Future therapeutic treatment of COPD

Citation for published version:

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
International Journal of Chronic Obstructive Pulmonary Disease

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Future therapeutic treatment of COPD: Struggle between oxidants and cytokines

Willem I de Boer 1
Hongwei Yao 2
Irfan Rahman 2

1 Netherlands Asthma Foundation, Leusden, The Netherlands; 2 Department of Environmental Medicine, Lung Biology and Disease Program, University of Rochester Medical Center, Rochester NY, USA

Abstract: Chronic obstructive pulmonary disease (COPD) is a global health problem. Being a progressive disease characterized by inflammation and predominantly caused by tobacco smoking, it deteriorates pulmonary and skeletal muscle functioning, and reduces physical behavior, societal participation and quality of life. During the last two decades studies were focused on the airway and systemic inflammation, oxidative stress, and airway and/or parenchymal remodeling. Macrophages, neutrophils and T cells are thought to be important key players, as well as structural cells like fibroblasts, epithelial, endothelial and smooth muscle cells. Mediators and proteins including cytokines, chemokines, growth factors, proteinases, and oxidants seem to be involved differentially in its pathogenesis. Current pharmacological treatments are directed to reducing airway inflammation, boosting the endogenous levels of anti-oxidants and relieving airway contraction and sputum production. Most agents were primarily used for treating asthma. But in contrast to asthma, these treatments are not very effective in COPD. As a result, novel more specifically acting interventional drugs with less side effects are being developed to treat chronic inflammatory diseases, including COPD. This review highlights studies on novel or potent drug antioxidants such as dietary antioxidants supplementation, N-acetyl-L-cysteine, N-acystelyn, endosteine, antioxidant enzyme mimetics, and anti-inflammatory agents like antagonists of cytokines, such as tumor necrosis factor (TNF)-α, CXCL8, and CCL2, and inhibitors of signal transduction proteins including phosphodiesterase 4, MAPK p38, PI-3k, and NFκB.

Keywords: COPD, inflammation, remodeling, cytokines, antioxidants, antagonists

Introduction
Chronic obstructive pulmonary disease (COPD) is a global health problem. According to estimates of the World Health Organization (WHO) about 80 million patients over the world suffered from COPD in 2005, whereas it accounts for 5% of all deaths. COPD is currently the 4th major cause of death worldwide. The prevalence is estimated to be about 1% worldwide, but is about 2 times higher in Western societies, and often underestimated due to un- or misdiagnosis (Pauwels and Rabe 2004; Menezes et al 2005; Lindberg et al 2006). The burden of COPD for the patient is high as patients experience a poorer quality of life, suffer from comorbidites (3.7 comorbidities per patient), and direct healthcare costs range from 0.28 billion euros in the Netherlands (in 2000) to 20.9 billion dollars in the USA (in 2004) (Hynninen et al 2005; Hoogendoorn et al 2006; Jones 2006; Sin et al 2006).

COPD is a progressive disease which is yet not curable. In Western countries the major cause is tobacco smoking, whereas in developing countries also indoor pollution eg, from cooking fuel biomass burning is a cause. Other risk factors for COPD include genetic predisposition, occupational and environmental exposure, and asthma. More than 90% of patients with COPD are smokers (Snider 1989), but at least 10%–20% of the smokers develop COPD pointing at an additional risk factor, eg, gene susceptibility. Among the genetic susceptibility factors are polymorphisms in genes coding for (anti-) proteases like alpha-1 antitrypsin (A1AT) (accounting for at least 5% of COPD cases) and a disinte-
grin and metalloproteinase 33 (ADAM33), genes coding for antioxidant enzymes like glutamate cysteine ligase, epoxide hydrolase, glutathione-S-transferase, and superoxide dismutase (SOD) 3, or genes coding for cytokines like tumor necrosis factor alpha (TNFα) and transforming growth factor beta 1 (TGFβ) (Harrison et al 1997; Keatings et al 2000; Sandford et al 2001; Kucukaycan et al 2002; Celedon et al 2004; Gosman, Boezen et al 2006; Young et al 2006).

Since chronic pulmonary inflammation and oxidative stress are important characteristics in the pathogenesis of COPD, this paper discusses the role of inflammatory mediators and oxidants and rational of anti-inflammatory and anti-oxidant therapeutic intervention in the management of COPD.

Pathophysiology
COPD is an inflammatory chronic lung disease that shows a largely irreversible lung function decline, and can be classified into 4 classes of severity based on lung function [GOLD Guidelines]. Emphysema, chronic bronchitis with airway obstruction, and small airways disease are the distinct phenotypes of COPD, but most patients show a combination. Emphysema is characterized by a Th1-type inflammation, destruction of the alveolar septa, loss of elastic recoil, airspace enlargement and hence loss of gas diffusion capacity (Wright and Churg 2006). Chronic bronchitis affects the airways by airway inflammation, goblet cell hyperplasia and mucus hypersecretion. In addition to decreased lung function, these patients experience chronic sputum production, coughing and often dyspnoea. Ciliated airway epithelium lining the airway lumen is impaired in its function due to the hypersecreted mucus, and in activation and damage by tobacco compounds. As a result, the ciliated epithelium may be partly replaced by other epithelial cell types like squamous and goblet cells, further contributing to the airway dysfunction. The inflammatory phenotype in chronic bronchitis is neutrophilic or eosinophilic. In eosinophilic chronic bronchitis the sputum or bronchoalveolar lavage (BAL) fluid presents more eosinophils whereas in neutrophilic airway inflammation predominates the neutrophilic phenotype. The eosinophilic phenotype exhibits a higher reversible forced expiratory volume in 1 second (FEV1) as compared to the neutrophilic phenotype, and is more responsive to corticosteroid treatment (Chanez et al 1997; Pizzichini et al 1998; Brightling et al 2005). Small airways disease mainly affects the bronchioles featuring airway inflammation and metaplasia of Clara cells.

Pathogenesis
The pathogenesis of COPD is not known yet. However, pathological features of COPD include lung tissue and vascular remodeling, and pulmonary and systemic inflammation (Barnes et al 2003; Langen et al 2003; Hogg 2004; De Boer 2005; Wright and Churg 2006; De Boer et al 2007). Clinical research in patients with established COPD showed inflammation with cells involved in innate immunity including macrophages, neutrophils, and T cells (predominantly CD8+, but less prominent in severe COPD) (Grashoff et al 1997; Di Stefano et al 2001; Hogg 2004; Barnes and Stockley 2005; De Boer 2005). Some studies also showed higher lung tissue numbers of mast cells, and also eosinophils during exacerbations or in patients with COPD showing reversible lung function (Grashoff et al 1997; Papi et al 2000; Barnes and Stockley 2005; De Boer 2005). Differences between the outcomes of studies may be due to inclusion criteria, numbers of participating patients, the COPD phenotype studied or the pulmonary location of sampled tissue. Recent studies also point to a link between the innate and acquired immune system. Studies with COPD patients demonstrated the presence of B-cell follicles in lung tissue (Gosman, Willemse et al 2006; van der Strate et al 2006) while studies on smoke-exposed mice show a role for dendritic cells in the pathogenesis of emphysema (Bracke et al 2006). A limited number of studies suggested an autoimmune component in emphysema (Agusti et al 2003). This suggestion was based on observations showing the presence of T cells (CD8+ and CD4+), B cell follicles in the airway walls, and clonal expansion of CD4+ T cells in lung tissue, the presence of anti-idiotypic antibodies with tobacco-like activity, and induction of emphysema in rats via a CD4+ and in mice via a CD8+ cell-specific mechanism (Saetta et al 1999; Koethe et al 2000; Sullivan et al 2005; Taraseviciene-Stewart et al 2005; Gosman, Willemse et al 2006; van der Strate et al 2006; Maeno et al 2007).

Oxidants and proteinases
Mechanisms that may contribute to the pathogenesis involve disturbed oxidant-antioxidant and proteinase-antiproteinase balances. Lungs are continuously exposed to oxidants, either generated endogenously by metabolic reactions (eg, from mitochondrial electron transport during respiration or during activation of phagocytes) or exogenously, such as air pollutants or cigarette smoke. Tobacco smoke constitutes about 5000 compounds and 1017 free radicals per puff, many of which are able to induce reactive oxygen or nitrogen species that can be inactivated by endogenous molecules like SOD, catalase, cytochrome P450, flavanoids and glutathione. In patients with
COPD this balance may be disturbed due to mutations in genes encoding these enzymes or enzymes related to these pathways (Langen et al 2003; Rahman and Adcock 2006). Production of reactive oxygen species (ROS) has been directly linked to oxidation of proteins, DNA, and lipids, which may cause direct lung injury or induce a variety of cellular responses, through the generation of secondary metabolic reactive species (Repine et al 1997). ROS may alter remodeling of extracellular matrix (ECM) and blood vessels, stimulate mucus secretion, inactivate antiproteases, cause apoptosis, and regulate cell proliferation (Rahman and MacNee 1996a, 1996b, 1999). Furthermore, increased levels of ROS have been implicated in initiating inflammatory responses in the lungs through the activation of transcription factors such as nuclear factor-kappaB (NF-κB) and activator protein-1 (AP-1), signal transduction, chromatin remodeling and gene expression of pro-inflammatory mediators (Rahman and MacNee 1998).

4-Hydroxy-2-nonenal (4-HNE) is a highly reactive and specific diffusible end-product of lipid peroxidation, and is found to induce the COX-2 genes in RAW264.7 cells (Kumagai et al 2004), thus reflecting the potential role of 4-HNE as perpetrator of inflammation. Furthermore exogenous micromolar levels of 4-HNE increases the expression of several genes eg, heme oxygenase-1, collagen α1(I), and aldose reductase (Parola et al 1993; Basu-Modak et al 1996; Spycher et al 1997). Also 4-HNE has been reported to have chemotactic, cytotoxic and immunogenic properties both in vitro and in vivo (Schauf et al 1994; Steinerova et al 2001), and these effects were achieved in vitro with 4-HNE concentrations as low as 2.5 μM (Muller et al 1996). Data from the authors’ laboratories indicate increased 4-HNE-modified protein levels in airway and alveolar epithelial cells, endothelial cells and neutrophils in subjects with airway obstruction compared to subjects without airway obstruction (Rahman and MacNee 2000b; Rahman et al 2002). An important outcome of 4-HNE generation is its interaction with the important thiol antioxidant glutathione (GSH) (Tjalkens et al 1999). The conjugation of 4-HNE with GSH might be one of the important mechanism whereby a cell may lose its antioxidant pool leading to oxidative stress. Interestingly, increased formation of 4-HNE has also been reported to induce expression of glutamyl cysteine ligase (GCL) gene which increases synthesis of GSH. This might be an important cellular antioxidant adaptation during oxidative stress. Inhibition of lipid peroxidation, specifically the pathways leading to the production of 4-HNE and F₂-isoprostane, may therefore be important and novel targets for antioxidant therapy in inflammation and injury in patients with COPD.

