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Cerebral malaria in children: using the retina to study the brain

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Cerebral malaria is a dangerous complication of Plasmodium falciparum infection, which takes a devastating toll on children in sub-Saharan Africa. Although autopsy studies have improved understanding of cerebral malaria pathology in fatal cases, information about in vivo neurovascular pathogenesis is scarce because brain tissue is inaccessible in life. Surrogate markers may provide insight into pathogenesis and thereby facilitate clinical studies with the ultimate aim of improving the treatment and prognosis of cerebral malaria. The retina is an attractive source of potential surrogate markers for paediatric cerebral malaria because, in this condition, the retina seems to sustain microvascular damage similar to that of the brain. In paediatric cerebral malaria a combination of retinal signs correlates, in fatal cases, with the severity of brain pathology, and has diagnostic and prognostic significance. Unlike the brain, the retina is accessible to high-resolution, non-invasive imaging. We aimed to determine the extent to which paediatric malarial retinopathy reflects cerebrovascular damage by reviewing the literature to compare retinal and cerebral manifestations of retinopathy-positive paediatric cerebral malaria. We then compared retina and brain in terms of anatomical and physiological features that could help to account for similarities and differences in vascular pathology. These comparisons address the question of whether it is biologically plausible to draw conclusions about unseen cerebral vascular pathogenesis from the visible retinal vasculature in retinopathy-positive paediatric cerebral malaria. Our work addresses an important cause of death and neurodisability in sub-Saharan Africa. We critically appraise evidence for associations between retina and brain neurovasculature in health and disease, and in the process we develop new hypotheses about why these vascular beds are susceptible to sequestration of parasitized erythrocytes.

Keywords: cerebral malaria; cerebral microvasculature; retinal microvasculature; haemorheology; surrogate marker
Abbreviation: CI = confidence interval; CM = cerebral malaria; CMRgluc = cerebral metabolic rate (glucose); CMRO2 = cerebral metabolic rate (oxygen); CNP = capillary non-perfusion; CRAE = central retinal artery equivalent; CRVE = central retinal vein equivalent; DD = disc diameter; FA = fluorescein angiogram; FAZ = foveal avascular zone; ICAM-1 = intercellular adhesion molecule 1; LDR = length to diameter ratio; MRI = magnetic resonance imaging; OR = odds ratio; PfEMP-1 = Plasmodium falciparum erythrocyte membrane protein 1; SD = standard deviation

Introduction

Paediatric cerebral malaria is a clinical syndrome that kills and disables children through mechanisms that remain incompletely understood. Adhesion of parasitized erythrocytes to the microvascular endothelium, leading to their sequestration in the brain, is the pathological signature of both adult and paediatric cerebral malaria, and is thought to be the chief cause of injury (Taylor et al., 2004; Ponsford et al., 2012). Several hypothetical mechanisms linking sequestration, which is entirely intravascular, to extravascular parenchymal damage have been proposed (Van der Heyde et al., 2006) but questions remain about which of these mechanisms are most important, how they might interact, and ultimately where new therapies should be directed. One of the reasons why such questions remain over 100 years after sequestration was first identified is because in vivo access to the brain is difficult, and advances in knowledge have relied on post-mortem and in vitro studies. Improved understanding of in vivo neurovascular pathogenesis and the development of better treatments will be facilitated by disease models or surrogate markers.

The retina may be a good source of surrogate markers of cerebrovascular injury because paediatric cerebral malaria is associated with a retinopathy (‘malarial retinopathy’) that accurately predicts cerebral sequestration (Taylor et al., 2004), correlates with severity of brain involvement (White et al., 2001), and is associated with mortality (Beare et al., 2004). Unlike the brain, the eye allows non-invasive access for structural and functional imaging of the microcirculation, which is thought to be the major site of sequestration.

The concept that neurovascular injury observed in the retina resembles neurovascular injury lying unseen in the brain is based on the assumption that the two circulations are analogous in ways that are relevant to the pathogenesis of paediatric cerebral malaria. Such assumptions should be supported by a biologically plausible rationale before specific retinal vascular features are considered as surrogate markers of cerebrovascular damage (International Conference on Harmonisation, 1998).

Is such a rationale likely? It is well known that the retina and brain have many similarities. Both are part of the CNS, with common embryological origins, vascular structure and metabolic demands. The relevance of these similarities for potential surrogate markers has been recognized (Patton et al., 2005, 2006), especially for stroke (Doubal et al., 2009, 2010). Despite this, detailed comparisons of the microvasculature of the two organs are rare (Cogan and Kuwabara, 1984; Patton et al., 2005).

Our objective was to discover how likely it is that the retinal vascular damage responsible for malarial retinopathy reflects analogous cerebrovascular damage in retinopathy-positive paediatric cerebral malaria. To do this we compared the manifestations of paediatric cerebral malaria in retina and brain, and then compared retina and brain in terms of vascular features likely to be important for cerebral malaria pathogenesis. Concluding that retina and brain are analogous in ways relevant to this disease would justify further investigation to see whether specific retinal signs predict both focal brain damage detectable by MRI and the patient’s response to treatment. The results of such investigations would address further criteria of surrogacy (International Conference on Harmonisation, 1998), and in the context of a strong biological rationale could shed light on the dynamics of cerebral malaria neurovascular pathogenesis in the period between coma onset and recovery or death.

Malarial retinopathy can occur in parasitaemic children without cerebral malaria (Beare et al., 2004), and indeed, some features of malarial retinopathy occur in conditions that don’t involve malaria at all (e.g. white-centred haemorrhages). It is not clear if retinopathy predicts cerebral sequestration in malarial syndromes besides paediatric cerebral malaria, or how often retinopathy might occur in severely ill children in general.

We therefore discuss retinopathy in the specific clinical context of paediatric cerebral malaria. In this particular population malarial retinopathy has high positive and negative predictive value to distinguish between the presence and absence of cerebral sequestration in fatal cases (Taylor et al., 2004). Until associations between retinopathy and cerebral sequestration are known for malaria in general, extrapolation from our review to severe malaria syndromes other than paediatric cerebral malaria may not be appropriate.

Our review is divided into four sections. In the first section we describe paediatric cerebral malaria, including the typical clinical presentation, manifestations of cerebral malaria in the brain, and manifestations in the retina. The neurovascular effects of cerebral malaria on retina and brain are then compared.

In the second section we introduce the concept that patterns of neurovascular pathology in retina and brain may be understood in terms of haemorheological dysfunction involving microvascular haematocrit, blood viscosity and shear stress. These factors depend on interactions between intrinsic properties of blood and structural properties of microvascular networks, and this suggests that microvascular architecture may be a useful point of comparison for a cerebral malaria-specific analogy between retina and brain.

The third section describes microvascular architecture in the human retina and brain, and compares features that may be relevant to the pathogenesis of cerebral malaria.

We end by discussing how our comparison of retina and brain provides a biologically plausible rationale for considering retinal signs as potential surrogates of brain damage in retinopathy-positive paediatric cerebral malaria. We consider how observations of retinal vessel structure and function might allow inference of in vivo cerebrovascular pathogenesis.
Paediatric cerebral malaria

Definition

Several species of *Plasmodium* parasite cause malaria in humans. *P. falciparum* is responsible for the majority of severe malaria and malaria-associated deaths worldwide, particularly in sub-Saharan Africa where severe malaria has a disproportionate impact on children <5 years of age (WHO, 2012). In children, severe malaria predominantly involves one or more of three syndromes: cerebral malaria, severe malarial anaemia and metabolic acidosis. Other manifestations of paediatric severe malaria include convulsions, hypoglycaemia, hyperparasitaemia and prostration (WHO, 2000). Cerebral malaria is defined as coma, with *P. falciparum* peripheral parasitaemia, in the absence of another identifiable cause of coma, such as hypoglycaemia or meningitis (Newton et al., 1998). This definition is broad, and misclassifies almost 25% of fatal paediatric cases as cerebral malaria when compared with histopathological identification of sequestration in cerebral vessels as the reference standard. The presence of malarial retinopathy on pre-mortem fundus examination accurately distinguishes histopathological cerebral malaria from cases that meet the clinical definition but actually have another cause of death (Taylor et al., 2004). Although the strong association between retinopathy and intracerebral sequestration has been established only in fatal cases, it is likely that the association also prevails in those who recover, and that the presence of retinopathy improves the accuracy of diagnosis in children with cerebral malaria (Beare et al., 2011).

Typical presentation and clinical course

In an area with intense transmission of *P. falciparum*, the typical history of paediatric cerebral malaria involves a young child who rapidly develops coma after a short prodrome of fever and generalized illness. The coma may be preceded or accompanied by convulsions, from which the child does not wake. On examination the child is usually febrile, and may have deep or rapid breathing. Coma is distinguished from prolonged post-ictal state by duration >30 min after seizure. Depth of coma is assessed using a modification of the Glasgow Coma Scale, known as the Blantyre Coma Scale (Molyneux et al., 1989; WHO, 2000, 2010). Convulsions or posturing, including opisthotonus, may be present. Ophthalmoscopy through dilated pupils reveals signs of malarial retinopathy: white patchy discoloration of the macula and/or peripheral retina; orange or white discoloration of retinal vessels; and/or retinal haemorrhages, typically with white centres. Papilloedema may be seen, but in isolation does not distinguish cerebral malaria from other causes of coma (Lewallen et al., 1999, 2008; Harding et al., 2006). Investigation may reveal concomitant metabolic acidosis and/or severe anaemia. Peripheral *P. falciparum* asexual parasitaemia is present by definition. Other treatable causes of coma in this setting must be ruled out and include malarial-associated hypoglycaemia and meningitis. Bacteraemia may be found, especially in infants and in the presence of severe anaemia (Bronzan et al., 2007).

The duration of illness is usually short, and most patients recover or die within 48 h. Death is typically by respiratory arrest, and case fatality with treatment is ~15%. Those who recover are at risk of neurodisability (Molyneux et al., 1989) and epilepsy (Birbeck et al., 2010). Guidelines for treatment have been published (WHO, 2010).

