Genomic Prediction of Resistance to Pasteurellosis in Gilthead Sea Bream (Sparus aurata) Using 2b-RAD Sequencing

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ABSTRACT Gilthead sea bream (*Sparus aurata*) is a species of paramount importance to the Mediterranean aquaculture industry, with an annual production exceeding 140,000 metric tonnes. Pasteurellosis due to the Gram-negative bacterium *Photobacterium damselae* subsp. *piscicida* (Phdp) causes significant mortality, especially during larval and juvenile stages, and poses a serious threat to bream production. Selective breeding for improved resistance to pasteurellosis is a promising avenue for disease control, and the use of genetic markers to predict breeding values can improve the accuracy of selection, and allow accurate calculation of estimated breeding values (EBV) of non-challenged animals. In the current study, a population of 825 sea bream juveniles, originating from a factorial cross between 67 broodfish (32 sires; 35 dams), were challenged by 30 min immersion with 1 × 10^5 CFU virulent Phdp. Mortalities and survivors were recorded and sampled for genotyping by sequencing. The 2b-RAD sequencing approach was used to generate genome-wide SNP genotypes for all samples. A high-density linkage map containing 12,085 SNPs grouped into 24 linkage groups (consistent with the karyotype) was constructed. The heritability of surviving days (censored data) was 0.22 (95% highest density interval: 0.11-0.36) and 0.28 (95% highest density interval: 0.17-0.4) using the pedigree and the genomic relationship matrix respectively. A genome wide association study did not reveal individual SNPs significantly associated with resistance at a genome wide significance level. Genomic prediction approaches were tested to investigate the potential of the SNPs obtained by 2bRAD for estimating breeding values for resistance. The accuracy of the genomic prediction models (r = 0.38 - 0.46) outperformed the traditional BLUP approach based on pedigree records (r = 0.30). Overall results suggest that major quantitative trait loci (QTL) affecting resistance to pasteurellosis were not present in this population, but highlight the effectiveness of 2b-RAD genotyping by sequencing for genomic selection in a mass spawning fish species.

KEYWORDS 2b-RAD, Aquaculture, Breeding, High-Throughput sequencing

INTRODUCTION Infectious diseases are a major threat to the profitability, sustainability and welfare status of farmed fish production (Yáñez et al. 2014). Gilthead sea bream (*Sparus aurata*) is one of the most important farmed fish in Mediterranean countries, with an annual production of approximately 146,000 metric tonnes (FEAP http://www.feap.info/default.asp?SHORTCUT=582). Pasteurellosis due to *Photobacterium damselae* subsp. *piscicida* is one of the primary disease problems faced by the sea bream aquaculture industry. High levels of mortality (90 – 100 %) are frequently observed, especially in periods where water temperature rise above 18°C, with larvae and juveniles being most susceptible (Noya et al. 1995;
Selection for improved genetic resistance in aquaculture breeding schemes is a valuable tool to help prevent or reduce disease outbreaks, especially where effective therapeutic agents or vaccines are lacking (Bishop and Woolliams 2014). Moderate to high heritabilities have been estimated for resistance to many common diseases, indicating that rapid genetic progress can be made through selective breeding (Odegård et al. 2011). In addition, recent technological advances in genome-wide sequencing and genotyping technology offer the potential of deriving more accurate estimated breeding values for individual selection candidates, as compared to the classical breeding approach where breeding values are typically estimated at a family level (Goddard and Hayes 2009). The application of genomic data to breeding is particularly valuable for disease resistance, which is typically expensive or impossible to measure on the selection candidates themselves. While marker-assisted selection for major disease resistance loci has been well documented in Atlantic salmon (Salmo salar) breeding programmes (Houston et al. 2008, 2010; Moen et al. 2009), few successful examples exist for other farmed finfish species. Genomic prediction uses genome-wide markers to estimate breeding values, and can deliver significant improvements in selection accuracy compared to traditional pedigree-based approaches, even for traits with a polygenic architecture (Odegård et al. 2014; Tsai et al. 2015, 2016; Vallejo et al. 2016).