One of deleterious outcomes of oxidative stress is the remodeling of ECM leading to lung injury. ROS activate latent proforms of matrix metalloproteinase (MMP) (Lindholt et al 2003), and antioxidant species decrease MMP expression and activation (Rajagopalan et al 1996). Cigarette smoke treatment of alveolar macrophages from subjects with COPD induced increased release of MMP-9 compared to that of non-smokers. MMP-9 has an ECM degrading activity, thus suggesting the role of oxidative components of cigarette smoke in increased elastolytic enzyme activity (Russell et al 2002). Increased proteolytic load due to MMP-9 has been attributed to increased neutrophil recruitment in the lungs that triggers degradation of ECM and basement membrane in the airways and lungs. Antioxidants N-acetylcysteine and pyrrolidine dithiocarbamate, and the NADPH oxidase inhibitors diphenylene iodonium chloride and apocynin decreased the production of MMP-2 and -9 in alveolar macrophages from surfactant protein D-deficient mice (Yoshida et al 2001). However, a recent study showed that mice lacking gp91phox, a phagocyte-specific component of the NADPH oxidase, developed extensive, spontaneous emphysematous destruction of their peripheral air spaces (Kassim et al 2005). In addition, peritoneal macrophages from gp91phox-null mice had greater MMP-12 activity than macrophages from wild type mice (Kassim et al 2005). These findings indicate that reactive intermediates provide a physiological mechanism to protect tissues from excessive macrophage-mediated damage during inflammation. Factors other than oxidative stress, such as ozone and lipid peroxides also induce collagen I and MMP-1 gene expression (Choi et al 1994). Other forms of oxidative stress derived from tert-butyl hydroperoxide and iron can also modify collagen synthesis, by a mechanism presumably involving redox sensor/receptor.

The proteinase-antiproteinase dysbalance is thought to be related to the increased proteolytic activity or protease expression observed in sputum, BAL fluid or tissue of patients with COPD, and tissue remodeling or destruction as seen in emphysema (Barnes et al 2003; Hogg 2004). Several studies reported enhanced levels or gene mutations of MMPs like MMP-1, MMP-9 or MMP-12 associated with COPD and lung function decline (Joos et al 2002; Culpitt et al 2005; Demedts et al 2006), the presence of fragments of ECM proteins like elastin or collagen (Dillon et al 1992; Stone et al 1995; Weathington et al 2006), and/or altered levels of ECM molecules in sputum, BAL fluid or lung tissue of patients with COPD (Lang et al 1994; Dentener et al 2005; Kranenburg et al 2006; Martin-Mosquero et al 2006). Extracellular matrix hyaluronan (HA) has a pro-inflammatory role and HA levels were found to be increased in sputum of COPD patients (Dentener et al 2006). Hyperactivity of MMPs also leads to the generation of secondary metabolic reactive species (Repine et al 1997). ROS may alter remodeling of extracellular matrix (ECM) and blood vessels, stimulate mucus secretion, inactivate antiproteases, cause apoptosis, and regulate cell proliferation (Rahman and MacNee 1996a, 1996b, 1999). Furthermore, increased levels of ROS have been implicated in initiating inflammatory responses in the lungs through the activation of transcription factors such as nuclear factor-kappaB (NF-κB) and activator protein-1 (AP-1), signal transduction, chromatin remodeling and gene expression of pro-inflammatory mediators (Rahman and MacNee 1998).
ment with HA partially blocked LPS (1 ng/ml) induced TNFα release by blood cells from COPD patients (Dentener et al. 2006). Thus the high levels of HA in COPD subjects would be a consequence of degradation of ECM, which in turn can bind to lung elastic fibers, thereby adaptively preventing their further degradation by protease (Cantor et al. 1997, 2000). Targeted deletion of neutrophil elastase or MMP-12 protects from the development of cigarette smoke or gp91 deficiency-induced emphysema (Hautamaki et al. 1997; Shapiro et al. 2003; Kassim et al. 2005). Furthermore, the structural alterations in ECM proteins may provoke an immune reaction, whereas degradation fragments generated during extensive tissue remodeling may lead to antigenic fragments also provoking an immune reaction. More specifically, exposure to reactive oxygen or nitrogen intermediates or aldehydes present in smoke or produced by inflammatory cells may lead to adduct formation of structural (lipo)proteins in the lung eg, as a result of lipid peroxidation or protein carbonyl formation (Rahman and Adcock 2006). In turn, this may result in the formation of neo-epitopes of ECM proteins (Kirkham et al. 2004; Morquette et al. 2006). The induction of neo-antigens upon tobacco smoke exposure may lead to specific B and T cell clones that provoke autoimmune reactions. Alternatively, during embryogenesis specific B and T cells are being selected and generated by peptide fragments of extracellular matrix components secreted by lung epithelial cells. At that moment, the immune system is tolerant to these peptides. Upon smoke exposure, these B and/or T cells may be activated causing the inflammatory phenotype and specific lung parenchymal destruction as seen in emphysema. Some of the mechanisms are outlined in Figure 1. As COPD is detected in an advanced form only at later age, other models are needed to study these and other pathogenetic mechanisms of COPD.

Cytokines and chemokines

Inflammation is one of the hallmarks of COPD. Inflammatory cells are known to migrate to chemotactic gradients, or to express and produce pro- or anti-inflammatory proteins, MMPs and other proteases, growth factors, antibacterial proteins like α-defensins, β-defensins and cathelicidins, and/or to degranulate by a plethora of molecules including cytokines, chemokines, growth factors, and arachidonic acid derivatives (like prostaglandins, leukotrienes and thromboxanes) (De Boer 2002; Bals and Hiemstra 2006; Rahman and Adcock 2006; Rolin et al. 2006). Classical cytokines include TNFα, interleukins (ILs) and interferons (IFNs). Chemokines can be subdivided into four subfamilies based on their structural homology around 4 cysteine residues: -C-, -CC-, -CXC-, and -CXXXC-, in which X substitutes for any amino acid. The major subfamilies are the -CXC- (or: α chemokines, and the -CC- chemokines (or: β-) chemokines) (De Boer 2002). Gene expression of cytokines and chemokines is regulated by transcription factors including AP-1 and Fos, and signal transduction kinases like mitogen activated protein kinase (MAPK) p38, Jun kinase (JNK), and NF-κB, and are synthesized by both structural cells (such as fibroblasts, epithelial, endothelial, and muscle cells) and inflammatory cells of the innate and the adaptive immune system. They act via specific membrane-bound receptors resulting in cell specific reactions. In patients with COPD, protein and/or mRNA levels of different cytokines and chemokines have been found to be altered compared with subjects without COPD (Table 1).

Among these, TNFα or TNFR levels, soluble IL-1 receptor antagonist (sIL-1Ra), CCL2 (monocyte chemotactant protein 1, MCP-1) and its receptor CCR2, CCL3 (macrophage inflammatory protein 1α, MIP-1α) and CCL4 (MIP-1β) and their receptor CCR5, CXCL8 (IL-8), and CXCL10 (interferon-inducible protein 10, IP-10) can be discerned as pro-inflammatory factors. In addition to the inflammatory effects, recent studies provided more evidence that cytokines and chemokines are also involved in tissue remodeling apart from growth factors, pointing to cytokine-driven effects of inflammatory cells on epithelial wound repair (De Boer et al 2007) (Figure 1).

CCL2 is produced by a variety of cells including macrophages, endothelial and epithelial cells. It binds to CCR2, a receptor that is highly expressed on circulating blood DCs and monocytes, and thus primarily controls their recruitment from the blood vessels to the tissue. CCL2 is also involved in tissue remodelling. In vitro, CCL2 and its receptor CCR2 were demonstrated to be directly involved in endothelial and lung epithelial cell proliferation, migration and wound closure (De Boer et al 2007). In addition, CCL2 was found to stimulate collagen synthesis in rat lung fibroblasts via a TGFβ1-dependent pathway, hence potentially contributing to
Future antioxidant and anti-cytokine therapy in COPD

Tobacco smoke

Figure 1 Simplified summary of inflammatory and remodeling mechanisms in the airways in COPD. Exposure to cigarette smoke in susceptible individuals leads to an abnormal inflammation and tissue remodeling. This appears to be self-perpetuating and may be linked to infection. Tobacco smoke activates different cell types including macrophages, epithelial and smooth muscle cells to produce cytokines, growth factors or proteases. Reactive molecules in tobacco smoke stimulate airway macrophages to produce cytokines and reactive oxygen or nitrogen species. Activated macrophages and epithelial cells attract and activate inflammatory cells including monocytes, macrophages, neutrophils and T cells. Alternatively, reactive species may react with extracellular matrix (ECM), and lipid moieties causing cell damage, gene expression or oxidative stress in different cell types. Chemokines like CXCL-8 and CXCL-1 cause T cell and neutrophil chemotaxis and activation of neutrophils to degranulate proteases like elastase and MMPs and produce reactive oxygen species like hydrogen peroxide or O₂⁻. Radicals may activate proteases that in turn fragment ECM molecules and/or form ECM neo-epitopes. Oxygen radicals may also react with ECM leading to adducts or neo-epitopes. Altered or fragmented ECM molecules may stimulate inflammation and auto-immune-like reactions. Tobacco smoke may also activate smooth muscle cells and fibroblasts to produce pro-inflammatory cytokines and growth factors (GF) like VEGF leading to Th1-mediated inflammation and vascular remodeling. Loss of epithelial cells due to direct toxicity of smoke, TNFα-induced apoptosis, or degradation of ECM, induces a repair process. Growth factors like EGF, FGF, TGFβ1 and VEGF stimulate tissue repair and vascular remodeling seen in COPD. Epithelial remodelling (squamous or mucous metaplasia, hyperplasia) may be due to excessive growth factor production or by TNFα resulting in a loss of lung clearance function and mucus hyperproduction. A-HNE, 4-hydroxy-2-nonenal; ROS, reactive oxygen species; RNS, reactive nitrogen species.

a fibrogenetic remodelling as seen in COPD. In turn, TGFβ1 was reported to induce CCL2 protein levels via downstream intracellular mechanisms including ROS, and MAPK p38 and p42/44 in mesangial cells (Cheng et al 2005). Results from studies in mice and cell lines suggest that oxidative stress activates MAPK p42/44 and p38 which stimulates the expression of TNFα, IL-1β, CCL2 and CXCL10 (Nishi et al 2005; Guest et al 2006; Huang et al 2006; Loke et al 2006). Oxidative stress led to an influx of macrophages and increased expression of proteins like NADPH oxidase, thioredoxin-1 and inducible nitric oxide synthase, via a CCR2-mediated mechanism (Hayasaki et al 2006). These effects were inhibitable by antioxidants including N-acetylcysteine, ascorbic acid, quercetin or overexpression of SOD1, deletion of CCR2, or by inhibition of MAPK p42/44 or p38 (Nishi et al 2005; Guest et al 2006; Hayasaki et al 2006; Huang et al 2006; Loke et al 2006). Hence the CCL2-CCR2-TGFβ axis may potentially contribute to tissue remodeling and inflammation as seen in COPD.

