Efficient nonmeiotic allele introgression in livestock using custom endonucleases

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We have expanded the livestock gene editing toolbox to include transcription activator-like (TAL) effector nuclease (TALEN)- and clustered regularly interspaced short palindromic repeats (CRISPR)/Cas9-stimulated homology-directed repair (HDR) using plasmid, rAAV, and oligonucleotide templates. Toward the genetic dehorning of dairy cattle, we introgressed a bovine POLLED allele into horned bull fibroblasts. Single nucleotide alterations or small indels were introduced into 14 additional genes in pig, goat, and cattle fibroblasts using TALEN mRNA and oligonucleotide transfection with efficiencies of 10–50% in populations. Several of the introduced edits mimic naturally occurring or disease-resistance alleles, including alteration of single base pairs. Up to 70% of the fibroblast colonies propagated without selection harbored the intended edits, of which more than one-half were homozygous. Edited fibroblasts were used to generate pigs with knockout alleles in the DAZL and APC genes to model infertility and colon cancer. Our methods enable unprecedented meiosis-free intraspecific and interspecífic introgression of select alleles in livestock for agricultural and biomedical applications.

**Results**

**Introgression of the Celtic POLLED Allele into the Genome of Horned Dairy Cattle Breeds.** We previously reported an efficient system for deriving TALEN-mediated NHEJ knockouts in livestock fibroblasts and embryos (15), and theorized that similar methods would be suitable for derivation of cells with HDR-mediated genetic changes. Accordingly, we used TALENs to stimulate HDR with plasmid and recombinant adeno-associated virus (rAAV) donor templates designed to introduce a naturally occurring 11-bp deletion or the Belgian Blue mutation (23–25) at position 821 (821del11) of growth differentiation factor 8 (GDF8) (SI Appendix, Fig. S1). Individual colonies with the intended mutation were recovered in 1% of colonies treated with the plasmid template and in 13% of colonies treated with the rAAV template (SI Appendix, Fig. S1D). These results inspired us to attempt introgression of more complex alleles into livestock.

To protect the welfare of dairy farm operators and cattle, horns are routinely manually removed from the majority of dairy cattle in the United States and Europe. Dehorning is painful, elicits a temporary elevation in animal stress, and adds expense to animal production (26). Moreover, despite the intent of protecting animals from subsequent injury, some view the practice as inhumane. Some beef breeds are naturally horn-free (e.g.,

**Significance**

Selective breeding has long been practiced to enrich for desirable DNA variation that influences livestock traits. We demonstrate that genetic variants can be directly introgressed into livestock genomes using a modified transcription activator-like effector nuclease system. The transient exposure of livestock cells to sequence-targeted editors stimulates homology-directed repair to levels that eliminate the need for transgene-dependent selection. Use of oligonucleotide template enables efficient single nucleotide changes to the genome and permits the transmission of both natural and novel DNA sequence variants into naive livestock breeds and species. Gene editing offers a powerful method for accelerating the genetic improvement of livestock for food and also for developing swine as a resource for regenerative medicine and models of human disease.
Angus), a dominant trait referred to as POLLED (27). Two allelic variants conferring polledness have been identified on chromosome 1 (28). Meliotic introgression of the POLLED trait into horned dairy breeds can be accomplished by traditional cross-breeding, but the genetic merit of the resulting animals would rank lower owing to the admixture of unselected (inferior) alleles for net merit (i.e., milk production) into the population. We undertook the nonmeliotic introgression of the Celtic POLLED allele (duplication of 212 bp that replaces 10 bp), referred to as Pc, into fibroblasts derived from horned dairy bulls. We constructed a plasmid HDR template containing a 1,594-bp fragment including the Pc allele from the Angus breed (Fig. 1A). TALENs were designed such that they could cleave the HORNED allele but leave the Pc allele unaffected. In addition, after finding that one pair of TALENs delivered as mRNA had similar activity as plasmid DNA (SI Appendix, Fig. S2), we chose to deliver TALENs as mRNA to eliminate the possible genomic integration of TALEN expression plasmids. Five of 226 colonies (2%) passed each PCR test shown to confirm introgression of Pc (Fig. 1B). Three of the five clones were homozygous for Pc introgression and were confirmed by sequencing (SI Appendix, Table S1).

