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Citation for published version:
Goldmann, W, Marier, E, Stewart, P, Konold, T, Street, S, Langeveld, J, Windl, O & Ortiz-Pelaez, A 2016, 'Prion protein genotype survey confirms low frequency of scrapie-resistant K222 allele in British goat herds' Veterinary Record, vol. 178, 168. DOI: 10.1136/vr.103521

Digital Object Identifier (DOI):
10.1136/vr.103521

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Veterinary Record

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Prion protein genotype survey confirms low frequency of scrapie-resistant K222 allele in British goat herds

W. Goldmann, E. Marier, P. Stewart, T. Konold, S. Street, J. Langeveld, O. Windl, A. Ortiz-Pelaez

Scrapie in goats is a transmissible, fatal prion disease, which is endemic in the British goat population. The recent success in defining caprine PRNP gene variants that provide resistance to experimental and natural classical scrapie has prompted the authors to conduct a survey of PRNP genotypes in 10 goat breeds and 52 herds to find goats with the resistant K222 allele. They report here the frequencies in 1236 tested animals of the resistance-associated K222 and several other alleles by breed and herd. Eight animals were found to be heterozygous QK222 goats (0.64 per cent genotype frequency, 95 per cent CI 0.28 to 1.27 per cent) but no homozygous KK222 goats were detected. The K222 allele was found in Saanen, Toggenburg and Anglo-Nubian goats. The fact that only a few goats with the K222 allele have been identified does not preclude the possibility to design and implement successful breeding programmes at national level.
others 2013, Lacroux and others 2014, Aguilar-Calvo and others 2015). The protective effect of K222 is supported by epidemiological evidence confirming the scarcity of K222 carriers in natural scrapie cases (Acuitis and others 2006, 2012, Vaccari and others 2006, Barillet and others 2009), although there may be exceptions to this association (Fraugia and others 2011, Kanata and others 2014). The K222 allele is present in the most important dairy breeds (e.g. Saanen) although at rather low frequencies in most European countries, and it was particularly low in a survey of British goats conducted between 2007 and 2009 (Goldmann and others 2011). Strong evidence from case-control studies of Cypriot goat herds support the protection against classical scrapie conferred by the S146 and D146 alleles. (Papasavva-Stylianou and others 2007, 2011, Ortiz-Pelaez and others 2014). In the UK, only S146 has been found, and in larger numbers only in Boer goats (Goldmann and others 2011). White and others (2012) have reported that after oral scrapie challenge of NS146 heterozygotes, all animals survived without developing disease for 40 per cent longer (at which point the experiment was terminated) than wild-type homozygotes, which had a 100 per cent attack rate, suggesting that the protective effect of this amino acid change may not be restricted to Cypriot scrapie strains. Moreover, the amino acid variation asparagine or serine in codon 146 has been found in sheep, goats and cattle. Association of amino acid changes with scrapie disease susceptibility in one species may also apply to another species, which makes the codon 146 polymorphism particularly interesting.

The goat population in Great Britain is small compared with other EU countries and contributes a small but growing fraction to the total livestock production. The commercial goat sector is specialised with most of the large holdings producing milk for on-farm processing or collection mainly for pasteurisation and cheese production. A few larger pedigrees maintain and supply purebred males to commercial herds that have developed and specialised with most of the large holdings producing milk for on-farm processing or collection mainly for pasteurisation and cheese production. For purposes other than farming (Ortiz-Pelaez and others 2012), the majority of goats in Great Britain are family pets with one or two individuals kept according to their production traits. The majority of goats in Great Britain are family pets with one or two individuals kept for purposes other than farming (Ortiz-Pelaez and others 2012). Scrapie became a notifiable disease in 1993 in the UK in accordance with EU Council Directive 91/68/EC. Passive surveillance systems, that is, reporting of clinical suspect animals, led to the histopathological confirmation of 28 UK goat scrapie cases between 1976 and 2002 (Dustan and others 2008), while in the 12-year period from 2002 to 2013 with additional active surveillance, a total of 204 cases were pathologically confirmed, all belonging to the classical disease form (Ortiz-Pelaez and Arnold 2014). This number of British scrapie cases supports the consideration of selection programmes based on PRNP genetics similar to sheep.

The purpose of this study was to ascertain by means of a survey the current frequency of the resistance-associated K222 allele in British goats. This survey was also used to record other PRNP polymorphisms, which allowed a comparison of PRNP allele and genotype frequencies in the British goat population to a similar survey conducted by the authors five years ago.