CXCL1 (GROα) and CXCL8 are produced by both structural and inflammatory cells including macrophages. Both bind to their receptor CXCR2 whereas CXCL8 binds also to CXCR1. Both receptors are expressed on neutrophils while CXCR2 is also expressed on other inflammatory cells including a subset of CD8⁺ T cells, mast cells and macrophages. CXCL1 and CXCL8 are chemotactic and activate inflammatory cells while CXCL8 also induces neutrophils to degranulate and causes an oxidative burst. Recent studies demonstrated that cigarette smoke-conditioned medium induces human monocytes or macrophages in vitro to release CXCL8 (Karimi et al 2006; Yang et al 2006). As inhibitors of Toll-like receptor (TLR)-4, NF-κB and IKK-2
impaired the CXCL8 expression this points to a TLR4 and NF-κB dependent mechanism. In addition, CXCL8 release was significantly inhibited by the anti-oxidant glutathione pointing to a regulatory effect of the chemokine expression by the redox status of the cells (Yang et al 2006). In contrast to monocytes, cigarette smoke conditioned medium did not affect CXCL8 or IL-6 expression nor NF-κB activation in human lung epithelial cells in vitro (Moodie et al 2004). This suggests that oxidants exert differential effects on NF-κB signalling and chemokine expression depending on the cell type, and that inflammatory cells produce CXCL8 upon oxidant exposure. The study showed that also epithelial cells express CXCR2 suggesting a role for CXCL8/CXCR2 in wound repair (De Boer 2002). In vivo, impaired CXCR2 expression in mice resulted in delayed skin wound repair as compared to wild type mice, which cannot be restored by exogenous CXCL8 (Devalaraja et al 2000; Milatovic et al 2003). In contrast, topical application of CXCL1 or CXCL8 stimulated human skin wound repair in chimeric mice (Rennekampff et al 1997, 2000), and stimulated human colon, skin and lung epithelial proliferation and/or migration in vitro (De Boer et al 2007). Inhibition of CXCL8 or CXCL1 signaling or expression as a treatment target in COPD may hence inhibit inflammatory cell activation and tissue degradation, but may potentially delay wound repair in COPD.

Cigarette smoke has been shown in vivo to be a cause of increased adherence of leukocytes to vascular endothelium (Noguera et al 1998). Shen and co-workers (1996) have shown that cigarette smoke condensate induces the expression of a subset of cell adhesion molecules, such as intercellular adhesion molecule (ICAM-1), endothelial leukocyte adhesion molecule 1 (ELAM-1), and vascular cell adhesion molecule (VCAM-1) in human umbilical vascular endothelial cells associated with an increase in the binding activity of NF-κB suggesting the increased transendothelial migration of monocytes by cigarette smoking. The release of proinflammatory mediators, such as IL-1β and soluble ICAM-1, was increased by cigarette smoke exposure in bronchial epithelial cells cultured from biopsy materials obtained from patients with COPD compared to smokers (Rusznak et al 2000). Moreover, Scott and coworkers (2000) demonstrated a clear dose-dependent relationship between smoke intake and sICAM-1 concentrations and sICAM-1 concentrations substantially reduced in those who stopped smoking for a year but remained elevated in continuing smokers. These results suggest that patients with COPD have a greater susceptibility to the effects of cigarette smoke.

<table>
<thead>
<tr>
<th>Product</th>
<th>Type</th>
<th>Compartment</th>
<th>Level in COPD</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNFα</td>
<td>mRNA, protein</td>
<td>serum, sputum, leg muscle</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td></td>
<td>gene polymorphism</td>
<td>blood</td>
<td>Asian: yes; Caucasian: no</td>
</tr>
<tr>
<td>TNFR1</td>
<td>protein</td>
<td>sputum</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>TNFR2</td>
<td>protein</td>
<td>serum</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>CCL2</td>
<td>mRNA, protein</td>
<td>sputum, BAL, lung tissue, BEC</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>CCR2</td>
<td>mRNA, protein</td>
<td>lung tissue, BEC</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>CXCL1</td>
<td>protein</td>
<td>sputum, lung tissue</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>CXCL8</td>
<td>mRNA, protein</td>
<td>serum, sputum, BAL, lung tissue</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>CXCR2</td>
<td>mRNA, protein</td>
<td>BAL, PBL</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>TGFβ1</td>
<td>mRNA, protein</td>
<td>lung tissue, BEC</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td></td>
<td>protein</td>
<td>lung tissue</td>
<td>lower (emph)</td>
</tr>
<tr>
<td></td>
<td>gene polymorphism</td>
<td>blood</td>
<td>yes (COPD)</td>
</tr>
<tr>
<td>TGFβR1</td>
<td>protein</td>
<td>lung tissue</td>
<td>lower (emph)</td>
</tr>
<tr>
<td>TGFβR2</td>
<td>protein</td>
<td>lung tissue (arteries)</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>VEGF</td>
<td>protein</td>
<td>sputum, lung tissue</td>
<td>lower (emph)</td>
</tr>
<tr>
<td>VEGFR2</td>
<td>protein</td>
<td>lung tissue</td>
<td>lower (emph)</td>
</tr>
<tr>
<td>4HNE</td>
<td>protein adducts</td>
<td>lung tissue</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>NF-κB</td>
<td>protein</td>
<td>lung tissue</td>
<td>higher (COPD)</td>
</tr>
<tr>
<td>HDAC2</td>
<td>mRNA, protein</td>
<td>lung tissue</td>
<td>lower (COPD)</td>
</tr>
<tr>
<td>HDAC5</td>
<td>mRNA</td>
<td>lung tissue</td>
<td>lower (COPD)</td>
</tr>
<tr>
<td>HDAC8</td>
<td>mRNA</td>
<td>lung tissue</td>
<td>lower (COPD)</td>
</tr>
</tbody>
</table>

Abbreviations: BAL, bronchoalveolar lavage; BEC, bronchial epithelial cells; emph, emphysema; PBL, peripheral blood lymphocytes.
Growth factors: VEGF and TGFβ1

Growth factors can be divided into different superfamilies based on structural and functional homology. These families include vascular endothelial growth factor (VEGF), TGF-β, epidermal growth factor (EGF)-like growth factors, fibroblast growth factor (FGF) and insulin-like growth factor (IGF) (De Boer et al 2007). With regard to COPD several studies suggest the involvement of these families in either pulmonary inflammation like for VEGF and TGFβ1 (De Boer et al 1998; Takizawa et al 2001; Postma and Timens 2006), vascular or tissue remodeling like for EGF-like growth factors, FGFs and VEGFs (Kraenenburg et al 2002, 2005; De Boer et al 2006; Postma and Timens 2006), or oxidative stress as with TGFβ1 or FGF-7 (Rahman et al 2000; Rahman et al 2002; Ray et al 2003) (Table 1). A review on growth factors as a potential target for drug therapy is presented elsewhere (De Boer et al 2007).

VEGF receptor impairment, VEGF gene deletion or generation of antibodies against VEGF receptors all cause airspace enlargement in rodents without airway inflammation (Kasahara et al 2000). In addition, in murine models tobacco smoke exposure leads to decreased expression of VEGF and VEGF receptors as well as emphysematous lesions, as has also been observed in smokers with emphysema. Furthermore, blockade of VEGF receptors was shown to induce oxidative stress and alveolar cell apoptosis that was inhibited by exogenous administration of the SOD mimetic M40419 (Tuder et al 2003). These data link oxidative stress with development of emphysema and abrogated VEGF signaling rather than alveolar damage induced by inflammation alone. Tuder and coworkers proposed a disturbed balance between oxidative stress, proteinases, antiproteinases and apoptosis, and lung inflammation as a non-specific reaction secondary to alveolar tissue damage (Tuder et al 2006). However, these data may not be applied to COPD as a whole as VEGF and VEGFR expression was observed to be enhanced in relation to vascular remodeling in non-emphysematous patients making these patients less eligible for VEGF therapy (Kraenenburg et al 2005; De Boer et al 2007).

