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An Investigation of the Mineral in Ductile and Brittle Cortical Mouse Bone

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ABSTRACT
Bone is a strong and tough material composed of apatite mineral, organic matter, and water. Changes in composition and organization of these building blocks affect bone’s mechanical integrity. Skeletal disorders often affect bone’s mineral phase, either by variations in the collagen or directly altering mineralization. The aim of the current study was to explore the differences in the mineral of brittle and ductile cortical bone at the mineral (nm) and tissue (μm) levels using two mouse phenotypes. Osteogenesis imperfecta model, oim−/−, mice have a defect in the collagen, which leads to brittle bone; PHOSPHO1 mutants, Phospho1−/−, have ductile bone resulting from altered mineralization. Oim−/− and Phospho1−/− were compared with their respective wild-type controls. Femora were defatted and ground to powder to measure average mineral crystal size using X-ray diffraction (XRD) and to monitor the bulk mineral to matrix ratio via thermogravimetric analysis (TGA). XRD scans were run after TGA for phase identification to assess the fractions of hydroxyapatite and β-tricalcium phosphate. Tibiae were embedded to measure elastic properties with nanoindentation and the extent of mineralization with backscattered electron microscopy (BSE SEM). Results revealed that although both pathology models had extremely different whole-bone mechanics, they both had smaller apatite crystals, lower bulk mineral to matrix ratio, and showed more thermal conversion to β-tricalcium phosphate than their wild types, indicating deviations from stoichiometric hydroxyapatite in the original mineral. In contrast, the degree of mineralization of bone matrix was different for each strain: brittle oim−/− were hypermineralized, whereas ductile Phospho1−/− were hypomineralized. Despite differences in the mineralization, nanoscale alterations in the mineral were associated with reduced tissue elastic moduli in both pathologies. Results indicated that alterations from normal crystal size, composition, and structure are correlated with reduced mechanical integrity of bone. © 2014 American Society for Bone and Mineral Research.

KEY WORDS: GENETIC ANIMAL MODELS; MATRIX MINERALIZATION; OSTEOGENESIS IMPERFECTA

Introduction

Cortical bone is a tough material; however, bone’s ability to resist fracture often deteriorates because of aging and/or skeletal diseases. The understanding of bone’s toughening mechanisms requires the interpretation of mechanical and structural properties at multiple scales.1–4 At the whole-bone level (macroscale), cortical bone is the compact bone in the diaphysis and the bony part of the outer shell of the epiphyses. At the tissue level (microscale), lamellar bone is built up of collagen fibers, which are composed of collagen fibrils and mineral crystals (nanoscale). The constituent elements of bone material include apatite mineral, primarily impure forms of hydroxyapatite (HA); organic matter, composed of collagen and noncollagenous proteins; and water, which resides on the surface, within mineral crystals, and between collagen fibers.

Because of this complex hierarchical structure, there are many determinants of bone’s fracture toughness. At the nanoscale, the composite nature of mineralized collagen fibrils and, thus, the mineral and collagen as well as the interaction between them contribute to bone-toughening mechanisms.5,6 Although collagen is accepted to play a major role in bone toughness,5–7 it is also known that bone mineral is altered in skeletal disorders, which lead to bone brittleness.8–12 The objective of this study was to evaluate the mineral properties at the μm-length (tissue) and nm-length (mineral) scales in brittle and ductile cortical bone. Two mouse phenotypes were chosen, based on known fracture toughness values (Fig. 1) to represent brittle and ductile bone.13,14 We used the mouse osteogenesis imperfecta model

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(oim<sup>-/-</sup>), which replicates the moderate to severe condition of osteogenesis imperfecta in humans. Osteogenesis imperfecta, also called brittle bone disease, is primarily caused by mutations in type 1 collagen genes, which lead to bone fragility.<sup>11,15,16</sup> Oim<sup>-/-</sup> bone has decreased ultimate stress<sup>11</sup> and toughness<sup>13,14</sup> at the whole-bone level, higher mineral density measured by electron backscattering,<sup>11,17</sup> and smaller and less arranged apatite crystals.<sup>8,11,17,18</sup> For ductile bones, we used PHOSPHO1 (<i>Phospho1</i><sup>-/-</sup>). These mice lack a phosphatase, which is required for the generation of inorganic phosphate for bone mineralization.<sup>19–21</sup> <i>Phospho1</i><sup>-/-</sup> mice have more ductile bones at the macroscopic scale<sup>13,19</sup> and reduced mineral to matrix ratio as shown by Raman experiments.<sup>19</sup>