Cerebral malaria also occurs in adults (Table 1). The clinical presentation is different from paediatric cerebral malaria (WHO, 2000), but it is not clear if this is because of age or differences in immunology related to transmission intensity (Idro et al., 2005). The geographical distribution is different from that of paediatric cerebral malaria. Adult cerebral malaria occurs primarily in South and South East Asia, where transmission of *P. falciparum* is endemic and seasonal and most children and adults have no immunity; whereas paediatric cerebral malaria occurs mainly in sub-Saharan Africa where transmission is endemic and where both older children and adults have acquired partial immunity. Adult travellers who are *P. falciparum* naive may develop cerebral malaria in endemic countries. Patterns of mortality and comorbidity vary with age (Dondorp et al., 2008).

Malarial retinopathy has been described in adults since at least the 1800s (Mackenzie, 1877). Maude et al. (2009) found retinopathy in 14/20 (70%) adults with cerebral malaria, 17/20 (85%) with other types of severe malaria, and 9/15 (60%) with uncomplicated malaria. The retinopathy was most severe in those with cerebral malaria. Abu Sayeed et al. (2011) found retinopathy in 31/75 (41%) with cerebral malaria, 17/20 (85%) with other types of severe malaria, and 1/31 (3%) with uncomplicated malaria. Number of retinal haemorrhages was an independent predictor of death. Both studies used sensitive retinal imaging techniques and a standardized grading scheme (Harding et al., 2006), but neither found the characteristic vessel discoloration seen in up to 30% of children with cerebral malaria (Beare et al., 2004). Compared with the adult literature, few data exist on the presence of malarial retinopathy in children with uncomplicated malaria, as consistent retinal examination of conscious children is difficult—particularly so for the retinal periphery. Further research is needed.

Manifestations of retinopathy-positive cerebral malaria in the paediatric brain

Sequestration

Sequestration is the histopathological hallmark of paediatric cerebral malaria (Taylor et al., 2004). Sequestration results from the binding of parasitized erythrocytes to vascular endothelium. Parasitized erythrocytes also bind *in vitro* to other erythrocytes (rosetting), and to platelets (clumping, or auto-agglutination). Sequestration is mediated by adhesion between malarial antigens on the surface of the infected erythrocyte and several host receptors on the vascular endothelium. The *P. falciparum* surface antigen most studied is *P. falciparum* erythrocyte membrane protein 1.
Table 1 Manifestations of cerebral malaria in the retina and brain

<table>
<thead>
<tr>
<th></th>
<th>Paediatric retina</th>
<th>Paediatric brain</th>
<th>Adult brain</th>
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<tbody>
<tr>
<td><strong>Sequestration</strong></td>
<td>Frequency: Always present in fatal cerebral malaria (Lewallen et al., 2000).</td>
<td>Always present in fatal cerebral malaria, and absent in fatal coma of other cause (Taylor et al., 2004; Dorovini-Zis et al., 2011).</td>
<td>Always present in fatal cerebral malaria (MacPherson et al., 1985; Oo et al., 1987; Pongponratn et al., 1991; Sein et al., 1993). In cerebral malaria density is greater in brain than other organs (MacPherson et al., 1985; Pongponratn et al., 1991). Significant sequestration may be present in fatal non-cerebral malaria (MacPherson et al., 1985; Silamut et al., 1999). The percentage of vessels with sequestration is greater in cerebral malaria than non-cerebral malaria (Ponsford et al., 2012)</td>
</tr>
<tr>
<td><strong>Location</strong></td>
<td>Patchy distribution within capillary network (Lewallen et al., 2000). Variatio</td>
<td>Most microvessels, and the margins of pial and larger vessels (Dorovini-Zis et al., 2011). Grey and white matter of cerebrum, subcortex, brainstem and cerebellum (Armah et al., 2005; Dorovini-Zis et al., 2011).</td>
<td>Occurs in capillaries, venules, and very occasional arterioles (MacPherson et al., 1985). Occurs in grey and white matter, but most dense in cerebral white matter (Nagatake et al., 1992). Density reduces from cerebrum to cerebellum to brainstem (Pongponratn et al., 2003). Density greater in cerebellum than cerebrum (Sein et al., 1993).</td>
</tr>
<tr>
<td><strong>Vessels involved</strong></td>
<td>Capillaries and margins of larger vessels (Lewallen et al., 2000). Vessel discol</td>
<td>Occurs in brain microvessels, pial and larger vessels (Dorovini-Zis et al., 2011).</td>
<td>Predominant site is the capillary bed, but also occurs in larger pial and subarachnoid vessels (Spitz, 1946). Uncommon in arterioles (MacPherson et al., 1985).</td>
</tr>
<tr>
<td><strong>Haemorrhages</strong></td>
<td>Type: White-centred, blot (White et al., 2001). Parasitized erythrocytes rarely s</td>
<td>Ring (Dorovini-Zis et al., 2011). Parasitized erythrocytes rarely seen outside vessel (White et al., 2001; Dorovini-Zis et al., 2011).</td>
<td>Ring, perivascular (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993; Turner, 1997). Parasitized erythrocytes are seen outside vessel (Sein et al., 1993; Turner, 1997). Ring haemorrhages present in up to 30% of cases of fatal cerebral malaria (Spitz, 1946). No significant difference in haemorrhage frequency between cerebral malaria (~60% of cases) and non-cerebral malaria (~40% of cases) (Medana et al., 2011). Usually occur in cerebral white matter; also reported in pons, medulla, cerebellum, and cortical grey matter (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993; Turner, 1997). No difference in haemorrhage frequency between cortex, diencephalon, and brainstem (Medana et al., 2011).</td>
</tr>
<tr>
<td></td>
<td>Frequency: Gross haemorrhages present in 78% fatal cerebral malaria, 7% fatal coma of other cause (White et al., 2009).</td>
<td>Any type present in 80% fatal cerebral malaria (Dorovini-Zis et al., 2011).</td>
<td></td>
</tr>
<tr>
<td><strong>Location</strong></td>
<td>All retinal quadrants. Usually restricted to inner retinal layers, with extension to subretinal haemorrhage in severe cases (White et al., 2009). Common in white matter, rare in grey matter except in the cerebellum (White et al., 2001; Dorovini-Zis et al., 2011).</td>
<td></td>
<td>Usually occur in cerebral white matter; also reported in pons, medulla, cerebellum, and cortical grey matter (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993; Turner, 1997). No difference in haemorrhage frequency between cortex, diencephalon, and brainstem (Medana et al., 2011).</td>
</tr>
<tr>
<td><strong>Vessel leakage</strong></td>
<td>Type: Fibrinogen leakage along vessels with and without associated haemorrhage (White et al., 2009). Fibrinogen leakage often associated with haemorrhage, can be independent of haemorrhage (Dorovini-Zis et al., 2011).</td>
<td>Fibrinogen leakage often associated with haemorrhage, can be independent of haemorrhage (Dorovini-Zis et al., 2011).</td>
<td>Rarefaction of the perivascular space, perivascular pools of proteinaceous material, vascular parenchymal oedema, oedema between fibres of white matter tracts, fluid-filled spaces between myelin fibres. No difference between fatal cerebral and non-cerebral malaria (Medana et al., 2011).</td>
</tr>
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Table 1 Continued

<table>
<thead>
<tr>
<th>Paediatric retina</th>
<th>Paediatric brain</th>
<th>Adult brain</th>
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<tr>
<td><strong>Frequency</strong></td>
<td></td>
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<tr>
<td>Fibrinogen leakage in 31% cases fatal cerebral malaria, 7% fatal coma of other cause (White et al., 2009). Average (SD) number of foci is 1.2 (2.6) in fatal cerebral malaria and 0.21 (1.1) in coma of other cause (White et al., 2009). Associated with vessels but not defined in terms of retinal quadrants or vessel type (White et al., 2009). Angiographic fluorescein leakage predominately affects venules (Beare et al., 2009).</td>
<td>Unclear how many cases of fatal cerebral malaria have leakage of any type. Leakage greater in white than grey matter (associated with haemorrhages) (Dorovini-Zis et al., 2011). Cerebral white and grey matter, subcortex, brainstem and cerebellum (Dorovini-Zis et al., 2011).</td>
<td>At least one type of oedema present in all cases of both cerebral and non-cerebral malaria (Medana et al., 2011). Oedema between white matter tracts is most common in: brainstem &gt; diencephalon &gt; cortex (Medana et al., 2011). Brainstem, diencephalon, cerebral cortex (Medana et al., 2011).</td>
</tr>
<tr>
<td><strong>Location/vessels involved</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Regions vulnerable to presumed ischaemia on imaging</td>
<td>Regions where MRI brain signal changes distinguish between retinopathy-positive and negative cerebral malaria (highest to lowest odds ratio): basal ganglia, corpus callosum, cerebral cortex, thalamus, cerebral white matter, posterior fossa (Potchen et al., 2012).</td>
<td>Regions reported to be involved: Brain stem, thalamus, cerebellum, corpus callosum, cerebral white matter (Yadav et al., 2008; Raalkar et al., 2011).</td>
</tr>
<tr>
<td><strong>Regions reported to be involved</strong></td>
<td></td>
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<tr>
<td>Retinal whitening and capillary non-perfusion appears to be especially prominent at the foveal avascular zone, horizontal raphe, and retinal periphery. All are watershed regions (Beare et al., 2009).</td>
<td>Associated with vessels but not defined in terms of retinal quadrants or vessel type (White et al., 2009).</td>
<td>Related to white matter, with subcortical involvement, extending to periventricular white matter and subcallosal areas (Nikolov et al., 2007).</td>
</tr>
</tbody>
</table>

**Haemorrhage**

Histopathology reveals subtypes of cerebral malaria within fatal retinopathy-positive paediatric cerebral malaria. In Malawi, 75% of cases coming to autopsy have sequestration, and intravascular microthrombi perivascular haemorrhages and intravascular microthrombi, whereas 25% have sequestration but no haemorrhages or oedema. A frequency of cases coming to autopsy have sequestration but no haemorrhages or oedema.

Marginal foci of retinal whitening and capillary non-perfusion appear to be especially prominent at the foveal avascular zone, horizontal raphe, and retinal periphery. All are watershed regions (Beare et al., 2009).