TGFβ1 has been associated with COPD either as a result of oxidative stress or an imbalance in proteinases and antiproteinases, but may also be related to an aberrant repair process and hence progression of COPD (Postma and Timens 2006; Rahman and Adcock 2006). TGFβ1 expression was demonstrated to be enhanced in patients with COPD (De Boer et al 1998) but decreased in patients with emphysema (Zandvoort et al 2006). The expression of intracellular inhibitory signaling proteins of TGFβ1, Smad6 and Smad7, was observed to be decreased both in bronchial and alveolar tissue from patients with COPD, whereas in expression of stimulatory Smad molecules including Smad3 was unaltered (Springer et al 2004; Zandvoort et al 2006). Smad7 is involved in the regulation of expression of inflammatory proteins. In vivo wound healing study with mice demonstrated that overexpression of Smad7 inhibits TGFβ1, CCL2, VEGF, MMP-9 and TIMP-2 protein and mRNA expression (Saika et al 2005). Reducing overexpression of Smad7 in patients with inflammatory bowel disease (IBD) using antisense Smad7 oligonucleotides caused a decreased production of proinflammatory cytokines IFNγ and TNFα upon treatment of intestinal tissue explants and cells with TGFβ1 (Monteleone et al 2001). With regard to COPD, it is not known whether Smad7 downregulation is intrinsic or due to inflammation, oxidative stress, or other factors, and what the consequences are of differential expression of TGFβ1 in patients with COPD or emphysema alone. An alternative hypothesis is that tobacco smoke exposure causes excessive growth factor production resulting in tissue remodeling, independent of inflammation. Recent data from a murine study (Churg et al 2006) provided support for this idea. Their study demonstrated that short-term smoke exposure for 2 hours stimulated early growth factor expression including TGFβ1 and type 1 procollagen synthesis before the onset of inflammation. Upon chronic smoke exposure for up to 6 months profibrotic growth factor expression continued as well as tissue remodeling characterized by enhanced collagen deposition, while other studies showed the development of airway inflammation and emphysema in rodents in this period. Taken together, the balance between TGFβ1 and Smad7 expression in pulmonary cells of patients with COPD seems to be delicate and may affect tissue remodeling and inflammation differently depending on the COPD phenotype. Targeting TGFβ1 as a therapy in COPD requires more studies on the precise role of these factors in the pathogenesis of COPD.

Figure 1 outlines briefly the proposed remodeling and inflammatory mechanisms in COPD, whereas Figure 2 summarizes potential intervention strategies. Based on this, specific anti-inflammatory therapies are being developed for COPD (De Boer 2005).

Current therapies

Therapies for COPD are mainly based on anti-inflammatory drugs for treating asthma, including corticosteroids or theophylline with or without bronchodilators like β2-agonists. Some studies reported reduction of the number
of exacerbations, improved quality of life and an decline in FEV₁ after short- or long-term treatment with inhaled corticosteroids, or no effect on lung function (Gartlehner et al 2006). Though some recent studies using higher doses or longer duration of treatment showed reduced airway inflammation, steroid treatment of patients with COPD is rather ineffective in reducing the decline in lung function (Barnes and Stockley 2005; Gan et al 2005). Adverse effects of steroids include increased risk of hip fractures and osteoporosis, skin bruising and candidiasis (Gartlehner et al 2006), and the airway remodeling is not positively affected by the current treatment. Anti-oxidant therapy by mucolytics such as N-acetylcysteine is also being used as a treatment reducing acute exacerbation frequency, but generally fails to reduce airway inflammation or decline in lung function (Poole and Black 2006; Sadowska et al 2007). Adverse effects of these mucolytic agents are rarely seen. The final part of this review focuses on the recent developments and advances in potential anti-oxidant and anti-cytokine treatment (Table 2).

**Development of antioxidant agents and anti-inflammatory therapies**

**Development of antioxidant therapies**

Systemic and local antioxidant capacity and antioxidant vitamins

Smoking and exacerbations of COPD result in decreased antioxidant capacity in plasma in association with depleted protein sulphhydrlys in the plasma (Rahman et al 1996, 1997; Corradi et al 2003). The decrease in antioxidant capacity...
Future antioxidant and anti-cytokine therapy in COPD

Antioxidant capacity in smokers occurs transiently during smoking and resolves rapidly after smoking cessation. In exacerbations of COPD, however, antioxidant capacity remains low for several days after the onset of the exacerbation, tending to return towards normal values at the time of recovery from the exacerbation (Rahman et al 1997). The depletion of antioxidant capacity could in part be explained by the increased release of ROS from peripheral blood neutrophils, as shown by a significant negative correlation between neutrophil superoxide anion release and plasma antioxidant capacity (Rahman et al 1996). Thus there is clear evidence that oxidants in cigarette smoke markedly decreases plasma antioxidants, which may play an important role in the pathogenesis of COPD. Furthermore, it is possible that inter-individual differences in antioxidant capacity may contribute to the differences in susceptibility to cigarette-smoke induced COPD. Thus, it is imperative to propose the rationale for antioxidant therapy ameliorating the increased oxidative stress and consequently the inflammatory response in COPD.

<table>
<thead>
<tr>
<th>Target</th>
<th>Compound</th>
<th>Mechanism</th>
<th>Status for disease</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>TNFα</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Infliximab</td>
<td>chimeric IgG1 TNFα antibody</td>
<td>scavenging TNFα</td>
<td>A (2) RA, CD, psor. (COPD, SA)</td>
</tr>
<tr>
<td>Etanercept</td>
<td>hu TNFR2-Fc IgG1 fusion protein</td>
<td>scavenging TNFα</td>
<td>A (2) RA, psor. (SA)</td>
</tr>
<tr>
<td>Adalimumab</td>
<td>hu IgG1 TNFα antibody</td>
<td>scavenging TNFα</td>
<td>A RA</td>
</tr>
<tr>
<td>PEG-sTNFRα</td>
<td>PEGylated hu TNFRα</td>
<td>scavenging TNFα</td>
<td>2 RA</td>
</tr>
<tr>
<td>Afelimomab</td>
<td>murine TNFα antibody</td>
<td>scavenging TNFα</td>
<td>3 sepsis</td>
</tr>
<tr>
<td>ISIS04838</td>
<td>antisense TNFα oligonucleotide</td>
<td>impaired TNFα translation</td>
<td>2 RA</td>
</tr>
</tbody>
</table>

| **CCL2/CCR2** | | | |
| MLN1202 | humanized CCR2 antibody | inhibits CCL2 binding | 2 ART, MS |
| CCX915 | small molecule | CCR2 antagonist | 1 MS |
| ABN912 | CCL2 antibody | scavenging CCL2 | 1,2 COPD, RA |
| INCB3284 | small molecule | CCR2 antagonist | 2 RA |

| **CCL2/CXCL8/CXCR1/CXCR2** | | | |
| SB656933 | small molecule | CXCR2 antagonist | preclinical |
| AZD8309 | pyrimidine derivate | CXCR2 antagonist | 1,2 COPD, RA |
| SCH527123 | cyclobutenedione derivate | CXCR2 antagonist | preclinical |
| | Thiazolpyrimidine derivate | CXCR2 antagonist | experimental |
| | Imidazolopyrimidine derivate | CXCR2 antagonist | experimental |

| **PDE4 inhibitors** | | | |
| Roflumilast | methoxy-N-pyridyl-benamide derivate | inhibition of PDE4 | 3 COPD, asthma |
| Cilomilast | cyano-cyclohexanecarboxylate derivate | inhibition of PDE4 | 3 COPD |
| Oglumilast | small molecule | inhibition of PDE4 | 2 COPD |
| Tetomilast | thiazole derivate | inhibition of PDE4 | 2 (3) COPD (IBD) |
| SB256066 | inhaled PDE4 inhibitor | 1 COPD, asthma, AR |

| **Signal transduction** | | | |
| BMS345541 | antisense NFκB oligonucleotide | impaired NFκB translation | experimental |
| SPC839 | imidazquinolinone derivate | IKK-1 and -2 inhibitor | preclinical |
| RV676757 | quinazoline derivate | IKK-2 inhibitor | experimental |
| SB856553 | pyridylimidazolbutyr-1-ol | JNK2, MAPK p38 inhibitor | 1 RA, CD, AR, psor |
| SB681323 | oral MAPK p38 inhibitor | 1 COPD, RA, ART |
| BIRB976 | diaryl urea derivate | MAPK p38 inhibitor | 3 inflam, RA, CD |
| SD-282 | indole-5-carboxamide | oral MAPK p38 inhibitor | preclinical |

| **Anti-oxidants** | | | |
| M40403 | Mn-pentaazatetracyclohexacosatriene | soluble SOD mimic | 2/3 pain |
| M40419 | small molecule | soluble SOD mimic | 1 pain |

Status indicates the experimental, preclinical or clinical trial phase for the mentioned disease.

Abbreviations: A, approved by US Federal drug administration or European union; AR, allergic rhinitis; ART, artherosclerosis; CD, Crohn’s diseases; IBD, inflammatory bowel disease; inflam, inflammation; MS, multiple sclerosis; psor, psoriasis; RA, rheumatoid arthritis; SA, severe asthma.
et al 1990; Bridges et al 1993; van Antwerpen et al 1993; Mezzetti et al 1995; Tug et al 2004). Moreover, decreased vitamin E and vitamin C levels were reported in leukocytes and BAL fluids from smokers. Ascorbate appears to be a particularly important antioxidant in the plasma. Cigarette smoke-induced lipid peroxidation of plasma in vitro is decreased by ascorbate (Cross et al 1994). Reduced levels of vitamin E and a marginal increase in vitamin C in the BAL fluid of smokers, compared to nonsmokers have been shown (Rahman and MacNee 1996a). Similarly, alveolar macrophages from smokers have both increased levels of ascorbic acid and augmented uptake of ascorbate, suggesting that these cells are trying to redress their antioxidant balance (Rahman and MacNee 1996a).

Dietary antioxidants supplementation is one of the simplest approaches to boost antioxidant defense systems. Supplementation of vitamin C, vitamin E and β-carotene has been attempted in cigarette smokers and patients with COPD (Cross et al 1993; Rautalahti et al 1997; Steinberg and Chait 1998; Aghdassi et al 1999; Habib et al 1999; Lykkesfeldt et al 2000; Uneri et al 2006). In the general population there is a positive association between dietary intake of antioxidant vitamins and lung function. Epidemiological studies have demonstrated negative associations of dietary antioxidant intake with pulmonary function and with obstructive airway disease (Grievinke et al 1998). Britton and co-workers (Britton et al 1995) showed a positive association between dietary intake of the antioxidant vitamin E and lung function in a population of 2,633 subjects, supporting the hypothesis that this antioxidant may have a role in protecting against the development of COPD. Another study has suggested that antioxidant levels in the diet could be a possible explanation for differences in COPD mortality in different populations (Sargeant et al 2000). Dietary polyunsaturated fatty acids may also protect cigarette smokers against the development of COPD (Shahar et al 1999). These studies support the concept that dietary antioxidant supplementation including polyphenols may be a possible therapy to prevent or inhibit the oxidative stress and inflammatory responses, which are key features in the development of COPD. However, robust clinical trials using dietary antioxidant vitamins and polyphenols are urgently needed to address the beneficial effects of these antioxidants in COPD.