Previous studies into the genetic resistance to pasteurellosis in sea bream have detected resistance QTL using microsatellite markers (Antonello et al. 2009; Massault et al. 2010). However, these studies were restricted by the low resolution of microsatellite markers, as compared to the high density SNP genotypes offered by SNP arrays (Houston et al. 2014) or genotyping by sequencing approaches (Davey et al. 2011). Restriction-site associated DNA (RAD) sequencing is a reduced representation high-throughput sequencing technique for the concurrent detection and genotyping of SNP markers in multiplexed samples with a unique nucleotide barcode (Baird et al. 2008). RAD sequencing and similar genotyping by sequencing techniques rely on digestion of the genomic DNA with a restriction enzyme, and subsequent high-depth sequencing of the flanking regions. These techniques have been applied in several studies of aquaculture species to generate high-density linkage maps (Gonen et al. 2014; Palaiokostas et al. 2013b,a, 2015a,b) and perform genome wide association studies (GWAS) in a cost-efficient manner (Campbell et al. 2014). A flexible and easily-streamlined variation of RAD sequencing named 2b-RAD sequencing utilises type IIB restriction enzymes to cleave genomic DNA upstream and downstream of the target site (Wang et al. 2012). In theory, 2b-RAD samples all the endonuclease recognition sites for sequencing, circumventing potential biases that may result from the size selection step in the original RAD protocol (Patriz et al. 2014). 2b-RAD data have also been applied for genetics studies in aquaculture species, for example to test genetic prediction in a limited number of Yesso scallop (Patinopecten yessoensis) families (Dou et al. 2016).

In this study, we used 2b-RAD sequencing to identify and genotype genome-wide SNPs in juvenile sea bream challenged with virulent Phdp bacteria, and recorded for survival time. A high-density SNP linkage map was constructed and a GWAS was performed to test the association between individual loci and resistance to pasteurellosis. Finally, genomic prediction of resistance was tested using several genomic selection models and marker densities to evaluate its potential in selection for improved resistance to pasteurellosis in sea bream.

### MATERIALS AND METHODS

#### Sample collection and preparation

The experimental population used in the present experiment was part of a larger group of juvenile sea bream that were subjected to an experimental challenge with Phdp to estimate heritability of disease resistance as reported (Antonello et al. 2009). Fish were provided by the fish farm Valle Ca’ Zuliani (Monfalcone, Italy). All broodstock fish were originally sampled from wild populations. Fertilized eggs were collected on the same day (year 2006) from natural mass spawning events occurring in four different broodstock tanks, therefore all fish had approximately the same age. Each broodstock tank contained 50-60 fish with a sex ratio 3:1 females:males. Approximately 10,000 eggs were collected, pooled, and kept in a separate tank without any size sorting until 110 days old. All fish were then transferred to the Istituto Zooprofilattico Sperimentale delle Venezie (Legnaro, Italy) for the experimental challenge. Fish were divided into two aerated tanks (A and B) each with 800 l of re-circulating seawater (salinity 35 ppt). Water temperature was maintained at 19°C. After 1 week of acclimation, fish were experimentally infected with a highly virulent strain of Phdp (strain 249/ittio99) as described in Antonello et al. (2009). Mortality was monitored daily for 19 days (Table S1). Mortality levels were nearly identical for both tanks, and only fish from tank A were included in the current study. Fish used in the challenge originated from 67 broodfish (32 sires; 35 dams). As already described (Antonello et al. 2009; Massault et al. 2010), parentage analysis was carried out using a panel of nine microsatellite loci (Table S2).

