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Evaluation of method of preparation of passive diffusion tubes for measurement of ambient nitrogen dioxide

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This study was carried out in response to suggestions that the measurement of NO2 by Palmes-type passive diffusion tubes (PDT) is affected by the method of preparation of the triethanolamine (TEA) absorbent coating on the grids. The following combinations of factors were investigated: TEA solvent (acetone or water), volume composition of TEA in solvent (50% or 20%), and grid coating method (dipping in solution prior to assembly, or pipetting solution on after assembly). Duplicate PDTs prepared by each of the 8 methods were exposed in parallel, in urban air, for a total of 80 separate 1 week exposures. NO2 concentrations derived from PDTs prepared by pipetting methods were significantly less precise than concentrations from dipping methods, with mean RSDs for duplicate measurements of 13.8% and 8.5%, respectively (n = 316 each category). Pipetting methods using solutions of 50% TEA composition were particularly imprecise (mean RSD 17.2%). Data from PDTs prepared by pipetting methods were systematically more poorly correlated with each other and with data from co-located chemiluminescence analysers, than corresponding data from PDTs prepared by dipping methods, indicating that more consistent accuracy was also obtained by the latter PDTs. The statistical evidence suggested that PDTs prepared by pipetting 50% TEA in water generally gave lower NO2 concentrations. Although this is in agreement with a previous study, it is also possible that such an observation here may be a statistical artefact given the demonstrably poorer precision of this method. The general tendency of PDTs to show positive bias in NO2 measurement in urban air in 1 week exposures was again evident in this study (mean biases at roadside and urban centre locations of +35% (n = 475) and +18% (n = 112), respectively) consistent with augmentation of within-tube NO2 flux by chemical reaction between co-diffusing NO and O3. Overall, it is recommended that the pipetting method of PDT grid preparation is avoided, or at least investigated further, because of the apparent degradation in precision and accuracy of NO2 measurement. Potential reasons for the effect are discussed.

1. Introduction

Nitrogen dioxide (NO2) is regarded as an air pollutant because of its adverse effect on human health and its contribution to the formation of low-level ozone. Consequently, in most countries, permitted concentrations of ambient NO2 are subject to legislation. The UK, following the EU and the WHO, has an annual average limit for NO2 of 40 μg m⁻³, equivalent to 21 ppb (parts per billion by volume) at ambient air pressure and temperature. Routine monitoring of NO2 is therefore required to assess compliance with air quality standards. Such monitoring is particularly important for NO2 because the majority of NO2 in ambient air is formed from chemical oxidation of primary emissions of nitric oxide, NO. The rate of this oxidation is influenced by many factors ranging from local-scale meteorological dispersion and mixing, to regional scale transport of ozone (O3) oxidant, making it difficult to predict NO2 spatial fields.

Continuous analysers, such as the chemiluminescence analyser, are usually specified as the reference method for measurement of NO2, but passive diffusion samplers (particularly the Palmes-type passive diffusion tube,1 PDT) remain widely used for indicative assessment of spatial variations of longer-term average NO2 concentrations because of their ease of use and low cost. For example, an NO2 concentration map for the entire UK is interpolated from a national network of PDTs.2

The fundamental principle of PDT measurement is that NO2 molecules are captured by triethanolamine (TEA) absorbent, N(CH2CH2OH)3, coating the grids inside the closed end of the tube, with the rate of capture of NO2 determined by the diffusive flux along the tube. Since TEA is extremely viscous at room temperature, it is dissolved in a solvent for application to the PDT grids. After exposure, trapped NO2 is extracted into aqueous solution as nitrite ions, NO2⁻, and quantified.

While it is accepted that NO2 PDTs are not as precise as continuous analysers, considerable debate remains regarding the accuracy of NO2 PDTs in the field. This debate surrounds the significance or not of specific factors giving rise to specific biases in deriving NO2 concentrations. Some investigators have concluded that NO2 PDTs are accurate within acceptable tolerances of precision,3 whilst others have argued that specific phenomena such as wind-induced turbulence at the entrance to the tube,4 or chemical production of additional NO2 by co-diffusing NO and O3 within the tube5,6 contribute to an effective over-measurement of NO2 by PDTs, or that an exposure-duration related loss of trapped NO2 contributes to an effective under-measurement.7