We compared the mineral of brittle <i>oim</i><sup>-/-</sup> and ductile <i>Phospho1</i><sup>-/-</sup> mice using bulk techniques to measure overall differences at the nanoscale. After grinding the femur to powder, we used X-ray diffraction (XRD) to determine the average mineral crystal size and thermogravimetric analysis (TGA) to compare the bulk mineral to matrix ratio. TGA monitors the loss of bone mass with temperature from which the fractions of total mineral content and organic content can be calculated. This method complements Raman or Fourier transform-infrared spectroscopy (FTIR) analyses,<sup>11,14,19,22</sup> which measure certain components of the mineral (Phosphate) and collagen (Amide I). TGA measures total mineral and organic contents, including carbonated mineral and noncollagenous proteins, providing additional insight into bulk composition. Changes in composition were investigated with a second XRD analysis of the bone powder after the TGA. Heating the mineral increases its crystallinity,<sup>23</sup> and thus differences in composition and structure become more apparent. The ramifications of the changes in mineral structure and composition were evaluated at the tissue level by analyzing the mineralized matrix. The extent of mineralization of the bone matrix was compared using backscattered electron scanning electron microscopy (BSE SEM). Although density is often used as a correlate of elastic modulus, this is not always true in pathologic bone, where altered mineral organization and structure also affect tissue modulus.<sup>11,24</sup>

We used nanoindentation to identify changes in elastic properties. These multiscale techniques were combined to analyze the mineral of brittle <i>oim</i><sup>-/-</sup> bones, which have a defect in the collagen, and ductile <i>Phospho1</i><sup>-/-</sup> bones, where the mineralization process is deteriorated. We hypothesized that these two pathologies, which map the extremes of whole-bone mechanics (brittle and ductile), will have altered mineral crystal size, composition, mineral fraction, degree of mineralization of the bone matrix, and tissue elastic properties compared with normal bone.

Materials and Methods

Specimens

Bones considered in this study belonged to two mouse strains that show the extremes of toughness (Fig. 1). At the whole-bone level, bones of B6C3Fe-a/aCol1a2oim/oim (<i)oim</i><sup>-/-</sup>) are brittle, whereas Phospho1-R74X null mutant (<i>Phospho1</i><sup>-/-</sup>) mice have ductile bones.<sup>13,14</sup> Both pathologic bones (<i>oim</i><sup>-/-</sup> and <i>Phospho1</i><sup>-/-</sup>) were compared with their corresponding wild-type controls: <i>oim</i><sup>+/+</sup> and <i>Phospho1</i><sup>+/+</sup>. In total, 24 male mice of aged 7 weeks were used (six per group). Femora were utilized to analyze the mineral (nanoscale), whereas tibiae were used to explore bone tissue (microscale) (Fig. 2).

Mineral characterization

Right femora of 6 mice per group (total 24 femora) were used for XRD and TGA analysis. Bones were cleaned of soft tissue and bone marrow, and both epiphyses were cut off with a water-cooled low-speed diamond saw (Isomet, Buehler GmbH, Dusseldorf, Germany). Femora were then defatted using first a 2:1 and then a 1:2 chloroform/ethanol solution for 48 hours each. The specimens were dehydrated in increasing
concentrations of ethanol (70% to 100%) for 10 minutes each. Each bone was wet ground in acetone until a uniform and homogeneous powder was obtained and the vials were left under the fume hood until the evaporation of the acetone. The same femur powder was used for the subsequent XRD-TGA-XRD analysis of each sample.

**X-ray diffraction of bone powder (XRD)**

XRD patterns were obtained using a PANalytical (PANalytical, Almelo, The Netherlands) XRD X’Pert Pro diffractometer operated at 40 kV and 40 mA with no spinning. The initial data collection was in the 2θ range of 20° to 80°, with a step size of 0.033°/2θ (kept constant) and a count time at each step of 35 seconds. A set of slower scans was carried out from 24° to 28° with 250 s/step to better capture the diffraction peak at 2θ = 26° (Fig. 3). This peak corresponds to the (0 0 2) c-axis and does not exhibit overlapping, which is why it was used to measure the average crystal size along the c-axis.\(^{(25)}\) The Scherrer equation was used with the FWHM method (full width at half maximum) to calculate the mean crystal size, \(B\):\(^{(26)}\)

\[
B(2\theta) = \frac{k\lambda}{L\cos\theta}
\]

where \(k\) is the shape factor, \(\lambda\) is the wavelength of the X-ray, \(L\) is the peak width at half maximum, and \(\theta\) is the Bragg angle where the peak is located. The peak at 26° is used to determine the average crystal size in the c-direction according to Eq. 8. \(L\) is the peak width at half maximum and \(\theta\) is the Bragg angle where the peak is located.