#### 2b-RAD library preparation and sequencing

A total of 892 2b-RAD libraries (67 parents and 825 juveniles) were constructed by following the protocol reported by Wang et al. 2012 with some modifications (Pecoraro et al. 2016). Template DNA for each individual (500 ng) was digested in 6 µl reaction volume using 1 U AluI at 37°C for 1 h followed by enzyme heat inactivation at 65°C for 20 min. The ligation reaction was performed by combining 5 µl of digested DNA with 20 µl ul of a ligation master mix containing 0.4 µM each of two library-specific adapters (with fully degenerate cohesive ends 5‘-NN-3’) 0.2 mM ATP (NEB) and 1000 U T4 DNA ligase (CABRU, Arcore, Italy). Ligation was carried out at 16°C for 3 h with subsequent heat inactivation for 10 min at 65°C.

Sample-specific barcodes were designed through a Barcode generator program (http://comalab.genomcenter.ucdavis.edu/index.php/Barcode_generator). 50 µl PCR reactions were prepared containing 12 µl of ligated DNA product, 0.2 µl M of each primer, 0.3 mM dNTP, 5X Phusion HF buffer and 2 U Phusion high-fidelity DNA polymerase (NEB). Each library was PCR amplified using the following conditions: 13 cycles of 95°C for 5 s, 60°C for 20 s and 72°C for 5 s.

Adaptor and primer sequences were those reported in Wang et al. (2012). PCR products were purified using the SPRSelect purification kit (Beckaman Coulter, Pasadena, CA, USA) and quantified through a Qubit 2.0 Fluorometer (Invitrogen). The quality of all amplicon libraries was checked at 1.8% agarose gel. Additionally, the quality of 10% of randomly-selected libraries, was also assessed by running them on an Agilent 2100 Bioanalyzer.

Individual libraries were pooled into equimolar amounts by adopting two different multiplexing strategies for parents (24 libraries / pool) and offspring (48 libraries / pool). The quality of each pool was verified on Agilent 2100 Bioanalyzer. Finally, pooled libraries were sequenced on an Illumina HiSeq2500 platform (Illumina, San Diego, CA, USA) using 50 base single-end sequencing.
Table 1  Gilthead sea bream linkage map

<table>
<thead>
<tr>
<th>LG</th>
<th>No. markers</th>
<th>Length (cM)</th>
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<tbody>
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<td>607</td>
<td>202</td>
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<tr>
<td>2</td>
<td>602</td>
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<tr>
<td>3</td>
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<td>24</td>
<td>366</td>
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</tr>
<tr>
<td>Total</td>
<td>12,089</td>
<td>3,899</td>
</tr>
</tbody>
</table>

(v3 chemistry).

Genotyping RAD alleles
Quality and adapters trimming of sequenced reads were performed by running a customized script (Pauletto et al. 2016; Pecoraro et al. 2016), obtaining 34-bp long fragments. SNP calling was performed using STACKS v1.23 (Catchen et al. 2011).

For each family/cross, individual genotypes were constructed using components of the STACKS pipeline as follows: i) for each individual, ustacks program was employed for building loci from all QC passed reads using the following parameters −m10 − M2 − N3 for parents and −m5 − M2 − N3 for offspring, ii) a catalog of loci unique for all families/crosses was constructed by using all parents’ reads on cstacks program, then iii) each set of parents / offspring per cross was matched separately against such a catalogue (sstacks program) followed by genotype assignment by setting the following parameters on genotypes program: −c − min_hom_segs 7 − max_het_seqs 0.05.

Unique tags created by STACKS were mapped against a draft assembly of Sparus aurata genome (L. Bargelloni, personal communication). Mapping analysis was carried out by means of CLC Genomic Workbench 7.5 with stringent criteria (length fraction = 0.9; similarity fraction = 0.9; nonspecific match handling= ignore).

