Long-term responders on olaparib maintenance in high-grade serous ovarian cancer: Clinical and molecular characterization

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Long-term responders on olaparib maintenance in high-grade serous ovarian cancer: 
Clinical and molecular characterization


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*Contributed equally to this work

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Keywords: Olaparib, maintenance, gynaecological cancers: ovarian, BRCA1/BRCA2, biomarkers and intervention

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Conflicts of interest: AMO has acted as principal investigator on an AstraZeneca-sponsored trial. JL, UM, CG, and SK have participated in AstraZeneca advisory boards. JL has received travel grants from AstraZeneca. CG has received honoraria from AstraZeneca. ZL, BD, SR, DRH, MJO’C, CJB, and TWH are employees of AstraZeneca and own AstraZeneca stock. None of the other authors have conflicts of interest to declare.

Word count: 3146/5000
Statement of translational relevance: This is the first study to report long-term responders to a poly(ADP-ribose) polymerase (PARP) inhibitor, with response durations of >2 years, in the context of platinum-sensitive, high-grade serous ovarian cancer. Based on extensive molecular profiling, the durable long-term responses were multifactorial and driven by germline and somatic BRCA1/2 mutations. The majority of patients in the long-term responders group harboured homologous recombination repair deficiency, with particular enrichment for mutations in BRCA2, compared with short-term responders. This pilot study also highlights potential non-BRCA1/2-mutated patients who demonstrated durable clinical benefit with the PARP inhibitor olaparib and, together with an ongoing larger cohort study (NCT02489058), seeks to identify additional molecular characteristics that can predict susceptibility to olaparib. Further investigation may allow outreach to more patients for treatment with olaparib.

Statement of translational evidence word count: 125/150
ABSTRACT

Purpose: Maintenance therapy with olaparib has improved progression-free survival in women with high-grade serous ovarian cancer (HGSOC), particularly those harboring BRCA1/2 mutations. The objective of this study was to characterize long-term (LT) versus short-term (ST) responders to olaparib.

Design: A comparative molecular analysis of Study 19 (NCT00753545), a randomized Phase II trial assessing olaparib maintenance after response to platinum-based chemotherapy in HGSOC, was conducted. LT response was defined as response to olaparib/placebo >2 years, ST as <3 months. Molecular analyses included germline BRCA1/2 status, three-biomarker homologous recombination deficiency (HRD) score, BRCA1 methylation, and mutational profiling. Another olaparib maintenance study (Study 41; NCT01081951) was used as an additional cohort.

Results: 37 LT (32 olaparib) and 61 ST (21 olaparib) patients were identified. Treatment was significantly associated with outcome (P<0.0001), with more LT patients on olaparib (60.4%) than placebo (11.1%). Long-term sensitivity to olaparib correlated with complete response to chemotherapy (P<0.05). In the olaparib LT group, 244 genetic alterations were detected, with TP53, BRCA1 and BRCA2 mutations being the commonest (90%, 25% and 35%, respectively). BRCA2 mutations were enriched among the LT responders. BRCA methylation was not associated with response duration. High Myriad HRD score (>42) and/or BRCA1/2 mutation was associated with LT response to olaparib. Study 41 confirmed the correlation of LT response with olaparib and BRCA1/2 mutation.

Conclusions: Findings show that LT response to olaparib may be multifactorial and correlated with homologous recombination repair deficiency, particularly BRCA1/2 defects. The type of BRCA1/2 mutation warrants further investigation.
Abstract word count: 246 (max 250)
INTRODUCTION