**Thermogravimetric analysis (TGA)**

TGA was carried out on the Netzsch STA 449 C Jupiter simultaneous thermal analyzer at a constant heating rate of 10 °C/min in controlled air atmosphere. Femur powder, which weighted 8 to 12 mg depending on the sample, was heated to the required temperature in a platinum crucible. In bone, the change in mass monitored by TGA is considered to be the result of loss of water for temperatures up to 200 °C, organic content from 200 °C to 600 °C, and carbonate content above 600 °C.\(^{(27-29)}\) Three of the samples per group were heated from room temperature to 800 °C and the other three to 1200 °C. Mineral to matrix ratio was calculated as the ratio between the percentages of mass remaining after heating to 600 °C and the organic mass loss between 200 °C and 600 °C.\(^{(28)}\)

\[
\frac{\text{Mineral}}{\text{Matrix}} = \frac{m_{600°C}(%)}{m_{200°C}(%)} - m_{600°C}(\%)
\]

The percentage of mass at 600 °C represents the weight percentage of mineral content, and it depends on the amount of mass lost because of moisture, which might be altered in pathologic bones. To avoid this influence, the mass percentage at 600 °C was translated to dry weight percentage for the calculation of the mineral to matrix ratio:

\[
m_{600°C}(\%\text{dryweight}) = \frac{m_{600°C}(\%\text{initialweight})}{m_{200°C}(\%\text{initialweight})} \times 100
\]

**Tissue characterization**

Right tibiae of 4 mice per group (total 16 tibiae) were used for nanoindentation and BSE SEM. Tibiae were cleaned of surrounding soft tissue and fixed in 70% ethanol for 48 hours. They were dehydrated in a series of increasing concentrations of ethanol (80%, 90%, and 100% for 24, 24, and 72 hours, respectively) and changed to a xylene solution (48 hours). The samples were then infiltrated in pure methyl methacrylate monomer mixed with \(\alpha\)-azo-iso-butyronitrile under vacuum and polymerized to PMMA at 30 °C (chemicals bought from VRW, Lutterworth, UK). Tibia blocks were cut transversally at the mid-diaphysis and the cross sections polished with graded silicon carbide papers (from P800 to P4000), and cleaned ultrasonically with distilled water between polishing steps.
Nanoindentation (NI)

Nanoindentation was performed on mid-diaphyseal medial cross sections of the PMMA-embedded tibias (Fig. 2) using a TI700 UBI system (Hysitron, Minneapolis, MN, USA). Indentation tests were carried out with the specimen in the dry condition using a 55-μm-radius sphere tip. A trapezoidal loading protocol was applied longitudinally to a maximum load of 8 mN with a rising time of 10 seconds and a holding of 30 seconds. Nine indents were made in each specimen with a minimum spacing of 15 μm between indents (Fig. 2). Viscoelastic analysis was used to evaluate elastic properties.\(^{33-35}\) The time-displacement data \(h(t)\) were fitted to the viscoelastic Boltzmann integral equation:

\[
h^{3/2}(t) = \frac{3}{8\sqrt{R}} \int J(t-u) \frac{dP(u)}{du} \, du \tag{4}
\]

where \(R = 55 \text{ μm}\) is the radius of the tip; \(P\) is the applied load; \(u\) is the dummy variable of integration for time; and \(J(t)\) is the material creep function. The creep function is defined as a function of the creep coefficients \(C_0, C_i\) and the material time constants \(T_i:\)

\[
J(t) = C_0 - \sum_{i=1}^{2} C_i \exp\left(-t/T_i\right) \tag{5}
\]