Reliable Oligonucleotide-Templated Allele Introgression in Pig, Goat, and Cattle Genomes. Although plasmid templates are effective for introgression of Pc and GDF8 alleles, many desirable alleles correspond to single nucleotide polymorphisms (SNPs) or small indels that ideally do not rely on the use of double-stranded DNA templates that could randomly integrate into the genome. In addition, although introgression of the GDF8 S211del11 allele with rAAV was efficient, viral methods are unlikely to be popular with consumers of animal protein. Recent studies have demonstrated that single-stranded oligonucleotides (oligos) 40–100 nt in length could serve as effective templates for HDR when stimulated by double-strand breaks (DSBs) in DNA (20, 21, 29–31). Accordingly, we sought to determine whether this approach could stimulate HDR in primary fibroblasts from pigs and cattle, cellular substrates suitable for the creation of gene-edited animals by cloning. We designed oligo templates to overlap their cognate TALEN-binding sites and to introduce a 4-bp indel into the spacer region for restriction fragment length polymorphism (RFLP) analysis. Primary fibroblasts were transfected with either TALEN-encoding plasmids (3 μg) or mRNA (1 μg) along with 3 μM of their cognate oligo-homologous template (SI Appendix, Table S4). Cells were then incubated at 37 °C or 30 °C for 3 d before expansion at 37 °C until day 10. TALEN activity was measured by the Surveyor assay at day 3 (Day3 Surveyor), and HDR was measured at days 3 and 10 by RFLP analysis (Day3 %HDR and Day10 %HDR). Each bar displays the average and SEM from three replicates.
(Fig. 2). The consistently high rate (25–50%) and stability of gene edits at all four loci suggest that edited cells should be recoverable by dilution cloning without selective enrichment. Indeed, analysis of ~650 colonies for intended indels at eight separate loci revealed a recovery rate of 10–64% (average, 45%), with up to 32% of the colonies homozygous for the edit (Table 1).

**Differential Stability of Gene Edits.** We were extremely successful in isolating individual colonies with custom indel alleles; however, introgression of SNPs presented a greater challenge. Both day 3 levels of HDR (7–18%) and the isolation of cellular clones with the intended SNP alleles (3–15%) in pig and cow GDF8 or pig p65 were significantly lower compared with indel HDR (day 3 level, 10–53%; clone isolation, 10–64%) (Table 1). We hypothesized that indels likely were more stable than SNP because introduction of indels into the TALEN spacer region would be expected to reduce recleavage of the locus, consistent with known constraints on TALEN spacer length (17). Accordingly, the introgression of large insertions (loxP) was extremely efficient and stable, and comparison of HDR frequencies with oligos within the same locus suggested that even a 4-bp insertion allele was more efficient than SNP alleles (SI Appendix, Fig. S4). Sequence analysis also revealed that nearly one-half of the isolated SNP-positive colonies for GDF8 or pig p65 harbored concomitant indels expected to change TALEN spacing (SI Appendix, Table S2). Regardless, we were able to recover colonies with homogeneous conversion of G938A in GDF8 (both pigs and cattle) and T1591C in pig p65 at up to nearly a 5% level without any additional changes to the locus (Table 1 and SI Appendix, Table S2). We also introgressed SNP alleles for the sheep FecB and Callipyge loci into the goat genome. This ability to precisely alter a single nucleotide is significant and unprecedented.

For comparison, we designed CRISPR gRNAs that overlapped the T1591C site of p65, and we evaluated introgression using the two platforms. Despite efficient production of DSBs at the intended site, CRISPR/Cas9-mediated HDR was <6% at day 3 and below the limit of detection at day 10 (SI Appendix, Fig. S5). In addition, recovery of modified colonies was lower with CRISPR-mediated HDR than with TALENs, even though the TALENs cut 35 bp away from the SNP site (Table 1). Analysis of CRISPR/Cas9-induced targeting at a second locus, ssAPC14.2, was much more efficient, but still did not reach the level of HDR induced by TALENs at this site (~30% vs. 60%; SI Appendix, Fig. S6).