*These points illustrate how tissue damage associated with sequestration may be caused through synergistic mechanisms, including endothelial dysfunction, altered microvascular coupling and perivascular haemorrhages.

**Capillary non-perfusion**

Reduced capillary perfusion may be caused through several molecular cascades, including endothelial dysfunction, altered microvascular coupling, tissue damage, and sequestration. These points illustrate how tissue damage associated with sequestration may be caused through synergistic mechanisms, including endothelial dysfunction, altered microvascular coupling and perivascular haemorrhages.
perivascular pathology visible on routine haematoxylin and eosinstained sections (Dorovini-Zis et al., 2011).

Petechial haemorrhages on the cut surface of fresh brain correspond to microscopic ring haemorrhages. Ring haemorrhages occur frequently in the white matter of the cerebral hemispheres, and extend to the grey–white border. They are rare in cortical and subcortical grey matter, but occur throughout the cerebellum and brainstem (White et al., 2001). Diffuse axonal injury follows the same distribution (Dorovini-Zis et al., 2011).

In general, ring haemorrhages in paediatric cerebral malaria consist of extravasated uninfected erythrocytes; each haemorrhage is centred on a distended capillary containing infected erythrocytes and commonly a microthrombus (Dorovini-Zis et al., 2011).

**Brain swelling and vessel leakage**

Studies of paediatric cerebral malaria where retinopathy status was either unknown or a combination of retinopathy-positive and retinopathy-negative cases, have found raised opening pressure at lumbar puncture (Newton et al., 1991), clinical signs consistent with brain herniation (12/12 fatal cases, 17/49 survivors) (Newton et al., 1994), papilloedema (Beare et al., 2004), and enlarged optic nerve sheath diameter (Beare et al., 2012).

Raised brain weight and extravasation of fibrinogen from microvessels occurs in histopathologically confirmed paediatric cerebral malaria, although these do not distinguish cerebral malaria from fatal coma of other cause. Leakage is most often associated with vascular pathology such as haemorrhage, but fibrinogen is also seen in cerebral grey matter where haemorrhages are absent (Dorovini-Zis et al., 2011). Focal loss of blood–brain barrier cell junction proteins (ZO1, occludin, vinculin) occurs adjacent to sequestration. The distribution of fibrinogen, IgG and C5b-9 in these cases did not indicate widespread leakage into brain tissue (Brown et al., 2001).

**Brain imaging**

Consistent with reports of raised intracranial pressure in clinically defined paediatric cerebral malaria, MRI in retinopathy-positive paediatric cerebral malaria reveals moderate to severe brain swelling in 47% (57/120) of cases. This is significantly more common than in paediatric cerebral malaria without retinopathy [7/32 cases, OR 3.2, 95% CI 1.3–8] (Potchen et al., 2012).

Discrete lesions are seen in basal ganglia, thalamus, corpus callosum, and cerebral grey and white matter. All are significantly more frequent in retinopathy-positive cerebral malaria than retinopathy-negative cases. Cortical abnormalities can be predominantly frontal or posterior. Lesions do not respect arterial watersheds (Potchen et al., 2012), but may reflect territories of venous drainage (Meder et al., 1994; Andeweg, 1996). Distribution of lesions according to venous territories is consistent with sequestration occurring primarily on the venous side of the microvasculature.

**Manifestations of cerebral malaria in the paediatric retina**

Several clinical (Harding et al., 2006) and fluorescein angiographic features of malarial retinopathy (Beare et al., 2009) are illustrated in Fig. 1. Standard images of all features are in Harding et al., (2006).

**Sequestration in retinal vessels**

Sequestration is distributed unevenly in capillaries and the margins of larger retinal vessels (Lewallen et al., 2000). Sequestration and vessel discoloration tend to occur at vascular branch points (Lewallen et al., 2000), where greater turbulence exists to disrupt laminar flow (Nagaoka and Yoshida, 2006). It has been proposed that de-haemoglobinization of parasitized erythrocytes is responsible for the clinically observed white or orange discolouration of retinal vessels seen in paediatric cerebral malaria (Lewallen et al., 2000). Retinal vessels, particularly venules, can have a mottled appearance on fluorescein angiogram, which may represent sequestration (Beare et al., 2009). The discolouration and mottled appearance of retinal vessels in malarial retinopathy appears to be pathognomonic for paediatric severe malaria, but is reported in severe malarial anaemia as well as cerebral malaria (Beare et al., 2004).

**Retinal vessel leakage**

Fluorescein leakage is seen in ~40% of cases with malarial retinopathy and indicates blood–retinal barrier disruption. It does not co-localize with vessel mottling (Beare et al., 2009). White et al. (2009) reported leakage of fibrinogen from retinal vessels in 31% (11/35) of cases with cerebral malaria versus 7% (2/29) coma of other cause. Leakage was not reported in terms of vessel type, but fluorescein angiogram imaging suggests that leaking is predominantly from venules (Beare et al., 2009) (Fig. 1). Again, this is consistent with cerebral malaria pathogenesis centring on capillaries and post-capillary venules.

**Retinal whitening**

Retinal whitening appears as discrete areas of pale discolouration of the retina (Fig 1A), which correspond to areas of capillary non-perfusion seen on fluorescein angiogram. Retinal whitening and capillary non-perfusion seem to occur earliest and most severely in watershed regions such as the margin of the foveal avascular zone, horizontal raphe and retinal peripheries (Beare et al., 2009). In contrast to the scattered geometric lesions seen in the macula, whitening and capillary non-perfusion in the retinal periphery can occur in large tide-mark distributions that cut across arterioles and venules of varying sizes (Beare et al., 2009) (Fig. 1B).

Whitening is likely to be caused by oncost cell swelling in response to reduced perfusion (Beare et al., 2009). Other possible mechanisms include metabolic steal by large numbers of parasites (Hero et al., 1997), and occlusion caused by microthrombi (White et al., 2009). These theories are not mutually exclusive.
Areas of pale retinal discolouration are seen in other retinal conditions including arterial occlusion, venous occlusion (Browning, 2004), and Purtscher’s retinopathy (Agrawal and McKibbin, 2006). Mild whitening has also been reported in paediatric severe malaria without coma (Beare et al., 2004; Burton et al., 2004). The severity and pattern of whitening seen in malarial retinopathy—around the fovea and extending into the horizontal raphe—appears to be specific to cerebral malaria. Whitening in retinal vein occlusion is associated with swelling of the inner nuclear and outer plexiform layers of the retina (Sarda et al., 2011). In paediatric cerebral malaria there is a significant inverse correlation between electroretinographic cone b-wave amplitude and severity of retinal whitening (Lochhead et al., 2010), indicating dysfunction of bipolar cells in the inner nuclear layer. The inner

Figure 1 The features of paediatric malarial retinopathy are: retinal haemorrhages (often white-centred), retinal whitening, and orange or white discolouration of vessels. Papilloedema is often seen but is not specific for cerebral malaria. Angiographic signs include capillary non-perfusion, vessel mottling, and leakage. (A) Colour retinal image showing white-centred haemorrhages and retinal whitening extending from the macula into the temporal periphery (horizontal raphe). (B) Fluorescein angiography shows severe capillary non-perfusion in the retinal periphery (marked in yellow). Capillary non-perfusion typically coincides with retinal whitening. (C) Leakage of fluorescein from retinal venules. (D) Vessel mottling can be seen on a magnified fluorescein angiogram image. Images are from different subjects.
nuclear and outer plexiform layers are supplied by the deep capillary plexus, which forms the superficial half of a watershed with the underlying choriocapillaris (McLeod, 2010). This suggests that patterns of retinal whitening in cerebral malaria may correspond to venous congestion in areas with limited collateral drainage as well as high metabolic demand.

Retinal haemorrhages

The haemorrhages of malarial retinopathy are often white-centred. Retinal vessel thrombi are more common in fatal paediatric cerebral malaria than in coma of other cause. Microthrombi are sometimes associated with haemorrhage, but often occur independently (White et al., 2009). White-centred haemorrhages also occur in bacterial endocarditis, leukaemia, and a range of other conditions in which capillary fragility and elevated venous pressure seem to be common factors (Duane et al., 1980; Ling and James, 1998; Zehetner and Bechrakis, 2011). Retinal haemorrhages have also been observed in children with severe malarial anaemia without profound coma, and severity seems to increase with decreasing consciousness (Beare et al., 2004). In paediatric cerebral malaria, the number of retinal haemorrhages at autopsy correlates significantly with the number of brain haemorrhages (White et al., 2001). Retinal haemorrhages usually involve the inner retinal layers, but can extend to all layers. Different locations within the retina give rise to the appearance of blot or flame haemorrhages. Subretinal haemorrhage with secondary retinal detachment is seen with unusually large retinal haemorrhages (White et al., 2009).

Manifestations of cerebral malaria: paediatric retina and brain compared

Several similarities exist between paediatric retina and brain vascular pathology (Table 1).

Microvascular sequestration is a defining histopathological feature of paediatric cerebral malaria in both retina and brain, and is thought to be the principal cause of tissue injury in both sites. Whenever brain and retinal histology has been compared in cases of fatal paediatric cerebral malaria, brain sequestration is always associated with retinal sequestration (Lewallen et al., 2000; White et al., 2009). On the other hand retinal vessel discoloration is not seen in fatal coma of other cause, implying absence of retinal sequestration in these cases (White et al., 2009). Sequestration appears to be patchy within microvascular networks of each organ, but is more widespread in brain than retina.

Whereas the density of sequestration in paediatric cerebral malaria appears to be roughly equal between cerebral white and grey matter, cerebellum and brainstem (Armah et al., 2005), the distribution and density of sequestration in the eye as a whole has not yet been formally evaluated. Pilot data from a small number of eyes from children with cerebral malaria suggest that both the percentage of parasitized vessels and the intensity of sequestration are higher in the retina than in the adjacent choroid (Hiscott and Barrera, unpublished observations).

The intensity of tissue specific sequestration within the eye might be explained by the distribution of endothelial receptors in different vascular beds. ICAM1 is constitutively expressed by retinal vascular and choroidal endothelium at low levels (Duguid et al., 1992), and in the choriocapillaris expression is greatest at the macula (Mullins et al., 2006). Expression of retinal endothelial ICAM1 increases in infectious (Toxoplasma gondii) (Smith et al., 2007) and non-communicable diseases (Funatsu et al., 2005), and in response to vascular endothelial growth factor (Lu et al., 1999).