The solution of Eq. 4 for the holding period of trapezoidal loading results in the following expression:

\[
h^{3/2}(t) = \frac{3}{8\sqrt{R}} P_{\text{max}} \left\{ C_0 - \sum_{i=1}^{2} C_i \exp\left(-t/T_i\right) \frac{T_i}{t_0} \left[\exp(t/T_i) - 1\right]\right\} \tag{6}
\]

where the maximum applied load \(P_{\text{max}} = 8 \text{ mN}\), and rising time \(t_0 = 10 \text{ s}\). The values of \(C_0, C_i\), and \(T_i\) were obtained using a nonlinear least-squares curve-fit function in MATLAB (Mathworks, Natick, MA, USA).\(^{36}\)

The instantaneous \(G_0\) and equilibrium \(G_\infty\) shear modulus are calculated from the creep coefficients:

\[
G_0 = \frac{1}{2(C_0 - C_1 - C_2)} \tag{7}
\]

\[
G_\infty = \frac{1}{2C_0} \tag{8}
\]

The ratio \(G_\infty/G_0\) gives insight into the viscoelastic behavior of the material: It ranges from 0, in perfectly viscous materials, to 1, in perfectly elastic materials. The instantaneous shear modulus was used to calculate the plane strain modulus, \(E'\):

\[
E' = 4 \times G_0 \tag{9}
\]

Young’s modulus \(E\) was computed assuming a Poisson’s ratio of \(ν = 0.3\) in cortical bone, following:

\[
E' = \frac{E}{(1 - ν^2)} \tag{10}
\]

Backscattered electron microscopy (BSE SEM)

After the indentations, the PMMA blocks were repolished and carbon coated. Samples were analyzed using EVO MA10 scanning electron microscope (Zeiss UK Ltd, Cambridge, UK) operated at 20 kV, with a beam current of 1.0 nA and at a working distance of 12 mm using monobromo and monoiodo dimethacrylate standards.\(^{11,37}\) Images\(^{38}\) was employed to plot the combined histogram of the grey values of all the bones to identify the lower (A, 125) and upper (B, 235) bounds across histograms (Fig. 4). These bounds were used to normalize the grey values of each pixel according to:

\[
p_n = \frac{(p-A)}{(B-A)} \times r \tag{11}
\]

where \(p_n\) is the normalized pixel value, \(p\) is the current pixel, and \(r\) is the bin range, in this case 255. For visualization purposes, the grey-level range of the normalized histogram was divided into eight equal-size classes of different colors ranging from nonmineralized (black) to highly mineralized (white) bone matrix. The results reported hereafter are the ones corresponding to the normalized distribution in Fig. 4.

Statistical analysis

Mean values and standard deviations were calculated for the measured parameters. Independent \(t\)-tests were used to compare crystal size, mineral/matrix ratio, elastic properties, and mean BSE intensity values of pathologic versus healthy bone (\(\text{oim}^-\) versus \(\text{oim}^+\); \(\text{Phospho}1^-\) versus \(\text{Phospho}1^+\)). Equality of variances was assumed when Levene’s test gave values of \(p > 0.05\). Mann-Whitney U test was used in the cases where the data were not normally distributed according to the Shapiro-Wilk test. Differences were considered significant at \(p < 0.05\). Statistical analysis was performed using SPSS (v.21, SPSS Inc., Chicago, IL, USA).

Results

Average crystal size

Pathologic bones had smaller crystal size than their controls (Fig. 5). In brittle bones, the average crystal size decreased from 25.0 ± 0.6 nm (\(\text{oim}^+\)) to 17.8 ± 0.9 nm (\(\text{oim}^-\)) (\(p < 0.001\)). In ductile bones, it decreased from 22.9 ± 0.5 nm (\(\text{Phospho}1^+\)) to 21.6 ± 0.6 nm (\(\text{Phospho}1^-\)) (\(p = 0.001\)).

Fig. 4. Combined histogram of grey values of all the bones used to identify the lower (A) and upper limits (B). And the normalized combined histogram between A and B.
Bulk mineral content

Representative TGA curves of weight loss with temperature are plotted in Fig. 6A. The percentages of weight associated with organic content ($m_{200}^\text{C} (%) - m_{600}^\text{C} (%)$), mineral phase ($m_{600}^\text{C} (%)$) and carbonate content ($m_{600}^\text{C} (%) - m_{800}^\text{C} (%)$) are shown in Table 1. The weight loss associated with moisture is not shown because it might have been influenced by sample preparation. Brittle $oim^{-/-}$ and ductile $Phospho1^{-/-}$ bones exhibited increased organic content ($p < 0.001$) compared with their controls. No differences were found in the loss of carbonate content to $800 \, ^\circ C$. Mineral/matrix ratios, calculated as the percentage of dry mass remaining at $600 \, ^\circ C$ divided by the loss of organic matter (Eq. 2, 3), indicated that both pathologic bones had smaller mineral/matrix ratio compared with their controls ($p < 0.001$) (Fig. 6B).