**Directly increasing lung antioxidant capacity**

The development and progress of COPD is associated with increased oxidative stress or decreased antioxidant resources (Boots et al 2003). The most direct way to redress the oxidant/antioxidant imbalance in COPD would be to increase the pulmonary capacity by antioxidants (Table 3). A variety of means by which to do this have been attempted with varying success.

**Glutathione and its biosynthesis**

The thiol antioxidant glutathione (GSH) is concentrated in epithelial lining fluid compared with plasma and has an important protective role in the airspaces and intracellularly in epithelial cells. Several studies have suggested that GSH homeostasis may play a central role in the maintenance of the integrity of the lung airspace epithelial barrier. Decreasing the levels of GSH in epithelial cells leads to loss of barrier function and increased permeability (Morrison et al 1999). Human studies have shown elevated levels of glutathione in epithelial lining fluid in chronic cigarette smokers compared with non-smokers (Morrison et al 1999). However, this increase is not present immediately after acute cigarette smoking (Morrison et al 1999). The two-fold increase in BALF GSH in chronic smokers may not be sufficient to deal with the excessive oxidant burden during smoking, when acute depletion of GSH may occur (Harju et al 2002). In addition, the immunoreactivity of γ-glutamylcysteine synthetase (γ-GCS; now called as glutamate cysteine ligase, GCL), the rate limiting enzyme in GSH synthesis, was decreased in the airways of smokers compared to non-smokers, suggesting that cigarette smoke predisposes lung cells to ongoing oxidant stress (Harju et al 2002). Neurohr and colleagues recently showed that decreased GSH levels in BALF cells of chronic smokers were associated with a decreased expression of γ-GCS/GCL-light subunit without a change in γ-GCS/GCL-heavy subunit expression (Neurohr et al 2003). Increasing the activity of γ-GCS/GCL, would be expected to increase cellular GSH levels. The induction of γ-GCS/GCL by molecular means to increase cellular GSH levels or γ-GCS/GCL gene therapy also holds great promise in protection against chronic inflammation and oxidant-mediated injury in COPD.

Direct increase of lung cellular levels of GSH would be a logical approach to enhance the antioxidant potential in the treatment of COPD. In fact, extracellular augmentation of GSH has been tried through intravenous administration of GSH, oral ingestion of GSH, and aerosol inhalation of nebulized GSH in an attempt to reduce inflammation in various lung diseases (Rahman and MacNee 1999, 2000a, 2000b). However, these routes of administration lead to undesirable effects suggesting that direct GSH therapy may not be an appropriate way of increasing GSH levels in lung epithelial lining fluid and cells in COPD. The bioavailability of
Future antioxidant and anti-cytokine therapy in COPD

GSH, pH, osmolility in the inflammatory micro-environment and the resultant formation of toxic products (GSSG and GSH-adducts) are further challenges for direct GSH administration. Alternative formulations may address bioavailability, such as liposomal delivery, but at present it seems that direct administration of GSH will not be successful in treating COPD.

**N-acetyl-L-cysteine (NAC)**

NAC, a cysteine-donating reducing compound, acts as a cellular precursor of GSH and becomes de-acetylated in the gut to cysteine following oral administration (Cotgreave 1997; Repine et al 1997). NAC may also reduce cystine to cysteine, which is an important mechanism for intracellular GSH elevation in vivo in lungs. It reduces disulphide bonds (a property of a good reducing agent), but also has the potential to interact directly with oxidants. NAC is also used as a mucolytic agent, to reduce mucus viscosity and to improve mucociliary clearance.

Pharmacological administration of NAC has been used in an attempt to enhance lung GSH in patients with COPD with varying success (Rasmussen and Glennow 1988; Bridgeman et al 1994). Schooten et al have reported that in a randomized, double-blind, placebo-controlled Phase II trial, a six month oral dose of 600 mg twice a day (b.i.d), reduced various plasma and BAL fluid oxidative biomarkers in smokers (Van Schooten et al 2002). Similarly, it has been shown that treatment with NAC 600 mg once daily for 12 months also reduced the concentration of H₂O₂ in EBC compared to placebo in stable COPD patients (Kasielski and Nowak 2001). A more recent clinical

<table>
<thead>
<tr>
<th>Name/Company</th>
<th>Antioxidant</th>
<th>Disease and phase of clinical trials</th>
<th>Mechanism of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>AstraZeneca</td>
<td>N-Acetyl-L-cysteine – AstraZeneca (Mucomyst®; AstraZeneca)</td>
<td>Bronchiectasis; COPD; Cystic fibrosis</td>
<td>Antioxidant</td>
</tr>
<tr>
<td>Zambon (Italy)</td>
<td>N-acetyl-L-cysteine (Flumucil; NAC; NSC-11118)</td>
<td>Pulmonary fibrosis; COPD</td>
<td>Reducing agent; Oxygen radical formation antagonist</td>
</tr>
<tr>
<td>Galephar; Cystic Fibrosis Foundation Therapeutics; SMB Laboratories</td>
<td>Nacystelyn</td>
<td>COPD; Cystic fibrosis Phase II trial</td>
<td>Antioxidant; Mucolytic</td>
</tr>
<tr>
<td>Refarmed</td>
<td>Erdosteine</td>
<td>Bronchitis; Cough; Cystic fibrosis</td>
<td>Mucolytic agent;</td>
</tr>
<tr>
<td>Nattermann</td>
<td>EbseleN</td>
<td>Asthma; Atherosclerosis; Myocardial ischaemia Phase I trial</td>
<td>Antioxidant agent</td>
</tr>
<tr>
<td>Redox Bio Science Inc</td>
<td>Recombinant human thioredoxin</td>
<td>Lung injury; ARDS, COPD</td>
<td>Thioredoxin stimulator;</td>
</tr>
<tr>
<td>OXIS International</td>
<td>Glutathione peroxidase mimetics – Alteon (Organoselenium compounds)</td>
<td>Inflammation, COPD Preclinical</td>
<td>Antioxidant</td>
</tr>
<tr>
<td>Sami Labs</td>
<td>Curcumin C3 Complex Curcumin</td>
<td>Cystic fibrosis, COPD, Cancer</td>
<td>Antioxidant and anti-inflammatory</td>
</tr>
<tr>
<td>Royalmount Pharma</td>
<td>Resveratrol and its analogs</td>
<td>Infection, COPD, Cancer Phase II</td>
<td>Anti-inflammatory; Anti-cancer; Free radical scavenger</td>
</tr>
<tr>
<td>US Department of Health and Human Services</td>
<td>Tempol (Spin trap)</td>
<td>Reperfusion injury; Inflammation; Cancer; ARDS, COPD Preclinical</td>
<td>Apoptosis stimulator; Free radical scavenger; Anti-inflammatory; Antioxidant agent</td>
</tr>
<tr>
<td>CereMedix</td>
<td>CereMedixResearch programme: COPD therapy</td>
<td>COPD Preclinical</td>
<td>Antioxidant</td>
</tr>
<tr>
<td>Aeolus Pharmaceuticals Inc Corp</td>
<td>Catalytic antioxidants</td>
<td>Reperfusion injury, Asthma, COPD, Cancer, ARDS No development on trials reported</td>
<td>Free radical scavenger; Antioxidant agent</td>
</tr>
<tr>
<td>Savient Pharmaceuticals Inc</td>
<td>OxSODrol (Superoxide dismutase)</td>
<td>Asthma, COPD, ARDS</td>
<td>Superoxide dismutase stimulator; Antioxidant agent</td>
</tr>
</tbody>
</table>

ARDS: acute respiratory distress syndrome
trial also proves that oral administration of NAC 600 mg b.i.d. for 2 months rapidly reduces the oxidant burden in airways of stable COPD patients (De Benedetto et al 2005). This reduction in oxidative biomarkers results in clinical benefit such as reduction in bronchial hypersecretion, in addition to decline in FEV$_1$ and in exacerbations (Stey et al 2000). Orally dosed NAC has been shown to increase phagocytic activity of BAL macrophages from healthy smokers, but similar results were not seen in COPD patients, possibly due to active concentrations of NAC not reaching the lung, as in vitro analysis of cells support an induction of phagocytosis by NAC. It has also been reported recently that orally dosed NAC increased the quadriceps endurance time of severe COPD patients (Koechlin et al 2004), thus suggesting that NAC administration may have beneficial effects on the systemic oxidative stress associated with COPD. However, a multi-centre study using NAC delivered by metered dose inhalers in patients with chronic cough failed to show a positive effect on well being, sensation of dyspnoea, cough or lung function (Dueholm et al 1992).

Whilst there is a body of evidence that the administration of NAC provides benefit for COPD patients, it is not clear whether this represents a maintenance therapy (Decramer et al 2001, 2005). A phase III multicentre Bronchitis Randomized on NAC Cost-Utility Study (BRONCUS) has recently been completed, with the aim of addressing this question and determining whether the effectiveness of NAC as an “antioxidant” results in an alteration in the rate of decline in FEV$_1$, exacerbation rate and quality of life in patients with moderate to severe COPD and hence support NAC administration as a maintenance therapy for COPD (Gerrits et al 2003; Dekhuijzen 2004). The results of the phase III BRONCUS trials showed no effect on decline in FEV$_1$, but a reduction in hyperinflation in patients with severe COPD and exacerbation rate in patients who are not treated with inhaled glucocorticoids (Decramer et al 2001, 2005).

N-acystelyn (NAL)

NAL is a lysine salt of N-acetyl-L-cysteine, and is a mucolytic and antioxidant (reducing) thiol compound. The advantage of NAL over NAC is that it has a neutral pH in solution, whereas NAC is acidic. NAL can be aerosolized into the lung without causing significant side effects (Gillissen et al 1997). Gillissen and co-workers compared the effect of NAL and NAC and found that both drugs enhance intracellular glutathione in alveolar epithelial cells and inhibited hydrogen peroxide and O$_2^•$ released from human blood-derived polymorphonuclear cells (PMN) from smokers with COPD (Gillissen et al 1997). NAL also inhibited ROS generation induced by serum-opsonised zymosan by human neutrophils. This inhibitory response was comparable to the effects of NAC (Gillissen et al 1997). Antonicelli and colleagues have shown that NAL inhibited oxidant-mediated CXCL8 release in alveolar epithelial A549 cells suggesting an anti-inflammatory effect of NAL (Antonicelli et al 2002). Therefore, NAL may represent an interesting alternative approach to augment the antioxidant screen and thereby inhibiting inflammatory responses in the lungs and has the advantage over other antioxidant agents in that it may be administered by inhalation. The clinical trials using NAL in the treatment of COPD are in progress.

