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Cerebral malaria in children: using the retina to study the brain

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Cerebral malaria is a dangerous complication of Plasmodium falciparum infection, which takes a devastating toll on children in sub-Saharan Africa. Although autopsy studies have improved understanding of cerebral malaria pathology in fatal cases, information about in vivo neurovascular pathogenesis is scarce because brain tissue is inaccessible in life. Surrogate markers may provide insight into pathogenesis and thereby facilitate clinical studies with the ultimate aim of improving the treatment and prognosis of cerebral malaria. The retina is an attractive source of potential surrogate markers for paediatric cerebral malaria because, in this condition, the retina seems to sustain microvascular damage similar to that of the brain. In paediatric cerebral malaria a combination of retinal signs correlates, in fatal cases, with the severity of brain pathology, and has diagnostic and prognostic significance. Unlike the brain, the retina is accessible to high-resolution, non-invasive imaging. We aimed to determine the extent to which paediatric malarial retinopathy reflects cerebrovascular damage by reviewing the literature to compare retinal and cerebral manifestations of retinopathy-positive paediatric cerebral malaria. We then compared retina and brain in terms of anatomical and physiological features that could help to account for similarities and differences in vascular pathology. These comparisons address the question of whether it is biologically plausible to draw conclusions about unseen cerebral vascular pathogenesis from the visible retinal vasculature in retinopathy-positive paediatric cerebral malaria. Our work addresses an important cause of death and neurodisability in sub-Saharan Africa. We critically appraise evidence for associations between retina and brain neurovasculature in health and disease, and in the process we develop new hypotheses about why these vascular beds are susceptible to sequestration of parasitized erythrocytes.

Keywords: cerebral malaria; cerebral microvasculature; retinal microvasculature; haemorheology; surrogate marker
Introduction

Paediatric cerebral malaria is a clinical syndrome that kills and disables children through mechanisms that remain incompletely understood. Adhesion of parasitized erythrocytes to the microvascular endothelium, leading to their sequestration in the brain, is the pathological signature of both adult and paediatric cerebral malaria, and is thought to be the chief cause of injury (Taylor et al., 2004; Ponsford et al., 2012). Several hypothetical mechanisms linking sequestration, which is entirely intravascular, to extravascular parenchymal damage have been proposed (Van der Heyde et al., 2006) but questions remain about which of these mechanisms are most important, how they might interact, and ultimately where new therapies should be directed. One of the reasons why such questions remain over 100 years after sequestration was first identified is because in vivo access to the brain is difficult, and advances in knowledge have relied on post-mortem and in vitro studies. Improved understanding of in vivo neurovascular pathogenesis and the development of better treatments will be facilitated by disease models or surrogate markers.

The retina may be a good source of surrogate markers of cerebrovascular injury because paediatric cerebral malaria is associated with a retinopathy (‘malarial retinopathy’) that accurately predicts cerebral sequestration (Taylor et al., 2004), correlates with severity of brain involvement (White et al., 2001), and is associated with mortality (Beare et al., 2004). Unlike the brain, the eye allows non-invasive access for structural and functional imaging of the microcirculation, which is thought to be the major site of sequestration.

The concept that neurovascular injury observed in the retina resembles neurovascular injury lying unseen in the brain is based on the assumption that the two circulations are analogous in ways that are relevant to the pathogenesis of paediatric cerebral malaria. Such assumptions should be supported by a biologically plausible rationale before specific retinal vascular features are considered as surrogate markers of cerebrovascular damage (International Conference on Harmonisation, 1998).

Is such a rationale likely? It is well known that the retina and brain have many similarities. Both are part of the CNS, with common embryological origins, vascular structure and metabolic demands. The relevance of these similarities for potential surrogate markers has been recognized (Patton et al., 2005, 2006), especially for stroke (Doubal et al., 2009, 2010). Despite this, detailed comparisons of the microvasculature of the two organs are rare (Cogan and Kuwabara, 1984; Patton et al., 2005).

Our objective was to discover how likely it is that the retinal vascular damage responsible for malarial retinopathy reflects analogous cerebrovascular damage in retinopathy-positive paediatric cerebral malaria. To do this we compared the manifestations of paediatric cerebral malaria in retina and brain, and then compared retina and brain in terms of vascular features likely to be important for cerebral malaria pathogenesis. Concluding that retina and brain are analogous in ways relevant to this disease would justify further investigation to see whether specific retinal signs predict both focal brain damage detectable by MRI and the patient’s response to treatment. The results of such investigations would address further criteria of surrogacy (International Conference on Harmonisation, 1998), and in the context of a strong biological rationale could shed light on the dynamics of cerebral malaria neurovascular pathogenesis in the period between coma onset and recovery or death.

Malarial retinopathy can occur in parasitaemic children without cerebral malaria (Beare et al., 2004), and indeed, some features of malarial retinopathy occur in conditions that don’t involve malaria at all (e.g. white-centred haemorrhages). It is not clear if retinopathy predicts cerebral sequestration in malarial syndromes besides paediatric cerebral malaria, or how often retinopathy might occur in severely ill children in general.

We therefore discuss retinopathy in the specific clinical context of paediatric cerebral malaria. In this particular population malarial retinopathy has high positive and negative predictive value to distinguish between the presence and absence of cerebral sequestration in fatal cases (Taylor et al., 2004). Until associations between retinopathy and cerebral sequestration are known for malaria in general, extrapolation from our review to severe malaria syndromes other than paediatric cerebral malaria may not be appropriate.

Our review is divided into four sections. In the first section we describe paediatric cerebral malaria, including the typical clinical presentation, manifestations of cerebral malaria in the brain, and manifestations in the retina. The neurovascular effects of cerebral malaria on retina and brain are then compared.

In the second section we introduce the concept that patterns of neurovascular patholgy in retina and brain may be understood in terms of haemorheological dysfunction involving microvascular haematocrit, blood viscosity and shear stress. These factors depend on interactions between intrinsic properties of blood and structural properties of microvascular networks, and this suggests that microvascular architecture may be a useful point of comparison for a cerebral malaria-specific analogy between retina and brain.

The third section describes microvascular architecture in the human retina and brain, and compares features that may be relevant to the pathogenesis of cerebral malaria.

We end by discussing how our comparison of retina and brain provides a biologically plausible rationale for considering retinal signs as potential surrogates of brain damage in retinopathy-positive paediatric cerebral malaria. We consider how observations of retinal vessel structure and function might allow inference of in vivo cerebrovascular pathogenesis.
Paediatric cerebral malaria

Definition

Several species of Plasmodium parasite cause malaria in humans. P. falciparum is responsible for the majority of severe malaria and malaria-associated deaths worldwide, particularly in sub-Saharan Africa where severe malaria has a disproportionate impact on children <5 years of age (WHO, 2012). In children, severe malaria predominantly involves one or more of three syndromes: cerebral malaria, severe malarial anaemia and metabolic acidosis. Other manifestations of paediatric severe malaria include convulsions, hypoglycaemia, hyperparasitaemia and prostration (WHO, 2000). Cerebral malaria is defined as coma, with P. falciparum peripheral parasitaemia, in the absence of another identifiable cause of coma, such as hypoglycaemia or meningitis (Newton et al., 1998). This definition is broad, and misclassifies almost 25% of fatal paediatric cases as cerebral malaria when compared with histopathological identification of sequestration in cerebral vessels as the reference standard. The presence of malarial retinopathy on pre-mortem fundus examination accurately distinguishes histopathological cerebral malaria from cases that meet the clinical definition but actually have another cause of death (Taylor et al., 2004). Although the strong association between retinopathy and intracerebral sequestration has been established only in fatal cases, it is likely that the association also prevails in those who recover, and that the presence of retinopathy improves the accuracy of diagnosis in children with cerebral malaria (Beare et al., 2011).

Typical presentation and clinical course

In an area with intense transmission of P. falciparum, the typical history of paediatric cerebral malaria involves a young child who rapidly develops coma after a short prodrome of fever and generalized illness. The coma may be preceded or accompanied by convulsions, from which the child does not wake. On examination the child is usually febrile, and may have deep or rapid breathing. Coma is distinguished from prolonged post-ictal state by duration >30 min after seizure. Depth of coma is assessed using a modification of the Glasgow Coma Scale, known as the Blantyre Coma Scale (Molyneux et al., 1989; WHO, 2000, 2010). Convulsions or posturing, including opisthotonus, may be present. Ophthalmoscopy through dilated pupils reveals signs of malarial retinopathy: white patchy discoloration of the macula and/or peripheral retina; orange or white discoloration of retinal vessels; and/or retinal haemorrhages, typically with white centres. Papilloedema may be seen, but in isolation does not distinguish cerebral malaria from other causes of coma (Lewallen et al., 1999, 2008; Harding et al., 2006). Investigation may reveal concomitant metabolic acidosis and/or severe anaemia. Peripheral P. falciparum asexual parasitaemia is present by definition. Other treatable causes of coma in this setting must be ruled out and include malaria-associated hypoglycaemia and meningitis. Bacteraemia may be found, especially in infants and in the presence of severe anaemia (Bronzan et al., 2007).

The duration of illness is usually short, and most patients recover or die within 48 h. Death is typically by respiratory arrest, and case fatality with treatment is ~15%. Those who recover are at risk of neurodisability (Molyneux et al., 1989) and epilepsy (Birbeck et al., 2010). Guidelines for treatment have been published (WHO, 2010).

Cerebral malaria also occurs in adults (Table 1). The clinical presentation is different from paediatric cerebral malaria (WHO, 2000), but it is not clear if this is because of age or differences in immunology related to transmission intensity (Idro et al., 2005). The geographical distribution is different from that of paediatric cerebral malaria. Adult cerebral malaria occurs primarily in South and South East Asia, where transmission of P. falciparum is endemic and seasonal and most children and adults have no immunity; whereas paediatric cerebral malaria occurs mainly in sub-Saharan Africa where transmission is endemic and where both older children and adults have acquired partial immunity. Adult travellers who are P. falciparum naïve may develop cerebral malaria in endemic countries. Patterns of mortality and comorbidity vary with age (Dondorp et al., 2008).