Epithelial ovarian cancer remains the leading cause of death from gynecologic malignancies, with high-grade serous ovarian cancer (HGSOC) the most common histological subtype (1). Following cytoreductive surgery and platinum-based chemotherapy, ~70% of HGSOC patients relapse despite an initial response to therapy (2). Given the practical certainty of recurrence in relapsed HGSOC, a maintenance approach was proposed to delay subsequent progression via continuation of treatment beyond cytotoxic chemotherapy. The poly(ADP-ribose) polymerase inhibitor (PARPi) olaparib (Lynparza™) is the first validated molecularly targeted agent for HGSOC. Ledermann et al demonstrated the benefit of maintenance olaparib after response to platinum-based chemotherapy in women with HGSOC recurrence (NCT00753545) (3). Patients were randomized to placebo or olaparib, introduced within 8 weeks of completion of the last dose of platinum-based chemotherapy and maintained until progression. Olaparib maintenance was associated with significantly longer progression-free survival (PFS; median 8.4 vs 4.8 months; hazard ratio [HR] 0.35; 95% confidence interval [CI] 0.25 to 0.49; \( P < 0.001 \); data maturity 58%). Patients with a BRCA1 mutation (BRCA1m) or a BRCA2 mutation (BRCA2m) had significantly greater PFS benefit with olaparib maintenance than those receiving placebo (median 11.2 vs 4.3 months; HR 0.18; 95% CI 0.10 to 0.31; \( P < 0.0001 \); data maturity 52%) (4). Overall survival improved by 3 months, which was not significant; possibly because of post-trial cross-over whereby 22.6% of placebo patients received PARPi in subsequent clinical trials following progression on placebo. Olaparib is now approved in Europe, Australia and Canada for the maintenance treatment of women with relapsed, BRCA1/2m-positive (germline or somatic) HGSOC who have had complete or partial response to platinum-based chemotherapy (5). Olaparib is also approved in the USA for monotherapy in patients with germline BRCA1/2m advanced HGSOC who have been treated with ≥3 prior lines of chemotherapy (6). Proteins encoded by the BRCA1/2 genes are crucial effectors of double-strand-break DNA repair (7); as a
result, \textit{BRCA1/2}m carriers are known to be highly responsive to DNA-damaging agents, including platinum-based chemotherapies (8, 9) and PARPi (8), although mechanisms of action and resistance to PARPi are not fully understood (10). Other than deleterious \textit{BRCA1}m or \textit{BRCA2}m, there are no predictive biomarkers for sensitivity to olaparib. Data from The Cancer Genome Atlas (TCGA) and functional studies indicate that approximately half of HGSOC are homologous recombination repair (HR) defective (11-13). PARP is involved in the repair of single-strand DNA breaks, and its inhibition can result in replication-associated double-strand breaks. Such HR-deficient cells as those found in \textit{BRCA1/2}-mutated tumors, whether repaired via error-prone pathways or persistent without repair, cause further genetic instability and can lead to cell death (14).

We hypothesize that studying HGSOC patients with prolonged response to olaparib may identify additional biomarkers of response beyond \textit{BRCA1/2}m. The objective of this study was to identify and characterize long-term responders to olaparib maintenance in comparison with short-term responders in terms of clinical and molecular profile to pinpoint additional markers of response and explore potential resistance factors.
METHODS

Patient population

We investigated the molecular and clinical characteristics of long- and short-term responders randomized to maintenance olaparib or placebo in the phase II, randomized, double-blind study of olaparib in patients with platinum-sensitive relapsed high-grade serous ovarian cancer following treatment with ≥2 lines of platinum-based chemotherapy (NCT00753545; Study 19) (3). This trial enrolled 265 patients, with 136 patients assigned to olaparib and 129 to placebo. Given that the greatest PFS benefit was at 11.2 months, we defined long-term (LT) responders, whatever their BRCA1/2 status, as having a double PFS benefit, i.e. of >2 years. Short-term (ST) responders were defined as having PFS of <3 months, given the PFS observed in the placebo group of 4.8 months.

A second comparison/confirmation cohort from the open-label, randomized, phase II study of olaparib/carboplatin/paclitaxel with olaparib maintenance versus carboplatin/paclitaxel/observation in patients with platinum-sensitive recurrent HGSOC (NCT01081951; Study 41) was evaluated (15). In the combination phase, 156 patients were treated (81 in the olaparib-plus-chemotherapy group and 75 in the chemotherapy-alone group) and 121 continued to maintenance or no further treatment (66 in the olaparib-plus-chemotherapy group and 55 in the chemotherapy-alone group). Given that patients in this study received olaparib with chemotherapy, the ST responders were defined as patients with PFS of <6 months, taking into account the time over which olaparib was administered with chemotherapy to assess the maintenance period.

Data collection

Clinical trial data were prospectively obtained for all treated patients (Table 1) and archival tumor specimens, predominantly taken at the time of initial diagnosis, from patients who
consented to further use of their specimens. Previously performed germline *BRCA1/2* testing (Myriad Genetics®) results were obtained.