Linkage map construction
Linkage map construction was performed using Lep-Map v2 (Rastas et al. 2013). Quality control was performed for each full sibling family by excluding SNPs with minor allele frequency below 0.05 and those deviating from expected Mendelian segregation (P < 0.001). Linkage groups were formed using a minimum LOD threshold value of 8 in the ‘SeparateChromosomes’ module, allowing a maximum distance between consecutive SNPs of 50 cM. Marker order within each linkage group was performed using the ‘OrderMarkers’ module, where the likelihood of marker order is computed by using a hidden Markov model (Rastas et al. 2013). Map distances were calculated in centiMorgans (cM) using the Kosambi mapping function.
Trait definition and heritability estimation

Heritability of surviving days was estimated with the R/BGLR software (Pérez and de Los Campos 2014) using both the pedigree-based and the genomic relationship matrix. The animal model was applied:

\[ y = \mu + Zu + \epsilon, \]

where \( y \) vector of recorded phenotypes (days to death; animals surviving at the end of the experiment treated as missing values sampled from corresponding truncated normal distribution with the resulting values being higher or equal to 20), \( \mu \) vector of the intercept, \( Z \) incidence matrix relating phenotypes with the random animal effects, \( u \) is the vector of animal effects \( N(0, \sigma^2_G) \) with either \( A \) corresponding to the pedigree-based relationship matrix or \( G \) the corresponding genomic relationship matrix, and \( \epsilon^2 \) the additive genetic variance. Finally, \( \epsilon \) is a vector of residuals. The \( G \) matrix was estimated according to VanRaden (2008) using the kin function of the R/synbreed package (Wimmer et al. 2012) (File S1). The additive genetic variance was estimated by applying Markov Chain Monte Carlo (MCMC) using a prior following the inverse-scaled \( \chi^2 \) distribution (df=5), using 10M iterations out of which the first 10% were discarded, and values were stored every 1,000 iterations thereafter. Convergence of the resulting posterior distribution was assessed both visually (inspecting the resulting MCMC plots) and analytically with the Geweke diagnostic using R/boa v1.1.7 (Smith 2007). Heritability for the number of surviving days was estimated using the following formula:

\[ h^2 = \frac{\sigma^2_G}{\sigma^2_G + \sigma^2_e}, \]

where \( \sigma^2_G \) the estimated additive genetic variance and \( \sigma^2_e \) the residual variance.

Genome wide association analysis (GWAS)

To test the association between individual SNPs and resistance to pasteurellosis (measured as surviving days), a GWAS was performed using R/rrBLUP (Endelman 2011). The mixed model applied (Yu et al. 2006) had the following format:

\[ y = Xa + Zu + \epsilon, \]

where \( y \) is the vector of the phenotypes (surviving days or overall survival), \( a \) is the vector of unknown marker effects, \( u \) is the vector of random animal effects \( N(0, \sigma^2_G) \) and \( \epsilon \) is the vector of residuals. The matrix \( G \) represents the genomic relationship matrix as described above, and \( \sigma^2_G \) the additive genetic variance estimated using REML. \( X \) and \( Z \) are incidence matrices relating \( y \) to \( a \) and \( u \), respectively. According to the above model additive SNP effects are treated as fixed effects, with the inclusion of the random animal effect to decrease spurious associations due to (genomic) relationships between the animals (Yu et al. 2006). The genome-wide significance threshold for the estimated additive SNP effects was calculated using a Bonferroni correction (0.05 / N), where \( N \) represents the number of QC-filtered SNPs across the entire genome.

Genomic Prediction

A genomic prediction approach was conducted to quantify the accuracy of the breeding values estimated using the SNP markers to predict the phenotypic trait values (surviving days). SNPs with > 15% missing genotypes were removed in order to minimize impact of imputed genotypes, since used software cannot handle missing genotypes. Missing values of the remaining SNPs were imputed using R/synbreed (Wimmer et al. 2012). Genomic breeding values were estimated using rrBLUP, BayesA, BayesB (Meuwissen et al. 2001) and BayesC (Habier et al. 2011) models using the R/BGLR (Pérez and de Los Campos 2014) software. The above models differ in regard to the prior distribution of the marker effects. Briefly, rrBLUP by using the Gaussian distribution, induces homogenous shrinkage across markers, while in BayesA the usage of a scaled-t distribution induces marker size effect shrinkage allowing for variable marker effect sizes. Models BayesB and BayesC also perform variable selection, with the difference between the two being the usage of a scaled-t or a Gaussian prior density respectively (Meuwissen et al. 2001; de los Campos et al. 2013). To compare the accuracy of genomic EBVs to the pedigree-based EBVs, pedigree-based BLUP (PBLUP) (Henderson 1975) was applied to calculate breeding values using the same software. The general form of the fitted models was the following:

\[ y = \eta + \epsilon, \]