**Strategies for Stabilizing Introgressed SNP Alleles.** Given the conservation of the 5′-thymidine nucleotide immediately preceding TAL-binding sites (32, 33), we reasoned that altering these bases in the oligo [known as blocking mutations (BMs)] would inhibit recleavage of edited alleles. Surprisingly, we found that BMs had no significant impact on the maintenance of SNP alleles at the pig LDLR or GDF8 loci (Fig. 3A). This suggests that either the conversion tract for oligo-templated HDR is quite short and does not incorporate the BM or that altering the 5′-thymidine does not completely abolish TALEN activity.

To examine this effect in greater detail, we conducted Illumina next-generation sequencing of 200–250-bp amplicons flanking the target sites from populations of cells transfected with oligos and TALEN mRNA. The results from five loci in pigs and cattle show that insertion alleles generally were more prevalent and stable in the population (Fig. 4). Whereas BMs had little influence on the preservation of intended alleles in culture, there was a slight bias toward incorporation of BMs in SNP-edited alleles compared with insertional edits (SI Appendix, Fig. S7). Consistent with our colony analysis, reads sorted on the basis of incorporating the intended SNP (iSNP), G938A, or T1591C conversion in bgGDF8 and p65 revealed that nearly one-half of the reads with the iSNP had an additional mutation (iSNP Mut) (Fig. 4B), the majority of which were indels (SI Appendix, Figs. S7 and S8). The majority of iSNP bgGDF8 reads with indels in the spacer also contained one or both BMs (SI Appendix, Fig. S8), demonstrating that modification of the conserved 5′-thymidine was not able to suppress recleavage and subsequent indel generation. Thus, this base must be less critical to TALEN binding than suggested by conservation, and provides a molecular basis for the inability of BMs to preserve alleles as described above.

Another strategy to reduce recutting of the SNP edits is to design TALENs such that their binding sites overlap the target SNPs. We evaluated the influence of such repeat variable dinucleotide

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**Table 1. Frequencies of recovery of colonies with HDR alleles**