**Molecular investigations**

Assessment of somatic *BRCA1/2* mutations (Myriad® tumor testing and Foundation Medicine®), determination of the three-biomarker HR deficiency (HRD) score (myChoice® HRD test, Myriad Genetics) (16), and *BRCA1* methylation (Myriad tumor testing) were conducted (17). HRD scores were determined using an assay that calculates scores for whole-genome tumor loss of heterozygosity (LOH), telomeric allelic imbalance (TAI) and large-scale state transition (LST). The HRD score is the sum of the three scores, where ≥42 is a predefined threshold validated as a high HRD score (16). The Methyl-Profiler DNA Methylation PCR Assay System (SABiosciences, Valencia, CA, USA) was used to quantify methylation levels, with a used quantitative cut-off of promoter methylation of >10%. HR deficiency status was determined on the basis of the combination of the dichotomized HRD score using the predefined HRD threshold and tumor *BRCA1/2* status (scored as mutated if deleterious or suspected deleterious mutations in germline or somatic *BRCA1/2* were present). Therefore, HR deficiency was defined as a high HRD score (≥42) and/or *BRCA1/2* mutated tumor (16).

Mutational profile determined by the Foundation Medicine T5 panel (entire coding sequence of 287 cancer-related genes plus select introns from 27 genes) and other genetic alterations (deletions and functional rearrangements) were assessed for archival tumor tissue (18). Classification of *TP53* mutations (*TP53m*) was determined using the WHO International Agency for Research on Cancer Database.

**Pathology review**
Pathology review for tumor cell content was performed for all patients with molecular analyses, whereas pathology review of histological subtype by an expert pathologist in gynecologic cancer, blinded to cohort or outcome, was only performed for patients with remaining tissue.

**Statistical analysis**

Descriptive analyses from all LT and ST responders to olaparib/placebo were assessed for statistical significance. Fisher’s exact or chi-squared tests were used to test for associations between individual explanatory variables and response duration (long vs short term) as appropriate. SPSS and Excel were used for all analyses. Given the exploratory data analysis, no type 1 error correction was performed.
RESULTS

Clinical data

Data were collected from 265 total patients on study for the patients with LT and ST responses to olaparib therapy or placebo as part of Study 19. Thirty-seven patients were identified as LT responders, of whom 32 (86.5%) had received olaparib. Of the 61 ST patients identified, 21 (34.4%) had received olaparib. The main characteristics are summarized in Table 1 and Table 2. LT responders on olaparib (32 patients) had a median of three prior lines of therapy, with one-third having relapsed within 6–12 months of their penultimate platinum-based chemotherapy. Of the LT responders on olaparib, 44% (14/32) had partial responses to their most recent platinum-based chemotherapy, with residual disease evident at the time of olaparib maintenance (Table 1). The other 18 of 32 (56%) LT responders on olaparib had complete response to the most recent platinum-based chemotherapy, in comparison with only five of 21 (24%) olaparib ST responders (Table 1). Complete response to platinum-based chemotherapy at the time of olaparib maintenance was associated with LT response to olaparib ($P=0.026$, univariate analysis). The treatment-free interval following the penultimate platinum-based chemotherapy did not correlate with LT response (Supplementary Table 1). Sixty-five of 98 archival tumour samples (37 LT and 61 ST) were from primary tumours and nine of 98 were from metastases (this information was unavailable for the other 24 patients).

Receipt of olaparib over placebo was significantly associated with LT response ($P<0.0001$; 32/53, 60.4% vs 5/45, 11.1%). More patients treated with olaparib were LT than ST responders ($P=0.052$).
**Molecular analysis**

**BRCA1/2 status (germline and somatic)**

From the LT responders, 27/37 (73%) had loss-of-function mutations in *BRCA1/2*. Among the 32 LT olaparib responders, *BRCA1*m and *BRCA2*m were found in 10 and 13 patients, respectively, with one patient showing deleterious mutations in both *BRCA1* and *BRCA2* (Table 2 and Table 3). All five LT responders receiving placebo were *BRCA1/2*m positive (Table 2). A greater number of *BRCA1/2*m was found in LT responders compared with ST responders (Table 3). In contrast, among the 21 olaparib ST responders, 10 were found to carry a deleterious *BRCA1/2*m (7 of 10 were in *BRCA1*; Table 2 and Table 3).