Malarial retinopathy has been described in adults since at least the 1800s (Mackenzie, 1877). Maude et al. (2009) found retinopathy in 14/20 (70%) adults with cerebral malaria, 17/20 (85%) with other types of severe malaria, and 9/15 (60%) with uncomplicated malaria. The retinopathy was most severe in those with cerebral malaria. Abu Sayeed et al. (2011) found retinopathy in 31/75 (41%) with cerebral malaria, 16/64 (25%) with other types of severe malaria, and 1/31 (3%) with uncomplicated malaria. Number of retinal haemorrhages was an independent predictor of death. Both studies used sensitive retinal imaging techniques and a standardized grading scheme (Harding et al., 2006), but neither found the characteristic vessel discoloration seen in up to 30% of children with cerebral malaria (Beare et al., 2004). Compared with the adult literature, few data exist on the presence of malarial retinopathy in children with uncomplicated malaria, as consistent retinal examination of conscious children is difficult—particularly so for the retinal periphery. Further research is needed.

Manifestations of retinopathy-positive cerebral malaria in the paediatric brain

Sequestration

Sequestration is the histopathological hallmark of paediatric cerebral malaria (Taylor et al., 2004). Sequestration results from the binding of parasitized erythrocytes to vascular endothelium. Parasitized erythrocytes also bind in vitro to other erythrocytes (rosetting), and to platelets (clumping, or auto-agglutination). Sequestration is mediated by adhesion between malarial antigens on the surface of the infected erythrocyte and several host receptors on the vascular endothelium. The P. falciparum surface antigen most studied is P. falciparum erythrocyte membrane protein 1.
Table 1 Manifestations of cerebral malaria in the retina and brain

<table>
<thead>
<tr>
<th>Sequestration</th>
<th>Paediatric retina</th>
<th>Paediatric brain</th>
<th>Adult brain</th>
</tr>
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<tbody>
<tr>
<td><strong>Frequency</strong></td>
<td>Always present in fatal cerebral malaria (Lewallen et al., 2000). Unclear if absent in fatal coma of other cause or severe malarial anaemia.</td>
<td>Always present in fatal cerebral malaria, and absent in fatal coma of other cause (Taylor et al., 2004; Dorovini-Zis et al., 2011). Commonly associated with sequestered leucocytes (Brown et al., 2001; Armah et al., 2005)</td>
<td>Always present in fatal cerebral malaria (MacPherson et al., 1985; Oo et al., 1987; Pongponratn et al., 1991; Sein et al., 1993). In cerebral malaria density is greater in brain than other organs (MacPherson et al., 1985; Pongponratn et al., 1991). Significant sequestration may be present in fatal non-cerebral malaria (MacPherson et al., 1985; Silamut et al., 1999). The percentage of vessels with sequestration is greater in cerebral malaria than non-cerebral malaria (Ponsford et al., 2012)</td>
</tr>
<tr>
<td><strong>Location</strong></td>
<td>Patchy distribution within capillary network (Lewallen et al., 2000). Variation between retinal regions not yet defined.</td>
<td>Most microvessels, and the margins of pial and larger vessels (Dorovini-Zis et al., 2011). Grey and white matter of cerebrum, subcortex, brainstem and cerebellum (Armah et al., 2005; Dorovini-Zis et al., 2011).</td>
<td>Occurs in capillaries, venules, and very occasional arterioles (MacPherson et al., 1985). Occurs in grey and white matter, but most dense in cerebral white matter (Nagatake et al., 1992). Density reduces from cerebrum to cerebellum to brainstem (Pongponratn et al., 2003). Density greater in cerebellum than cerebrum (Sein et al., 1993).</td>
</tr>
<tr>
<td><strong>Vessels involved</strong></td>
<td>Capillaries and margins of larger vessels (Lewallen et al., 2000). Vessel discoloration affects capillaries, venules, and arterioles (personal observation)</td>
<td>Occurs in brain microvessels, pial and larger vessels (Dorovini-Zis et al., 2011)</td>
<td>Predominant site is the capillary bed, but also occurs in larger pial and subarachnoid vessels (Spitz, 1946). Uncommon in arterioles (MacPherson et al., 1985).</td>
</tr>
<tr>
<td><strong>Haemorrhages</strong></td>
<td><strong>Type</strong></td>
<td>White-centred, blot (White et al., 2001). Parasitized erythrocytes rarely seen outside vessel (White et al., 2001).</td>
<td>Ring (Dorovini-Zis et al., 2011). Parasitized erythrocytes rarely seen outside vessel (White et al., 2001; Dorovini-Zis et al., 2011).</td>
</tr>
<tr>
<td><strong>Frequency</strong></td>
<td>Gross haemorrhages present in 78% fatal cerebral malaria, 7% fatal coma of other cause (White et al., 2009).</td>
<td>Any type present in 80% fatal cerebral malaria (Dorovini-Zis et al., 2011).</td>
<td></td>
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<tr>
<td><strong>Location</strong></td>
<td>All retinal quadrants. Usually restricted to inner retinal layers, with extension to subretinal haemorrhage in severe cases (White et al., 2009).</td>
<td>Common in white matter, rare in grey matter except in the cerebellum (White et al., 2001; Dorovini-Zis et al., 2011).</td>
<td>Usually occur in cerebral white matter; also reported in pons, medulla, cerebellum, and cortical grey matter (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993; Turner, 1997). No difference in haemorrhage frequency between cortex, diencephalon, and brainstem (Medana et al., 2011).</td>
</tr>
<tr>
<td><strong>Vessel leakage</strong></td>
<td><strong>Type</strong></td>
<td>Fibrinogen leakage along vessels with and without associated haemorrhage (White et al., 2009).</td>
<td>Fibrinogen leakage often associated with haemorrhage (Dorovini-Zis et al., 2011).</td>
</tr>
<tr>
<td>Paediatric retina</td>
<td>Paediatric brain</td>
<td>Adult brain</td>
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<tr>
<td><strong>Frequency</strong></td>
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<tr>
<td>Fibrinogen leakage in 31% cases fatal cerebral malaria, 7% fatal coma of other cause (White et al., 2009).</td>
<td>Unclear how many cases of fatal cerebral malaria have leakage of any type.</td>
<td>At least one type of oedema present in all cases of both cerebral and non-cerebral malaria (Medana et al., 2011).</td>
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<tr>
<td>Average (SD) number of foci is 1.2 (2.6) in fatal cerebral malaria and 0.21 (1.1) in coma of other cause (White et al., 2009).</td>
<td>Leakage greater in white than grey matter (associated with haemorrhages) (Dorovini-Zis et al., 2011).</td>
<td>Oedema between white matter tracts is most common in: brainstem &gt; diencephalon &gt; cortex (Medana et al., 2011).</td>
<td></td>
</tr>
<tr>
<td>Associated with vessels but not defined in terms of retinal quadrants or vessel type (White et al., 2009).</td>
<td>Cerebral white and grey matter, subcortex, brainstem and cerebellum (Dorovini-Zis et al., 2011).</td>
<td>Brainstem, diencephalon, cerebral cortex (Medana et al., 2011).</td>
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<tr>
<td>Angiographic fluorescein leakage predominantly affects venules (Beare et al., 2009).</td>
<td>Regions where MRI brain signal changes distinguish between retinopathy-positive and negative cerebral malaria (highest to lowest odds ratio): basal ganglia, corpus callosum, cerebral cortex, thalamus, cerebral white matter, posterior fossa (Potchen et al., 2012).</td>
<td>Regions reported to be involved: Brain stem, thalamus, cerebellum, corpus callosum, cerebral white matter (Yadav et al., 2008; Raalkar et al., 2011).</td>
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<tr>
<td><strong>Regions vulnerable to presumed ischaemia on imaging</strong></td>
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<td>Retinal whitening and capillary non-perfusion appears to be especially prominent at the foveal avascular zone, horizontal raphe, and retinal periphery. All are watershed regions (Beare et al., 2009).</td>
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**Haemorrhage**

Histopathology reveals subtypes of cerebral malaria within fatal retinopathy-positive paediatric cerebral malaria. In Malawi, 75% of cases coming to autopsy have sequestration and intravascular microthrombi, whereas 25% have sequestration but no haemorrhages or other manifestations of retinopathy.

Histopathology reveals subtypes of cerebral malaria within fatal retinopathy-positive paediatric cerebral malaria. In Malawi, 75% of cases coming to autopsy have sequestration and intravascular microthrombi, whereas 25% have haemorrhages.
perivascular pathology visible on routine haematoxylin and eosin-stained sections (Dorovini-Zis et al., 2011).

Petechial haemorrhages on the cut surface of fresh brain correspond to microscopic ring haemorrhages. Ring haemorrhages occur frequently in the white matter of the cerebral hemispheres, and extend to the grey–white border. They are rare in cortical and subcortical grey matter, but occur throughout the cerebellum and brainstem (White et al., 2001). Diffuse axonal injury follows the same distribution (Dorovini-Zis et al., 2011).

In general, ring haemorrhages in paediatric cerebral malaria consist of extravasated uninfected erythrocytes; each haemorrhage is centred on a distended capillary containing infected erythrocytes and commonly a microthrombus (Dorovini-Zis et al., 2011).

**Brain swelling and vessel leakage**

Studies of paediatric cerebral malaria where retinopathy status was either unknown or a combination of retinopathy-positive and retinopathy-negative cases, have found raised opening pressure at lumbar puncture (Newton et al., 1991), clinical signs consistent with brain herniation (12/12 fatal cases, 17/49 survivors) (Newton et al., 1994), papilloedema (Beare et al., 2004), and enlarged optic nerve sheath diameter (Beare et al., 2012).