<table>
<thead>
<tr>
<th>Reagent</th>
<th>ID</th>
<th>Species</th>
<th>Mutation type</th>
<th>Nucleotide change</th>
<th>Amino acid change</th>
<th>Day3% HDR</th>
<th>HDR+ (%)</th>
<th>Biallelic HDR+ (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TALEN</td>
<td>ssLDLR2.1*</td>
<td>Pig ♀</td>
<td>Ins/FS</td>
<td>141 (ins4)</td>
<td>47PTC</td>
<td>38</td>
<td>55/184 (30)</td>
<td>4/184 (2)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssDAZL3.1*</td>
<td>Pig ♂</td>
<td>Ins/FS</td>
<td>173 (ins4)</td>
<td>57PTC</td>
<td>25</td>
<td>34/92 (37)</td>
<td>8/92 (9)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssAPC4.2*</td>
<td>Pig ♀</td>
<td>Ins/FS</td>
<td>2703 (ins4)</td>
<td>902aPTC</td>
<td>48</td>
<td>22/40 (55)</td>
<td>4/40 (10)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssTpt53</td>
<td>Pig ♂</td>
<td>Ins/FS</td>
<td>463 (ins4)</td>
<td>154aPTC</td>
<td>22</td>
<td>42/71 (59)</td>
<td>12/71 (17)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssRAG2.1</td>
<td>Pig ♂</td>
<td>Ins/FS</td>
<td>228 (ins4)</td>
<td>76aPTC</td>
<td>47</td>
<td>32/77 (42)</td>
<td>13/77 (17)</td>
</tr>
<tr>
<td>TALEN</td>
<td>btRosa1.2</td>
<td>Cow ♂</td>
<td>Ins/mloxP</td>
<td>Ins34</td>
<td>NA</td>
<td>45</td>
<td>14/22 (64)</td>
<td>7/22 (32)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssSRY3.2</td>
<td>Pig ♂</td>
<td>Ins/mloxP</td>
<td>Ins34</td>
<td>NA</td>
<td>30</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssKissR3.2</td>
<td>Pig ♀</td>
<td>Ins/FS</td>
<td>322 (ins6) 323 (del2)</td>
<td>107aPTC</td>
<td>53</td>
<td>57/96 (59)</td>
<td>17/96 (18)</td>
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<tr>
<td>TALEN</td>
<td>btGDF83.1</td>
<td>Cow ♂</td>
<td>del/FS</td>
<td>821 (del11)</td>
<td>F5</td>
<td>&lt;10</td>
<td>7/72 (10)</td>
<td>2/72 (3)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssEif4G14.1</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>G2014A T2017C C2019T</td>
<td>N672D L673F</td>
<td>52</td>
<td>68/102 (67)</td>
<td>40/102 (39)</td>
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<td>btGDF83.6N</td>
<td>Cow ♂</td>
<td>SNP</td>
<td>G938A T945C</td>
<td>C313Y</td>
<td>18</td>
<td>8/94 (9)</td>
<td>3/94 (3)</td>
</tr>
<tr>
<td>TALEN</td>
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<td>Cow ♂</td>
<td>SNP</td>
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<td>C313Y</td>
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<td>7/105 (7)</td>
<td>2/105 (2)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssP65.8</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>T1591C</td>
<td>S531P</td>
<td>18</td>
<td>6/40 (15)</td>
<td>3/40 (8)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssP65.8</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>T1591C</td>
<td>S531P</td>
<td>7</td>
<td>6/63 (14)</td>
<td>5/63 (8)</td>
</tr>
<tr>
<td>TALEN</td>
<td>ssGDF83.6i</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>G938A C313Y</td>
<td>C313Y</td>
<td>NA</td>
<td>3/90 (3)</td>
<td>1/90 (1)</td>
</tr>
<tr>
<td>TALEN</td>
<td>caFecB6.1</td>
<td>Goat ♂</td>
<td>SNP</td>
<td>A747G</td>
<td>Q249R</td>
<td>17</td>
<td>17/72 (24)</td>
<td>3/72 (4)</td>
</tr>
<tr>
<td>TALEN</td>
<td>caCLPG1.1</td>
<td>Goat ♂</td>
<td>SNP</td>
<td>A--G</td>
<td>Extragenic</td>
<td>4</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>CRISPR</td>
<td>ssP65 G1s</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>T1591C</td>
<td>S531P</td>
<td>6</td>
<td>6/96 (6)</td>
<td>2/96 (2)</td>
</tr>
<tr>
<td>CRISPR</td>
<td>ssP65 G2a</td>
<td>Pig ♂</td>
<td>SNP</td>
<td>T1591C</td>
<td>S531P</td>
<td>5</td>
<td>2/45 (4)</td>
<td>0/45</td>
</tr>
<tr>
<td>CRISPR</td>
<td>ssAPC14.2 G1a</td>
<td>Pig ♀</td>
<td>Ins/FS</td>
<td>2703 (ins4)</td>
<td>902aPTC</td>
<td>32</td>
<td>ND</td>
<td>ND</td>
</tr>
</tbody>
</table>

*ss, Sus scrofa; bt, Bos taurus; ca, Capra aegagrus; FS, frame shift; NA, not applicable; ND, not determined; PTC, premature termination codon; R, replicate.

*Homologous null LDLR<sup>loxP</sup> could not be propagated in culture.

<sup>a</sup>Modiﬁed loxP site (mloxP) was inserted into btROSA26 and ssSRY; positive colonies were identiﬁed by PCR.

<sup>§</sup>Only the target SNP was introduced. A triprimer PCR combined with Sanger sequencing was used to identify positive colonies (SI Appendix, Table S2).
piglets were heterozygous for SNP introgression using oligo donors. (founders).

Values are indicated.

<table>
<thead>
<tr>
<th>Founder</th>
<th>Allele (btGDF83.6-A)</th>
<th>Allele (btGDF83.6-G)</th>
</tr>
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<tr>
<td></td>
<td></td>
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</tbody>
</table>

