)00897-8.
 45. Berryer MH., Chattopadhyaya B., Xing P., Riebe I., Bosoi C., Sanon N., et al. Decrease of SYNGAP1 in GABAergic cells impairs inhibitory synapse connectivity, synaptic inhibition and cognitive function. *Nat Commun* 2016;**7**. Doi: 10.1038/ncomms13340.
 46. Mignot C., von Stülpnagel C., Nava C., Ville D., Sanlaville D., Lesca G., et al. Genetic

and neurodevelopmental spectrum of SYNGAP1-associated intellectual disability and epilepsy. *J Med Genet* 2016;**53**(8):511–22. Doi: 10.1136/jmedgenet-2015-103451.

47. Vlaskamp DRM., Shaw BJ., Burgess R., Mei D., Montomoli M., Xie H., et al. SYNGAP1 encephalopathy: A distinctive generalized developmental and epileptic encephalopathy. *Neurology* 2019;**92**(2):E96–107. Doi: 10.1212/WNL.0000000000006729.
48. Weldon M., Kilinc M., Lloyd Holder J., Rumbaugh G. The first international conference on SYNGAP1-related brain disorders: A stakeholder meeting of families, researchers, clinicians, and regulators. *J Neurodev Disord* 2018;**10**(1):1–6. Doi: 10.1186/s11689-018-9225-1.