Raised brain weight and extravasation of fibrinogen from microvessels occurs in histopathologically confirmed paediatric cerebral malaria, although these do not distinguish cerebral malaria from fatal coma of other cause. Leakage is most often associated with vascular pathology such as haemorrhage, but fibrinogen is also seen in cerebral grey matter where haemorrhages are absent (Dorovini-Zis et al., 2011). Focal loss of blood–brain barrier cell junction proteins (ZO1, occludin, vinculin) occurs adjacent to sequestration. The distribution of fibrinogen, IgG and C5b-9 in these cases did not indicate widespread leakage into brain tissue (Brown et al., 2001).

**Brain imaging**

Consistent with reports of raised intracranial pressure in clinically defined paediatric cerebral malaria, MRI in retinopathy-positive paediatric cerebral malaria reveals moderate to severe brain swelling in 47% (57/120) of cases. This is significantly more common than in paediatric cerebral malaria without retinopathy (7/32 cases, OR 3.2, 95% CI 1.3–8) (Potchen et al., 2012).

Discrete lesions are seen in basal ganglia, thalamus, corpus callosum, and cerebral grey and white matter. All are significantly more frequent in retinopathy-positive cerebral malaria than retinopathy-negative cases. Cortical abnormalities can be predominantly frontal or posterior. Lesions do not respect arterial watersheds (Potchen et al., 2012), but may reflect territories of venous drainage (Meder et al., 1994; Andeweg, 1996). Distribution of lesions according to venous territories is consistent with sequestration occurring predominantly on the venous side of the microvasculature.

**Manifestations of cerebral malaria in the paediatric retina**

Several clinical (Harding et al., 2006) and fluorescein angiographic features of malarial retinopathy (Beare et al., 2009) are illustrated in Fig. 1. Standard images of all features are in Harding et al., (2006).

**Sequestration in retinal vessels**

Sequestration is distributed unevenly in capillaries and the margins of larger retinal vessels (Lewallen et al., 2000). Sequestration and vessel discoloration tend to occur at vascular branch points (Lewallen et al., 2000), where greater turbulence exists to disrupt laminar flow (Nagaoka and Yoshida, 2006). It has been proposed that de-haemoglobinization of parasitized erythrocytes is responsible for the clinically observed white or orange discoloration of retinal vessels seen in paediatric cerebral malaria (Lewallen et al., 2000). Retinal vessels, particularly venules, can have a mottled appearance on fluorescein angiogram, which may represent sequestration (Beare et al., 2009). The discoloration and mottled appearance of retinal vessels in malarial retinopathy appears to be pathognomonic for paediatric severe malaria, but is reported in severe malarial anaemia as well as cerebral malaria (Beare et al., 2004).

**Retinal vessel leakage**

Fluorescein leakage is seen in ~40% of cases with malarial retinopathy and indicates blood–retinal barrier disruption. It does not co-localize with vessel mottling (Beare et al., 2009). White et al. (2009) reported leakage of fibrinogen from retinal vessels in 31% (11/35) of cases with cerebral malaria versus 7% (2/29) coma of other cause. Leakage was not reported in terms of vessel type, but fluorescein angiogram imaging suggests that leaking is predominantly from venules (Beare et al., 2009) (Fig. 1). Again, this is consistent with cerebral malaria pathogenesis centring on capillaries and post-capillary venules.

**Retinal whitening**

Retinal whitening appears as discrete areas of pale discoulouration of the retina (Fig. 1A), which correspond to areas of capillary non-perfusion seen on fluorescein angiogram. Retinal whitening and capillary non-perfusion seem to occur earliest and most severely in watershed regions such as the margin of the foveal avascular zone, horizontal raphe and retinal peripheries (Beare et al., 2009). In contrast to the scattered geometric lesions seen in the macula, whitening and capillary non-perfusion in the retinal periphery can occur in large tide-mark distributions that cut across arterioles and venules of varying sizes (Beare et al., 2009) (Fig. 1B).

Whitening is likely to be caused by oncotic cell swelling in response to reduced perfusion (Beare et al., 2009). Other possible mechanisms include metabolic steal by large numbers of parasites (Hero et al., 1997), and occlusion caused by microthrombi (White et al., 2009). These theories are not mutually exclusive.
Areas of pale retinal discolouration are seen in other retinal conditions including arterial occlusion, venous occlusion (Browning, 2004), and Purtscher’s retinopathy (Agrawal and McKibbin, 2006). Mild whitening has also been reported in paediatric severe malaria without coma (Beare et al., 2004; Burton et al., 2004). The severity and pattern of whitening seen in malarial retinopathy—around the fovea and extending into the horizontal raphe—appears to be specific to cerebral malaria. Whitening in retinal vein occlusion is associated with swelling of the inner nuclear and outer plexiform layers of the retina (Sarda et al., 2011). In paediatric cerebral malaria there is a significant inverse correlation between electroretinographic cone b-wave amplitude and severity of retinal whitening ( Lochhead et al., 2010), indicating dysfunction of bipolar cells in the inner nuclear layer. The inner

Figure 1 The features of paediatric malarial retinopathy are: retinal haemorrhages (often white-centred), retinal whitening, and orange or white discolouration of vessels. Papilloedema is often seen but is not specific for cerebral malaria. Angiographic signs include capillary non-perfusion, vessel mottling, and leakage. (A) Colour retinal image showing white-centred haemorrhages and retinal whitening extending from the macula into the temporal periphery (horizontal raphe). (B) Fluoresein angiography shows severe capillary non-perfusion in the retinal periphery (marked in yellow). Capillary non-perfusion typically coincides with retinal whitening. (C) Leakage of fluorescein from retinal venules. (D) Vessel mottling can be seen on a magnified fluorescein angiogram image. Images are from different subjects.
nuclear and outer plexiform layers are supplied by the deep capillaryplexus, which forms the superficial half of a watershed with the underlying choriocapillaris (McLeod, 2010). This suggests that patterns of retinal whitening in cerebral malaria may correspond to venous congestion in areas with limited collateral drainage as well as high metabolic demand.

**Retinal haemorrhages**

The haemorrhages of malarial retinopathy are often white-centred. Retinal vessel thrombi are more common in fatal paediatric cerebral malaria than in coma of other cause. Microthrombi are sometimes associated with haemorrhage, but often occur independently (White et al., 2009). White-centred haemorrhages also occur in bacterial endocarditis, leukaemia, and a range of other conditions in which capillary fragility and elevated venous pressure seem to be common factors (Duane et al., 1980; Ling and James, 1998; Zehetner and Bechrakis, 2011). Retinal haemorrhages have also been observed in children with severe malarial anaemia without profound coma, and severity seems to increase with decreasing consciousness (Beare et al., 2004). In paediatric cerebral malaria, the number of retinal haemorrhages at autopsy correlates significantly with the number of brain haemorrhages (White et al., 2001). Retinal haemorrhages usually involve the inner retinal layers, but can extend to all layers. Different locations within the retina give rise to the appearance of blot or flame haemorrhages. Subretinal haemorrhage with secondary retinal detachment is seen with unusually large retinal haemorrhages (White et al., 2009).

**Manifestations of cerebral malaria: paediatric retina and brain compared**

Several similarities exist between paediatric retina and brain vascular pathology (Table 1).

Microvascular sequestration is a defining histopathological feature of paediatric cerebral malaria in both retina and brain, and is thought to be the principal cause of tissue injury in both sites. Whenever brain and retinal histology has been compared in cases of fatal paediatric cerebral malaria, brain sequestration is always associated with retinal sequestration (Lewallen et al., 2000; White et al., 2009). On the other hand retinal vessel discolouration is not seen in fatal coma of other cause, implying absence of retinal sequestration in these cases (White et al., 2009). Sequestration appears to be patchy within microvascular networks of each organ, but is more widespread in brain than retina.

Whereas the density of sequestration in paediatric cerebral malaria appears to be roughly equal between cerebral white and grey matter, cerebellum and brainstem (Armah et al., 2005), the distribution and density of sequestration in the eye as a whole has not yet been formally evaluated. Pilot data from a small number of eyes from children with cerebral malaria suggest that both the percentage of parasitized vessels and the intensity of sequestration are higher in the retina than in the adjacent choroid (Hiscott and Barrera, unpublished observations).

The intensity of tissue specific sequestration within the eye might be explained by the distribution of endothelial receptors in different vascular beds. ICAM1 is constitutively expressed by retinal vascular and choriald endothelium at low levels (Duguid et al., 1992), and in the choriocapillaris expression is greatest at the macula (Mullins et al., 2006). Expression of retinal endothelial ICAM1 increases in infectious (Toxoplasma gondii) (Smith et al., 2007) and non-communicable diseases (Funatsu et al., 2005), and in response to vascular endothelial growth factor (Lu et al., 1999).

Physiological differences between retinal and choroidal vascular beds may also influence sequestration, as they differ significantly in terms of capillary width, blood flow volume and oxygen extraction. In isolated rat microvessels sequestration density is inversely related to venule diameter, suggesting flow velocity and shear rate may be important (Kaul et al., 1991). Several authors suggest that microvascular architecture may contribute to differential sequestration rates in various organs (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993).

White-centred and ring haemorrhages are common in the retina and brain, respectively. Presumably ring haemorrhages are spherical before histopathological sectioning, which like retinal imaging, provides a 2D view of the observed tissue. The haemorrhages of paediatric cerebral malaria generally affect inner retinal layers and cerebral white matter. In both sites long vessel segments are present, and these may be important in determining the localization of haemorrhages (Spitz, 1946). Clinically, haemorrhages in malarial retinopathy may occur without white centres (i.e. blot haemorrhages), or may develop white centres over time. At histopathology retinal haemorrhages often appear similar to ring haemorrhages—centred on a small thrombosed vessel with a halo of non-parasitized erythrocytes (White et al., 2009). Variations in appearance result from the histological section and differences between retinal and cerebral cellular architecture.