**Materials and methods**

**Herd selection and sample collection**

The initial selection was based on ≥10 goats per holding using the list of herds in the Sheep and Goat Inventory 2012 in Great Britain, applying a multistage random sampling strategy. Within the herds, all males were sampled and additional females were chosen by the goat keeper to reach the maximum number of animals as determined by the sample size calculation (see below). Animals older than six months and intended to be kept as breeding animals were eligible for the survey.

The calculation for the number of herds to sample was based on the assumption that 10 per cent of the herds contained at least one animal with the 222K allele, a confidence level of 95 per cent confidence and 5 per cent desired accuracy of the expected prevalence. To calculate the sample size in each herd, a 2 per cent prevalence of the 222K in the whole population of goats was assumed with a 95 per cent confidence level and 5 per cent desired accuracy. Assumptions on animal and herd prevalence of the 222K allele in the British goat population were based on published (Goldmann and others 2011) and unpublished data (W. Goldmann, unpublished observations). With these assumptions, a maximum of 50 animals per herd (including all males) and 117 herds were required for this survey. With the current distribution of herd size in the sampling frame as per holdings with >10 goats listed in the Sheep and Goat Inventory 2012 (Defra UK website), approximately 2300 samples (one sample per animal) were required.

The survey was presented at the Goat Veterinary Society Annual Meeting in November 2012 (Ortiz-Pelaez and others 2013) and two calls for expressions of interest were published in the British Goat Society Journal (Clayton 2015, Ortiz-Pelaez 2015) in an attempt to increase participation. Participation in the survey was voluntary. In return, recruited goat keepers were offered free sampling, genotyping and a report with genotype results and animal details. Participants were advised to retain animals with resistant genotypes with the view to promote breeding for scrapie resistance in their herd. The selection of the farmers was adapted due to the difficulty in recruiting randomly selected farms. Farmers were first recruited from those who expressed an interest and met the minimum herd size. Subsequently, goat keepers were selected at random from the database of holdings (inventory 2012) and along with goat breeder associations and veterinarians contacted directly with the view to increase the returns, which resulted in recruitment of three farms with <10 goats. During the sampling visit, owner’s consent was obtained via a signed acceptance of the sampling procedure in the conditions described earlier. They were also asked to fill in a questionnaire to obtain basic epidemiological data of the herd: production type and number of homebred and purchased goats by age, sex and breed.

Initially, a blood sample was collected from the jugular vein (max. 10 ml, in EDTA) for DNA extraction. It was later replaced by nasal swabbing whereby a cotton bud-sized sponge was gently rubbed along the nasal mucosa and placed in a liquid-containing tube to stabilise the DNA (Performagene Nasal PG-100 collection kit, DNA Genotek, Ontario, Canada). After a trial on several farms, the nasal swabbing was found to cause no harm and compared with insertion of a hypodermic needle was considered a refinement that did not constitute a regulated procedure under the Animal (Scientific Procedures) Act. It could therefore be carried out by farmers.

Farmers were asked to fill in a questionnaire to obtain basic epidemiological data of the herd: production type and number of homebred and purchased goats by age, sex and breed.

Samples and paperwork to the project office.

**Genotyping**

DNA was extracted from blood using the Qiagen DNeasy blood & tissue kit (Qiagen, Crawley, UK) or from the Performagene nasal swap (DNA Genotek, Ontario, Canada) as recommended by the manufacturers. PCR amplification of PrP coding region and sequencing were performed as described in Goldmann and others (2011). Two alternative PCR reactions were performed depending on the quality of recovered DNA, one resulting in the full open reading frame (ORF) sequence and one producing codons 194 to 256 only to allow genotyping of codons 211 and 222.

**Results**

A total of 52 goat farms were included in the survey conducted between August 2013 and December 2014, which were located in England (49), Scotland (2) and Wales (1). Figure 1 shows the distribution of goat herds across Great Britain that were sampled in relation to the overall distribution of goat herds based on the 2013 goat inventory. Herd sizes ranged from 8 to 6651 goats (mean herd size 513, median 54). The goats were kept as breeding (12 herds, 22.6 per cent), production (56 herds, 71.6 per cent)
or companion animals (3 herds, 5.7 per cent) encompassing 10 breeds (Saanen, Toggenburg, Alpine, Anglo-Nubian, Boer, Cashmere, Angora, pygmy, Golden-Guernsey and Bagot) and crossbreeds (see Tables 1 and 2). The production animal herds were split by breed type into 24 dairy (63.2 per cent), 8 meat (21 per cent) and 6 fibre (15.8 per cent) herds.