We further analyzed the type and location of *BRCA1/2*m in the olaparib cohort between LT and ST responders (Figure 1 and Supplementary Figure 1). Of the 17 patients on olaparib who had *BRCA1*m (LT and ST), nine had founder mutations (E23fs* or Q1756fs*), whereas only one patient had a founder *BRCA2*m (S1982fs*; ST responder group). Interestingly, among patients on olaparib, mutations in the DNA-binding domains of *BRCA1* (n=1) or *BRCA2* (n=4) were only seen in the LT responder group.

*BRCA1* methylation status was available for 27/37 (73%) LT responders, all of whom were negative. In contrast *BRCA1* methylation status was available for 42/61 (69%) ST responders, eight of whom (19%) had *BRCA1* methylation. Methylation of *BRCA1* was not associated with LT olaparib response.

**Homologous recombination repair deficiency**

No significant difference was seen between LT and ST responders according to the HRD score (Table 3). Most of the patients enrolled in the study were characterized by HR-deficient status (81% of the LT and ST on olaparib). Among data available for 26 of the 32 (81%) LT responders on olaparib, 25 patients (96%) had HR deficiency, in contrast to 76% of the ST
olaparib responders (Table 3). Of the 21 patients with high HRD score in the LT responder group, three were BRCA1/2 wild type. Of the nine ST patients with a high HRD score, two (22%) were BRCA1/2 negative in the ST responder group. However, a significant number of HRD scores are missing for the BRCA1/2 wild-type group (Table 2).

Taking together high HRD score (>42) and/or BRCA1/2 mutated tumor, there was a trend towards HR deficiency in LT versus ST responders, with 96% of LT responders and only 76% of ST responders showing HR deficiency (P=0.07).

**Next-generation-sequencing analysis**

Next-generation sequencing was performed on archival tumor DNA from 44 patients (28 [87%] LT and 16 [76%] ST responders on olaparib). From 44 patients, over 600 identified alterations were classified by type and likely functional significance by Foundation Medicine (Supplementary Table 2). A total of 242 likely functional generic alterations in 99 genes were found, with TP53, BRCA1 and BRCA2 mutations being the most common (89%, 34% and 36% of patients, respectively; Figure 2 and Supplementary Table 2).

TP53 signalling was considered aberrant in most cases (90%), as expected for the HGSOC histology subtype (27 of 28 patients with available data analyzed in the LT responder group, 12 of 16 in the ST responder group; Supplementary Figure 1). The patient with no TP53m in the LT responder group had pathology reviewed and HGSOC confirmed. Of the four patients with no TP53m in the ST responder group, two had confirmed HGSOC. Other types of mutations and gene amplifications were observed, particularly in genes encoding proteins involved in DNA repair and damage response, regulation of cell cycle, apoptosis, and MAPK/PI3K signalling (Figure 2 and Supplementary Table 2). Furthermore, four patients had alterations in PTEN (2 homozygous deletions, 1 somatic variant and 1 functional intergenic truncation), all of whom were LT responders on olaparib. These PTEN alterations co-occurred
with $BRCA1/2$ mutation for three of the patients, only one LT patient had a $PTEN$ mutation in the absence of a $BRCA1/2$ mutation. In contrast, no $PTEN$ alterations were seen in the ST responder group (Supplementary Table 3). Interestingly, three patients randomized to the placebo arm had $PTEN$ alterations and were ST responders (Supplementary Table 3).

**Validation cohort**

LT and ST responders from Study 41 were identified and analyzed (Figure 3 and Supplementary Table 4). In total, 19 LT responders were identified and all were in the olaparib arm. Olaparib maintenance was also significantly associated with LT responders ($P<0.0001$). $BRCA1/2m$ was statistically correlated with LT response to olaparib (Supplementary Table 4). $BRCA1m$ and $BRCA2m$ were observed in six and five LT responders, respectively. The 11 ST patients were all $BRCA1/2$ wild-type.
DISCUSSION