Some areas of retina and brain appear to be affected more frequently by perfusion abnormalities than others. In the retina, fluorescein angiogram imaging suggests that watershed regions such as the horizontal raphe and margin of the foveal avascular zone are especially susceptible to capillary non-perfusion (Beare et al., 2009). In the brain, patterns of T2 and diffusion-weighted imaging signal changes on MRI (Potchen et al., 2012) could represent boundaries of venous territories. Analysis of venous watershed regions in retina and brain may identify vessel properties that are important for the microvascular pathogenesis of cerebral malaria.

**Haemorheology and neurovascular manifestations of cerebral malaria**

Blood flow characteristics such as viscosity, hematocrit, and shear stress are relevant to paediatric cerebral malaria because they are likely to influence both the delivery of parasitized erythrocytes to
organ regions and the propensity for adherence to the endothelium and other erythrocytes. Shear stress strongly influences endothelial binding, for both leucocytes (Xu et al., 2003; Crane and Liversidge, 2008) and parasitized erythrocytes (Fedosov et al., 2011b), and is related to the deformability of parasitized and uninfected erythrocytes in severe adult falciparum malaria. Admission erythrocyte deformability is reduced in adult severe malaria compared with uncomplicated cases, and is associated with mortality (Dondorp et al., 1997). Platelet-mediated auto-agglutination (clumping) of erythrocytes is also associated with severity of malaria in adults (Pain et al., 2001; Chotivanich et al., 2004).

The movement of blood in small vessels is complex and depends on the character of blood and the vascular network it flows through. Unlike water (a Newtonian fluid), blood is a suspension of cells in plasma, and is an example of a shear thinning non-Newtonian fluid. The movement of blood in small vessels therefore varies with viscosity, haematocrit, blood cell deformability, aggregation and interaction with the endothelium (Schmid-Schönbein, 1999; Baskurt and Meiselman, 2003; Lipowsky, 2005; Popel and Johnson, 2005).

Blood viscosity decreases with increasing shear rate (Fig. 2), and as shear rate is related to blood velocity and vessel width, blood moves more easily at greater velocities in vessels of lower calibre (Baskurt and Meiselman, 2003). Erythrocyte aggregation and deformations are major determinants of shear thinning (Popel and Johnson, 2005), and both are altered in P. falciparum infection (Dondorp et al., 1997; Pain et al., 2001; Chotivanich et al., 2004; Fedosov et al., 2011b). Erythrocyte stiffness is particularly important under high shear conditions, whereas non-streamlined aggregates increase resistance at low shear rates that are insufficient to bond between erythrocytes (Baskurt and Meiselman, 2003). Consistent with this, experimentally induced rosetting of P. falciparum infected erythrocytes is seen in venules, but not arterioles where shear rates are likely to be higher (Kaul et al., 1991). Besides shear rate, viscosity depends on the volume fraction of erythrocytes in plasma (i.e. haematocrit). Rising haematocrit is associated with an exponential increase in viscosity.

Variable viscosity

Under normal conditions blood viscosity decreases as vessel width reduces. This phenomenon is known as the Fähraeus-Lindqvist effect, and is thought to result from migration of erythrocytes away from the vessel wall and into a central column—reducing resistance to flow by creating a lubricating cell-depleted layer next to the endothelium. Apparent viscosity reaches a minimum at an internal diameter of 5–7 μm (close to erythrocyte dimensions: ~6–8 μm diameter, 2 μm thick) after which it rises steeply (Popel and Johnson, 2005). Apparent viscosity is greater in vivo than in equivalent glass tubes of the same internal diameter, and this is thought to result from resistance to flow produced by an endothelial lining (the endothelial surface layer or glycocalyx) (Pries et al., 2000; Popel and Johnson, 2005). Protrusion of endothelial cell pseudopods into the vessel lumen during endothelial activation can almost double capillary resistance to flow (Schmid-Schönbein, 1999), and sequestered erythrocytes, or associated inflammation, may produce a similar effect.

Erythrocytes are by far the most common suspended component of blood, and so under physiological conditions, local haematocrit and shear rate are the major determinants of apparent viscosity in microvessels (Lipowsky, 2005). Leucocytes make up a relatively small fraction of total blood volume (~1/600). Nonetheless, as a typical inactivated neutrophil is ~8 μm wide, temporary obstruction of capillaries (internal diameter 4–8 μm) is common, and leucocytes can increase resistance to blood flow within an organ even without binding to endothelium, depending on leucocyte count, haematocrit and the capillary length of the organ involved. The effect is greater in organs with long capillary segments than in those with short segments such as the pulmonary circulation, and probably results from reductions in capillary erythrocyte velocity to match that of the slower leucocytes (Schmid-Schönbein, 1999). Leucocytes, which are relatively large and stiff, also strip the endothelial surface layer from capillary endothelium as they pass. This effect may persist into post-capillary venules and result in increased exposure of endothelial receptors such as ICAM1 (Popel and Johnson, 2005), presumably...
facilitating endothelial binding and subsequent migration through the vessel wall. Parasitized erythrocytes are similar to leucocytes in that they are relatively stiff and bind to ICAM1 (Fedosov, et al., 2011a; Moxon et al., 2011), and this may contribute to microvascular congestion, whereas exposure of capillary and post-capillary endothelial receptors as a result of stripping of the endothelial surface layer may be an important step in sequestration. Post-capillary venules may favour sequestration because shear stress is lower than in arterioles (Nagaoka and Yoshida, 2006). Microvascular resistance is likely to be raised by high proportions of inflexible erythrocytes, sequestration and auto-agglutination, leading to reduced velocity, increased viscosity, and lower shear stress. Increased viscosity can have a dramatic effect on the retinal circulation, and ultimately lead to venous stasis or occlusion (Pourrara, et al., 2008). Congestion and blockage of venules and capillaries secondary to high blood viscosity is therefore consistent with clinical signs of malarial retinopathy such as haemorrhage and capillary non-perfusion.

**Variable haematocrit**

Local microvascular haematocrit is not the same as systemic haematocrit. The haematocrit of blood flowing into a small tube is significantly less than that measured in the static effluent exiting the tube. This is known as the Fåhræus effect, and—as with the Fåhræus–Lindqvist effect—is thought to result from the tendency of flowing erythrocytes to migrate axially to form a central column (Lipowsky, 2005; Popel and Johnson, 2005).

Microvessel haematocrit is not distributed evenly between vessels. When blood meets a bifurcation there is unequal division of erythrocytes into the daughter vessels. Distribution depends on parent vessel haematocrit, branching angle and daughter vessel flow rates. Flow rate is especially influential. This effect is known as phase separation and, again, is thought to result from the axial position of erythrocytes (Fig. 2). As a result microvessel haematocrit is heterogeneous across a network (Popel and Johnson, 2005; Hirsch et al., 2012), and can show great variation (Ganesan et al., 2010; Guibert et al., 2010). As haematocrit is a major determinant of blood viscosity, the concept of phase separation and heterogeneous haematocrit within microvascular networks is crucial to understanding the possible role of haemorheological factors in the retinal and cerebral manifestations of retinopathy-positive paediatric cerebral malaria.

**Computational models of blood flow**

The close relationship between haemorheology and microvascular anatomy and physiology. The choroid is made up of an outer layer of large vessels (Haller’s layer), a middle layer of smaller vessels (Satler’s layer), and an innermost layer of capillaries (the choriocapillaris). The range of capillary width in the choriocapillaris is large compared to inner retina (3–50 μm versus 3.5–6 μm) (Anand-Apte and Hollyfield, 2010). Although anatomical studies classically suggested that the...
Table 2 Comparing vascular features between retina and brain that are likely to be important in cerebral malaria pathogenesis