There were 1293 collected samples with an average of 24.8 goat samples per herd (male to female ratio: 1:5). A total of 791 samples (61.2 per cent) were taken as blood by veterinary or scientific staff and 502 samples (38.8 per cent) were collected as nasal swaps by goat keepers. The success rate for genotyping was 95.3 per cent and 97.4 per cent for blood and swap samples, respectively. However, the need to repeat the analysis was higher than for swap samples.

Out of the 1293 samples, 1256 had a genotype result available (95.6 per cent of collected samples) of which 8 animals were found to be heterozygous QK222 goats (0.64 per cent genotype frequency, 95 per cent CI 0.28 to 1.27 per cent) while no homozygous KK222 goats were detected in the survey (Table 1). This gives a total K222 allele frequency of 0.32 per cent for this herd population. The authors genotyped codon 146 for 172 Boer goats across nine herds and found 105 carrying the SI46 allele, 81.6 per cent were SI146 heterozygotes and 18.4 per cent SI146 homozygotes (Table 4); none carried the DI46 allele. The allele frequency of 50.5 per cent in Boer goats is significantly different (P<0.0001) from that of the other breeds in this study, which was 1.3 per cent of 405 analysed genotypes. Genotypes for codons 142 and 211 were established for 742 and 784 goats, respectively. The M142 and Q211 alleles confer partial resistance and their frequency was therefore of interest. The M142 allele had high frequencies in Saanen (30.1 per cent, 12 herds) and Toggenburg (34.4 per cent, 7 herds) while the crossbreed, with a substantial number of MM142 homozygotes (9 per cent and 13 per cent, respectively). All other goats together reached a 3 per cent allele frequency. The Q211 allele frequencies were particularly high in the Alpine and Golden Guernsey breeds with 15.6 per cent (three herds) and 68.1 per cent (three herds), respectively; all other breeds together had an allele frequency of only 0.9 per cent.

Discussion

Breeding for scrapie resistance in goats is a long held aim for countries in which this prion disease has significant impact on their livestock production and animal welfare. The implementation of classical scrapie eradication programmes for sheep in European countries proved that it is possible to significantly reduce the prevalence of scrapie using PRNP genetic approaches in conjunction with herd cull protocols (EFSA 2014). This has been demonstrated by applying the appropriate dissemination of resistance alleles from the male breeding population, resulting in a large increase in the frequency of resistance alleles. For example, in Cyprus 95 per cent of the sheep population are now homozygous or heterozygous carriers of the resistant ARR allele, a massive increase from its 2005 level of 54 per cent (EU TSE Annual Report 2013).

In goats, there are two strongly protective PRNP alleles based on the K222 and SI46/DI46 polymorphisms. The authors’ survey has shown again that for the British goat population the SI46 PrP allele is mostly limited to the Boer breed, where it is, however, found at a high frequency. In contrast, the K222 allele has been found in all major dairy breeds in the GB population, but at a very low frequency. Their analysis of 1236 goats from different regions of Britain for the K222 allele revealed only eight carriers in 52 herds, a very low allele frequency of 0.6 per cent.
and a herd frequency of 11.5 per cent. No KK222 homozygotes were found, which is in line with the low allele frequency. When compared with the British goat survey conducted in 2007–2009 (Goldmann and others 2011), Saanen goats and their crossbreds had a very similar allele frequency of 0.7 per cent, implying that without active PRNP genotyping and selection there may not be a significant increase in the K222 allele frequency.

Classical scrapie is commonly found in animals with homozygous IRRQ allele carriers, for example, L218 and H154 in Cashmere and G102 in Golden Guernsey. To accurately assess the allele frequency of these alleles, a larger number of herds and animals would have to be collected.

This survey has shown that the potentially scrapie-protective K222 allele is very rare in British goats and that there has not been a major change since the last survey six years ago when the breed composition of the goat sampling is taken into account. However, there were a larger proportion of herds containing at least one goat with the K222 allele. Breeding for K222-associated resistance to scrapie will be challenging and may demand a strict breeding programme using natural mating in those breeds where the K222 allele was found. Alternatively, the use of artificial insemination has to be contemplated with imported semen from K222 males of the targeted breeds. For those breeds where no K222 allele holders were found, private initiatives from breeding societies to genotype billy goats and/or importing semen or breeding material may be considered.