Fig. 3. SNP introgression using oligo donors. (A) The influence of BMs on maintenance of HDR alleles was evaluated. Each oligo was designed to introduce the same SNPr restriction sites with or without BMs. HDR was quantified by RFLP assay in transfected populations initially cultured at 30 °C for 3 d and then maintained at 37 °C up to day 12. Values are averaged (n = 3). (B) Introgression of myostatin C313Y into Wagyu fibroblasts. The C313Y missense mutation is caused by a G-to-A transition (indicated by oversized text) at nucleotide 938 of bovine myostatin (23–25). The HDR template also includes a T-to-C transition (red) to introduce a novel EcoRI site for RFLP screening. Two left TALENs were designed against the locus: btGDF83.6-G, targeting the WT allele (WT), and btGDF83.6-A targeting the mutant allele (C313Y). The two share a common right TALEN. Transfection, culture, and measurement were conducted as described above. The average and SEM values for btGDF83.6-G (n = 30) and btGDF83.6-A (n = 5) represent 12 and 3 biological replicates, respectively. A two-sided Student t test was used to compare averages between groups; P values are indicated.

(RVD)/nucleotide mismatches for introgression of G938A SNP into cattle GDF8. Two pairs of TALENs were generated, one that bound the WT “G” allele (btGDF83.6-G) and another that bound the intended “A” allele (btGDF83.6-A) (Fig. 3B). HDR with each TALEN pair was similar at day 3, whereas levels measured at day 12 were significantly higher using the TALENs that bound the WT G allele, indicating that recleavage was more prevalent with btGDF83.6-A, which targets the repaired allele perfectly. Different RVD/nucleotide mismatches likely will have a greater influence on maintenance of HDR alleles, because the Asn-Asn (NN) RVD used for the WT G TALEN is able to bind both G and A nucleotides. For modification of porcine EIF4G1, we found that three RVD/nucleotide mismatches were sufficient for protection of the HDR edit; nearly 70% of isolated colonies contained an edited allele, more than half of which were homozygotes (Table 1 and SI Appendix, Fig. S9). Thus, the intentional alteration of the target locus to resist recleavage is an effective strategy for preserving edits.

Ultimately, gene editing is a dynamic process. TALEN cleavage and recleavage are in flux with repair by NHEJ, HDR with an oligo template, and HDR with the sister chromatin as a template. We hypothesized that the observed loss of SNP alleles might be reduced by extending the hypothermic treatment, slowing cell proliferation long enough to outlast the burst of TALEN activity from TALEN mRNA transfection. Indeed, this extension almost tripled the level of SNP HDR-edited alleles recovered after extended culture (SI Appendix, Fig. S10).

Production of Biomedical Model Pigs with Gene-Edited Alleles. We chose two gene-edited loci in the porcine genome, deleted in azoospermia-like (DAZL) and adenomatous polyposis coli (APC), to carry through to live animals. Colonies with HDR- or NHEJ-edited alleles of DAZL or APC were pooled for cloning by chromatin transfer (CT). Each pool yielded two pregnancies from three transfers, of which one pregnancy each was carried to term. A total of eight piglets were born from DAZL-modified cells, each of which reflected genotypes of the chosen colonies consistent with either the HDR allele (founders 1650, 1651, and 1657) or deletions resulting from NHEJ (Fig. 5A). Three of the DAZL piglets (founders 1655–1657) were stillborn. Of the six piglets from APC-modified cells, one was stillborn, three died within 1 wk, and another died after 3 wk, leaving only founder 1661 alive. The lack of correlation between genotype and survival suggests that the early deaths were related to cloning rather than to gene edits (34). All six APC piglets were heterozygous for the intended HDR-edited allele, and all but one piglet had either an in-frame insertion or deletion of 3 bp on the second allele (Fig. 5A and B). The remaining animals are being raised for phenotypic analyses of spermatogenesis arrest (DAZL+/- founders) or development of colon cancer (APC+/− founders).

Discussion

The data presented in Table 1 demonstrate that combining mRNAs encoding TALENs and oligo templates for directing HDR achieves several key benchmarks for a precise genome-editing strategy: (i) only target nucleotides were changed, and mRNA transfection avoids unintended integration of plasmid DNA; (ii) selection markers are unnecessary, an important factor in the acceptance of edited livestock for human consumption; (iii) gene edits were efficient, ranging from approximately 10% for SNPs to >50% for some larger alterations; and (iv) the method was highly reliable; at each locus tested, targeted alteration of 16 of 16 sites (15 genes)