The linear predictor \( \eta \) in the case of rrBLUP, BayesA, BayesB and BayesC had the following general form:

\[ \eta = 1\mu + X_1\beta_1, \]

where \( \mu \) is the intercept, \( X_1 \) the design matrix relating the phenotypes to the markers, \( \beta_1 \) is the vector of marker effects with corresponding priors depending on the model used. Marker coding followed the format where the heterozygotes were coded as 1 and the two alternate homozygotes as 0 and 2. The linear predictor \( \eta \) in the case of pedigree BLUP had the following form:

\[ \eta = 1\mu + u, \]

where \( u \) is the animal random effect vector \( N(0, \sigma^2_G) \) with the matrix \( A \) representing the pedigree estimated relationship matrix. The parameters of the above models were estimated through MCMC (110,000 iterations; burn-in: 10,000; thin: 100).

Assessment of the accuracy of breeding value predictions was conducted according to the following procedure. The data set was randomly split into a training set (\( n = 578 \) animals) and a validation set (\( n = 200 \)). The above was repeated 100 times with the obtained prediction accuracies being adjusted for the trait heritability for each tested model. The genomic estimated breeding values (GEBV) for each replicate of the validation data set were estimated as:

\[ \text{GEBV} = Xu, \]

where \( X \) the incidence matrix relating GEBV with SNP genotypes and \( u \) the vector of estimated SNP effects from the corresponding training data set. The accuracy of the estimated GEBV was approximated as:

\[ r = (\text{GEBV}, y) / h, \]

where \( y \) the vector of recorded phenotypes and \( h \) the square root of the heritability. In all tested scenarios the heritability estimated using the genomic relationship matrix was used. Reported accuracies for each tested model refer to the mean accuracy of the above-mentioned 100 replicates of validation data sets. In order to test the predictive ability of varying SNP densities, the above procedure was followed using (i) SNPs spaced > 1 cM apart (2,614 SNPs) or (ii) SNPs spaced > 5 cM apart (705 SNPs) on the linkage map.

Data Availability

Raw reads were deposited in EBI repository under project ID SRP081498. Table S1 contains the phenotypic data. Table S2 contains the pedigree. Table S3 and S4 contain summary of obtained reads for parental and offspring samples respectively. Table S5 contains the location of the linkage map SNPs. Table S6 contains the genotypic data. File S1 contains customed R script used for genomic prediction.
**RESULTS**

**Disease challenge**

The challenged population consisted of 75 full sibling families with a mean family size of 10, originating from a factorial cross between 67 broodfish (32 sires; 35 dams). The largest full-sib family consisted of 114 animals, while the smallest had only 2 animals (3 full-sib families). The overall survival at the end of the pasteurellosis challenge was 4.7%. Observed mortality levels showed three distinct peaks on day 7 (10.4 % loss), day 11 (14 % loss) and day 15 (5.7 % loss) followed by a steady reduction in daily mortality rate (Figure 1).

![Figure 1](image1.png)

**Genotyping RAD alleles**

The mean number of raw reads was 8.76 million (M) and 4.51 M while the number of reads passing QC was 7.74 M (88 %) and 3.69 M (81 %) for parents (Table S3) and offspring (Table S4), respectively. The STACKS catalogue consisted of 202,598 unique 2b-RAD loci, of which 73,876 contained at least one SNP in the parents (EBI repository SRP081498). To confirm the identity of loci created by STACKS, the 202,598 tags of the catalog were mapped against a *Sparus aurata* draft reference genome assembly (unpublished data). A high percentage of 2b-RAD loci were successfully mapped, with 93.5 % of tags showing a unique match to the reference genome. In order to maximise the number of informative SNPs and minimise the amount of missing or erroneous data, RAD-tags that were retrieved in at least 75% of the samples, and that carried only one or two SNPs were retained. 48 animals with missing data > 30% were excluded from subsequent analysis. A total of 21,974 putative SNPs were finally used for construction of the genetic map from 777 disease challenged offspring (genotypic missing data < 30%) and their corresponding 67 parents.