The difficulty in recruiting the target number of herds has resulted in less precise estimates, adding a greater degree of uncertainty on the actual allele frequencies in the goat population, especially to the number of herds holding goats with potentially resistant alleles (up to 20 per cent). Goat keepers interested in producing breeding stock and interested in adding potentially resistant alleles (up to 20 per cent) of this allele in this breed. Several of the other variant alleles appear to be breed specific, for example, L218 and H154 in Cashmere and G102 in Golden Guernsey. To accurately assess the allele frequency of these alleles, a larger number of herds and animals would have to be collected.

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The difficulty in recruiting the target number of herds has resulted in less precise estimates, adding a greater degree of uncertainty on the actual allele frequencies in the goat population, especially to the number of herds holding goats with potentially resistant alleles (up to 20 per cent). Goat keepers interested in producing breeding stock and interested in adding value to their goats may have been more likely to participate. On the other hand, farmers may have been less keen to participate with suspicion that the samples could be used to test for scrapie or reluctance to sample goats without a diagnostic

### Table 2: Frequencies of QK222 PRNP genotypes in goat breeds and other PRNP gene variation by breed

<table>
<thead>
<tr>
<th>Breed</th>
<th>Alpine</th>
<th>Anglo-Nubian</th>
<th>Angora</th>
<th>Boer</th>
<th>Cashmere</th>
<th>Golden-Guernsey</th>
<th>Pygmy</th>
<th>Saanen</th>
<th>Toggenburg</th>
<th>SaanenX</th>
<th>ToggenburgX</th>
<th>Others*</th>
<th>All</th>
</tr>
</thead>
<tbody>
<tr>
<td>Herds n</td>
<td>11</td>
<td>15</td>
<td>5</td>
<td>14</td>
<td>1</td>
<td>4</td>
<td>2</td>
<td>24</td>
<td>17</td>
<td>10</td>
<td>5</td>
<td>1236</td>
<td>5</td>
</tr>
<tr>
<td>Goats n</td>
<td>51</td>
<td>120</td>
<td>140</td>
<td>29</td>
<td>40</td>
<td></td>
<td>29</td>
<td>211</td>
<td>173</td>
<td>148</td>
<td>39</td>
<td>1236</td>
<td>5</td>
</tr>
<tr>
<td>QK222 n (%)</td>
<td>0</td>
<td>2 (1.7)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td></td>
<td>0</td>
<td>4 (1.9)</td>
<td>1 (0.5)</td>
<td>1 (0.7)</td>
<td>0</td>
<td>8 (0.6)</td>
<td>5</td>
</tr>
</tbody>
</table>

**Other detected polymorphisms**

| Polymorphism | S127 | R143 | M142 | G102 | S146 | H154 | L218 | Q211 | P2405 | P2405 | P2406 | P2405 | P2405 |
|--------------|------|------|------|------|------|------|------|------|-------|-------|-------|-------|-------|-------|
| S127         | S27  | S127 | S127 | G102 | S127 | S127 | S127 | S127 | S127  | S127  | S127  | S127  |       |
| R143         | R143 | R143 | R143 | S146 | R143 | R143 | R143 | R143 | R143  | R143  | R143  | R143  |       |
| M142         | M142 | M142 | M142 | M142 | M142 | M142 | M142 | M142 | M142  | M142  | M142  | M142  |       |
| S146         | S146 | S146 | S146 | S146 | S146 | S146 | S146 | S146 | S146  | S146  | S146  | S146  |       |
| H154         | H154 | H154 | H154 | H154 | H154 | H154 | H154 | H154 | H154  | H154  | H154  | H154  |       |
| L218         | L218 | L218 | L218 | L218 | L218 | L218 | L218 | L218 | L218  | L218  | L218  | L218  |       |
| Q211         | Q211 | Q211 | Q211 | Q211 | Q211 | Q211 | Q211 | Q211 | Q211  | Q211  | Q211  | Q211  |       |
| P2405        | P2405| P2405| P2405| P2405| P2405| P2405| P2405| P2405| P2405  | P2405  | P2405  | P2405  |       |

*One herd represented by 30 animal samples could not be specifically classified as the information given by farmer was “Saanen, Toggenburg, Alpine, Anglo-Nubian”; six animals were described as ‘British goat’; one as ‘mix’ and two were Bagot.

†The variation in codon 240 occurs in different combinations with the other polymorphisms and is also present in all breeds as a polymorphism of the wild-type allele: IRRQ-S240 or IRRQ-P240.