Fig. 4. Sequence analysis of TALEN-stimulated HDR alleles. PCR amplicons flanking the target site (200–250 bp total) derived from TALEN mRNA and oligo-transfected cell populations were sequenced by illumina sequencing. The total read count ranged from 10,000 to 400,000 per sample. (A and B) The counts of perfect, intended HDR reads versus the WT reads are plotted for insertion (A) and SNP alleles (B). The target locus, time point, and whether or not BMs were included in the oligo are indicated. (C) Reads from btGDF8 and p65 populations were sorted for incorporation of the target SNP and then classified into intended (iSNP only) vs. those with the target SNP plus an additional mutation (iSNP+Mut) and plotted against the total number of reads.
Various objectives were achieved by precise gene genome sequencing coupled with detailed phenotyping. At similar frequencies, we integrated a modifiedloxP(loxP) site into ROSA26, a presumptive safe harbor locus, and SRY loci in cattle and pigs that can serve as a landing pad for insertion of novel sequences in livestock via recombinase-mediated cassette exchange (36, 37). Previously, only NHEJ edits had been demonstrated for the Y chromosome of livestock (15); however, TALENs are clearly suitable for direct stimulation of knockout/knockin, a capability that we are currently exploiting in livestock and that was recently demonstrated in mice (38).

Of more immediate value, we achieved efficient nonmeiotic introgression of native alleles between species or breeds. Interspecies introgression is impossible by breeding, and intraspecies crossbreeding for allele introgression is either insufficient or deleterious to other performance features owing to the genetic admixture. Our nonmeiotic introgression includes the double-muscling mutations of GDF8 [SNP G938A from Piedmontese cattle (23, 25) and 821del11 (23–25) from Belgian Blue cattle] into the genome of Wagyu cattle and Landrace pigs. For improvement of animal welfare, we transferred the Pc allele for polledness from a beef-producing breed into cells from horned dairy cattle. We also transferred a candidate SNP allele for African swine fever resistance virus T1591C of65 (39) from warthog to the genome of conventional swine cells and introgressed sheep SNPs responsible for elevated fecundity (FecB; BMPR-IB) (40) and parent-of-origin dependent muscle hypertrophy (Callipyge) (41) into the goat genome. Up to now, nonmeiotic allele introgression has not been possible without selective enrichment, and our efficiencies are at least 10^2-fold greater than those observed previously with selection (42, 43). Such high levels of unscreened single-allele introgression suggest the feasibility of altering multiple alleles in a single generation of farm animals, decreasing the impact of long generation intervals on genetic improvement.

**Implications for Animal Breeding and Human Medicine.** High-throughput genome sequencing coupled with detailed phenotyping provides unprecedented opportunities for identifying specific alleles that affect livestock performance, and for locus-centric or whole-genome–based selection for improvement of animal genetics (44). Our results suggest that gene editing can be incorporated into selection programs to accelerate genetic improvement when selective breeding is either inefficient or impossible. Nonmeiotic introgression provides a genetic method for precisely crossbreeding for large effect alleles without compromising the genetic merit of indigenous or purpose-bred populations by whole-genome admixtures (2). By allowing such single-generation introgression of select alleles, it will accelerate genetic improvement based on local ecologies and needs that may derive from climate change and emerging diseases.

We used customized endonucleases to generate live animals with precise edits at two independent loci. Pigs edited to disrupt the DAZL gene can serve as a model for studying the restoration of human fertility by germ cell transplantation or for producing genetically modified offspring by transfer of genetically modified germline stem cells, as has been demonstrated in pigs (45), goats (46), and rodents (47, 48). Gene-edited alleles of APC also could provide a size-relevant model of colon cancer for preclinical evaluation of therapeutics, surgical intervention, or detection modalities. These results, coupled with our previous findings (15), demonstrate relative groundbreaking ease in introducing genetic modifications that mimic natural polymorphisms or human disease alleles into livestock.