N-isobutrylcysteine (NIC)

Because NAC becomes hydrolyzed in biological systems, the measured bioavailability of the drug is low. Thus, it was speculated that a drug might be synthesized that possessed greater bioavailability than NAC, and could be used as a more effective treatment for chronic bronchitis. N-isobutrylcysteine (NIC) is a NAC-like thiol compound that does not undergo effective first-pass hydrolysis and hence has a higher oral bioavailability than NAC (Ekberg-Jansson et al 1999). The oral bioavailability can be as high as 80%, dependent on food intake. However, when evaluated as a therapy for exacerbations of chronic bronchitis, NIC performed no better than placebo drugs, and not as well as NAC (Gillissen et al 1997). Furthermore, a study of NIC also failed to reduce exacerbation rates in patients with COPD (Ekberg-Jansson et al 1999).

Erdosteine

Erdosteine is a new thiol compound that also acts as an anti-oxidant, but in addition has mucoactive properties and reduces bacterial adheriveness. In the “Equalife” randomized placebo controlled clinical study, erdosteine was dosed orally 300 mg twice a day (b.i.d) for a period of 8 months (Moretti et al 2004). Patients receiving erdosteine had significantly fewer exacerbations and spent less days in hospital than the placebo group. Moreover, patients receiving erdosteine showed no reduction in lung function over this period and showed a significant improvement in health related quality of life. It is not clear whether this clinical benefit is due to antioxidant effects of erdosteine. The mucolytic effect of erdosteine is perhaps due to the presence of a sulphhydryl group. It may be possible that erdosteine may reduce bacterial colonization through a direct effect on adhesion.
Procysteine
Procysteine (L-2-oxothiazolidine-4-carboxylate), is a cysteine donating compound which increases the cysteine levels of the cells and has a greater bioavailability than NAC. This thiol compound is well tolerated has been shown to increase mitochondrial levels of GSH in alveolar type II cells (Guidot and Brown 2000). Glutathione esters, particularly GSH monoethyl esters can increase the GSH levels of the cells by cleavage of ester bond (an ethyl group esterified to glycine). GSH esters have been shown to increase GSH levels in the lungs of rats, however, this compound can be cytotoxic and variation in the uptake levels of GSH has been shown in various cellular models (Butterworth et al 1993).

Antioxidant enzyme mimetics and spin traps
Increased activity of antioxidant enzymes (superoxide dismutase and catalase) in alveolar macrophages from young smokers have been reported (McCusker and Hoidal 1990). However, Kondo and co-workers (Kondo et al 1994) found that the increased superoxide generation by alveolar macrophages in elderly smokers was associated with decreased antioxidant enzyme activities when compared with non-smokers. The activities of CuZnSOD, glutathione-S-transferase and glutathione peroxidase (GP) are all decreased in alveolar macrophages from elderly smokers (Gilks et al 1998).

The activities of SOD and glutathione peroxidase have been shown to be higher in the lungs of rats exposed to cigarette smoke. McCusker and Hoidal (1990) have also demonstrated enhanced antioxidant enzyme activity in alveolar macrophages from hamsters following cigarette smoke exposure, which resulted in reduced mortality when the animals were subsequently exposed to >95% oxygen. They speculated that alveolar macrophages undergo an adaptive response to chronic oxidant exposure that ameliorates potential damage to lung cells from further oxidant stress. The mechanism(s) for the induction of antioxidant enzymes in erythrocytes, alveolar macrophages, and lungs, by cigarette smoke exposure are currently unknown.

Spin traps such as α-phenyl-N-tert-butylnitrone react directly with reactive oxygen and reactive nitrogen species at the site of inflammation (Chabrier et al 1999). In a recent study, Smith and colleagues have shown that intratracheal instillation of a catalytic antioxidant, manganese (III) mesotetakis (N,N'-diethyl-1,3-imidazolium-2-yl) porphyrin (AEOL 10150 and AEOL 10113) inhibited the cigarette smoke-induced inflammatory response (decreased number of neutrophils and macrophages) in rats after 2 d or 8 weeks (6 hours/day, 3 days/week) exposure (Smith et al 2002).

These compounds also mimic extracellular SOD and catalase, scavenging both lipid peroxides and peroxynitrite, and have been shown to be effective in a number of animal models of lung disease.

It has been shown that SOD mimetic M40419 blocked the development of emphysema and significantly reduced lung markers of oxidative stress in an animal model (Tuder et al 2003). Animal studies have shown that recombinant SOD treatment can prevent the neutrophil influx to the airspaces and CXCL8 release induced by cigarette smoking through a mechanism involving down regulation of NF-κB (Nishikawa et al 1999). This further substantiate the idea that generation of compounds with anti-oxidant enzyme properties may be able to act as novel anti-inflammatory drugs by regulating the molecular events in COPD.

Development of anti-inflammatory therapies

NF-κB inhibitors
Studies with IκBα mutants (Baldwin 1996; Ghosh et al 1998) gave the first evidence that NF-κB pathway could be specifically inhibited. Signal-induced phosphorylation and degradation of cytoplasmic IκBα is required for NF-κB pathway activation. However, an IκBα protein with mutations at serine-32 and 36 is not phosphorylated by IKK (IκBα kinase) and therefore not degraded by the proteasome. This IκBα mutant or super-repressor exerts its negative effect by sequestering NF-κB in the cytoplasm and thus prevents the induction of specific NF-κB target genes.

Another novel way whereby NF-κB activity may be regulated is by the use of inhibitors of proteasome function, which can reduce the degradation of IκB and thus prevent NF-κB activation (Baldwin 1996; Ghosh et al 1998). A series of peptide aldehydes such as MG101, MG132, and MG115, make up a family of agents that inhibit the protease activity of the proteasome. Lactacystin, another class of proteasome inhibitor, blocks proteolytic activity by acylating a threonine residue in one of the key proteasome subunits. Furthermore, a group of boronic acid peptides, including PS-341, are extremely potent inhibitors of proteasome function (Adams et al 1999), thus inhibiting activation of the NF-κB pathway. It is also possible that inhibitors of the ubiquitin ligase that mediates IκB ubiquitination may be a useful target in preventing proteasome degradation of IκB. Thus, a wide variety of potential inhibitors of proteasome function may have a therapeutic role in anti- NF-κB pathway dependent strategies.

Certain natural antioxidants/products such as flavonoids/polyphenols quercetin, curcumin, resveratrol, and myricetin...
are also known to mediate their anti-inflammatory properties through down-regulation of the NF-κB pathway (Tsai et al 1999; Holmes-McNary and Baldwin 2000). For example, resveratrol, which is found in red wine, can inhibit NF-κB activity and induce apoptosis in transformed cells, which may reduce mortality from coronary heart diseases, certain cancers and inflammatory diseases (Holmes-McNary and Baldwin 2000). Resveratrol has strong inhibitory effects on iNOS expression and NO generation in activated macrophages (Tsai et al 1999). Since treatment of macrophages with resveratrol blocks LPS-induced phosphorylation and degradation of IκBα to decrease NF-κB DNA binding activity, is suggestive of the fact that its anti-inflammatory effects may be due at least in part to the inhibition of NF-κB-dependent NO synthesis (Tsai et al 1999). Thus several of the biological activities of flavonoids may be mediated by their inhibition of the NF-κB pathway.

Thus it is evident that there are several possible approaches to inhibition of NF-κB, including gene transfer of IκB, inhibitors of IκB kinases (IKK), NF-κB-inducing kinase and IκB ubiquitin ligase, which regulate the activity of NF-κB, and inhibit the degradation of IκB (Delhase et al 2000). The most promising approach however, may be the inhibition of IKK-2 by small molecule inhibitors (Castro et al 2003) (Table 2), which suppress the release of inflammatory cytokines and chemokines from alveolar macrophages (Jazrawi et al 2003). This in particular might be more effective in COPD, particularly since alveolar macrophages are resistant to the anti-inflammatory actions of corticosteroids (see HDACs modifiers). It is however, of concern that long-term inhibition of NF-κB, with effective inhibitors may result in immune suppression and therefore impair host defenses. This concern is validated from a study that mice lacking NF-κB genes succumb to septicemia. However, alternative modulation of pathways of NF-κB activation via kinases other than IKK might be a more safer approach in inflammatory disease and would have less potential effect on innate and adaptive immune responses (Nasuhara et al 1999).

**PDE4 inhibitors**

Phosphodiesterase 4 (PDE4) is the predominant PDE isoenzyme in most inflammatory cells thought to have a role in the pathogenesis of COPD (Figure 2). Its activity is elevated in lung macrophages from COPD patients (Barber et al 2004). In contrast to steroids that have a limited anti-inflammatory efficacy in cigarette smoke models both in the mouse and guinea pig, there are increasing numbers of studies documenting the in vivo efficacy of PDE4 inhibitor in animal models of COPD. There are at least currently five oral PDE4 inhibitors in clinical development for COPD, one of which is suspended (C1393 in phase II, from Merck) (see Table 2). A major hurdle in their development has been to overcome their side effects which include nausea, emesis, and headache.

In 24 weeks Phase multi-center III trails in COPD patients (RECORD trial), oral administration of roflumilast or cilomilast improved pre- and post-bronchodilator FEV₁, (Rabe et al 2005; Rennard et al 2006). The health-related quality of life (SGRQ) was also improved when compared with the placebo control. In addition, exacerbation frequency was lower in drugs group than in the placebo group.

The relationship between these improvements in clinical outcome and potential anti-inflammatory activity has been investigated in a single study (Gamble et al 2003; Grootendorst et al 2005). After a 4-week treatment with roflumilast post-bronchodilator FEV₁ improved by 68.7 ml compared with placebo. Treatment with roflumilast significantly reduced the absolute numbers of neutrophils and eosinophils of sputum. These were paralleled with a reduction in CXCL8 and neutrophil elastase. Although a 12 weeks treatment with cilomilast had no effect on sputum neutrophils, macrophages, elastase, CXCL8 or lung function, bronchial biopsies demonstrated that cilomilast treatment was associated with significant reductions in CD8⁺ T lymphocyte and CD68⁺ cells. The results showed that related outcomes observed in longer term trials could be due, at least in part, to anti-inflammatory activity of drugs.