<table>
<thead>
<tr>
<th>Area of comparison</th>
<th>Similarities / differences</th>
<th>Discussion</th>
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<tbody>
<tr>
<td>Vascular geometry</td>
<td><strong>Similarities</strong></td>
<td>First and second generation retinal arterioles are ~100-μm wide (Nagaoka and Yoshida, 2006), deep white matter arterioles are 100 to 170-μm wide (Nonaka et al., 2003b), arterioles in the putamen are ~100 to 150-μm wide (Nonaka et al., 1998). Retinal perifoveal capillaries are ~5-4-μm wide (Wang et al., 2011), cerebral grey matter capillaries are ~6.5-μm wide (Lauwers et al., 2008), capillaries in the putamen ~5 to 7-μm wide (Wolfram-Gabel and Maillot, 1994). The largest retinal venules are 130-μm to 150-μm wide (Nagaoka and Yoshida, 2006), cerebral grey and white matter venules range up to 125 μm (Duvernoy et al., 1981). Retina (Pournaras et al., 2008), cerebral grey (Cassot et al., 2010) and white matter (Figs 7 and 9 in Nonaka et al., 2003b) all have ~90° branches from relatively long straight trunks. Caudate and putamen have retrograde arteriolar branching, Basal ganglia venous branches join at right angles (Nonaka et al., 1998).</td>
</tr>
<tr>
<td>Vascular topology</td>
<td><strong>Differences</strong></td>
<td>First generation retinal arterioles are ~100-μm wide (Nagaoka and Yoshida, 2006), cerebral grey matter penetrating arterioles are 20- to 65-μm wide (Duvernoy et al., 1981; Reina-de LaTorre et al., 1998). The largest retinal venules are 130- to 150-μm wide (Nagaoka and Yoshida, 2006), principal veins in the putamen can be up to ~500-μm wide (Wolfram-Gabel and Maillot, 1994). Retinal arteriolar and venular length between bifurcations is similar to the length of entire penetrating arterioles or venules in grey matter.</td>
</tr>
<tr>
<td>Watershed regions</td>
<td><strong>Similarities</strong></td>
<td>Strahler order in the macula is ~3.5, in cerebral grey matter it is 3 to 5 (Cassot et al., 2010; Yu et al., 2010). Capillary density immediately around the human foveal avascular zone is similar to primate cortex (Tam et al., 2010).</td>
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<tr>
<td>Metabolic demand</td>
<td><strong>Differences</strong></td>
<td>Human macular superficial and deep plexus have density 40% and 20% per unit area, whereas human grey matter has density ~1.5 to 2% brain volume (Cassot et al., 2006; Lauwers et al., 2008; Mendis et al., 2010). Arteriole/venule ratio in retina is 1:1; in cerebral grey matter it is 2:1; in basal ganglia it is up to 5:1 (Wolfram-Gabel and Maillot, 1994; Cassot et al., 2010).</td>
</tr>
<tr>
<td>Blood flow</td>
<td><strong>Similarities</strong></td>
<td>Both brain and retina have arterial and venous watershed regions. Insufficient venous outflow can cause oedema, haemorrhage, and ischaemia in brain (Teksam et al., 2008) and retina (Browning, 2004).</td>
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<td><strong>Differences</strong></td>
<td>Retinal arteriolar and venular watersheds tend to have the same distribution, e.g. the edge of the foveal avascular zone, and horizontal raphe. In the brain arteriolar and venular watersheds cover different anatomical territories (Miyawaki and Statland, 2003a,b). In the retina venous drainage almost always follows arterioles. Variation in cerebral venous drainage is common in children (Widjaja and Griffiths, 2004).</td>
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<td>Metabolic demand per unit tissue for retina and brain is comparable, and higher than any other organ (Wong-Riley, 2010). Both retina and brain depend on a constant supply of oxygen and glucose (Mckenna et al., 2006). Both inner retina and brain vessels have an arterio-venous O₂ difference of ~40–50% (McLeod, 2010; Seifert and Secher, 2011).</td>
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<td>Retinal metabolism is greatest around the fovea and in retinal layers rich in synapses (Yu and Cringle, 2001; Birol et al., 2007). Cerebral metabolism is greatest in grey matter than white matter (Sokoloff, 2003). Cerebral metabolic demand peaks in childhood: cerebral metabolic rate for O₂ is 4.3 to 6.2 ml O₂/100 g/min (3 to 6 years, whole brain) (Kennedy and Sokoloff, 1957); cerebral metabolic rate for glucose is &gt;30 μmol/100 g/min (1 to 2 years, calcarine cortex, transverse temporal cortex, lenticular nuclei) (Chugani et al., 1987). It is not clear if retinal metabolic demand changes significantly after birth.</td>
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<td>Both retina and brain receive high blood flow volume per unit tissue. Inner retinal blood flow volume is roughly half that of the adult brain (25-50 ml/100 g/min, inner retinal circulation to total brain) (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003; Pournaras et al., 2008). Cerebral blood flow is much higher in early childhood compared with adulthood (130 ml/100 g/min, age 2 to 4 years) (Wintermark et al., 2004). Paediatric peak systolic cerebral blood flow velocity is ~95 cm/s in the middle cerebral artery and ~4.5 cm/s in the central retinal artery (Geeraerts et al., 2005).</td>
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choroid has multiple anastomoses, in vivo functional imaging reveals end-arterial segmental perfusion with associated watershed regions (Hayreh, 1990, 2010b). In the macaque, choroidal blood flow volume is roughly 20 times greater than in the retina, and nine times greater than cerebral grey matter, per weight of tissue (Alm and Bill, 1973; Hayreh 2010b). Wide capillaries and high volumetric flow rate may help to explain why the choriocapillaris is less susceptible to sequestration than inner retinal vessels (Hiscott and Barrera, unpublished observations).

**Geometry of the inner retinal circulation**

The central retinal artery usually divides to produce four branches that extend from the optic disc into the four quadrants of the
Further branching is either at right angles to the main trunk or dichotomous (i.e. two daughter branches at approximately right angles to each other) (Berntson, 1995; Pournaras et al., 2008).

In healthy adults the central retinal artery is ~160 µm wide (Dorner et al., 2002). First and second generation arterioles are ~100 µm wide, whereas first and second-generation venules are ~150 and 130 µm, respectively (Nagaoka and Yoshida, 2006). Retinal arteriolar cross-sectional profiles tend to be circular, whereas venule lumens may be circular but tend to collapse (Feke et al., 1989).

Vascular segments extend close to the anterior limit of the retina, leaving a peripheral avascular zone ~1.5 mm wide. Each terminal arteriole gives rise to a network of 10 to 20 interconnected capillaries (Hayreh, 2010a). Capillary arrangement varies between retinal locations. In general there are two layers: a superficial plexus between the nerve fibre layer and ganglion cell layer, and a deep plexus between the inner nuclear and outer plexiform layers (McLeod, 2010). Only one layer exists adjacent to the fovea and at the far periphery. The peripheral network is also relatively sparse. In the peripapillary region a third capillary layer extends radially from the optic disc for a distance of up to several millimetres (Hayreh, 2010a).

Capillaries are absent at the foveal avascular zone, and adjacent to retinal arterioles (Fig. 3). The foveal avascular zone is supplied by diffusion from the underlying choriocapillaris and in adults is ~400 µm wide and 350 µm high (Yu et al., 2010). Periarteriolar capillary free zones are between 50 and 120 µm wide (Kuwabara and Cogan, 1960). They reflect the combined radius of oxygen diffusion from both artery segment and adjacent capillaries. Capillary-free zones are narrower next to venules, reflecting the high oxygen extraction (~50%) of the retinal circulation and reducing oxygen diffusion radius from post-capillary venules (McLeod, 2010).

Draining venules rise obliquely from the capillary plexuses to the nerve fibre layer and combine to form vascular patterns similar to retinal arterioles. Venules are generally slightly wider than arterioles, with shorter distances between bifurcations, narrower branching angles and less tortuosity (Hughes et al., 2009). Postcapillary venules interdigitate with precapillary arterioles in an alternating pattern (Bek and Jensen, 1993); this can become strikingly apparent in malarial retinopathy when leakage affects venules more than arterioles (Beare et al., 2009) (Fig. 1C). Venules follow arterioles to converge at the optic disc, where they drain into the central retinal vein. Venous blood from the inner retina then flows into the cavernous sinus, either directly or by way of the superior ophthalmic vein. There are no valves (Hayreh, 2010a; Semmer et al., 2010).

Some information exists about vessel geometry in children (Table 3). Paediatric central retinal artery equivalent and central retinal vein equivalent are calculated values based on the widths of first generation retinal vessels, and seem to be similar to adult values. Arteriolar bifurcation angle (the angle subtended between two branches from a parent arteriole) seems to be greater in children than adults, as does arteriolar length to diameter ratio. It is not clear how these differences might affect sequestration.

The macula

The macula is a unique region within the CNS situated temporally to the optic disc and specialized for fine resolution colour vision.
Using the retina to study the brain

Table 3 Retinal vessel geometry in children and adults

<table>
<thead>
<tr>
<th>Measurement</th>
<th>Children</th>
<th>Adults</th>
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<tr>
<td></td>
<td>Age</td>
<td>Value</td>
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<tr>
<td>Central retinal artery</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7–9 y</td>
<td>760</td>
<td>165.2 (164.2–166.3) μm</td>
</tr>
<tr>
<td>43–86 y</td>
<td>4231</td>
<td>165.2 (164.2–166.3) μm</td>
</tr>
<tr>
<td>Central retinal vein</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7–9 y</td>
<td>760</td>
<td>225.4 (224.2–226.8) μm</td>
</tr>
<tr>
<td>43–86 y</td>
<td>4231</td>
<td>242.0 (222.8–223.9) μm</td>
</tr>
<tr>
<td>Arteriolar bifurcation angle</td>
<td>12 y</td>
<td>78.6 (78.3–78.9) %</td>
</tr>
<tr>
<td>Arteriolar simple tortuosity</td>
<td>12 y</td>
<td>0.025 (0.020–0.030)</td>
</tr>
<tr>
<td>Arteriolar length to diameter</td>
<td>12 y</td>
<td>13.1 (11.5, 14.5) mm</td>
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Several subregions exist within the macula, for which adult vessel topology has been described (Yu et al., 2010). Macular arterioles and venules are paired, with an average of nine pairs converging radially towards the fovea. Only three pairs enter the fovea itself to supply the terminal capillary ring marking the edge of the foveal avascular zone. Macular capillaries arise at right angles from parent arterioles and venules (Yu et al., 2010)—an arrangement consistent with significant phase separation and variation in viscosity. Mean perfused capillary width has been measured at 5.4 μm using ultra-high resolution optical coherence tomography (Wang et al., 2011). These capillaries are slightly narrower than capillaries in post-mortem sections of human temporal cortex (6.5 μm) (Lauwers et al., 2008).

The retinal periphery

At the far periphery arterioles form looping arcades with adjacent venules, which may also be traversed by bridging vessels. Trypsin digest reveals peripheral loops with a diameter greater than capillaries at the posterior pole (up to 30 μm versus 5 μm). This arrangement varies between quadrants within eyes, and between individuals (Spitznas and Bornfeld, 1977). If retinal haematocrit is indeed concentrated towards the periphery, these vessels may facilitate flow of blood with relatively higher viscosity than found at the macula. These vascular features are visible in fluorescein angiogram images in paediatric subjects (Penman et al., 1994), though the optical properties of the eye mean that appearances on fluorescein angiogram are likely to be magnified compared to histopathology. In malarial retinopathy normal arteriovenous loops should not be confused with capillary non-perfusion in the far periphery.