Our work also has implications for personalized medicine, demonstrating the ability to develop precisely engineered large animals to serve as tailored biomedical models for testing drug, device, and cellular therapeutics, and potentially as resources for xenogeneic and autologous therapeutic cells and organs. Finally, the precision and high efficiency that we have achieved was accomplished by interrupting coding sequences with 4-bp indels. This strategy was very reliable and generally resulted in HDR edits in approximately 40% of the clones (range, 26–60%), up to one-third of which were homozygotes. At similar frequencies, we integrated a modifiedloxP site into ROSA26, a presumptive safe harbor locus, and SRY loci in cattle and pigs that can serve as a landing pad for insertion of novel sequences in livestock via recombinase-mediated cassette exchange (36, 37). Previously, only NHEJ edits had been demonstrated for the Y chromosome of livestock (15); however, TALENs are clearly suitable for direct stimulation of knockout/knockin, a capability that we are currently exploiting in livestock and that was recently demonstrated in mice (38).

Of more immediate value, we achieved efficient nonmeiotic introgression of native alleles between species or breeds. Interspecies introgression is impossible by breeding, and intraspecies crossbreeding for allele introgression is either insufficient or deleterious to other performance features owing to the genetic admixture. Our nonmeiotic introgression includes the double-muscling mutations of GDF8 [SNP G938A from Piedmontese cattle (23, 25) and 821del11 (23–25) from Belgian Blue cattle] into the genome of Wagyu cattle and Landrace pigs. For improvement of animal welfare, we transferred the Pc allele for polledness from a beef-producing breed into cells from horned dairy cattle. We also transferred a candidate SNP allele for African swine fever resistance virus T1591C of65 (39) from warthog to the genome of conventional swine cells and introgressed sheep SNPs responsible for elevated fecundity (FecB; BMPR-IB) (40) and parent-of-origin dependent muscle hypertrophy (Callipyge) (41) into the goat genome. Up to now, nonmeiotic allele introgression has not been possible without selective enrichment, and our efficiencies are at least 10^2-fold greater than those observed previously with selection (42, 43). Such high levels of unscreened single-allele introgression suggest the feasibility of altering multiple alleles in a single generation of farm animals, decreasing the impact of long generation intervals on genetic improvement.

**Implications for Animal Breeding and Human Medicine.** High-throughput genome sequencing coupled with detailed phenotyping provides unprecedented opportunities for identifying specific alleles that affect livestock performance, and for locus-centric or whole-genome–based selection for improvement of animal genetics (44). Our results suggest that gene editing can be incorporated into selection programs to accelerate genetic improvement when selective breeding is either inefficient or impossible. Nonmeiotic introgression provides a genetic method for precisely crossbreeding for large effect alleles without compromising the genetic merit of indigenous or purpose-bred populations by whole-genome admixtures (2). By allowing such single-generation introgression of select alleles, it will accelerate genetic improvement based on local ecologies and needs that may derive from climate change and emerging diseases.

We used customized endonucleases to generate live animals with precise edits at two independent loci. Pigs edited to disrupt the DAZL gene can serve as a model for studying the restoration of human fertility by germ cell transplantation or for producing genetically modified offspring by transfer of genetically modified germline stem cells, as has been demonstrated in pigs (45), goats (46), and rodents (47, 48). Gene-edited alleles of APC also could provide a size-relevant model of colon cancer for preclinical evaluation of therapeutics, surgical intervention, or detection modalities. These results, coupled with our previous findings (15), demonstrate relative groundbreaking ease in introducing genetic modifications that mimic natural polymorphisms or human disease alleles into livestock.

Our work also has implications for personalized medicine, demonstrating the ability to develop precisely engineered large animals to serve as tailored biomedical models for testing drug, device, and cellular therapeutics, and potentially as resources for xenogeneic and autologous therapeutic cells and organs. Finally, the precision and high efficiency that we have achieved was accomplished by interrupting coding sequences with 4-bp indels. This strategy was very reliable and generally resulted in HDR edits in approximately 40% of the clones (range, 26–60%), up to one-third of which were homozygotes. At similar frequencies, we integrated a modifiedloxP site into ROSA26, a presumptive safe harbor locus, and SRY loci in cattle and pigs that can serve as a landing pad for insertion of novel sequences in livestock via recombinase-mediated cassette exchange (36, 37). Previously, only NHEJ edits had been demonstrated for the Y chromosome of livestock (15); however, TALENs are clearly suitable for direct stimulation of knockout/knockin, a capability that we are currently exploiting in livestock and that was recently demonstrated in mice (38).