Physiological differences between retinal and choroidal vascular beds may also influence sequestration, as they differ significantly in terms of capillary width, blood flow volume and oxygen extraction. In isolated rat microvessels sequestration density is inversely related to venule diameter, suggesting flow velocity and shear rate may be important (Kaul et al., 1991). Several authors suggest that microvascular architecture may contribute to differential sequestration rates in various organs (Spitz, 1946; Nagatake et al., 1992; Sein et al., 1993).

White-centred and ring haemorrhages are common in the retina and brain, respectively. Presumably ring haemorrhages are spherical before histopathological sectioning, which like retinal imaging, provides a 2D view of the observed tissue. The haemorrhages of paediatric cerebral malaria generally affect inner retinal layers and cerebral white matter. In both sites long vessel segments are present, and these may be important in determining the localization of haemorrhages (Spitz, 1946). Clinically, haemorrhages in malarial retinopathy may occur without white centres (i.e. blot haemorrhages), or may develop white centres over time. At histopathology retinal haemorrhages often appear similar to ring haemorrhages—centred on a small thrombosed vessel with a halo of non-parasitized erythrocytes (White et al., 2009). Variations in appearance result from the histological section and differences between retinal and cerebral cellular architecture.

Some areas of retina and brain appear to be affected more frequently by perfusion abnormalities than others. In the retina, fluorescein angiogram imaging suggests that watershed regions such as the horizontal raphe and margin of the foveal avascular zone are especially susceptible to capillary non-perfusion (Beare et al., 2009). In the brain, patterns of T2 and diffusion-weighted imaging signal changes on MRI (Potchen et al., 2012) could represent boundaries of venous territories. Analysis of venous watershed regions in retina and brain may identify vessel properties that are important for the microvascular pathogenesis of cerebral malaria.

Haemorheology and neurovascular manifestations of cerebral malaria

Blood flow characteristics such as viscosity, haematocrit, and shear stress are relevant to paediatric cerebral malaria because they are likely to influence both the delivery of parasitized erythrocytes to...
organ regions and the propensity for adherence to the endothelium and other erythrocytes. Shear stress strongly influences endothelial binding, for both leucocytes (Xu et al., 2003; Crane and Liversidge, 2008) and parasitized erythrocytes (Fedosov et al., 2011b), and is related to the deformability of parasitized and uninfected erythrocytes in severe adult falciparum malaria. Admission erythrocyte deformability is reduced in adult severe malaria compared with uncomplicated cases, and is associated with mortality (Dondorp et al., 1997). Platelet mediated auto-agglutination (clumping) of erythrocytes is also associated with severity of malaria in adults (Pain et al., 2001; Chotivanich et al., 2004).

The movement of blood in small vessels is complex and depends on the character of blood and the vascular network it flows through. Unlike water (a Newtonian fluid), blood is a suspension of cells in plasma, and is an example of a shear thinning non-Newtonian fluid. The movement of blood in small vessels therefore varies with viscosity, haematocrit, blood cell deformability, aggregation and interaction with the endothelium (Schmid-Schönbein, 1999; Baskurt and Meiselman, 2003; Lipowsky, 2005; Popel and Johnson, 2005).

Blood viscosity decreases with increasing shear rate (Fig. 2), and as shear rate is related to blood velocity and vessel width, blood moves more easily at greater velocities and in vessels of lower calibre (Baskurt and Meiselman, 2003). Erythrocyte aggregation and deformation are major determinants of shear thinning (Popel and Johnson, 2005), and both are altered in P. falciparum infection (Dondorp et al., 1997; Pain et al., 2001; Chotivanich et al., 2004; Fedosov et al., 2011b). Erythrocyte stiffness is particularly important under high shear conditions, whereas non-streamlined aggregates increase resistance at low shear rates that are insufficient to break bonds between erythrocytes (Baskurt and Meiselman, 2003). Consistent with this, experimentally induced rosetting of P. falciparum infected erythrocytes is seen in venules, but not arterioles where shear rates are likely to be higher (Kaul et al., 1991). Besides shear rate, viscosity depends on the volume fraction of erythrocytes in plasma (i.e. haematocrit). Rising haematocrit is associated with an exponential increase in viscosity.

**Variable viscosity**

Under normal conditions blood viscosity decreases as vessel width reduces. This phenomenon is known as the Fähræus-Lindqvist effect, and is thought to result from migration of erythrocytes away from the vessel wall and into a central column—reducing resistance to flow by creating a lubricating cell-depleted layer next to the endothelium. Apparent viscosity reaches a minimum at an internal diameter of 5–7 μm (close to erythrocyte dimensions: ~6–8 μm diameter, 2 μm thick) after which it rises steeply (Popel and Johnson, 2005). Apparent viscosity is greater in vivo than in equivalent glass tubes of the same internal diameter, and this is thought to result from resistance to flow produced by an endothelial lining (the endothelial surface layer or glycocalyx) (Pries et al., 2000; Popel and Johnson, 2005). Protrusion of endothelial cell pseudopods into the vessel lumen during endothelial activation can almost double capillary resistance to flow (Schmid-Schönbein, 1999), and sequestered erythrocytes, or associated inflammation, may produce a similar effect.

Erythrocytes are by far the most common suspended component of blood, and so under physiological conditions, local haematocrit and shear rate are the major determinants of apparent viscosity in microvessels (Lipowsky, 2005). Leucocytes make up a relatively small fraction of total blood volume (~1/600). Nonetheless, as a typical inactivated neutrophil is ~8 μm wide, temporary obstruction of capillaries (internal diameter 4–8 μm) is common, and leucocytes can increase resistance to blood flow within an organ even without binding to endothelium, depending on leucocyte count, haematocrit and the capillary length of the organ involved. The effect is greater in organs with long capillary segments than in those with short segments such as the pulmonary circulation, and probably results from reductions in capillary erythrocyte velocity to match that of the slower leucocytes (Schmid-Schönbein, 1999). Leucocytes, which are relatively large and stiff, also strip the endothelial surface layer from capillary endothelium as they pass. This effect may persist into post-capillary venules and result in increased exposure of endothelial receptors such as ICAM1 (Popel and Johnson, 2005), presumably
facilitating endothelial binding and subsequent migration through the vessel wall. Parasitized erythrocytes are similar to leucocytes in that they are relatively stiff and bind to ICAM1 (Fedosov, et al., 2011a; Moxon et al., 2011), and this may contribute to microvascular congestion, whereas exposure of capillary and post-capillary endothelial receptors as a result of stripping of the endothelial surface layer may be an important step in sequestration. Post-capillary venules may favour sequestration because shear stress is lower than in arterioles (Nagaoka and Yoshida, 2006). Microvascular resistance is likely to be raised by high proportions of inflexible erythrocytes, sequestration and auto-agglutination, leading to reduced velocity, increased viscosity, and lower shear stress. Increased viscosity can have a dramatic effect on the retinal circulation, and ultimately lead to venous stasis or occlusion (Pournaras et al., 2008). Congestion and blockage of venules and capillaries secondary to high blood viscosity is therefore consistent with clinical signs of malarial retinopathy such as haemorrhage and capillary non-perfusion.

Variable haematocrit

Local microvascular haematocrit is not the same as systemic haematocrit. The haematocrit of blood flowing into a small tube is significantly less than that measured in the static effluent exiting the tube. This is known as the Fåhraeus effect, and—as with the Fåhraeus–Lindqvist effect—is thought to result from the tendency of flowing erythrocytes to migrate axially to form a central column (Lipowsky, 2005; Popel and Johnson, 2005).

Microvessel haematocrit is not distributed evenly between vessels. When blood meets a bifurcation there is unequal division of erythrocytes into the daughter vessels. Distribution depends on parent vessel haematocrit, branching angle and daughter vessel flow rates. Flow rate is especially influential. This effect is known as phase separation and, again, is thought to result from the axial position of erythrocytes (Fig. 2). As a result microvessel haematocrit is heterogeneous across a network (Popel and Johnson, 2005; Hirsch et al., 2012), and can show great variation (Ganesan et al., 2010; Guibert et al., 2010). As haematocrit is a major determinant of blood viscosity, the concept of phase separation and heterogeneous haematocrit within microvascular networks is crucial to understanding the possible role of haemorheological factors in the retinal and cerebral manifestations of retinopathy-positive paediatric cerebral malaria.

Computational models of blood flow

The close relationship between haemorheology and microvascular architecture means that computational models of flow parameters can be extended from single vessels to entire networks. Models exist for mouse retina (Ganesan et al., 2010) and primate cerebral cortex (Guibert et al., 2010). The mouse model predicts high regional haematocrit in the retinal periphery, but varying up to a factor of four, likely owing to phase separation. Predicted viscosity is greatest in capillaries and peri-capillary vessels, but can also vary significantly (by a factor of three) owing to heterogeneous local haematocrit. Shear stress is lower in venules than arterioles of the same size, and again reduces towards the peripheral retina for vessels of a given width (Ganesan et al., 2010). Distribution of more erythrocytes to the retinal periphery than to the posterior pole may be important physiologically, because oxygen delivery depends on blood viscosity as well as on haematocrit (Cho and Cho, 2011). It may be that the macula benefits from greater oxygen transfer as a result of lower regional viscosity compared to the retinal periphery.

Estimations from the mouse are unlikely to correspond exactly to the human retina. However, the haemorheological principles behind the model may help to explain some features of malarial retinopathy. For example the peripheral retina may develop large zones of capillary non-perfusion that cut across arterioles and venules, while macular capillary non-perfusion tends to affect smaller patches of capillaries (Beare et al., 2009) (Fig. 1B). This may be because greater parasite delivery to the periphery promotes occlusion of wider vessels, compared to the macula, which has higher metabolic demands but lower microvascular haematocrit.