More recently, two studies8,9 have suggested that the accuracy of NO2 PDT measurement may also be affected by the way in which the TEA absorbent at the end of the tube is prepared. The observation has potentially major consequences for comparing NO2 PDT data since there is no generally-recognised standard method of tube preparation. The variables in preparation are that the TEA absorbent may be dissolved in acetone or deionised water, in volume ratios ranging from 20%–50%, and the solution applied to the grids either by dipping the grids into the solution prior to PDT assembly, or by pipetting a known volume of solution onto the grids after assembly. The report9 produced by NETCEN, the UK body currently responsible for collating and validating data from the UK national network of NO2 PDTs, recommended that the
preferred preparation method was to pipette an unspecified volume of a 20% TEA:water solution onto grids already assembled within the PDT cap. The recommendation was based on the observation that this preparation method yielded NO$_2$ measurements closest in value to those of a chemiluminescence analyser, whilst PDTs prepared by dipping grids into a 50% TEA:acetone solution (probably the most widespread method), or by pipetting a 50% TEA:water solution onto assembled grids, were reported to yield NO$_2$ measurements consistently higher, or lower, respectively, than the analyser. The study was conducted in a laboratory chamber. The field investigation by Kirby et al.$^8$ also reported that tubes prepared by pipetting 50% TEA:water solution onto assembled grids yielded lower NO$_2$ measurement but that there was no difference in NO$_2$ measurement from tubes prepared using smaller volume ratios of TEA in water or using grids dipped in 50% TEA in acetone. Conversely, a recent review of routine PDT data from UK local authority networks reported no discernible effect of tube preparation method on PDT measurements.$^{10}$

One difficulty in trying to rationalise observations concerning the effect, or not, of absorbent preparation method is uncertainty regarding the actual mechanism of NO$_2$ complexation by TEA. A mechanism proposed by Glasius et al.$^1$ that yields a 1:1 ratio between NO$_2$ in air and NO$_2^-$ in solution, is shown in eqn. (1):

$$2\text{NO}_2 + \text{N}(	ext{CH}_2\text{CH}_2\text{OH})_3 + 2\text{OH}^- \rightarrow 2\text{NO}_2^- + \text{O}^- \cdot \text{N}(	ext{CH}_2\text{CH}_2\text{OH})_3 + \text{H}_2\text{O} \quad (1)$$

The required hydroxyl ions are postulated to arise from the dissociation of TEA in water molecules present in the air, so the reaction will not take place in completely dry air, in accord with observations that TEA hydration is important for quantitative NO$_2$ sampling.$^{12}$ The OH$^-$ ions may also derive from water molecules in the absorbent solution itself (if present), and it has been suggested that the extent of this process may account for the apparent lower trapping efficiency of different molar ratio TEA:water absorbent solutions.$^8$

It is not clear from the studies cited above whether the physical process itself of coating grids (i.e. dipping or pipetting) contributes to differences in NO$_2$ measurement. It is also difficult to rationalise how different preparation methodologies may influence measurement. Given the ambiguity of the above studies and the continued importance of PDTs for ambient NO$_2$ measurement, the impetus for the current work was a detailed investigation of whether, and how, absorbent preparation method affects NO$_2$ concentrations derived from PDTs. Eight different grid preparation methods were compared in simultaneous exposures in urban air. Uniquely, this study also compared a sub-set of the PDT NO$_2$ measurements with the NO$_2$ concentration predicted to be measured by a PDT when within-tube production of NO$_2$ along the diffusion path of the tube is also taken into account.$^5$ To limit the impact on data interpretation of the further confounding issue of the loss of nitrite evident during longer exposure periods,$^7$ all exposures were of 1 week duration only.

### 2. Experimental section

Standard Palms-type acrylic diffusion tubes from Gradko International were used (length 7.1 cm, internal cross-sectional area 0.92 cm$^2$, two stainless steel grids per tube). Tubes, caps and grids were cleaned thoroughly before use.

Eight different methods of applying TEA to PDT grids were compared, comprising all combinations of two choices of three two-level factors: TEA solvent (acetone or deionised water), TEA concentration (50% or 20% by volume in the solvent), and application method of solution to grid (dipping or pipetting).

<table>
<thead>
<tr>
<th>Absorbent solution composition</th>
<th>Grid preparation method</th>
</tr>
</thead>
<tbody>
<tr>
<td>50% TEA, 50% acetone</td>
<td>A</td>
</tr>
<tr>
<td>20% TEA, 80% acetone</td>
<td>B</td>
</tr>
<tr>
<td>50% TEA, 50% deionised water</td>
<td>C</td>
</tr>
<tr>
<td>20% TEA, 80% deionised water</td>
<td>D</td>
</tr>
</tbody>
</table>

The preparation methods, and their labelling nomenclature, are summarised in Table 1. For dipped method preparations, grids were immersed in the appropriate solution for approximately 5 min, laid on tissue paper, and patted gently with another piece of tissue paper to remove excess solution. Tubes were then assembled and capped. For pipetted methods, a volume of 50 µl of solution was pipetted directly onto the two grids within the capped end of a tube already assembled. Tubes were left to stand for approximately 3 min before the remaining open end was capped.