Of more immediate value, we achieved efficient nonmeiotic introgression of native alleles between species or breeds. Interspecies introgression is impossible by breeding, and intraspecies crossbreeding for allele introgression is either insufficient or deleterious to other performance features owing to the genetic admixture. Our nonmeiotic introgression includes the double-muscling mutations of GDF8 [SNP G938A from Piedmontese cattle (23, 25) and 821del11 (23–25) from Belgian Blue cattle] into the genome of Wagyu cattle and Landrace pigs. For improvement of animal welfare, we transferred the Pc allele for polledness from a beef-producing breed into cells from horned dairy cattle. We also transferred a candidate SNP allele for African swine fever resistance virus T1591C of65 (39) from warthog to the genome of conventional swine cells and introgressed sheep SNPs responsible for elevated fecundity (FecB; BMPR-IB) (40) and parent-of-origin dependent muscle hypertrophy (Callipyge) (41) into the goat genome. Up to now, nonmeiotic allele introgression has not been possible without selective enrichment, and our efficiencies are at least 10^2-fold greater than those observed previously with selection (42, 43). Such high levels of unscreened single-allele introgression suggest the feasibility of altering multiple alleles in a single generation of farm animals, decreasing the impact of long generation intervals on genetic improvement.

Gene Editing for Single-Step Introgression of Valuable Alleles into Livestock. Various objectives were achieved by precise gene editing (Table 1). Knockout of genes of biomedical relevance was achieved. The high efficiency and precision reported here are astonishing compared with what was feasible only a few years ago (2).

There are two concerns with gene editing: stabilizing the changes at the targeted site and avoiding modification of unintended sites. Regarding the first concern, we found evidence that HDR edits directing single bp changes (i.e., SNPs) could be lost (Figs. 3 and 4B). Based on the prediction that a thymidine preceding the targeted DNA sequence influences TAL binding (32, 33), we attempted to block recleavage of introgressed alleles by introducing BMs; however, BMs did not prevent TALEN activity and recleavage of edited alleles (Figs. 3 and SI Appendix, Figs. S7 and S8). In contrast, introduction of multiple SNPs or additional sequences (Fig. 2 and SI Appendix, Fig. S9) resulted in more stable HDR edits. Extension of hypothermic culture also resulted in the stabilization of introgressed SNP alleles. Because hypothermia slows cell proliferation primarily by prolonging the G1 phase of the cell cycle (35), this treatment may differentially favor oligo-HDR versus sister chromatid-templated repair in a cell cycle-dependent manner. Regardless of the mechanism, this approach offers a straightforward strategy for recovering cells with precise introgression of SNP alleles. As to the second concern, the frequency of modification at unintended sites is exceedingly low and in the context of livestock breeding will be lost by independent segregation in subsequent generations (2).
in altering either single nucleotides or small sequences in live- stock fibroblasts is likely to be applicable to the correction of disease-causing mutations in patient-derived fibroblasts, a re- source for conversion to induced pluripotent stem cells for use in cellular therapy (49), without the use of exogenous selec- tion markers.

Materials and Methods

Detailed information is provided in SI Appendix and our previous paper (15). In brief, TALENs were assembled using the Golden Gate assembly protocol and library (50). TALEN-encoding mRNA was synthesized in vitro using the mMESSAGE mMACHINE T3 Kit (Ambion). The CRISPR/Cas9 endonucleases were generated based on the Church laboratory system and methods (8). The transfections were conducted using the Neon transfection system (Invitrogen) unless stated otherwise. The frequency of total mutations in a TALEN- or CRISPR/Cas9-transfected population was assessed by the Sur- veyor Mutation Detection Kit (Transgenomic), and rates of mutation (Day3 Surveyor) was calculated by the Guscin method (51); %HDR was evaluated by RFLP or PCR. The colonies listed in Table 1 were obtained by cloning dilution from corresponding populations without drug selection. Population samples were sequenced on a illumina MiSeq sequencer after PCR amplifi- cation. The RVD sequences of TALENs, oligo sequences for HDR, and PCR primers are listed in SI Appendix, Tables S1, S3, S4, S5, and S6. Pigs were cloned by CT under contract with Minitrust of America under its Animal Welfare Assurance no. A452001.

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