There are limited data describing ovarian cancer patients who experience prolonged benefit from PARP inhibition, other than evidence for the role played by deleterious mutations in *BRCA1/2*. Defective DNA repair via homologous recombination repair deficiency is a fundamental vulnerability in HGSOC and can be exploited with PARPi, such as olaparib, by induction of cancer-specific synthetic lethality (19). Examination of broader clinical and molecular data of extreme responders may uncover potential biomarkers of response (20). We identified LT and ST responders to PARPi from women with HGSOC entered into two olaparib maintenance trials, Studies 19 and 41, and have molecularly characterized these exceptional responders. We found that 32 and 19 patients with recurrent platinum-sensitive HGSOC achieved LT (defined as >2 years) response to olaparib maintenance as part of the studies, respectively. Germline and somatic *BRCA1/2*^m^ were observed to be associated with LT response to olaparib, with an enrichment of *BRCA2*^m^ in the LT olaparib responders, compared with the frequency of *BRCA1*^m^ and *BRCA2*^m^ in HGSOC detected at the start of the trial and the *BRCA1/2* Ratio observed in the general population (21). Our finding is in agreement with data suggesting differences in outcome and response to therapy between *BRCA1* and *BRCA2* genotypes (22). Previous studies have shown that *BRCA2*^m^ is associated with prolonged survival in invasive epithelial ovarian cancer (23). In 2012, Liu et al showed that the presence of a *BRCA2*^m^ was associated with longer survival and better therapy response than a *BRCA1*^m^ in HGSOC (24). Many LT olaparib responders had a *BRCA2*^m^ in the *RAD51*-binding domain, described as a frequent site of *BRCA2*^m^ by TCGA (11), but also in DNA-binding sites (Figure 3). As such, mutations in the *RAD51* region are expected to attenuate or abolish interactions with *RAD51*, resulting in failure to load *RAD51* to DNA-damage sites (24). Our data also suggest that silencing of *BRCA1* through promoter methylation does not result in improvement in response to platinum-based chemotherapy or to sequential chemotherapy and maintenance olaparib therapy,
as previously suggested by TCGA and other publications showing a lack of survival benefit and correlation with platinum sensitivity (11, 25).

However, our study did not identify a potential mechanism involved in the small group of BRCA1/2 wild-type patients who had a LT benefit to olaparib maintenance, currently not eligible for olaparib in clinical practice. Beyond BRCA1/2m, there have been a number of mechanisms of HR deficiency described that may correlate with platinum and PARP response (11), and newly developed homologous recombination repair panels have assessed several additional novel genes, including NBN, MRE11, RAD50, RAD51C, PALB2, BARD1, and BRIP1 (19, 26, 27). Our results show that the majority of patients enrolled in the study were HR deficient, a potential enrichment due to the selection of HGSOC patients based on platinum-sensitivity recurrence and objective response to platinum. The phase II study ARIEL2 investigating rucaparib (another PARPi) monotherapy in patients with recurrent platinum-sensitive HGSOC has confirmed BRCA1/2m as a biomarker of response, as well as genomic LOH, a potential predictive surrogate marker for HR deficiency (28). It was hypothesized that the inability of the cell to perform HR repair leads to genomic scarring and LOH, thus enabling the use of high LOH as a signature of HR deficiency. However, no data are currently available for LT responders and PARPi progression. Recently, the maintenance phase III study of niraparib, another PARPi, showed increased PFS in all patients – germline BRCA1/2 patients (the group with greatest benefit), and non-BRCA1/2 carriers (comprising of both HRD-positive as well as HRD-negative tumors) (29). Eligible patients had to have achieved response following 4 to 6 cycles of platinum-based chemotherapy with a CA-125 in the normal range or reduced by 90% for at least 7 days; an absence of measurable disease greater than 2cm at study commencement. Consistent with our findings, the PFS benefit to PARPi is driven by minimal residual disease (complete response prior maintenance), BRCA1/2 mutation and HRD-positive tumors, though not exclusively. The current
HR deficiency assays available did not completely identify biomarkers involved in response or resistance to PARPi.

Interestingly, four genetic alterations in the PTEN gene were observed, of which three were associated with BRCA1/2m in the LT responders on olaparib but none in the ST responder group. Moreover, in the placebo arm, three patients with PTEN alterations had disease progression within 3 months (Supplementary Table 3). The significance of this finding is not clear but warrants further investigation. While many LT responders to olaparib harbor a BRCA1/2m, our data show the occurrence of ST responders to olaparib in patients with BRCA1/2m. This finding highlights the limitation of the analysis on archival rather than tumor tissue at the time of disease progression. There is evidence that in cells deficient in DNA-damage repair, such as those with BRCA1/2m, additional mutations can restore function and allow effective DNA repair (30). Reversion mutations in BRCA1/2 have been described and associated with olaparib and platinum resistance (31), although this effect was supposed to be minimized by the selection of patients with response to platinum-based chemotherapy. Understanding therapeutic resistance requires comprehensive disease assessment at the specific time of therapeutic intervention; timing and treatment strategy are imperative to efficacy. Several mechanisms of resistance have been described related to the HR pathway (30). While tumors harboring a BRCA1/2m lack the HR repair pathway required for error-free repair of DNA double-strand breaks (32), other DNA-repair pathways exist that can become engaged, ultimately leading to olaparib resistance but platinum sensitivity, as with non-homologous end-rejoining alterations (33, 34). As such, determining the disease- and therapy-specific HR deficiency signature is important. Reports on this signature show varying gene lists, and these differences are likely attributed to variances in methodologies. Peng et al identified a HR defect gene signature that can be used to functionally assess HR status and predict clinical outcomes (27). Pennington et al reported that germline and somatic mutations in 13 HR genes predict platinum
response and survival in ovarian, fallopian tube and peritoneal carcinomas (26). These signatures need prospective validation.
CONCLUSION