Conversion to HA and TCP

Fig. 7 represents XRD patterns of $oim$ and $Phospho1$ bone powder in three stages: unheated, after TGA to $800 \, ^\circ C$, and after the heat treatment to $1200 \, ^\circ C$. The XRD spectra of unheated bones could not be distinguished among all four mouse models because of low crystallinity and peak overlap. XRD after thermal treatment induced an increase in crystallinity for all the samples and revealed differences in the amounts of HA and TCP among the bones (Fig. 8). Fig. 8 shows representative diffraction patterns of the samples heated to $1200 \, ^\circ C$, where the peaks corresponding to HA and TCP are identified. The heat treatment induced a bigger mass conversion to TCP for pathologic bones, which were heated to $1200 \, ^\circ C$ (42 ± 3% TCP in $oim^{-/-}$ and 31 ± 1% in $Phospho1^{-/-}$) compared with wild-type controls (25 ± 3% in $oim^{+/+}$, and 25 ± 1% in $Phospho1^{+/+}$). It must be noted that only half of the bones were heated to $1200 \, ^\circ C$, with pathologic bones having a bigger fraction of TCP mass (44 ± 6% in $oim^{-/-}$ and 23 ± 1% in $Phospho1^{-/-}$) than their wild-type controls (18 ± 1% in $oim^{+/+}$ and $Phospho1^{+/+}$).

Tissue elastic properties

Table 2 summarizes the means and standard deviations of elastic properties for $oim$ and $Phospho1$ tibiae. The plane strain modulus $E'$, instantaneous shear modulus $G_0$, and infinite shear modulus $G_\infty$ were significantly smaller in pathologic bones compared with wild-type controls. As expected for dry and PMMA-embedded samples, no significant differences were found in the extent of viscoelasticity, $G_\infty/G_0$. Pathologic bones had smaller Young’s modulus than their controls ($p < 0.001$ for $oim^{-/-}$; $p = 0.017$ for $Phospho1^{-/-}$) (Fig. 9).

Degree of mineralization of bone matrix

$Oim^{-/-}$ bones were more mineralized and $Phospho1^{-/-}$ bones were less mineralized than their controls (Fig. 10). Combined histograms were plotted per strain to compute average pixel distributions. The mean grey value was smaller for $Phospho1^{-/-}$ bones ($117 \pm 28$) and bigger for $oim^{-/-}$ ($168 \pm 29$) when compared with $Phospho1^{+/+}$ ($143 \pm 26$, $p < 0.001$) and $oim^{+/+}$ ($139 \pm 27$, $p < 0.001$), respectively.

Discussion

This study explored the mineral phase of brittle $oim^{-/-}$ and ductile $Phospho1^{-/-}$ bones. Interestingly, despite their extremely different mechanical behavior at the macroscale, the mineral of...
**Table 1.** Mean and standard deviations of weight % of organic (200–600 °C), mineral (at 600 °C), and carbonate content (600–800 °C) for oim and Phospho1 bone powder

<table>
<thead>
<tr>
<th></th>
<th>Oim+/+</th>
<th>Oim−/−</th>
<th>p-value</th>
<th>Phospho1+/+</th>
<th>Phospho1−/−</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Organic (wt %)</td>
<td>24.8 (1.0)</td>
<td>32.3 (1.9)</td>
<td>&lt;0.001*</td>
<td>22.1 (0.6)</td>
<td>25.6 (1.0)</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>Mineral (wt %)</td>
<td>65.6 (1.0)</td>
<td>55.0 (3.4)</td>
<td>&lt;0.001*</td>
<td>65.9 (1.2)</td>
<td>61.7 (1.5)</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>Carbonate (wt %)</td>
<td>1.1 (0.2)</td>
<td>1.0 (0.1)</td>
<td>0.310</td>
<td>0.9 (0.1)</td>
<td>0.9 (0.1)</td>
<td>0.628</td>
</tr>
</tbody>
</table>

*p < 0.05

oim−/− and Phospho1+/+ bones displayed some similar trends. Both pathologic bones had smaller mineral crystal size, less bulk mineral to matrix ratio, and bigger thermal conversion to TCP than controls. However, oim−/− bones were hypermineralized, whereas Phospho1−/− were hypomineralized. Despite the differences in the degree of mineralization of the bone matrix, both brittle and ductile bones had reduced tissue elastic moduli.