Tubes were exposed for 1 week at three sites in central Edinburgh: Princes Street gardens (PSt), Castle Street (CSt), and Haymarket (HMt). The PSt site is classified as an urban centre (city centre but not roadside), while CSt and HMt are roadside sites (1–5 m from a busy road).

PDTs were always deployed in duplicate. The eight pairs of tubes (plus a field blank of method A) were arranged in random order at each site, adjacent to the inlet of a NO$_x$ chemiluminescence analyser. The PSt site also had an O$_3$ analyser. Hourly averaged NO, NO$_2$ and O$_3$ data at this site were used as input to a numerical model$^{12,13}$ to calculate the amount of additional NO$_2$ created and trapped within the PDTs, during each exposure, by chemical reaction between NO and O$_3$ also diffusing inside the tube.

PDT exposures were carried out between November 2001 and March 2002 and again between November 2002 and March 2003, yielding data for a total of 30 1-week exposures for each preparation method at each site, except for the PSt site at which the continuous analyser ceased operation in December 2002.

After exposure, trapped nitrite in the tubes was extracted into deionised water and quantified by the standard sulfanilamide/NEDA colorimetric method (absorbance measurement at 540 nm). Independent duplicate sets of nitrite calibration standards were prepared each week and the average ambient [NO$_2$] during the exposure calculated from the NO$_2$ calibration curves using 0.154 cm$^2$ s$^{-1}$ as the diffusion coefficient of NO$_2$ in air.

### 3. Results

#### 3.1 Precision

The precision of the PDT measurements was assessed before addressing issues of PDT accuracy. Scatter plots of all the duplicate NO$_2$ measurements are shown in Figs. 1a and b, segregated into dipped and pipetted methods of preparation, respectively.

Fig. 1 clearly shows that measurement precision from tubes prepared by dipping methods A–D is considerably better than precision from tubes prepared by pipetting methods E–H. Precision is particularly poor for methods E and G (Fig. 1b) in which an absorbent solution of 50% TEA in acetone or water, respectively, is pipetted directly onto the grids after PDT assembly. The two other pipetting preparation methods F and H show better precision although instances of poor precision remain.

Table 2 summarises the precision RSD data by preparation
Method E has the worst precision, with mean RSD of 20.9%. The mean RSD of method G is also high (13.5%), although a median RSD of 8.2% indicates that the mean is adversely affected by some instances of very poor precision (visible in Fig. 1b). The three most precise methods are dipping methods. The mean and median RSD values for all dipping methods (8.5% and 6.7%, respectively) are significantly better than the corresponding values (13.8% and 9.0%) for all pipetting methods. With the exception of methods E and G, mean RSD values of 7.7–10.6% for each of the other 6 methods compare well with previously quoted mean RSD values of 8%14–10%15 and <4%6 for PDT precision.

### 3.2 Accuracy

In order to avoid any subjectivity, the mean of each duplicate pair was used as the measure of NO2 concentration for all PDT data presented in the rest of this paper. No data exclusion criteria were applied.

Time-series comparisons between NO2 concentrations derived from each PDT preparation method and the collocated chemiluminescence analyser for all exposure periods are shown in Fig. 2. The entire dataset comprises up to 80 exposures for each preparation method.

The first observation from Fig. 2 is the highly significant trend for PDT NO2 values to exceed the exposure-average chemiluminescence NO2 concentration (positive bias), regardless of the PDT preparation method (P < 0.001, paired t-tests). The mean overestimation of analyser NO2 by PDT across all data (n = 587) is 32%. The mean overestimations by PDT at each site are 18% for PST (n = 112), 41% for CSt (n = 238) and 29% for HM1 (n = 237).