The relationship between flow and microvascular networks in paediatric cerebral malaria

In summary, haemorheological factors may both influence and be influenced by P. falciparum. For example, organ regions with physiologically high microvascular viscosity and low shear stress may be especially susceptible to sequestration, while P. falciparum-associated erythrocyte stiffness, auto-agglutination and sequestration are likely to increase viscosity. As well as factors arising from blood itself, predisposition of microvascular regions to high or low haematocrit and viscosity depends on vessel network architecture. The combination of physiological heterogeneity within microvascular networks, which is influenced by network architecture, and pathological derangements of blood movement caused by P. falciparum, may help to explain manifestations of cerebral malaria in retina and brain. If so, common neurovascular network architecture could contribute to a biologically plausible rationale for inferring unseen cerebrovascular pathogenesis from the visible retina. Therefore we now compare retinal and cerebral microvasculature geometry and topology (Table 2).

The retina

Retinal microvasculature

The retina has a dual blood supply. The inner retinal circulation supplies the visible inner surface of the retina. The choroidal circulation, lying between the retinal pigment epithelium and sclera, supplies the outer retina, including the photoreceptors (Hayreh, 2010a; McLeod, 2010). Retinal and choroidal circulations have major differences in anatomy and physiology. The choroid is made up of an outer layer of large vessels (Haller’s layer), a middle layer of smaller vessels (Satler’s layer), and an innermost layer of capillaries (the choriocapillaris). The range of capillary width in the choriocapillaris is large compared to inner retina (3–50 µm versus 3.5–6 µm) (Anand-Apte and Hollyfield, 2010). Although anatomical studies classically suggested that the
choroid has multiple anastomoses, in vivo functional imaging reveals end-arterial segmental perfusion with associated watershed regions (Hayreh, 1990, 2010b). In the macaque, choroidal blood flow volume is roughly 20 times greater than in the retina, and nine times greater than cerebral grey matter, per weight of tissue (Alm and Bill, 1973; Hayreh 2010b). Wide capillaries and high volumetric flow rate may help to explain why the choriocapillaris is less susceptible to sequestration than inner retinal vessels (Hiscott and Barrera, unpublished observations).

### Geometry of the inner retinal circulation

The central retinal artery usually divides to produce four branches that extend from the optic disc into the four quadrants of the

<table>
<thead>
<tr>
<th>Area of comparison</th>
<th>Similarities / differences</th>
<th>Discussion</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vascular geometry</td>
<td>First and second generation retinal arterioles are (~100-\mu m) wide (Nagaoka and Yoshida, 2006), deep white matter arterioles are (100-170-\mu m) wide (Nonaka et al., 2003b), arterioles in the putamen are (~100-150-\mu m) wide (Nonaka et al., 1998). Retinal perivascular capillaries are (~5.4-\mu m) wide (Wang et al., 2011), cerebral grey matter capillaries are (~6.5-\mu m) wide (Lauwers et al., 2008), capillaries in the putamen (~5-7-\mu m) wide (Wolfram-Gabel and Maillot, 1994). The largest retinal venules are (130-\mu m) to (150-\mu m) wide (Nagaoka and Yoshida, 2006), cerebral grey and white matter venules range up to (125-\mu m) (Duvernoy et al., 1981). Retina (Pournaras et al., 2008), cerebral grey (Cassot et al., 2010) and white matter (Figs 7 and 9 in Nonaka et al., 2003b) all have (~90) branches from relatively long straight trunks. Caudate and putamen have retrograde arteriolar branching. Basal ganglia venous branches join at right angles (Nonaka et al., 1998).</td>
<td></td>
</tr>
<tr>
<td>Vascular topology</td>
<td>Strahler order in the macula is (~3.5), in cerebral grey matter it is (3) to (5) (Cassot et al., 2010; Yu et al., 2010). Capillary density immediately around the human foveal avascular zone is similar to primate cortex (Tam et al., 2010).</td>
<td></td>
</tr>
<tr>
<td>Watershed regions</td>
<td>Both brain and retina have arterial and venous watershed regions. Insufficient venous outflow can cause oedema, haemorrhage, and ischaemia in brain (Teksam et al., 2008) and retina (Browning, 2004).</td>
<td></td>
</tr>
<tr>
<td>Metabolic demand</td>
<td>Retinal metabolism is greatest around the fovea and in retinal layers rich in synapses (Yu and Cringle, 2001; Birol et al., 2007). Cerebral metabolism is greatest in grey matter than white matter (Sokoloff, 2003).</td>
<td></td>
</tr>
<tr>
<td>Blood flow</td>
<td>Both retina and brain receive high blood flow volume per unit tissue. Inner retinal blood flow volume is roughly half that of the adult brain (25-50 ml/100 g/min, inner retinal circulation to total brain) (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003; Pournaras et al., 2008). Cerebral blood flow is much higher in early childhood compared with adulthood (130 ml/100 g/min, age 2 to 4 years) (Wintermark et al., 2004).</td>
<td></td>
</tr>
</tbody>
</table>

It is not clear if retinal metabolic demand changes significantly after birth. Paediatric peak systolic cerebral blood flow velocity is \(~95\) cm/s in the middle cerebral artery and \(~4.5\) cm/s in the central retinal artery (Geeraerts et al., 2005).

### Table 2 Comparing vascular features between retina and brain that are likely to be important in cerebral malaria pathogenesis

<table>
<thead>
<tr>
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<th>Similarities / differences</th>
</tr>
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<tbody>
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</tr>
</tbody>
</table>
Further branching is either at right angles to the main trunk or dichotomous (i.e. two daughter branches at approximately right angles to each other) (Berntson, 1995; Pournaras et al., 2008).

In healthy adults the central retinal artery is ~160 μm wide (Dorner et al., 2002). First and second generation arterioles are ~100 μm wide, whereas first and second-generation venules are ~150 and 130 μm, respectively (Nagaoka and Yoshida, 2006). Retinal arteriolar cross sectional profiles tend to be circular, whereas venule lumens may be circular but tend to collapse (Feke et al., 1989).

Vascular segments extend close to the anterior limit of the retina, leaving a peripheral avascular zone ~1.5 mm wide. Each terminal arteriole gives rise to a network of 10 to 20 interconnected capillaries (Hayreh, 2010a). Capillary arrangement varies between retinal locations. In general there are two layers: a superficial plexus between the nerve fibre layer and ganglion cell layer, and a deep plexus between the inner nuclear and outer plexiform layers (McLeod, 2010). Only one layer exists adjacent to the fovea and at the far periphery. The peripheral network is also relatively sparse. In the peripapillary region a third capillary layer extends radially from the optic disc for a distance of up to several millimetres (Hayreh, 2010a).

Capillaries are absent at the foveal avascular zone, and adjacent to retinal arterioles (Fig. 3). The foveal avascular zone is supplied by diffusion from the underlying choriocapillaris and in adults is ~400 μm wide and 350 μm high (Yu et al., 2010). Periarteriolar capillary free zones are between 50 and 120 μm wide (Kuwabara and Cogan, 1960). They reflect the combined radius of oxygen diffusion from both artery segment and adjacent capillaries. Capillary-free zones are narrower next to venules, reflecting the high oxygen extraction (~50%) of the retinal circulation and reducing oxygen diffusion radius from post-capillary venules (McLeod, 2010).

Draining venules rise obliquely from the capillary plexuses to the nerve fibre layer and combine to form vascular patterns similar to retinal arterioles. Venules are generally slightly wider than arterioles, with shorter distances between bifurcations, narrower branching angles and less tortuosity (Hughes et al., 2009). Postcapillary venules interdigitate with precapillary arterioles in an alternating pattern (Bek and Jensen, 1993); this can become strikingly apparent in malarial retinopathy when leakage affects venules more than arterioles (Beare et al., 2009) (Fig. 1C). Venules follow arterioles to converge at the optic disc, where they drain into the central retinal vein. Venous blood from the inner retina then flows into the cavernous sinus, either directly or by way of the superior ophthalmic vein. There are no valves (Hayreh, 2010a; Semmer et al., 2010).

Some information exists about vessel geometry in children (Table 3). Paediatric central retinal artery equivalent and central retinal vein equivalent are calculated values based on the widths of first generation retinal vessels, and seem to be similar to adult values. Arteriolar bifurcation angle (the angle subtended between two branches from a parent arteriole) seems to be greater in children than adults, as does arteriolar length to diameter ratio. It is not clear how these differences might affect sequestration.

**The macula**

The macula is a unique region within the CNS situated temporally to the optic disc and specialized for fine resolution colour vision.
### Table 3  Retinal vessel geometry in children and adults

<table>
<thead>
<tr>
<th>Measurement</th>
<th>Children</th>
<th>Adults</th>
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</thead>
<tbody>
<tr>
<td>Central retinal artery equivalent</td>
<td>7–9 y</td>
<td>43–86 y</td>
</tr>
<tr>
<td>Age</td>
<td>670</td>
<td>4231</td>
</tr>
<tr>
<td>Value (μm; mean (95% CI))</td>
<td>156.4 (155.4–157.3)</td>
<td>165.29 (15.42)</td>
</tr>
<tr>
<td>Reference</td>
<td>Cheung et al., 2007</td>
<td>Lee et al., 2004</td>
</tr>
<tr>
<td>Central retinal vein equivalent</td>
<td>9 y</td>
<td>43–86 y</td>
</tr>
<tr>
<td>Age</td>
<td>266</td>
<td>4231</td>
</tr>
<tr>
<td>Value (μm; mean (95% CI))</td>
<td>168.41 (14.82)</td>
<td>165.29 (15.42)</td>
</tr>
<tr>
<td>Reference</td>
<td>Sun et al., 2009</td>
<td>Cheung et al., 2007</td>
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<tr>
<td>0.5–1.0 DD from optic disc margin</td>
<td>4–6 y</td>
<td>43–86 y</td>
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<tr>
<td>Age</td>
<td>385</td>
<td>4231</td>
</tr>
<tr>
<td>Value (μm; mean of groups)</td>
<td>159.08 (14.0)</td>
<td>165.29 (15.42)</td>
</tr>
<tr>
<td>Reference</td>
<td>Li et al., 2011</td>
<td>Cheung et al., 2007</td>
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<tr>
<td>0.5–1.0 DD from optic disc margin</td>
<td>6 y</td>
<td>43–86 y</td>
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<tr>
<td>Age</td>
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<td>4231</td>
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<tr>
<td>Value (μm; mean of groups)</td>
<td>163.2 (14.0)</td>
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<td>Central retinal vein equivalent</td>
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<tr>
<td>Age</td>
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<td>4231</td>
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<tr>
<td>Value (μm; mean of groups)</td>
<td>163.3–166.9</td>
<td>165.29 (15.42)</td>
</tr>
<tr>
<td>Reference</td>
<td>Taylor et al., 2007</td>
<td>Cheung et al., 2007</td>
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<tr>
<td>Central retinal vein equivalent</td>
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<td>43–86 y</td>
</tr>
<tr>
<td>Age</td>
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<td>4231</td>
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<tr>
<td>Value (μm; mean of groups)</td>
<td>228.8–234.6</td>
<td>242.08 (22.86)</td>
</tr>
<tr>
<td>Reference</td>
<td>Taylor et al., 2007</td>
<td>Cheung et al., 2007</td>
</tr>
<tr>
<td>Arteriolar bifurcation angle</td>
<td>12 y</td>
<td>45–75 y</td>
</tr>
<tr>
<td>Age</td>
<td>263</td>
<td>167</td>
</tr>
<tr>
<td>Value (mean of groups)</td>
<td>78.63</td>
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</tr>
<tr>
<td>Reference</td>
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<td>Arteriolar simple tortuosity</td>
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<tr>
<td>Age</td>
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<tr>
<td>Value (mean of groups)</td>
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<tr>
<td>Reference</td>
<td>Tapp et al., 2007</td>
<td>Hughes et al., 2009</td>
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<tr>
<td>Arteriolar length to diameter ratio</td>
<td>12 y</td>
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<tr>
<td>Age</td>
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<tr>
<td>Value (mean of groups)</td>
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<td>167</td>
</tr>
<tr>
<td>Reference</td>
<td>Tapp et al., 2007</td>
<td>Hughes et al., 2009</td>
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</table>