Topology of inner retinal vessels

Topological measurements exist for the human macula and fovea. Using the generation number and Strahler taxonomy schemes, average macular branching generation for both arterioles and venules is ~11.5, and average Strahler vessel order is ~3.5. This indicates a high number of bifurcations from each arteriole and venule within a relatively short vessel segment length (Yu et al., 2010). Strahler order in human cerebral grey matter is 3 to 5 (Cassot et al., 2010).

Macular capillary density 1500 μm from the centre of the foveal avascular zone is ~40% in the superficial capillary network and 20% in the deep network, calculated as percentage of sample area filled by vessel segments (Mendis et al., 2010).

The relatively sparse capillary network encircling the foveal avascular zone has been quantified as total capillary length/sample area. Average density ranges from 30 to 34 mm/mm² (Tam et al., 2010). Direct comparison of 2D retinal area with 3D brain volume is difficult, but capillary density in the human brain has been measured at ~250 mm/mm³ (visual cortex) (Bell and Ball, 1985).

Watershed regions

Excepting arteriovenous loops in the far periphery, the absence of arterial, venous, and arteriovenous anastomoses means that there
are several watershed zones in the inner retinal circulation (Fig. 3). These exist between:

(i) Terminal vessels of each of the four arcades, most notably the superior and inferior temporal branches where they form the horizontal raphe;
(ii) Vessels of the retina and vessels of the ciliary body at the anterior limit of the retina; and
(iii) The deep capillary plexus and choriocapillaris throughout the retina, because there is no communication between the inner retina and the choriocapillaris. As the foveal avascular zone is supplied solely by the choriocapillaris this watershed is demarcated clinically in central retinal artery occlusion as the edge of the classic ‘cherry red spot’.

It is important to note that arterial and venous watersheds in the retina are identical, because of pairing of arterioles and venules. This is not the case in the brain, where arterial and venous territories are different.

**Retinal metabolism and blood flow**

The retina is considered by many to be the most metabolically active tissue in the body (Wong-Riley, 2010; Kur et al., 2012). High demands may put it at greater risk of ischaemia in cerebral malaria. Perifoveal oxygen consumption at the dark-adapted macaque is 4.9 ml/100 g/min (Birol et al., 2007). Retinal oxygen consumption in man is estimated at 9.7 ml O$_2$/100 ml retina/min (Anderson and Saltzman, 1964) or 9–10 ml O$_2$/100 g retina/min (assuming retinal specific gravity of 1.0425) (Stefansson et al., 1987), compared with ~3 ml/100 g/min for brain (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003). In the macaque, retinal glucose consumption is among the highest in the CNS (Sperber and Bill, 1985). Estimates of energy expenditure based on ATP used per neuron suggest the human retina may consume ~11.75 mW/g, and the human brain ~10.5 mW/g (assuming retinal weight = 0.4 g, retinal energy = 4.7 mW; brain weight = 1400 g, brain energy = 14.6 W) (Sarpeshkar, 2010).

Only a small proportion of blood from the ophthalmic artery goes to the inner retinal circulation. The value often reported is ~4% (Williamson and Harris, 1994; Pournaras et al., 2008). It is not clear if this figure is from human or animal studies, but for comparison Alm and Bill (1973) found that retinal flow in the macaque makes up ~3.3% of blood to the retina and uvea combined (~27/811 mg/min). Consequently, inner retinal blood flow volume is a small fraction of ocular flow, which is itself a small fraction of blood flow in the internal carotid artery.

In non-human mammals, the retinal layers with highest metabolic rate are the inner segments of the photoreceptors, the outer plexiform layer, and the deeper region of the inner plexiform layer (Yu and Cringle, 2001). Oxygen reaches these layers by diffusion from the choriocapillaris (photoreceptor inner segments), and deep capillary plexus (outer plexiform and deep inner plexiform layers) (McLeod, 2010). In the macaque, demand for oxygen is highest at the perifovea (Birol et al., 2007), and is likely to decrease towards the periphery where photoreceptor density is substantially lower (Jonas et al., 1992). Patterns of metabolic demand may influence the susceptibility of neuronal tissue to the effects of sequestration, and resulting manifestations of malarial retinopathy. Retinal whitening is common in the highly metabolically active perifovea.

Measurements of human mean blood flow volume in the inner retinal circulation have been made using laser Doppler flowmetry, and vary between studies from ~30 to 80 μl/min/retina (reviewed in Pournaras et al., 2008), which is ~9 to 25 μl/100 mg/min, assuming a human retinal weight of 326 mg (Feke et al., 1989). The wide range may result from methodological differences. For comparison, inner retinal blood flow in the macaque, measured by radioactive microspheres, is ~18 μl/100 mg/min (assuming specific gravity of blood = 1.06 and macaque retinal weight of 128 mg) (Alm and Bill, 1973; Feke et al., 1989).

In humans retinal blood flow volume is likely to be approximately half that of the brain, by tissue weight (i.e. 25:50 ml/100 g/min, inner retinal circulation to total brain) (Sokoloff, 2003; Pournaras et al., 2008).

Mean perifoveal capillary flow velocity ranges from 1.37 to 3.3 mm/s. Leucocyte movement is pulsatile, and velocity in perifoveal vessels 7–11 μm wide is ~1.4 mm/s (Pournaras et al., 2008). Rather than passing equally through all capillary segments perifoveal leucocytes seem to follow preferred pathways (Tam et al., 2011). Similar patterns may exist for the movement of parasitized erythrocytes.

Estimated shear stress in healthy adults is highest in first generation arterioles (mean 54.0 dyne/cm²), 20% lower in second-generation arterioles, and 50% lower in both first and second-generation venules (Nagaoka and Yoshida, 2006; Nagaoka et al., 2009).

**The brain**

The circulation of the brain is substantially more complex than that of the retina. We give a brief overview before focusing in more detail on three brain areas typically involved in acute paediatric retinopathy-positive cerebral malaria: the basal ganglia, cerebral grey matter and cerebral white matter. All are affected significantly more often in children with retinopathy-positive cerebral malaria than in paediatric patients with cerebral malaria without retinopathy (Potchen et al., 2012). Retinopathy could therefore reasonably be associated with vascular disease processes in these areas, if microvascular characteristics are similar in both organs.

**Cerebral vasculature architecture and watershed regions**

Carotid and basilar arteries supply the variably anastomotic Circle of Willis, which sends off branches including the anterior, middle and posterior cerebral arteries. Arterial territories are exclusive and vary between individuals (Miyawaki and Statland, 2003a). As in the retina, arterial territories become apparent clinically in thrombo-embolic arterial occlusion when infarction occurs up to the watershed of that arterial territory.

Unlike the retina where arterioles and venules cover identical territories, cerebral venous drainage does not parallel the arterial supply. Several classifications exist (reviewed by Nowinski, 2012), but essentially venous drainage can be understood in terms of...
superficial and deep venous systems (Ono et al., 1984; Oka et al., 1985). Although venous anastomoses exist within each system, there is no physiological anastomosis between superficial and deep systems (Andeweg, 1996, 1999). They join at the torcular (confluens sinuorum), where the straight sinus (draining the deep system) meets the superior sagittal sinus and transverse sinus (draining the superficial system) (Miyawaki and Statland, 2003b). Anatomy is variable but the superficial venous system includes several cortical veins and sinuses that drain the exterior cortex, ranging in average width from ~0.5 to 5.0 mm (Oka et al., 1985). The deep system drains subcortical and periventricular structures such as the insular cortex and basal ganglia. The watershed between the superficial and deep venous systems lies between the periventricular and subcortical white matter (Andeweg, 1996). Ultimately venous blood leaves the cranial vault through the internal jugular or spinal veins, and a small number of emissary veins.

Major venous channels show significant variation (Miyawaki and Statland, 2003b). In a magnetic resonance venography series of healthy children the inferior sagittal sinus was not seen in 54%. The right or left of the following were absent in many cases (% cases absent): vein of Trolard (~80%), vein of Labbe (~50%), superficial petrosal sinus (66%), and inferior petrosal sinus (~33%) (Widjaja and Griffiths, 2004). Lack of major collateral channels would make associated brain regions more vulnerable to venous congestion in paediatric cerebral malaria. Variation in venous drainage could help to explain patterns of MRI signal changes in acute retinopathy-positive paediatric cerebral malaria in the same way that venous territories within the superficial system appear to influence the distribution of radiographic lesions in paediatric cerebral venous thrombosis (Meder et al., 1994; Teksam et al., 2008).

Vascular anatomy of the basal ganglia

The basal ganglia comprise several interrelated nuclei with different neuronal structures, vascular architectures, and embryological origins. The arterial supply of the basal ganglia arises mainly from the middle cerebral artery with some contributions from the anterior cerebral artery and anterior choroidal artery. Branches from the anterior cerebral and anterior choroidal arteries supply the head of the caudate nucleus and globus pallidus (Akima, 1993). Ten to 20 lenticulostriate arteries from the middle cerebral artery enter the putamen, where they spread out in a fan shape and divide into two to three branches, each ~100 to 150 μm wide. These form capillary networks within the putamen and cross the internal capsule to supply the caudate nucleus (Nonaka et al., 1998).

The vasculature of the putamen and caudate nucleus is distinct from that of the globus pallidus, internal and external capsules. Branching arterioles (~50 μm wide) of the putamen may reflect back and coil around the parent vessel before branching further. Precapillary arterioles of the putamen are ~20 μm wide (Wolfram-Gabel and Mailloot, 1994). Retrograde arteriolar branching is seen but it is not clear if this is a feature of old age or is universally present (Nonaka et al., 1998). If the latter they could produce heterogeneous viscosity from phase separation in children. In the globus pallidus, arterioles tend to be fewer and straighter, with no coiling. The end arteriolar branches (~40 μm wide) form arteriolar anastomoses as well as a capillary net (Wolfram-Gabel and Mailloot, 1994).