**Linkage Map**

The linkage map consisted of 12,085 SNPs that were grouped into 24 linkage groups, in accordance with gilthead sea bream karyotype, with a total map length of 3,899 cM (Table 1, Figure 2; Table S5). The remaining SNPs (9,889) either failed to pass QC filters, or were not placed on the resulting linkage groups during mapping, and these were discarded. The female and male maps were comparable with total lengths of 3,822 cM and 4,010 cM respectively. The number of SNPs per chromosome ranged between 366 to 607 (mean = 503; s.d = 53), while linkage group length ranged between 115 and 202 cM (mean = 162; s.d = 26). The correlation between number of SNPs and corresponding chromosome map length was 0.74 (n = 24 linkage groups).

**Heritability estimation and Genome wide association analysis (GWAS)**

The heritability of surviving days (censored data) was 0.22 (95% highest density interval: 0.11-0.36) and 0.28 (95% highest density interval: 0.17-0.4) using the pedigree and the genomic relationship matrix respectively of the 777 disease challenged offspring. No SNPs surpassed the Bonferroni-corrected genome-wide significance threshold (\(P = 4.1 \times 10^{-6}\)). The SNPs with the lowest p-values (\(P < 10^{-3}\)) were located in linkage groups 1 - 3, 10, 17, 20 and 21 (Figure 3).

![Figure 2](image2.png)

**Genomic Prediction**

Genomic prediction was tested as a means of obtaining breeding values, and compared to prediction using a pedigree-based approach (File S1). The prediction was conducted using genotype information from 11,239 SNP markers (passed QC filters set for genomic prediction) for the 777 disease challenged animals that were randomly split in training (n=578) and validation (n=200) datasets. The application of all the genomic prediction models resulted in higher accuracies than those achieved using pedigree-based BLUP (Table 2). Prediction accuracy with PBLUP was 0.3, and versus 0.38 to 0.46 for the genonic prediction models, with highest accuracy being observed using the BayesA method. Prediction accuracy dropped when more sparse SNP marker datasets were used with the last scenario (utilizing SNPs > 5cM apart) giving similar accuracies with the ones obtained by using the pedigree-based BLUP.

In the scenario of utilizing only SNPs located more than 1 cM apart on the linkage map (2,614 SNPs) accuracies ranged between 0.3 and 0.36, with the highest accuracy obtained using rrBLUP. For the dataset utilizing only SNPs located more than 5 cM apart (705 SNPs) accuracies ranged between 0.29 - 0.31 with highest accuracies obtained using rrBLUP and BayesA.

**DISCUSSION**

Gilthead sea bream (*Sparus aurata*) is a farmed species of paramount importance for Mediterranean aquaculture. While vaccines can offer some protection against pasteurellosis, the low immune competence observed in larval and juvenile stages renders this protection only temporary (Antonello et al. 2009). Breeding for...
The previous map of (Tsigenopoulos et al. 2014) has a total length of 1769.7 cM, which may reflect the larger number of markers used in the current study. This trend of increase in map distance with increased marker density was observed with previous sea bass (Dicentrarchus labrax) linkage maps (Chistiakov et al. 2005, 2008; Palaiokostas et al. 2015b).

The estimated heritability of resistance was moderate (0.22 and 0.28 for the different models) compared to those previously reported for disease resistance traits in various aquaculture species (Odegård et al. 2011). Nonetheless, successful implementation even in the case of low heritability traits in breeding programs is still possible, as demonstrated in livestock (Heringstad et al. 2003). Also, since heritability of mortality traits are frequency dependent, with maximal values reported at intermediate mortality levels (Bishop and Woolliams 2014), the low survival rate in the current study may have resulted in an underestimate, and analysis of additional challenge and field data is merited.