In an attempt to reduce the potential for systemic side effects and to administer relatively higher doses to the lung, inhaled PDE4 inhibitors are being developed. GSK842470 (AWD-12-281) was licensed from Elbion and reached Phase II for asthma and COPD but there have been unconfirmed reports that it had no advantage over oral PDE4 inhibitors. This compound no longer appears on GSK’s pipeline but remains in development for rhinitis by Elbion. Currently, GSK (SB256066, Phase I) and Pfizer (Phase II) are reported to have inhaled PDE4 inhibitors in clinical development for COPD.

Experimental data suggest that PDE4D inhibition is one likely cause of the side effects of the orally-delivered compounds, while PDE4B is a therapeutically relevant target. Therefore, PDE4 subtype inhibitors eg, PDE4B for treatment of COPD is being studied by Plexxikon.

**MAPK p38 inhibitors**

MAPKs play a key role in chronic inflammation and several complex enzyme cascades have now been defined (Johnson and Lapadat 2002). One of these, the p38 MAPK pathway, is
activated by cellular stress and regulates the expression of a wide variety of inflammatory cytokines that include CXCL8, TNFα and MMPs (Meja et al 2000). Small molecule inhibitors of MAP kinase p38, such as SB203580, SB 239063 and RWJ 67657 having a broad range of anti-inflammatory effects have been developed (Kumar et al 2003) (Table 2). Administration of SB203580 has beneficial effects in animal disease models such as collagen-induced arthritis and endotoxin-induced septic shock (Lee et al 1999). p38 has also been shown to upregulate cytokine production by several independent mechanisms, including direct phosphorylation of transcription factors, and direct or indirect (through downstream kinases such as MAPKAPK2) stabilization and increased translation of mRNAs containing 3% untranslated region adenylylate/uridylate-rich elements (AREs) by phosphorylation of ARE-binding proteins (Dean et al 2004; Briata et al 2005; Hitti et al 2006). These observations have attracted interest in p38 as a molecular target in the treatment of inflammatory human diseases. MAPK p38 has 4 isozymes. Each inhibitor has its own specificity towards one of more of these isozymes, causing differential effects Studies in healthy volunteers given p38α/p38β inhibitors found reductions in pro-inflammatory cytokine secretion from ex-vivo LPS-stimulated peripheral-blood mononuclear cells (PBMCs) (Parasrampuria et al 2003), and decreased LPS-induced pro-inflammatory cytokine production, neutrophil and endothelial-cell activation in vivo. SB239063 on the other hand reduces neutrophil infiltration and the concentrations of IL-6 and MMP-9 in BALF of rats after endotoxin inhalation, suggesting its potential as an anti-inflammatory agent in COPD (Underwood et al 2000).

The potential therapeutic utility of p38 MAPK inhibition in respiratory disease has been supported by data generated in a range of pulmonary inflammatory models in vivo including LPS induced pulmonary neutrophilia (Haddad et al 2001), bleomycin induced fibrosis (Matsuoka et al 2002), and antigen induced eosinophilia (Underwood et al 2000). A recent study demonstrated the efficacy of p38α MAPK inhibitor, SD282, in mouse COPD models (Fitzgerald et al 2006). In this model, SD-282 inhibited cigarette smoke induced pulmonary neutrophilia and macrophage recruitment. Although a number of oral p38 MAPK inhibitors are in clinical development for arthritis and cancer only two compounds are currently in development for COPD. GSK-681323 is currently in a 4 week Phase II trial where the efficacy outcome measures include lung function, sputum and serum biomarkers, including CRP. GSK-85633 is in Phase I. Although it is likely that such a broad spectrum anti-inflammatory drug might ensue some toxicity or impair natural immune responses, but inhalation route might be a feasible therapeutic approach.

**Elastase inhibitor**

Neutrophil elastase is a serine protease that is synthesized in the neutrophils and secreted following neutrophils activation. One of the major actions of neutrophil elastase is its ability to degrade matrix proteins and appears to be a key enzyme in the development of hereditary due to α1-antitrypsin deficiency. Although its role in the development of non hereditary emphysema is currently unclear it also has a broad range of actions that are consistent with it having a pro-inflammatory role in COPD pathogenesis (Shapiro et al 2003; Chughtai and O’Riordan 2004). Neutrophil elastase can increase MMPs activity by directly activating MMPs such as MMP-9 (Ferry et al 1997) and by inactivating the endogenous MMP inhibitor, TIMP (Okada et al 1988), thus potentially enhancing the role of MMPs in COPD. Neutrophil elastase has also been reported to stimulate mucin secretion and modulate apoptosis of human lung epithelial cells. Therefore, neutrophil elastase may play a role in emphysema and lung remodeling through matrix degradation and by inducing apoptosis.

In cigarette smoke models, neutrophil elastase knockout mice are protected from emphysema development and this effect is accompanied by an inhibition of both neutrophils and macrophages recruitment (Shapiro et al 2003). Treatment with α1-antitrypsin inhibited emphysema development and reduced recruitment of pulmonary neutrophils and macrophages (Churg et al 2003; Pemberton et al 2006). Similarly, a small molecule neutrophil elastase inhibitor, ZD0892, reduced cigarette smoke induced emphysema and pulmonary cells recruitment in a guinea pig model (Wright et al 2002). These studies suggest inhibition of neutrophil elastase can be anti-inflammatory in addition to preventing emphysema development.

The search for potent, safe oral inhibitors of neutrophil elastase has been going on for over 20 years and many of the compounds that progressed into clinical development failed because of poor pharmacokinetics and a low therapeutic index. Subsequently, tripeptidyl trifluoromethyl ketones were developed that had a better oral profile, although these molecules were not fully optimized for oral delivery (Edwards et al 1997). Also monocyclic b-lactam neutrophil elastase inhibitors were identified (Vincent et al 1997). More recently, both GSK (GSK-311366) and ONO Pharmaceuticals (ONO6818) have had oral compounds in Phase I/II. However, they failed in early clinical trials and thus far no oral elastase inhibitors have been evaluated fully in Phase II COPD trials. The AstraZeneca protease inhibitor AZD3342
(thought to be an elastase inhibitor) is scheduled to progress to Phase II studies in the second half of 2006 and Bayer GmbH has an inhibitor in Phase I but the outcomes of these trials are still awaited.

Protein inhibitors of neutrophil elastase administered via nebulisation are also being pursued and supportive data has been obtained for inhaled recombinant α1-antitrypsin in mouse cigarette smoke models. The probability of inhaled combination products consisting of elastase inhibitors with bronchodilators is low. However, the opportunity for safe long acting inhaled elastase inhibitors with low systemic bioavailability that are suitable for combination with bronchodilators is being pursued by Argenta Discovery.

**HDACs modifiers**

NF-κB being a pro-inflammatory transcription factor, on activation binds to specific recognition sequence motifs in DNA and subsequently interacts with a multitude of co-activator molecules, such as cAMP-response-element-binding protein (CREB), CREB-binding protein (CBP), p300 and p300/CBP-associated factor (pCAF). These co-activators act as molecular switches of transcription and acetylate histones by their intrinsic histone acetyltransferase (HAT) activity (Ogryzko et al 1996; Roth et al 2001). Acetylation of core histones allow DNA unwinding and thus access to wide variety of transcription factors (Roth et al 2001). The transcriptional process can be reversibly switched off by deacetylation of acetylated histones, thus rewinding the exposed DNA, consequently leading to gene silencing. The deacetylation of acetylated histones is brought about by histone deacetylases (HDACs), which act as co-repressors, in association with other co-repressor proteins that are concomitantly recruited. Since all inflammatory processes involve acetylation and deacetylation of histones (chromatin remodeling) (Barnes 2006), HDACs therefore are attractive targets for anti-inflammatory therapies.

Ito and co-workers have shown a role for histone acetylation and deacetylation in IL-1β-induced TNF-α release in alveolar macrophages derived from cigarette smokers (Ito et al 2001). They have also suggested that oxidants may play an important role in the modulation of HDAC and inflammatory cytokine gene transcription. Furthermore, we have shown that both cigarette smoke/H₂O₂ and TNF-α caused an increase in histone acetylation (HAT activity) leading to CXCL8 expression in monocytes and alveolar epithelial cells both in vitro and in vivo in rat lungs (Rahman et al 2002; Marwick et al 2004; Moodie et al 2004).

Glucocorticoid suppression of inflammatory genes requires recruitment of HDAC2 to the transcription activation complex by the glucocorticoid receptor (Ito et al 2001; Rahman et al 2004). This results in deacetylation of histones and a decrease in inflammatory gene transcription. A reduced level of HDAC2 was associated with increased proinflammatory response and reduced responsiveness to glucocorticoids in alveolar macrophages obtained from smokers (Ito et al 2001; Rahman et al 2002; Marwick et al 2004; Moodie et al 2004; Rahman et al 2004). Culpitt and co-workers have shown that cigarette smoke solution stimulated release of CXCL8 and GM-CSF, which was not inhibited by dexamethasone, in alveolar macrophages obtained from patients with COPD compared to that of smokers (Culpitt et al 2003). They suggested that the lack of efficacy of corticosteroids in COPD might be due to steroid insensitivity of macrophages in the respiratory tract. Thus, the cigarette smoke/oxidant-mediated reduction in HDAC2 levels in alveolar epithelial cells and macrophages will not only increase inflammatory gene expression but will also cause a decrease in glucocorticoid function in patients with COPD. HDAC2 activity has also been measured in bronchial biopsies and alveolar macrophages from COPD patients and smoking control, demonstrating a significant decrease in HDAC2 activity, the magnitude of which increased with severity of disease (Ito et al 2005) (Table 1). Moreover, protein expression of HDAC2 was decreased in a similar manner in COPD patients.

Consequently, a potential means by which to treat COPD would be to increase HDAC2 expression and activity such that steroids regain their anti-inflammatory activity. We have shown that co-incubation of cells with NAC and H₂O₂ protects HDAC2 from down-regulation and reduction of specific activity (Moodie et al 2004). In addition, it has been reported that theophylline has a similar effect in lung macrophage cells, increasing HDAC2 expression and re-sensitising the cells to steroids (Cosio et al 2004). Similar data were obtained for curcumin, a dietary polyphenol, in restoration of steroid efficacy (unpublished data). Alternative means of upregulating HDAC2 activity would be of great interest for potential combination therapies of the future.