At the mineral level, average size of oim−/− and Phospho1+/+ apatite crystals along the c-axis was reduced. This might be a consequence of the disrupted collagen fibril template in brittle bones[11] and altered mineral formation pathway owing to lack of phosphate in ductile bones.[39,40] Transmission electron microscopy (TEM) and small-angle X-ray scattering (SAXS) have shown that apatite crystals of oim−/− bone have significantly smaller thickness and are packed more tightly in a less organized manner when compared with oim+/+.[11,17,18] Smaller crystals have also been measured in children with one severe form of osteogenesis imperfecta.[41] In the current study, it was found that, although the differences were not as pronounced as in brittle bone, Phospho1−/− bones also had significantly reduced average mineral crystal size. Variations in the mineral crystal size have previously been found in bones exhibiting altered mechanical properties;[42] as the crystal size might influence how bone, as a composite material, responds to load. The current study shows that a reduction in the average crystal size is associated with both brittle oim−/− and ductile Phospho1−/− bone.

The bulk mineral content was reduced in both pathologic bones, which led to lower mineral/matrix ratios (Fig. 6B). It must be noted that differences in moisture content might have been affected by sample preparation, which is why the mineral to matrix ratio was calculated using the dry weight at 600 °C. In Phospho1−/− bones, the lower mineral/matrix ratio is in agreement with Raman experiments.[19] However, in oim−/− bones, Raman and FTIR studies have measured increased[11,14] and decreased[22] mineral/matrix ratios. The discrepancy between Raman, FTIR, and TGA in brittle bones is likely to be attributable to differences in how the organic and mineral contents are calculated: TGA provides a bulk measurement, whereas Raman and FTIR are local measurements. TGA measures the ratio of the total mineral content (including carbonated hydroxyapatite) to the total organic content (including bone matrix collagen, noncollagenous proteins, lattice water, blood vessels, and all other organic content in osteocytes and other cells in blood vessel canal spaces).[43,44] In contrast, in FTIR and Raman, the mineral content is represented only by the phosphate band and the organic content is limited to the collagen type I Amide I band.[45] Smaller crystals have also been measured in children with one severe form of osteogenesis imperfecta.[41] In the current study, it was found that, although the differences were not as pronounced as in brittle bone, Phospho1−/− bones also had significantly reduced average mineral crystal size. Variations in the mineral crystal size have previously been found in bones exhibiting altered mechanical properties;[42] as the crystal size might influence how bone, as a composite material, responds to load. The current study shows that a reduction in the average crystal size is associated with both brittle oim−/− and ductile Phospho1−/− bone.

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vessels, which are expected to influence the fracture behavior of bone.\(^{47-50}\)

After the heat treatment, all bones evolved to a biphasic mixture of HA and TCP, but pathologic samples showed a higher conversion to TCP (Fig. 8), indicating chemical deviations from the stoichiometric HA. The increased conversion to TCP, with a lower Ca/P ratio than HA, suggests that the mineral of the stoichiometric HA. The increased conversion to TCP, with a higher percentage of TCP (Fig. 8), indicating chemical deviations from Phillips and colleagues\(^{10}\) measured a reduction of Ca/P in the demineralized bone.\(^{36,54,55}\) Young’s modulus was reduced by 19% in oim\(^{-/-}\) bones, which is in accordance with a recent study.\(^{11}\) Thus, reductions in Ca/P in pathologic bones not readily evident from XRD without heat treatment.

A notable difference in the tissue properties of brittle and ductile bones in the present experiments resides in the mineralization of the tissue. Because our aim was to compare mineralization degrees among bones within the same study, mineralization was left in terms of normalized BSE SEM grey values, instead of translating these values to mineral density.\(^{27}\)

This was done with the purpose of avoiding the many assumptions required to convert grey values to mineral density.\(^{11}\) Mineralized matrix of oim\(^{-/-}\) bone was more mineralized, which is in agreement with previous BSE SEM studies on oim\(^{-/-}\) and human OI bone.\(^{11,17,33}\) In contrast, as expected from the lack of PHOSPHO1 enzyme and suggested from our previous studies,\(^{21}\) Phospho1\(^{+/+}\) bones were less mineralized than controls.