The GWAS results pointed to a polygenic or oligogenic genetic architecture for resistance to pasteurellosis, with no genome-wide significant QTL identified, with the lowest P values indicative of putative suggestive QTL on linkage groups 1 - 3, 10, 17, 20 and 21. Unfortunately the lack of an integrated linkage map for sea bream does not allow direct comparison of the previously described resistance QTL (Massault et al. 2010) with the current study. Furthermore no major QTL were identified in the above study, with the largest QTL for surviving days explaining approximately 4% of the phenotypic variance. The fact that genomic prediction using all tested models gave similar prediction accuracies, a phenomenon often observed in the study of polygenic traits (Meuwissen et al. 2001; Kizilkaya et al. 2010) seem to support the hypothesis of polygenic resistance. Nevertheless, since the selected priors will influence the output of the Bayesian models (Gianola 2013), interpretation of genetic architecture based on these results should be treated with caution. Additionally, the moderate sample size in the current study is likely to limit the statistical power to detect small to medium size effect QTL. Further, the low mortality level precludes effective estimation of genetic parameters for survival per se, and the genetic correlation between survival time (days to death) and overall survival is unknown.

The results from the genomic prediction approach were encouraging for practical implementation of selective breeding for genetic resistance in sea bream, with the genomic prediction models outperforming traditional BLUP. The advantage of the genomic-based models was retained also when only SNPs at 1cM location intervals were used, which may be useful for reducing genotyping costs and improving cost-effectiveness. However, due to the limited number

<table>
<thead>
<tr>
<th>Table 2 Genomic prediction accuracies for surviving days</th>
</tr>
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<tbody>
<tr>
<td>Model</td>
</tr>
<tr>
<td>pBLUP</td>
</tr>
<tr>
<td>rrBLUP</td>
</tr>
<tr>
<td>BayesA</td>
</tr>
<tr>
<td>BayesB</td>
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<tr>
<td>BayesC</td>
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</table>

<sup>a</sup> Analysis included 12,085 SNPs
<sup>b</sup> Analysis included 2,614 SNPs
<sup>c</sup> Analysis included 705 SNPs
<sup>d</sup> standard error
of families in the current study, the training and validation sets contain closely related animals, which will increase the accuracy of prediction. Additional testing of genomic prediction at varying marker densities on separate and preferably larger populations would be required to ascertain the appropriate density for commercial application of genomic selection. The benefit of genomic prediction over a pedigree-based approach is likely to be due to the ability to capture within-family genetic variation. In mass-spawning species such as sea bream, in which family size and structure is difficult to control, this approach is likely to be particularly advantageous. Overall, the current study demonstrates that SNP markers generated via 2b-RAD are effective at capturing the genetic variation in a complex trait in a sea bream breeding population. This approach is likely to be useful in other species with less-developed genomic tools, and provides further evidence that incorporation of genomic selection is likely to result in significant improvement in selection accuracy and genetic gain compared to traditional family selection in aquaculture breeding.

Conclusion

2b-RAD sequencing was applied to investigate genetic resistance to pasteurellosis in gilthead sea bream. The SNP data generated were applied to create the first high-density linkage map for sea bream. Only suggestive QTLS were detected, implying that resistance to pasteurellosis has an oligogenic-polygenic architecture for the studied population. Genomic prediction using the 2b-RAD genotype data was effective, with substantial improvement in prediction accuracy over the pedigree-based model. This highlights the utility of genotyping by sequencing for genomic prediction of disease resistance in aquaculture species, and its potential to apply genomic selection in commercial breeding programs.

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AUTHOR CONTRIBUTIONS

SF and RF carried out DNA extractions, 2bRAD library preparation and sequence data processing. LB conceived the study, contributed to designing the experiments, analysing the data. CP and RH designed the quantitative genetic analyses. CP conducted the genetic analysis. All authors contributed to drafting the ms. bibilography example bibliography

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