**Topology of inner retinal vessels**

The retinal periphery

At the far periphery arterioles form looping arcades with adjacent veins, which may also be traversed by bridging vessels. Using the generation number and Strahler taxonomy schemes, the average macular branching generation for both arterioles and venules is ~11.5, and average Strahler vessel order is ~3.5. This indicates a high number of bifurcations from each arteriole and vein within a relatively short vessel segment length (Yu et al., 2010). Strahler order in the human cerebral grey matter is 3 to 5 (Cassot et al., 2010). Mean perifoveal capillary width has been measured at 5.4 μm (Spitznas and Bornfeld, 1977); this average macular branching generation is indeed consistent with significant phase separation in the far periphery. Mean perifoveal capillary width has been measured in vivo using 54 fs and 2 ps optical coherence tomography (Wang et al., 2011). These capillaries are slightly narrower than capillaries in post-mortem sections of human temporal cortex.
are several watershed zones in the inner retinal circulation (Fig. 3). These exist between:

(i) Terminal vessels of each of the four arcades, most notably the superior and inferior temporal branches where they form the horizontal raphe;
(ii) Vessels of the retina and vessels of the ciliary body at the anterior limit of the retina; and
(iii) The deep capillary plexus and choriocapillaris throughout the retina, because there is no communication between the inner retina and the choriocapillaris. As the foveal avascular zone is supplied solely by the choriocapillaris this watershed is demarcated clinically in central retinal artery occlusion as the edge of the classic ‘cherry red spot’.

It is important to note that arterial and venous watersheds in the retina are identical, because of pairing of arterioles and venules. This is not the case in the brain, where arterial and venous territories are different.

**Retinal metabolism and blood flow**

The retina is considered by many to be the most metabolically active tissue in the body (Wong-Riley, 2010; Kur et al., 2012). High demands may put it at greater risk of ischaemia in cerebral malaria. Perifoveal oxygen consumption at the dark-adapted macaque is 4.9 ml/100 g/min (Birol et al., 2007). Retinal oxygen consumption in man is estimated at 9.7 ml O2/100 ml retina/min (Anderson and Saltzman, 1964) or 9 ml O2/100 g retina/min (assuming retinal specific gravity of 1.0425) (Stefánsson et al., 1987), compared with ~3 ml/100 g/min for brain (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003). In the macaque, retinal glucose consumption is among the highest in the CNS (Sperber and Bill, 1985). Estimates of energy expenditure based on ATP used per neuron suggest the human retina may consume ~11.75 mW/g, and the human brain ~10.5 mW/g (assuming retinal weight = 0.4 g, retinal energy = 4.7 mW; brain weight = 1400 g, brain energy = 14.6 W) (Sarpeshkar, 2010).

Only a small proportion of blood from the ophthalmic artery goes to the inner retinal circulation. The value often reported is ~4% (Williamson and Harris, 1994; Pournaras et al., 2008). It is not clear if this figure is from human or animal studies, but for comparison Alm and Bill (1973) found that retinal flow in the macaque makes up ~3.3% of blood to the retina and uvea combined (~27/811 ml/min). Consequently, inner retinal blood flow volume is a small fraction of ocular flow, which is itself a small fraction of blood flow in the internal carotid artery.

In non-human mammals, the retinal layers with highest metabolic rate are the inner segments of the photoreceptors, the outer plexiform layer, and the deeper region of the inner plexiform layer (Yu and Cringle, 2001). Oxygen reaches these layers by diffusion from the choriocapillaris (photoreceptor inner segments), and deep capillary plexus (outer plexiform and deep inner plexiform layers) (McLeod, 2010). In the macaque, demand for oxygen is highest at the perifovea (Birol et al., 2007), and is likely to decrease towards the periphery where photoreceptor density is substantially lower (Jonas et al., 1992). Patterns of metabolic demand may influence the susceptibility of neuronal tissue to the effects of sequestration, and resulting manifestations of malarial retinopathy. Retinal whitening is common in the highly metabolically active perifovea.

Measurements of human mean blood flow volume in the inner retinal circulation have been made using laser Doppler flowmetry, and vary between studies from ~30 to 80 µl/min/retina (reviewed in Pournaras et al., 2008), which is ~9 to 25 µl/100 mg/min, assuming a human retinal weight of 326 mg (Feke et al., 1989). The wide range may result from methodological differences. For comparison, inner retinal blood flow in the macaque, measured by radioactive microspheres, is ~1.8 µl/100 mg/min (assuming specific gravity of blood = 1.06 and macaque retinal weight of 128 mg) (Alm and Bill, 1973; Feke et al., 1989).

In humans retinal blood flow volume is likely to be approximately half that of the brain, by tissue weight (i.e. 25:50 µl/100 g/min, inner retinal circulation to total brain) (Sokoloff, 2003; Pournaras et al., 2008).

Mean perifoveal capillary flow velocity ranges from 1.37 to 3.3 mm/s. Leucocyte movement is pulsatile, and velocity in perifoveal vessels 7–11 µm wide is ~1.4 mm/s (Pournaras et al., 2008). Rather than passing equally through all capillary segments perifoveal leucocytes seem to follow preferred pathways (Tam et al., 2011). Similar patterns may exist for the movement of parasitized erythrocytes.

Estimated shear stress in healthy adults is highest in first generation arterioles (mean 540 dyne/cm²), 20% lower in second-generation arterioles, and 50% lower in both first and second-generation venules (Nagaoka and Yoshiida, 2006; Nagaoka et al., 2009).

**The brain**

The circulation of the brain is substantially more complex than that of the retina. We give a brief overview before focusing in more detail on three brain areas typically involved in acute paediatric retinopathy-positive cerebral malaria: the basal ganglia, cerebral grey matter and cerebral white matter. All are affected significantly more often in children with retinopathy-positive cerebral malaria than in paediatric patients with cerebral malaria without retinopathy (Potchen et al., 2012). Retinopathy could therefore reasonably be associated with vascular disease processes in these areas, if microvascular characteristics are similar in both organs.

**Cerebral vasculature architecture and watershed regions**

Carotid and basilar arteries supply the variably anastomotic Circle of Willis, which sends off branches including the anterior, middle and posterior cerebral arteries. Arterial territories are exclusive and vary between individuals (Miyawaki and Statland, 2003a). As in the retina, arterial territories become apparent clinically in thrombo-embolic arterial occlusion when infarction occurs up to the watershed of that arterial territory.

Unlike the retina where arterioles and venules cover identical territories, cerebral venous drainage does not parallel the arterial supply. Several classifications exist (reviewed by Nowinski, 2012), but essentially venous drainage can be understood in terms of
superficial and deep venous systems (Ono et al., 1984; Oka et al., 1985). Although venous anastomoses exist within each system, there is no physiological anastomosis between superficial and deep systems (Andeweg, 1996, 1999). They join at the torcula (confluens sinuum), where the straight sinus (draining the deep system) meets the superior sagittal sinus and transverse sinus (draining the superficial system) (Miyawaki and Statland, 2003b). Anatomy is variable but the superficial venous system includes several cortical veins and sinuses that drain the exterior cortex, ranging in average width from ~0.5 to 5.0 mm (Oka et al., 1985). The deep system drains subcortical and periventricular structures such as the insular cortex and basal ganglia. The watershed between the superficial and deep venous systems lies between the periventricular and subcortical white matter (Andeweg, 1996). Ultimately venous blood leaves the cranial vault through the internal jugular or spinal veins, and a small number of emissary veins.

Major venous channels show significant variation (Miyawaki and Statland, 2003b). In a magnetic resonance venography series of healthy children the inferior sagittal sinus was not seen in 54%. The right or left of the following were absent in many cases (% cases absent): vein of Trolard (~80%), vein of Labbe (~50%), superficial petrosal sinus (66%), and inferior petrosal sinus (~33%) (Widjaja and Griffiths, 2004). Lack of major collateral channels would make associated brain regions more vulnerable to venous congestion in paediatric cerebral malaria. Variation in venous drainage could help to explain patterns of MRI signal changes in acute retinopathy-positive paediatric cerebral malaria in the same way that venous territories within the superficial system appear to influence the distribution of radiographic lesions in paediatric cerebral venous thrombosis (Meder et al., 1994; Teksam et al., 2008).