Table 2. Means and standard deviations of elastic properties of oim and Phospho1 tibiae embedded in pmma, indented with a sphere in dry conditions

<table>
<thead>
<tr>
<th></th>
<th>Oim(^{-/-})</th>
<th>Oim(^{+/+})</th>
<th>p-value</th>
<th>Phospho1(^{-/-})</th>
<th>Phospho1(^{+/+})</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>(E'(GPa))</td>
<td>17.1 (2.3)</td>
<td>13.9 (2.5)</td>
<td>&lt;0.001*</td>
<td>12.0 (2.7)</td>
<td>10.3 (1.9)</td>
<td>0.014</td>
</tr>
<tr>
<td>(G_0(GPa))</td>
<td>4.27 (0.58)</td>
<td>3.47 (0.63)</td>
<td>&lt;0.001*</td>
<td>3.01 (0.68)</td>
<td>2.57 (0.48)</td>
<td>0.016</td>
</tr>
<tr>
<td>(G_{\infty}(GPa))</td>
<td>2.77 (0.54)</td>
<td>2.35 (0.53)</td>
<td>0.004*</td>
<td>2.03 (0.40)</td>
<td>1.64 (0.33)</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>(G_{\infty}/G_0)</td>
<td>0.65 (0.10)</td>
<td>0.68 (0.09)</td>
<td>0.487</td>
<td>0.68 (0.05)</td>
<td>0.65 (0.09)</td>
<td>0.440</td>
</tr>
</tbody>
</table>

\(^{1}E’\) is the plane strain modulus; \(G_0\) is the instantaneous shear modulus and \(G_{\infty}\) corresponds to the shear modulus at infinite time; \(G_{\infty}/G_0\) represents the elastic fraction (viscous \(0 \leq G_{\infty}/G_0 \leq 1\) elastic).

\(^{*}p < 0.05\)
are not visible in the SEM images. The differences between results derived from TGA and BSE SEM in both pathologic bones highlight the need to distinguish bulk bone mineral quantity (measured with TGA) from the extent of mineralization of the bone matrix (by BSE SEM).

Conclusions

The mineral phase of brittle and ductile bones was compared at the nano- and microscale. In brittle oim<sup>-/-</sup> bones, the mutation affecting collagen structure has a profound effect on mineralization, whereas in ductile Phospho1<sup>-/-</sup> bones, the mineral is directly affected by the lack of PHOSPHO1, which in turn might also affect collagen structure.<sup>(19)</sup> Although the consequences of these two defects are very different at the macroscale, the current study demonstrated that both pathologies had smaller apatite crystals, which were less stoichiometric than healthy bone mineral, and showed that pathologic bones had a lower weight % of bulk mineral content. In contrast, the extent of mineralization of the bone matrix was different for oim<sup>-/-</sup> and Phospho1<sup>-/-</sup> bones, as brittle bones were hypermineralized, whereas ductile bones were hypomineralized. Despite these differences in the mineralization, the tissue elastic modulus was reduced in both pathologies. This emphasizes that mineralization is not the only determinant of tissue elastic moduli and suggests that deviations in the size, composition, and organization of bone mineral affect bone micromechanics. The current study was limited to the analysis of the mineral in oim<sup>+</sup> and Phospho1<sup>+</sup> bone; however, future studies should examine the organic content, which was increased in both pathologies, to identify nanoscale alterations in the collagen composition, matrix architecture, and collagen cross-links, as well as microscale alterations in vascular porosity. A detailed multiscale analysis of pathologic bones is essential to characterize the properties that should be targeted in the development of new therapies for skeletal diseases affecting whole-bone mechanics.

Disclosures

All authors state that they have no conflicts of interest.

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**Fig. 9.** Means and standard deviations of Young’s modulus E, with pathologic bones exhibiting reduced Young’s modulus. *p < 0.05.

**Fig. 10.** Normalized backscattered electron intensity maps of tibial cross sections (left) and the combined histograms of pixel values for each strain (right) from nonmineralized (0, black) to high mineralization (255, white). Bone matrix was hypomineralized in Phospho1<sup>-/-</sup> and hypermineralized in oim<sup>-/-</sup>, as calculated from the mean grey values of the histograms. *p < 0.001.
analysis: AB and NR-F. Drafting manuscript: NR-F. Revising manuscript: all authors. Approving final version of manuscript: all authors. NR-F takes responsibility for the integrity of data analysis.

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