The capillary networks of the putamen and caudate nucleus have a similar density to the cerebral grey matter (600–650 capillaries per mm² in a 30 to 40 μm mesh), whereas the capillary density of the internal capsule approximates that of the cerebral white matter. The globus pallidus has capillary density between these extremes (200–350 per mm² in a 60–80 μm mesh) (Wolfram-Gabel and Mailloot, 1994; Nonaka et al., 1998). Capillary density seems to be proportional to the density of synapses (Akima, 1993). Capillaries of the putamen are ~5–7 μm wide (Wolfram-Gabel and Mailloot, 1994).

Venules and veins throughout the basal ganglia are generally wider than the arterial vessels, and smaller veins meet larger vessels at right angles (Nonaka et al., 1998). In the putamen, five arterioles are typically arranged around a single central ‘principal’ vein, which together form a vascular unit. These veins can be large (~500 μm wide) (Wolfram-Gabel and Mailloot, 1994). The high ratio of arterioles to veins suggests a similarity to cerebral cortical vessels, and an important difference with the retina where arterioles and venules are paired. Congestion of a principal vein is likely to have a disproportional impact on perfusion. This may be relevant to cerebral malaria since sequestration is thought to occur in venules to a greater extent than arterioles, although it is not clear how often congestion occurs in venules up to 500 μm wide.

Vascular anatomy of the cerebral grey matter

Branches from the anterior, middle, and posterior cerebral arteries produce a network of pial arterioles on the cerebral surface. Cortical arterioles (~20 to 90 μm wide) branch from the pial vessels and travel on the brain surface for ~2150 to 750 μm before turning 90° to penetrate the cortex (Reina-de La Torre et al., 1998).

There are no capillaries on the brain surface, and the absence of arterio-venous anastomoses at this point means that all arterial blood is forced into penetrating cortical arterioles and the capillaries of the cerebral cortex (Duvernoy et al., 1981).

Penetrating cortical arterioles reach depths of 150 μm to ~3 mm and have widths between ~20 and 65 μm. They branch to form capillaries and pre-capillary arterioles at several levels parallel with the brain surface. Mean arteriolar branching angle is 112 to 126°, depending on daughter vessel diameter (Cassot et al., 2010). Some pre-capillary branches are recurrent (Duvernoy et al., 1981; Reina-de La Torre et al., 1998).

The topology of cortical vessel segments can be thought of in terms of ‘tree-like’ penetrating vessels and ‘net-like’ capillaries (Lauwers et al., 2008). The tree-like arteriolar and venular segments have a long parent vessel ‘trunk’ from which smaller branches emanate (Cassot et al., 2010), and are suited to rapid movement of blood, whereas the capillary net is homogeneous and space-filling—suited to nutrient exchange (Lorthois and Cassot, 2010). Cortical penetrating arterioles and venules have a...
Strahler branching order from 3 to 5 between surface pial vessels and the capillary network (Cassot et al., 2010). Unlike capillaries of the inner retina, it seems there are no boundaries between capillaries arising from different arterioles.

Average total vascular density in temporal cortex is \( \sim 500 \text{ mm}^2 \), of which capillaries contribute \( \sim 50\% \) (Cassot et al., 2006). This is consistent with human visual cortex, which has an average capillary density of \( \sim 250 \text{ mm}^2/\text{mm}^3 \) (Bell and Ball, 1985). Narrow arterio-venous anastomoses may exist at the capillary junction, but it is difficult to distinguish these from larger capillaries (Duvernoy et al., 1981).

Ascending cortical venules have \( \sim 50\% \) more branches than arterioles; mean branching angle is 118 to 129° (Cassot et al., 2010). Ascending veins include principal veins (Group V5), which arise from subcortical white matter, and are 120–125 μm wide. The diameter of venules decreases as vessel origin becomes more superficial, ranging from 65 μm (Group V4) to 20 μm (Group VI) (Duvernoy et al., 1981). Once ascending venules reach the cortical surface they make a 90° turn to converge on surface venules (\( \sim 130 \mu \text{m} \) wide). The surface venous network is anastomotic with channels up to 180 μm wide, but has fewer anastomoses than the arteriolar network, and some lobules have no surface venous anastomoses. This drains into larger venules adherent to the arachnoid dura (Duvernoy et al., 1981). The largest veins (1.3 to 3.3 mm) cross the subdural space to meet sinuses incorporated into the dura mater (Oka et al., 1985). There are no venous anastomoses in the cerebral grey matter, and only rare reports of arteriovenous anastomoses (Duvernoy et al., 1981).

Occlusion of cortical vessels

The absence of anastomoses between penetrating vessels means that penetrating cortical arterioles and ascending venules represent a physiological bottleneck. Experimental occlusion of penetrating arterioles in rat cortex leads to severe reduction in blood flow velocity in vessels at least seven branches downstream from the occlusion (Nishimura et al., 2010).

In the rat, ascending venules are twice as prevalent as penetrating arterioles, and this is likely to explain the relatively low extent of flow disruption after ascending venule occlusion (Nguyen et al., 2011). In contrast human grey matter penetrating arterioles outnumber venules on average by 2:1 (Cassot et al., 2010), and principal veins are sometimes surrounded by several rings of arterioles. The tissue volume of such vascular units is variable (Duvernoy et al., 1981). The general implication is that, as in the basal ganglia, a low ratio of venules to arterioles in human cortex may mean venous congestion in cerebral malaria disrupts flow to disproportionately large areas of tissue.

Vascular anatomy of the cerebral white matter

The cerebral white matter is supplied by penetrating arterioles from the pial arterial network, which remain unbranched within the cerebral grey matter until its deepest layer. These arterioles enter the subcortical white matter perpendicular to the brain surface. Vessels express branches at \( \sim 90° \) from the main trunk—five to ten, depending on trunk length. Only a proportion of arterioles reach the periventricular zone, where they arborize. A few form arteriolar anastomoses (Nonaka et al., 2003a,b). Penetrating arterioles have an internal diameter of 30 to 40 μm in subcortical white matter, compared to 40 to 60 μm in the grey matter (Nonaka et al., 2003a), and 100 to 170 μm in deep white matter (Nonaka et al., 2003b).

There has been disagreement about whether the deep white matter also receives arterioles from ventricular sources (Van den Bergh and Vander Eecken, 1968). Moody et al. (1990), Nelson et al. (1991) and Nonaka et al. (2003b) found no evidence of periventricular arterioles directed towards the brain surface.

Cerebral white matter is drained by superficial medullary veins and deep medullary veins. The former drain subcortical white matter into superficial cerebral veins on the brain surface, and the latter drain periventricular white matter into subependymal veins at the lateral ventricle (Hassler, 1966; Meder et al., 1994; Andeweg, 1996; Nonaka et al., 2003b). Superficial medullary veins are thought to drain the external 1–2 cm of white matter (Meder et al., 1994), and the deep medullary veins the remainder (Hassler, 1966; Hooshmand et al., 1974). Trans-cerebral veins connecting superficial and deep venous systems have been reported (Huang and Wolf, 1964; Hassler, 1966; Meder et al., 1994; Nakamura et al., 1994; Schaller, 2004), but may be artefactual as functional anastomoses between superficial and deep venous systems are not consistent with patterns of injury seen after deep venous system obstruction (Hassler, 1966; Andeweg, 1999). The predominance of haemorrhages in white matter is consistent with relatively long venous drainage pathways (Spitz, 1946).

Cerebral blood flow and metabolism

Like the retina the brain is highly metabolically active (reviewed in Wong-Riley, 2010), and depends on a constant supply of oxygen and glucose (Mckenna et al., 2006).

Normal cerebral blood flow characteristics vary with age and between brain regions (Kety, 1950; Ide and Secher, 2000). Abnormal cerebral blood flow in the middle cerebral arteries has been reported in cases of paediatric cerebral malaria with seizure or raised intracranial pressure (Newton et al., 1996).

Total cerebral blood flow and metabolism changes dramatically during childhood (Kety, 1956; Kennedy and Sokoloff, 1957; Kinnala et al., 1996; Takahashi et al., 1999; Wintermark et al., 2004; Mckenna et al., 2006). Cerebral blood flow increases rapidly during the first year of life (Varela et al., 2012) and flow volume peaks at 55% cardiac output between age 2 to 4 (\( \sim 130 \text{ ml/100 g/min} \)) before declining to adult values (Wintermark et al., 2004). In comparison the brain of a resting adult receives \( \sim 16.5\% \) cardiac output (Kety, 1950), or \( \sim 50 \text{ ml/100 g/min} \) (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003).

Cerebral metabolic rate for oxygen is also highest from 3 to 6 years, ranging from 4.3 to 6.2 ml O₂/100 g/min (Kennedy and Sokoloff, 1957). The average of 5.25 ml would be \( >50\% \) total body oxygen (Mckenna et al., 2006). Adult resting cerebral metabolic rate for O₂ is \( \sim 3 \text{ ml/100 g/min} \) (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003), and accounts for \( \sim 20\% \) of total body oxygen consumption (Kety, 1950). Cerebral
arteriovenous oxygen difference is ~40% in adults (Seifert and Secher, 2011), comparable to retinal oxygen extraction of ~50% (McLeod, 2010). The developmental curve for cerebral metabolic rate for glucose follows a similar trajectory to cerebral blood flow, though with a more gradual decline to adult values at around age 17 (Chugani et al., 1987; Kinnala et al., 1996; Chugani, 1998).