### Table 3: PRNP allele frequencies (%) for two goat surveys in Great Britain

<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Animals</td>
<td>1195</td>
<td>748</td>
</tr>
<tr>
<td>Chromosomes</td>
<td>2390</td>
<td>1496</td>
</tr>
</tbody>
</table>

**Polymorphism**

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Wild-type</th>
<th>R101</th>
<th>G102</th>
<th>S127</th>
<th>M142</th>
<th>R143</th>
<th>S146</th>
<th>H154</th>
<th>Q211</th>
<th>L218</th>
<th>K222</th>
</tr>
</thead>
<tbody>
<tr>
<td>[ % ]</td>
<td>62.7</td>
<td>0.1</td>
<td>0.1</td>
<td>6.4</td>
<td>22.6</td>
<td>1</td>
<td>3.6</td>
<td>0.1</td>
<td>2.3</td>
<td>0.4</td>
<td>0.9</td>
</tr>
<tr>
<td>[ % ]</td>
<td>62.7</td>
<td>0.1</td>
<td>0.1</td>
<td>4.6</td>
<td>15.8</td>
<td>2.1</td>
<td>8.2</td>
<td>0.3</td>
<td>5.1</td>
<td>0.3</td>
<td>0.5</td>
</tr>
<tr>
<td>Change</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>−6.8</td>
<td>−6.8</td>
<td>+1.1</td>
<td>+6.6</td>
<td>+0.2</td>
<td>+2.8</td>
<td>−0.1</td>
<td>−0.4</td>
</tr>
</tbody>
</table>

### Table 4: PRNP genotype frequencies

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Goats (n)</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>G6127</td>
<td>640</td>
<td>90.6</td>
</tr>
<tr>
<td>G5127</td>
<td>61</td>
<td>8.6</td>
</tr>
<tr>
<td>S6127</td>
<td>6</td>
<td>0.8</td>
</tr>
<tr>
<td>I1142</td>
<td>540</td>
<td>72.8</td>
</tr>
<tr>
<td>I1412</td>
<td>167</td>
<td>22.5</td>
</tr>
<tr>
<td>M1412</td>
<td>35</td>
<td>4.7</td>
</tr>
<tr>
<td>NN146</td>
<td>78</td>
<td>41.9</td>
</tr>
<tr>
<td>NS146</td>
<td>90</td>
<td>48.4</td>
</tr>
<tr>
<td>SS146</td>
<td>18</td>
<td>9.7</td>
</tr>
<tr>
<td>RR211</td>
<td>725</td>
<td>92.5</td>
</tr>
<tr>
<td>RQ211</td>
<td>41</td>
<td>5.2</td>
</tr>
<tr>
<td>QQ211</td>
<td>18</td>
<td>2.3</td>
</tr>
<tr>
<td>QQ222</td>
<td>1228</td>
<td>99.3</td>
</tr>
<tr>
<td>QQ222</td>
<td>8</td>
<td>0.7</td>
</tr>
<tr>
<td>K222</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

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purpose. However, given the fact that breeding for resistance for scrapie in goats is an unknown practice in Great Britain, it is unlikely that the selection of farmers may have introduced bias in the authors’ estimates.

Despite the low frequency of the potentially resistant K222 allele and the apparent low incidence of scrapie in British goats, the potential for breeding resistance to scrapie in the goat population is an activity worth promoting among goat keepers, breeding societies and other stakeholders. It is important to note that the scrapie-infected herds detected in Great Britain have presented very high incidence of cases and suffered economic losses caused by decreased productivity, disruption of business operations due to official restrictions and compromised welfare. A genotype-based approach to eradicate scrapie is only an option for sheep according to current EU regulation (EC) No 999/2001 directive. However, given the fact that breeding for resistance for scrapie in goats has been identified does not preclude the possibility to design and implement successful breeding programmes at national level in goats.

Acknowledgements

The authors thank Dorothy Kisielewski for genotyping, Emma Wittmann for project management, Derek Clifford and Ian Dexter for sampling, Catherine McHugh and Stuart Ashfield for general support of the survey and the participating farmers.

Funding

The study was funded by Defra, UK (grant SE2018) to WG, PS, EM, TK, SS, OW and AO–E European Union projects (FODD-CT-2008-36353 [GoatBSE] to WG, PS, JF and 218235 ERA-Net EMID [ID/17–10–2012] to WG, EM, PS, TK, SS, JL, OW and AO–P Dutch Ministry of Economic Affairs (grant WOT-01-002-001:01) to JL), and the Biotechnology and Biological Sciences Research Council (strategic programme grant BB/J004322/1 to the Roslin Institute) to WG and PS.

Ethics approval

This regulated procedure was carried out following ethical advice to the Animal & Plant Health Agency and approval by the UK Home Office under the Animal (Scientific Procedures) Act 1986 (project licence 70/4645).

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Veterinary Record published online January 11, 2016

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