Fig. 2a also shows the model-derived NO2 concentration expected for a PDT at the PST site when additional within-tube chemical production of NO2 (from reaction between the known ambient concentrations of NO and O3 during each exposure period) is also included. The average ratio between the modelled PDT NO2 concentration and analyser NO2 concentration for this sub-set of exposures is 1.26 (n = 14, range 1.13–1.40), confirming again the intrinsic capacity for PDTs to significantly overestimate NO2 because of within-tube chemistry at locations near fluctuating strong sources of NO (e.g. near roads).5–7 The average PDT:analyser NO2 ratio, regardless of PDT preparation method, for exposures for which a modelled value is available is 1.18 (n = 112), which compares well with the mean ratio of 1.26 predicted by the chemical overestimation model. (Note that the observed ratio is likely to be slightly lower, on average, than the model-predicted ratio because of some exposure-duration decline in PDT efficacy even over the 1 week exposures used in this study7). In fact, for this sub-set of PDT data which can be compared with model-predicted values, NO2 concentrations derived from PDT methods A–F do not differ significantly from model concentrations, whilst concentrations from methods G and H are significantly lower (paired t-tests).

The Spearman rank correlation coefficients between NO2 concentrations derived from each PDT preparation method and the continuous analyser are shown in Table 3. The correlations within dipped PDT data (all r > 0.73) and between dipped PDT data and analyser data (r values 0.58–0.75) are consistently greater than the correlations within pipetted PDT data (all r < 0.67) or between pipetted PDT data and analyser data (r values 0.55–0.68). The poorer correlations associated with data from pipetting methods are probably partly a consequence of the lower precision associated with these data (Section 3.1), but must also reflect a greater inaccuracy of pipetting-method data to track the variation of NO2 concentration with exposure/location.

This latter observation is clearly evident in Fig. 2 which shows that, although PDTs prepared by all 8 methods show positive bias in NO2 concentration, there are systematic variations in the accuracy, i.e. the extent to which PDT measurements from different preparation methods are clustered together along a common trend. In general, NO2 data

![Fig. 1 Scatter plot of duplicate determinations of NO2 concentration for PDTs prepared by (a) all dipping methods, (b) all pipetting methods. Tubes in each replicate pair are randomly assigned the label 1 or 2.](image-url)

![Fig. 2 Scatter plot of duplicate determinations of NO2 concentration for PDTs prepared by (a) all dipping methods, (b) all pipetting methods. Tubes in each replicate pair are randomly assigned the label 1 or 2.](image-url)

### Table 2 Summary of duplicate precision data for NO2 measurement using different tube preparation methods

<table>
<thead>
<tr>
<th>Preparation method</th>
<th>Mean % RSD</th>
<th>Median % RSD</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>A (dip, 50%, acetone)</td>
<td>10.4</td>
<td>7.7</td>
<td>80</td>
</tr>
<tr>
<td>B (dip, 20%, acetone)</td>
<td>7.7</td>
<td>6.5</td>
<td>80</td>
</tr>
<tr>
<td>C (dip, 50%, water)</td>
<td>8.2</td>
<td>7.4</td>
<td>77</td>
</tr>
<tr>
<td>D (dip, 20%, water)</td>
<td>7.7</td>
<td>5.3</td>
<td>79</td>
</tr>
<tr>
<td>E (pip, 50%, acetone)</td>
<td>20.9</td>
<td>14.1</td>
<td>79</td>
</tr>
<tr>
<td>F (pip, 20%, acetone)</td>
<td>10.4</td>
<td>7.4</td>
<td>80</td>
</tr>
<tr>
<td>G (pip, 50%, water)</td>
<td>13.5</td>
<td>8.2</td>
<td>77</td>
</tr>
<tr>
<td>H (pip, 20%, water)</td>
<td>10.6</td>
<td>7.5</td>
<td>80</td>
</tr>
<tr>
<td>All dipped</td>
<td>8.5</td>
<td>6.7</td>
<td>316</td>
</tr>
<tr>
<td>All pipetted</td>
<td>13.8</td>
<td>9.0</td>
<td>316</td>
</tr>
<tr>
<td>All</td>
<td>11.2</td>
<td>7.8</td>
<td>632</td>
</tr>
</tbody>
</table>
derived from PDTs prepared by dipping methods A–D (solid symbols) are both more closely clustered to each other, and follow a more tightly-constrained trend of values with respect to the analyser values, than the NO₂ data from PDTs prepared by pipetting methods E–H (open symbols). These latter data are considerably more erratic in accuracy. Fig. 2 shows that data from method E (but also methods F and G) are particularly erratic. However, there needs to be a lot of caution in making statements concerning the accuracy of method E in particular, since the very poor precision associated with this method (Fig. 1b) implies very low confidence in the absolute values.