Vascular anatomy of the basal ganglia

The basal ganglia comprise several interrelated nuclei with different neuronal structures, vascular architectures, and embryological origins. The arterial supply of the basal ganglia arises mainly from the middle cerebral artery with some contributions from the anterior cerebral artery and anterior choroidal artery. Branches from the anterior cerebral and anterior choroidal arteries supply the head of the caudate nucleus and globus pallidus (Akima, 1993). Ten to 20 lenticulostriate arteries from the middle cerebral artery enter the putamen, where they spread out in a fan shape and divide into two to three branches, each ~100 to 150 μm wide. These form capillary networks within the putamen and cross the internal capsule to supply the caudate nucleus (Nonaka et al., 1998).

The vasculature of the putamen and caudate nucleus is distinct from that of the globus pallidus, internal and external capsules. Branching arterioles (~50 μm wide) of the putamen may reflect back and coil around the parent vessel before branching further. Precapillary arterioles of the putamen are ~20 μm wide (Wolfram-Gabel and Maillo, 1994). Retrograde arteriolar branching is seen but it is not clear if this is a feature of old age or is universally present (Nonaka et al., 1998). If the latter they could produce heterogeneous viscosity from phase separation in children. In the globus pallidus, arterioles tend to be fewer and straighter, with no coiling. The end arteriolar branches (~40 μm wide) form arteriolar anastomoses as well as a capillary net (Wolfram-Gabel and Maillo, 1994).

The capillary networks of the putamen and caudate nucleus have a similar density to the cerebral grey matter (600–650 capillaries per mm² in a 30 to 40 μm mesh), whereas the capillary density of the internal capsule approximates that of the cerebral white matter. The globus pallidus has capillary density between these extremes (200–350 per mm² in a 60–80 μm mesh) (Wolfram-Gabel and Maillo, 1994; Nonaka et al., 1998). Capillary density seems to be proportional to the density of synapses (Akima, 1993). Capillaries of the putamen are ~5–7 μm wide (Wolfram-Gabel and Maillo, 1994).

Venules and veins throughout the basal ganglia are generally wider than the arterial vessels, and smaller veins meet larger vessels at right angles (Nonaka et al., 1998). In the putamen, five arterioles are typically arranged around a single central ‘principal’ vein, which together form a vascular unit. These veins can be large (~500 μm wide) (Wolfram-Gabel and Maillo, 1994). The high ratio of arterioles to veins suggests a similarity to cerebral cortical vessels, and an important difference with the retina where arterioles and venules are paired. Congestion of a principal vein is likely to have a disproportionate impact on perfusion. This may be relevant to cerebral malaria since sequestration is thought to occur in venules to a greater extent than arterioles, although it is not clear how often congestion occurs in venules up to 500 μm wide.

Vascular anatomy of the cerebral grey matter

Branches from the anterior, middle, and posterior cerebral arteries produce a network of pial arterioles on the cerebral surface. Cortical arterioles (~20 to 90 μm wide) branch from the pial vessels and travel on the brain surface for ~150 to 750 μm before turning 90° to penetrate the cortex (Reina-de La Torre et al., 1998).

There are no capillaries on the brain surface, and the absence of arterio-venous anastomoses at this point means that all arterial blood is forced into penetrating cortical arterioles and the capillaries of the cerebral cortex (Duvernay et al., 1981).

Penetrating cortical arterioles reach depths of 150 μm to ~3 mm and have widths between ~20 and 65 μm. They branch to form capillaries and pre-capillary arterioles at several levels parallel with the brain surface. Mean arteriolar branching angle is 112 to 126°, depending on daughter vessel diameter (Cassot et al., 2010). Some pre-capillary branches are recurrent (Duvernay et al., 1981; Reina-de La Torre et al., 1998).

The topology of cortical vessel segments can be thought of in terms of ‘tree-like’ penetrating vessels and ‘net-like’ capillaries (Lauwers et al., 2008). The tree-like arteriolar and venular segments have a long parent vessel ‘trunk’ from which smaller branches emanate (Cassot et al., 2010), and are suited to rapid movement of blood, whereas the capillary net is homogeneous and space-filling—suited to nutrient exchange (Lorthois and Cassot, 2010). Cortical penetrating arterioles and venules have a
Strahler branching order from 3 to 5 between surface pial vessels and the capillary network (Cassot et al., 2010). Unlike capillaries of the inner retina, it seems there are no boundaries between capillaries arising from different arterioles.

Average total vascular density in temporal cortex is ~500 mm/mm², of which capillaries contribute ~50% (Cassot et al., 2006). This is consistent with human visual cortex, which has an average capillary density of ~250 mm/mm³ (Bell and Ball, 1985). Narrow artero-venous anastomoses may exist at the capillary junction, but it is difficult to distinguish these from larger capillaries (Duvernoy et al., 1981).

Ascending cortical venules have ~50% more branches than arterioles; mean branching angle is 118° to 129° (Cassot et al., 2010). Ascending veins include principal veins (Group V5), more superficial, ranging from 65 μm wide. The diameter of venules decreases as vessel origin becomes more superficial, ranging from 65 μm (Group V4) to 20 μm (Group VI) (Duvernoy et al., 1981). Once ascending venules reach the cortical surface they make a 90° turn to converge on surface venules (~130 μm wide). The surface venous network is anastomotic with channels up to 180 μm wide, but has fewer anastomoses than the arteriolar network, and some lobules have no surface venous anastomoses. This drains into larger veins adherent to the arachnoid dura (Duvernoy et al., 1981). The largest veins (1.3 to 3.3 mm) cross the subdural space to meet sinuses incorporated into the dura mater (Oka et al., 1985). There are no venous anastomoses in the cerebral grey matter, and only rare reports of arteriovenous anastomoses (Duvernoy et al., 1981).

Occlusion of cortical vessels

The absence of anastomoses between penetrating vessels means that penetrating cortical arterioles and ascending venules represent a physiological bottleneck. Experimental occlusion of penetrating arterioles in rat cortex leads to severe reduction in blood flow velocity in vessels at least seven branches downstream from the occlusion (Nishimura et al., 2010).

In the rat, ascending venules are twice as prevalent as penetrating arterioles, and this is likely to explain the relatively low extent of flow disruption after ascending venule occlusion (Nguyen et al., 2011). In contrast human grey matter penetrating arterioles outnumber venules on average by 2:1 (Cassot et al., 2010), and principal veins are sometimes surrounded by several rings of arterioles. The tissue volume of such vascular units is variable (Duvernoy et al., 1981). The general implication is that, as in the basal ganglia, a low ratio of venules to arterioles in human cortex may mean venous congestion in cerebral malaria disrupts flow to disproportionately large areas of tissue.

Vascular anatomy of the cerebral white matter

The cerebral white matter is supplied by penetrating arterioles from the pial arteriolar network, which remain unbranched within the cerebral grey matter until its deepest layer. These arterioles enter the subcortical white matter perpendicular to the brain surface. Vessels express branches at ~90° from the main trunk—five to ten, depending on trunk length. Only a proportion of arterioles reach the periventricular zone, where they arborize. A few form arteriolar anastomoses (Nonaka et al., 2003a,b). Penetrating arterioles have an internal diameter of 30 to 40 μm in subcortical white matter, compared to 40 to 60 μm in the grey matter (Nonaka et al., 2003a), and 100 to 170 μm in deep white matter (Nonaka et al., 2003b).

There has been disagreement about whether the deep white matter also receives arterioles from ventricular sources (Van den Bergh and Vander Eecken, 1968). Moody et al. (1990), Nelson et al. (1991) and Nonaka et al. (2003b) found no evidence of periventricular arterioles directed towards the brain surface.

Cerebral white matter is drained by superficial medullary veins and deep medullary veins. The former drain subcortical white matter into superficial cerebral veins on the brain surface, and the latter drain periventricular white matter into subependymal veins at the lateral ventricle (Hassler, 1966; Meder et al., 1994; Andeweg, 1996; Nonaka et al., 2003b). Superficial medullary veins are thought to drain the external 1–2 cm of white matter (Meder et al., 1994), and the deep medullary veins the remainder (Hassler, 1966; Hooshmand et al., 1974). Trans-cerebral veins connecting superficial and deep venous systems have been reported (Huang and Wolf, 1964; Hassler, 1966; Meder et al., 1994; Nakamura et al., 1994; Schaller, 2004), but may be artefactual as functional anastomoses between superficial and deep venous systems are not consistent with patterns of injury seen after deep venous system obstruction (Hassler, 1966; Andeweg, 1999). The predominance of haemorrhages in white matter is consistent with relatively long venous drainage pathways (Spitz, 1946).

Cerebral blood flow and metabolism

Like the retina the brain is highly metabolically active (reviewed in Wong-Riley, 2010), and depends on a constant supply of oxygen and glucose (Mckenna et al., 2006).

Normal cerebral blood flow characteristics vary with age and between brain regions (Kety, 1950; Ide and Secher, 2000). Abnormal cerebral blood flow in the middle cerebral arteries has been reported in cases of paediatric cerebral malaria with seizure or raised intracranial pressure (Newton et al., 1996).

Total cerebral blood flow and metabolism changes dramatically during childhood (Kety, 1956; Kennedy and Sokoloff, 1957; Kinnala et al., 1996; Takahashi et al., 1999; Wintermark et al., 2004; Mckenna et al., 2006). Cerebral blood flow increases rapidly during the first year of life (Varela et al., 2012) and flow volume peaks at 55% cardiac output between age 2 to 4 (~130 ml/100 g/min) before declining to adult values (Wintermark et al., 2004). In comparison the brain of a resting adult receives ~16.5% cardiac output (Kety, 1950), or ~50 ml/100 g/min (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003). Cerebral metabolic rate for oxygen is also highest from 3 to 6 years, ranging from 4.3 to 6.2 ml O₂/100 g/min (Kennedy and Sokoloff, 1957). The average of 5.25 ml would be ~50% total body oxygen (Mckenna et al., 2006). Adult resting cerebral metabolic rate for O₂ is ~3 ml/100 g/min (Kety and Schmidt, 1948; Madsen et al., 1993; Sokoloff, 2003), and accounts for ~20% of total body oxygen consumption (Kety, 1950). Cerebral
arteriovenous oxygen difference is \(\sim 40\%\) in adults (Seifert and Secher, 2011), comparable to retinal oxygen extraction of \(\sim 50\%\) (McLeod, 2010). The developmental curve for cerebral metabolic rate for glucose follows a similar trajectory to cerebral blood flow, though with a more gradual decline to adult values at around age 17 (Chugani et al., 1987; Kinnala et al., 1996; Chugani, 1998).