**Cytokine and chemokine antagonists**

Several recent reviews point to the development of novel antagonists of cytokines, chemokines or their receptors (De Boer 2005; Chung 2006; De Boer et al 2007). These molecules may reduce gene expression, impair production or secretion of mature proteins, antagonize binding of cytokines and chemokines to their receptors or inhibit receptor signal transduction (Table 2). Antibodies and solubilized receptors
like TNFR often scavenge solubilized cytokines and chemokines, or prevent binding of these proteins to their receptors. Small molecules 1) prevent binding of cytokines and chemokines to their receptors by non-activating mimicking of cytokines or chemokines, or 2) prevent intracellular signal transduction activation, or 3) interfere with gene expression and translation by direct inhibition of transcription factors (like IKK2 inhibition) or mRNA binding via small interference (si) RNA or antisense mRNA.

**TNFα and receptors antagonists**

As a major proinflammatory cytokine TNFα and its receptors TNFR1 (or: TNFR p55) and TNFR2 (or: TNFR p75) seem to play an important role in many chronic diseases including COPD and asthma. Therefore, several drugs have been developed to reduce TNFα levels, of which some have been approved by eg, the Federal Drug Administration (FDA) for treatment of RA, ankylosing spondylitis, Crohn’s disease, or psoriasis. These approved drugs include etanercept (soluble human TNFR2), infliximab (chimeric human/mouse IgG1 antibody against TNFα), and adalumimab (human IgG1 antibody against TNFα). Many others are being developed in order to enhance efficacy, reduce side effects due to frequent subcutaneous injection, increase bioavailability or protection to proteolytic degradation by coupling to polyethylene glycol (PEG) chains, or reduce immunogenicity by humanization of antibodies or designing small molecules. Some examples are given in Table 2. Recent clinical trial phase II studies demonstrated that patients with moderate to severe asthma may profit from treatment with either of these drugs. Three infusions of infliximab over 6 weeks reduced the number of exacerbations as well as sputum levels of TNFα, IL-6, CXCL8 and CXCL10 but not peak expiratory flow (PEF) or inflammatory cell count in sputum of patients with moderate asthma (Erin et al 2006). Other studies demonstrated that twice-weekly treatment with etanercept during 10 to 12 weeks improved the bronchial hyperresponsiveness (BHR, expressed as PC_{20}), post-bronchodilator FEV_{1}, and the quality of life of patients with refractory, severe asthmatic patients (Howarth et al 2005; Berry et al 2006). Treatment of asthmatics with Marimastat, an inhibitor of TNFα and MMP activation, also reduced BHR but failed to significantly reduce sputum inflammatory cell numbers, asthma symptoms, FEV_{1}, or bronchodilator use (Bruce and Thomas 2005). In contrast to asthma, 2 studies showed that treatment of COPD patients with 3 infusions of infliximab over 6 to 24 weeks did not result in any significant improvement of lung function, airway inflammation, or quality of life (Abdelhady et al 2005; van der Vaart et al 2005; Rennard et al 2007). Treatment may have some beneficial effect on physical endurance seen by the 6 minute walking distance test (Rennard et al 2007). This ineffectiveness may be due to a rather short treatment period, the choice of infliximab over other drugs, or to the complex or less significant role of TNFα in progressed COPD. Future studies should sort out whether long-term treatment or treatment with different TNFα antagonists are beneficial to all COPD patients or only a specific population of COPD patients (Rennard et al 2007).

**CXCL1, CXCL8, and receptors antagonists**

As previously mentioned (De Boer 2005), several CXCR2 and CXCL8 antagonists are available, some of which were in clinical trial for COPD. Updated information shows that either the testing of these drugs is discontinued (like the antibody ABX-IL-8 against human CXCL8) or is not to be found in the public domain. Hence, little is known yet on treatment of patients with COPD with CXCL8 or CXCR2 antagonists. The small molecule CXCR2 antagonist SB-656933 (by GSK) has recently been demonstrated to inhibit the CXCL8-induced expression of CD11b molecules on peripheral blood neutrophils from COPD patients (Nicholson et al 2007). The antagonist was mentioned to enter clinical trial studies for COPD in 2005, but is not so in GSK’s pipeline of 2006. AZD-8309 is a pyrimidine derivate currently in phase I clinical trial for COPD and phase II for RA. Data from these studies have not yet been published. SB-265610 is a small molecule inhibiting CXCR2. Studies demonstrated that hyperoxia in newborn rats led to pulmonary inflammation by neutrophils and the formation of ROS and RNS mediating impaired lung development and lipid peroxidation (Autten et al 2001; Liao et al 2006). Treatment with SB-265610 reduced airway neutrophilia, radical formation, lipid peroxidation and protein nitration, as well as improved conservation of lung development and lung function. This points to the importance of reducing neutrophilia in order to reduce reactive species formation, peroxidation or nitration and tissue destruction or alterations. Data from other studies supported the effectiveness of CXCL8 or CXCR2 antagonists in reducing neutrophilia in vivo in rodents and inhibition of neutrophil activation and degranulation in vitro (De Boer 2002, 2005). These data point to the potential need for development of novel antagonists of CXCR1, CXCR2 or their ligands CXCL1 and CXCL8. Recent studies showed that novel thiazolopyrimidine, cyclobutenedione (eg, SCH 527123), or imidazolopyrimidine CXCR2 antagonists had a good oral bioavailability in rats with reasonable pharmacokinetics (half life of at least 1.2h) (Baxter et al 2006; Dwyer...
et al 2006; Ho et al 2006), and inhibition of CXCL1- or CXCL8-induced chemotaxis of cells (Baxter et al 2006; Dwyer et al 2006).

CCL2 and CCR2 antagonists
The humanized monoclonal antibody MLN1202 (by Millemium) against CCR2 is currently in phase II for treatment of chronic inflammatory diseases like multiple sclerosis and artherosclerosis. Clinical data from these studies have not yet been published. Other inhibitors in clinical trial include the small molecule CCR2 inhibitors CCX915 (by ChemoCentryx) for multiple sclerosis (phase I), INCB3284 (by Incyte/Pfizer) which is in phase IIa for treatment of RA, and the monoclonal antibody ABN912 against CCL2 (by Novartis) for COPD (phase I) and RA (phase II). Recently, patients with RA did not clinically and immunohistologically respond to a 2-week treatment with ABN912 (Haringman et al 2006). Serum levels of CCL2 increased clearly after treatment (doses higher than 1 mg/kg bw), and chemotactic complexes of the antibody with CCL2 were formed. This may hamper the treatment of patients with COPD, however these results have to be awaited. Several other compounds based on butyramides or γ-aminoamides were recently developed as specific CCR2 antagonists with a good oral bioavailability in rats (Butora et al 2006; Pasternak et al 2006). Others are in preclinical phase, including INCB8696, JNI-27553292, SKL-2841 and INCB3344. INCB3344 is a rodent CCR2 small molecule antagonist (Brodmerkel et al 2005). The compound was shown to inhibit macrophage influx in a mouse model for delayed hypersensitivity.

Other anti-inflammatory agents for COPD include ion channel blocker (P2X7, Phase II, from AstraZeneca), monoclonal antibody to IL-1β (Phase I, from Novartis), inhibitors of 5-lipoxygenase (PEP03 inhaled, Phase II, from PharmaEngine) and iNOS inhibitors (GW-274150, Phase II, from GSK).

Target growth factors
Several studies pointed to the involvement of VEGF/VEGFR in pulmonary and vascular remodeling and inflammation. Therefore, VEGF inhibitors may be of potential use for treatment of vascularization, as seen in asthma or in specific subtypes of COPD, such as chronic bronchitis, but not emphysema (Marwick et al 2006; De Boer et al 2007). Several drugs have been developed to reduce tumor growth and metastasis by impairing the neovascularization. Antagonists include those in clinical trials and in preclinical investigations. Among those in clinical trial are VEGF-Trap, bevacizumab (or: Avastin), CEP-7055, VEGF Trap, and PTK787 (Ferrara et al 2003). However, little is known yet of side effects or their efficacy in the treatment of COPD. In addition, in patients with emphysema for example, inhibitors of apoptosis are needed whereas in contrast to chronic bronchitis, expression of hypoxia-inducible factor, VEGF, and VEGFR2 may be stimulated to overcome chronic and epithelial cell death and improve vascular restoration (Voelkel and Coll 2003). Hence, treatment with VEGF/VEGFR inhibitors should be further investigated and may be restricted to specific subtypes of chronic lung diseases.

As discussed aforementioned, TGFβ expression was increased in COPD as compared to smokers without COPD, inhibition of TGFβ-mediated pathway may hence be a target for therapeutic intervention. However, almost all of TGFβ antagonists are still in preclinical development. Hence, little can be discussed yet on clinical applicability.

Concluding remarks
In order to reverse continuing damage and remodeling needs stopping exposure to tobacco smoke by many patients with COPD. However, after smoking cessation inflammation goes on which may hamper tissue repair. The ongoing inflammation may be due to an autoimmune component and/or epigenetics alterations. However, not all patients with COPD are smokers but do suffer from tissue damage and/or remodeling. Hence, an effective treatment of patients with COPD not only needs stopping exposure to toxic compounds like smoke, but also needs reduction of excessive inflammation, oxidative status, and stimulation of parenchymal and/or airway repair (Leff 2006). This needs further development and testing of therapeutic drugs antagonizing a low antioxidant status, inflammation, and excessive remodeling. There are several small molecule compounds in clinical trials that target oxidant signaling or quench oxidants derived from inflammatory cells and cigarette smoke. Regarding inhibition of inflammation several drugs have been used successfully for treating different chronic inflammatory diseases, but not yet for COPD. As pointed out in this review, success may be hindered by the complexity of actions of eg, chemokines and antioxidants, or by a low efficacy and bioavailability of the drugs. Whereas more insight is needed in the pathogenesis of COPD or its subtypes, individualized combination therapy with such novel and specific anti-inflammatory and anti-oxidant drugs may be advantageous to patients with specific phenotypes of the disease.

References