The existence of differences in NO₂ concentration with tube preparation method is confirmed statistically \((P < 0.001)\) by the non-parametric Friedman test, in which the NO₂ values from the 8 preparation methods from each exposure are ranked from 1–8 and the sum of the ranks assigned to each method over the 76 complete sets compared. The preparation method(s) that differ significantly are determined by comparing the differences between the rank sums for the methods with an
appropriate critical value (Table 4). The table shows that NO₂ concentrations from preparation method G are consistently lower than for other methods and significantly lower than for methods F and B.

4. Discussion

The explanation of an intrinsic potential positive bias in PDT measurement of NO₂ in locations at which NO, NO₂ and O₃ are not at photostationary state is previously documented.5-7 This positive bias is again evident in the PDT data presented here (Fig. 2), and is again consistent with the chemical overestimation interpretation for the subset of data for which such a comparison can be made. It is important to remember that data here are from 1 week exposures. The fact that a positive bias often appears not to exist for PDTs deployed for longer exposures has been explained as the consequence of two opposing effects: a positive bias from chemical overestimation, and a negative bias from length of exposure.5-7 The latter was again demonstrated during this study through observation of significantly lower NO₂ concentrations from 4 week exposed PDTs compared with the average of the four corresponding 1 week exposed PDTs. (Data not presented since they are not relevant to the main study on effect of PDT preparation method).

The comparison of PDT absorbent preparation methods shows a very clear tendency for data from PDTs prepared by dipping the grids in TEA-solvent prior to tube assembly to be more precise than data from PDTs prepared by pipetting the TEA-solvent on to the grids after tube assembly (Fig. 1). There is also a persistent tendency for data from PDTs prepared by pipetting to show greater fluctuation in accuracy relative to each other and to continuous analyser measurements (Fig. 2).
entire grid surface with TEA will reduce the flux of captured NO₂, leading to a negative bias in derived NO₂ concentration, while any "creep" of solution up the inside wall of an assembled tube will increase the absorbent surface area and lead to a positive bias in derived NO₂ concentration. The latter may occur in the pipetting methodology if tubes are tilted or inverted too quickly after pipetting and solution runs down the inside walls of the tube. (It is assumed that the physical act of patting dry dipped grids prior to assembly means that there is no possibility of the above issues for dipped preparation methods). Thus it is proposed that variation from one PDT to the next in the practical action of assembling tubes via a pipetting methodology may explain the greater imprecision of data derived from these preparation methods. The two pipetting methods that yield least precision (methods E and G) both use 50% TEA solutions. Pure TEA is extremely viscous at room temperature, which is the reason it has to be dissolved in a solvent in the first place. A 50% TEA solution is still fairly viscous, so the greater difficulty in reproducibly dispensing such a viscous solution from the pipette may also contribute to the greater imprecision for these specific preparation methods. In the work presented here, tubes were prepared by two different analysts, so whatever the specific cause(s) of the greater imprecision in pipetting methods, it was not analyst specific.

Finally, it is important to emphasize that any effect of preparation method on PDT precision and accuracy will be independent of the effect of any other operational factor(s) that may influence PDT accuracy (for example, a decline in measured NO₂ from longer exposure, and/or an enhancement of measured NO₂ from within-tube chemistry, or wind-induced turbulence). The NO₂ measurement derived from a PDT exposure will be the composite of all relevant influences. The identified influences of these factors do not negate the continued use of PDTs for indicative NO₂ measurement, but it is important that PDT data are always interpreted with due consideration of all parameters associated with their exposures.

5. Conclusions

NO₂ concentrations derived from PDTs prepared by pipetting TEA-solvent solutions onto the grids after assembly are, in general, less precise than concentrations from PDTs prepared by dipping the grids into TEA-solvent solutions prior to tube assembly.

NO₂ concentrations derived from PDTs prepared by pipetting TEA-solvent solutions onto the grids after assembly are also, in general, more variable in accuracy than concentrations from PDTs prepared by dipped methods. There is only weak evidence that one or more method gives systematically different NO₂ across all exposures.

For 1 week exposures at these roadside and urban centre locations, NO₂ concentrations from PDTs prepared by all methods are greater than chemiluminescence analyser NO₂ concentrations, with greater discrepancy at the roadside locations.

Overall, it is concluded from this study that PDT performance is influenced by the physical method of grid preparation, whereby pipetting absorbent solution onto grids after tube assembly (particularly pipetting solutions of high (50%) TEA composition) leads to an apparent degradation in precision and accuracy of NO₂ measurement. It is recommended that these preparation methods be avoided, or at least investigated further.

Acknowledgements

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References