The highest regional peak in cerebral blood flow is in the calcarine cortex (area V1, 155 ml/100 g/min, 29 months). Basal ganglia, temporal, frontal, and occipital cortices have lower peaks (\(\sim 125\) to 140 ml/100 g/min) (Wintermark et al., 2004). The highest regional peak in the cerebral metabolic rate for glucose at this age is in the calcarine cortex, transverse temporal cortex, and lenticular nuclei (all \(>30\mu\text{mol}/100\text{g/min}\) between 1 and 2 years) (Chugani et al., 1987).

**Flow in cerebral microvessels**

Data on capillary flow dynamics in human brain are scarce, although many studies on animal models exist (Hudetz et al., 1996; Vovenko, 1999; Jespersen and Ostergaard, 2011). Capillary flow velocity is between 0.6 and 2.4 mm/s in rat cortex (Hudetz et al., 1996), which overlaps the lower range of velocities in human perifoveal vessels (Pouranaras et al., 2008).

Guibert et al. (2010) have modelled blood flow in primate cortex, which has a ratio of arterioles to venules similar to human brain. They found that local haematocrit could vary by approximately a factor of two owing to phase separation. Brain haematocrit measured by dual tracer single-photon emission computed tomography in adult humans is \(\sim 76\%\) systemic haematocrit (Sakai et al., 1985). Blood is thought to make up \(\sim 3-5\%\) of brain volume, and three quarters of this is venous (An and Lin, 2002).

Blood volume is greater in grey (\(\sim 5\text{ml}/100\text{g}\)) than white matter (\(\sim 2\text{ml}/100\text{g}\)) (Kuppusamy et al., 1996) consistent with greater capillary density in grey matter.

**Retina and brain compared: implications for studying neurovascular pathogenesis**

Retina and brain microvasculature have much in common, including circulation-tissue barriers and local control over blood flow (Patton et al., 2005). After reviewing microvascular haemorheology and the microvasculature of retina and brain regions, it seems that sequestration-induced injury could be influenced by several characteristics (Table 2).

**Vessel branching**

Retinal vessels are relatively long and straight, with right-angled or dichotomous branches. This is also true of cerebral grey matter, white matter and basal ganglia—although brain-specific features are also present. Haemodynamic models suggest near 90° arteriolar branching produces significant variation in haematocrit and viscosity (Ganesan et al., 2010; Guibert et al., 2010) and therefore may be important in distributing parasitized erythrocytes and fostering sequestration. Right-angled venular confluences may also provide opportunities for sequestration since cells may continue to roll along the endothelium when moving from a smaller to larger vessel before joining the axial flow stream, potentially stripping the endothelial surface layer and exposing ICAM1 (Schmid-Schönbein, 1999; Popel and Johnson, 2005). Local variation in haematocrit and viscosity associated with branching architecture may explain why retinal vessels full of parasitized erythrocytes can be found immediately adjacent to vessels unaffected by sequestration (Lewallen et al., 2000).

**Arteriole to venule ratio**

Both basal ganglia and cerebral grey matter have more arterioles than venules, whereas in the retina, arterioles and venules are paired. A high ratio of arterioles to venules in the absence of anastomoses may make these brain regions more vulnerable than the retina to the effects of post-capillary venous congestion with parasitized erythrocytes.

**Venous drainage and watershed regions**

Retina and brain both have vascular watersheds that are amenable to imaging. Although retinal arterial and venous watersheds are identical, retinal arterioles and venules can be distinguished using fluorescein angiogram. In the brain, arterial supply and venous drainage occur over different territories, and radiological changes to these anatomical regions can be assessed using MRI. Retinal capillary non-perfusion is most common in watershed regions, and brain lesions may also follow patterns of venous drainage into superficial and deep systems, especially those of the cerebral white matter (Meder et al., 1994; Andeweg, 1996). Sequestration is thought to mainly involve capillaries and post-capillary venules, and retinal white-centred haemorrhages occur in other conditions involving venous congestion (Duane et al., 1980). These points suggest comparison of retinal and cerebral venous watersheds in cerebral malaria is reasonable, and perhaps that the severity of non-perfusion in retinal watersheds might reflect the degree of \(T_2\) or diffusion-weighted imaging abnormalities in the cerebral white matter in acute cerebral malaria (Fig. 4). The consequences of venous congestion are much greater in brain than retina. For example, swelling of the splenium, or posterior cerebrum (Potchen et al., 2012) could compress the vein of Galen against the tor-torium, potentially obstructing outflow from the deep venous system and rapidly raising intracranial pressure (Andeweg, 1999).

**Blood flow and metabolism**

Adult inner retinal blood flow is roughly half that of the brain. To our knowledge there are no data available on developmental changes to retinal blood flow during childhood. However, cerebral blood flow varies dramatically during childhood development, peaking within the age range typically admitted with paediatric cerebral malaria in high transmission areas (Snow et al., 1997; Wintermark et al., 2004). The median age at admission with cerebral malaria in high transmission areas appears to coincide with the childhood peak in total cerebral blood flow and metabolism. For example, the mean
age of children admitted to the paediatric research ward in Blantyre, Malawi, ranged from 3 to 4 years between 2001–10 (Roca-Feltrer et al., 2012). Snow et al. (1997) found the median (IQR) age at admission with cerebral malaria was low in high transmission areas and increased with falling endemicity—ranging from from 2 (1.5–6.5) years (hyperendemic), 4 (3–5) (mesoendemic), to 6.5 (4–7) years (hypoendemic). A similar pattern is found using residential altitude as a proxy for transmission intensity (Reyburn et al., 2005). As suggested by others (Billig et al., 2012) high metabolic demands may make the paediatric brain more vulnerable to the effects of sequestration than other paediatric organs, and in the context of high transmission predispose to early cerebral complications rather than the acute renal failure or pulmonary oedema seen in adults (Thanachartwet et al., 2013).

Retina and brain are comparable in that they both have very high metabolic demands, and in early childhood the brain may be more susceptible than the retina (and other organs) to the effects of dysfunctional microvascular flow. However cerebral malaria is only one manifestation of severe malaria in children, and it is not clear how high paediatric cerebral metabolic rate for $O_2$ in isolation could influence which children develop cerebral malaria, and which develop severe malarial anaemia or metabolic acidosis.

**Complicating factors**

The clinical picture of paediatric cerebral malaria is complicated by the presence of multiple interrelated insults in addition to sequestration, including anaemia, hypoglycaemia, and metabolic acidosis.

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**Figure 4** Comparison of retinal and cerebral venous watershed regions. (A) Fluorescein angiogram showing capillary non-perfusion at the macula and horizontal raphe, which is an arteriolar and venular watershed between the supero-temporal and infero-temporal arcades. Axial (B) and coronal (C) MRI images show extensive high T2 signal in the subcortical white matter of a different child with cerebral malaria (white arrows). The cerebral white matter is the site of a venous watershed between superficial and deep venous systems.
Conclusions

Description of the whole brain in terms of greater or lesser similarity to the whole retina is difficult, and probably inappropriate. Rather, analysis of data from retina and brain should take account of anatomical variation within both organs, as neither retina nor brain are homogeneous units. Both have regions where microvasculature may, or may not, be comparable in specific respects that are relevant to a given neurovascular disease.

Retina and brain are similar in ways relevant to paediatric cerebral malaria. Similar vascular pathology suggests both organs experience similar disease processes. Similarities in terms of vessel network architecture, imaging abnormalities in venous watersheds, and high metabolic demand suggest that both organs may be vulnerable to sequestration for the same reasons. These factors may influence delivery of parasitized erythrocytes, the flow conditions necessary for sequestration, and the vulnerability of tissue to ischaemia associated with venous congestion or occlusion. If so, the impact of congestion on common venous anatomy may explain the distribution of MRI brain and retinal fluorescein angiogram lesions in paediatric cerebral malaria.

The retina and brain also have differences. Arteriovenous ratios vary between brain regions, while in the retina arterioles and venules are paired. Cerebral arterial and venous watersheds are different, while retinal watersheds overlap. The paediatric brain has a metabolic peak in early childhood, which is likely to raise cerebral metabolic demands significantly above retinal demands in the age range typically affected by cerebral malaria.

Considering these similarities and differences, a biologically plausible analogy can be made between the retinal periphery, or horizontal raphe, and the cerebral white matter, because these regions have similar vascular architecture. All receive blood from long vessels with near 90° branches, and so may contain blood with disproportionately high microvascular haematocrit and viscosity. The presence of venous watersheds gives little option for collateral drainage from these areas in the event of venous congestion. Given this background, similar degrees of coexistent injury to these retinal and brain areas in a population of children with retinopathy-positive cerebral malaria would suggest that retinal capillary non-perfusion and leakage could be surrogates for brain ischaemia and swelling, and might provide a useful tool to understand cerebral vascular pathogenesis.

Our review has limitations. Only three brain regions are compared with the retina, and comparable data are not available for all geometrical or topological variables at each site. We have resorted to measurements from adults where paediatric data are not available, and we have not attempted comparison of retinal or cerebral microvascular architecture with other organs in which sequestration is minimal or absent. We have not compared paediatric and adult malarial retinopathy—this deserves a separate review.

We have identified significant similarities and differences in retinal and cerebrovascular architecture, blood flow and metabolic demand. These comparisons inform new interpretations of retinal and cerebral data in retinopathy-positive paediatric cerebral malaria by incorporating information on haemorheology and vascular architecture from multiple regions of the CNS. Increasingly comprehensive comparisons of human neurovasculature will provide greater insights into how far retinal vessel changes reflect neurovascular pathology. Mapping the contours of warranted analogy between retina and brain has the potential to strengthen the biological rationale of retinal imaging in neurology, inform interpretation of neurovascular data, and stimulate further hypotheses—for retinopathy-positive paediatric cerebral malaria in particular, and for neurovascular disease in general.

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