The highest regional peak in cerebral blood flow is in the calcarine cortex (area V1, 155 ml/100 g/min, 29 months). Basal ganglia, temporal, frontal, and occipital cortices have lower metabolic rates for glucose (30 to 60 ml/100 g/min). Cerebral blood flow in the corpus callosum at this age is in the calcarine cortex, transverse temporal cortex, and lenticular nuclei (all 40% in adults (Seifert and Secher, 2011)).

Flow in cerebral microvessels

Data on capillary flow dynamics in human brain are scarce, although many studies on animal models exist (Hudetz et al., 1996; Vovenko, 1999; Jespersen and Ostergaard, 2011). Capillary flow velocity is between 0.6 and 2.4 mm/s in rat cortex (Hudetz et al., 1996), which overlaps the lower range of velocities in human perioveal vessels (Pourmaras et al., 2008).

Guibert et al. (2010) have modelled blood flow in primate cortex, which has a ratio of arterioles to venules similar to human brain. They found that local haematocrit could vary by approximately a factor of two owing to phase separation. Brain haematocrit measured by dual tracer single-photon emission computed tomography in adult humans is ~76% systemic haematocrit (Sakai et al., 1985). Blood is thought to make up ~3–5% of brain volume, and three quarters of this is venous (An and Lin, 2002). Blood volume is greater in grey (~5 ml/100 g) than white matter (~2 ml/100 g) (Kuppusamy et al., 1996) consistent with greater capillary density in grey matter.

Retina and brain and brain compared: implications for studying neurovascular pathogenesis

Retina and brain microvasculature have much in common, including circulation–tissue barriers and local control over blood flow (Patton et al., 2005). After reviewing microvascular haemorheology and the microvasculature of retina and brain regions, it seems that sequestration-induced injury could be influenced by several characteristics (Table 2).

Vessel branching

Retinal vessels are relatively long and straight, with right-angled or dichotomous branches. This is also true of cerebral grey matter, white matter and basal ganglia—although brain-specific features are also present. Haemodynamic models suggest near 90° arteriolar branching produces significant variation in haematocrit and viscosity (Ganesan et al., 2010; Guibert et al., 2010) and therefore may be important in distributing parasitized erythrocytes and fostering sequestration. Right-angled venular convergens may also provide opportunities for sequestration since cells may continue to roll along the endothelium when moving from a smaller to larger vessel before joining the axial flow stream, potentially stripping the endothelial surface layer and exposing ICAM1 (Schmid-Schönbein, 1999; Popel and Johnson, 2005). Local variation in haematocrit and viscosity associated with branching architecture may explain why retinal vessels full of parasitized erythrocytes can be found immediately adjacent to vessels unaffected by sequestration (Lewallen et al., 2000).

Arteriole to venule ratio

Both basal ganglia and cerebral grey matter have more arterioles than venules, whereas in the retina, arterioles and venules are paired. A high ratio of arterioles to venules in the absence of anastomoses may make these brain regions more vulnerable than the retina to the effects of post-capillary venous congestion with parasitized erythrocytes.

Venous drainage and watershed regions

Retina and brain both have vascular watersheds that are amenable to imaging. Although retinal arterial and venous watersheds are identical, retinal arterioles and venules can be distinguished using fluorescein angiogram. In the brain, arterial supply and venous drainage occur over different territories, and radiological changes to these anatomical regions can be assessed using MRI. Retinal capillary non-perfusion is most common in watershed regions, and brain lesions may also follow patterns of venous drainage into superficial and deep systems, especially those of the cerebral white matter (Meder et al., 1994; Andeweg, 1996). Sequestration is thought to mainly involve capillaries and post-capillary venules, and retinal white-centred haemorrhages occur in other conditions involving venous congestion (Duane et al., 1980). These points suggest comparison of retinal and cerebral venous watersheds in cerebral malaria is reasonable, and perhaps that the severity of non-perfusion in retinal watersheds might reflect the degree of $T_2$ or diffusion-weighted imaging abnormalities in the cerebral white matter in acute cerebral malaria (Fig. 4). The consequences of venous congestion are much greater in brain than retina. For example, swelling of the splenium, or posterior cerebrum (Potchen et al., 2012) could compress the vein of Galen against the tentorium, potentially obstructing outflow from the deep venous system and rapidly raising intracranial pressure (Andeweg, 1999).

Blood flow and metabolism

Adult inner retinal blood flow is roughly half that of the brain. To our knowledge there are no data available on developmental changes to retinal blood flow during childhood. However, cerebral blood flow varies dramatically during childhood development, peaking within the age range typically admitted with paediatric cerebral malaria in high transmission areas (Snow et al., 1997; Wintermark et al., 2004). The median age at admission with cerebral malaria in high transmission areas appears to coincide with the childhood peak in total cerebral blood flow and metabolism. For example, the mean
age of children admitted to the paediatric research ward in Blantyre, Malawi, ranged from \(~3\) to 4 years between 2001–10 (Roca-Feltrer et al., 2012). Snow et al. (1997) found the median (IQR) age at admission with cerebral malaria was low in high transmission areas and increased with falling endemicity—ranging from from 2 (1.5–6.5) years (hyperholoendemic), 4 (3–5) (mesoendemic), to 6.5 (4–7) years (hypoendemic). A similar pattern is found using residential altitude as a proxy for transmission intensity (Reyburn et al., 2005). As suggested by others (Billig et al., 2012) high metabolic demands may make the paediatric brain more vulnerable to the effects of sequestration than other paediatric organs, and in the context of high transmission predispose to early cerebral complications rather than the acute renal failure or pulmonary oedema seen in adults (Thanachartwet et al., 2013).

Retina and brain are comparable in that they both have very high metabolic demands, and in early childhood the brain may be more susceptible than the retina (and other organs) to the effects of dysfunctional microvascular flow. However cerebral malaria is only one manifestation of severe malaria in children, and it is not clear how high paediatric cerebral metabolic rate for \(O_2\) in isolation could influence which children develop cerebral malaria, and which develop severe malarial anaemia or metabolic acidosis.

Complicating factors

The clinical picture of paediatric cerebral malaria is complicated by the presence of multiple interrelated insults in addition to sequestration, including anaemia, hypoglycaemia, and metabolic acidosis.
Insults, seizures are common (Birbeck reviewed in Hochman and Kim, 2012). In addition to systemic insults, seizures are common (Birbeck et al., 2010) and could inflict brain injury without corresponding damage to the retina. Retinal signs could indicate a high tide mark of tissue injury common to particular regions of both retina and brain, which may or may not reach a threshold needed to induce coma, and thereafter trigger additional brain damage from other mechanisms, such as seizure. Alternatively, one or more co-insults may be necessary for development of the cerebral malaria phenotype, as malarial retinopathy is also seen in children with severe malarial anaemia without coma, albeit much less severely (Beare et al., 2004).

Conclusions

Description of the whole brain in terms of greater or lesser similarity to the whole retina is difficult, and probably inappropriate. Rather, analysis of data from retina and brain should take account of anatomical variation within both organs, as neither retina nor brain are homogeneous units. Both have regions where microvasculature may, or may not, be comparable in specific respects that are relevant to a given neurovascular disease.

Retina and brain are similar in ways relevant to paediatric cerebral malaria. Similar vascular pathology suggests both organs experience similar disease processes. Similarities in terms of vessel network architecture, imaging abnormalities in venous watersheds, and high metabolic demand suggest that both organs may be vulnerable to sequestration for the same reasons. These factors may influence delivery of parasitized erythrocytes, the flow conditions necessary for sequestration, and the vulnerability of tissue to ischaemia associated with venous congestion or occlusion. If so, the impact of congestion on common venous anatomy may explain the distribution of MRI brain and retinal fluorescein angiogram lesions in paediatric cerebral malaria.

The retina and brain also have differences. Arteriovenous ratios vary between brain regions, while in the retina arterioles and venules are paired. Cerebral arterial and venous watersheds are different, while retinal watersheds overlap. The paediatric brain has a metabolic peak in early childhood, which is likely to raise cerebral metabolic demands significantly above retinal demands in the age range typically affected by cerebral malaria.

Considering these similarities and differences, a biologically plausible analogy can be made between the retinal periphery, or horizontal raphe, and the cerebral white matter, because these regions have similar vascular architecture. All receive blood from long vessels with near 90° branches, and so may contain blood with disproportionately high microvascular haematocrit and viscosity. The presence of venous watersheds gives little option for collateral drainage from these areas in the event of venous congestion. Given this background, similar degrees of coexistent injury to these retinal and brain areas in a population of children with retinopathy-positive cerebral malaria would suggest that retinal capillary non-perfusion and leakage could be surrogates for brain ischaemia and swelling, and might provide a useful tool to understand cerebral vascular pathogenesis.

Our review has limitations. Only three brain regions are compared with the retina, and comparable data are not available for all geometrical or topological variables at each site. We have resort to measurements from adults where paediatric data are not available, and we have not attempted comparison of retinal or cerebral microvascular architecture with other organs in which sequestration is minimal or absent. We have not compared paediatric and adult malarial retinopathy—this deserves a separate review.

We have identified significant similarities and differences in retinal and cerebrovascular architecture, blood flow and metabolic demand. These comparisons inform new interpretations of retinal and cerebral data in retinopathy-positive paediatric cerebral malaria by incorporating information on haemorheology and vascular architecture from multiple regions of the CNS. Increasingly comprehensive comparisons of human neurovasculature will provide greater insights into how far retinal vessel changes reflect neurovascular pathology. Mapping the contours of warranted analogy between retina and brain has the potential to strengthen the biological rationale of retinal imaging in neurology, inform interpretation of neurovascular data, and stimulate further hypotheses—for retinopathy-positive paediatric cerebral malaria in particular, and for neurovascular disease in general.

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References


Billig EMW, O’Meara WP, Riley EM, McKenzie FE. Developmental allostery and paediatric malaria. Malar J 2012; 11: 64.