This is the first study contrasting LT with ST responders to PARPi in terms of clinical and molecular data. Our results show that LT response to olaparib has been observed in platinum-sensitive recurrent HGSOC. This durable response may be multifactorial and driven by germline and somatic BRCA1/2m. This pilot study warrants a larger cohort to characterize LT responders. A study is ongoing to identify LT and ST responders to olaparib and allows for additional tumor tissue collection for analysis (NCT02489058).

ACKNOWLEDGMENTS

This research was conducted in collaboration with AstraZeneca.
REFERENCES


28. McNeish IA, Oza AM, Coleman RL, Scott CL, Konecny GE, Tinker A, et al. Results of ARIEL2: a Phase 2 trial to prospectively identify ovarian cancer patients likely to


Table 1. Characteristics of patients on olaparib capsule maintenance therapy – Study 19

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<th>BRCA status</th>
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LT responders: >2

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(CR, complete response; FIGO, International Federation of Gynecology and Obstetrics; LT, long term; ST, short term; PR, partial response; pts, patients; RECIST, Response Evaluation Criteria in Solid Tumors; WT, wild type)
Table 2. Molecular profiling of patients on olaparib and on placebo – Study 19

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Placebo arm

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Data are number (percentage). Pts, patients; LT, long term; ST, short term; WT, wild type
Table 3. Analysis of *BRCA1* and *BRCA2* mutational frequency in the olaparib arm (unless otherwise stated), including total *BRCA* mutations and mutations stratified by somatic mutations – Study 19

<table>
<thead>
<tr>
<th>Number of patients with each mutation shown</th>
<th>LT responders</th>
<th>ST responders</th>
<th>P value (Fisher’s exact test)</th>
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<td>N=21</td>
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<tr>
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<td>N=19</td>
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<td>BRCA mutation</td>
<td>22(^b) (69%)</td>
<td>10 (53%)</td>
<td>0.2179(^a)</td>
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<td><em>BRCA1</em> mutation</td>
<td>10 (45%)</td>
<td>7 (70%)</td>
<td>1.0000(^a)</td>
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<tr>
<td><em>BRCA2</em> mutation</td>
<td>13 (59%)</td>
<td>3 (30%)</td>
<td>0.0631(^a)</td>
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<tr>
<td>BRCA WT</td>
<td>10 (31%)</td>
<td>9 (47%)</td>
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<tr>
<td>Unknown/missing</td>
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<td>2 missing</td>
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<tr>
<td>(excluded from analysis)</td>
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<tr>
<td>Somatic BRCA mutations available</td>
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<tr>
<td>Olaparib treated (N=53)</td>
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<td>3</td>
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<tr>
<td>BRCA mutation</td>
<td>6(^b) (20%)</td>
<td>3 (16%)</td>
<td>1.0000(^a)</td>
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<tr>
<td><em>BRCA1</em> mutation</td>
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<td>1</td>
<td>1.0000(^a)</td>
</tr>
<tr>
<td><em>BRCA2</em> mutation</td>
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<td>2</td>
<td>0.6909(^a)</td>
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<td>Placebo treated (N=45)</td>
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<td>Un-methylated</td>
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<td>12 (86%)</td>
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<td>7 missing</td>
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</table>
HRD scores are given for LT responders and ST responders, and HR deficiency status is defined as a high HRD score and/or presence of a \( BRCA1/2 \) mutation. \( ^{a} \)P value between groups compared with WT; \( ^{b} \)One LT responder had both \( BRCA1 \) and \( BRCA2 \) mutations. HRD, homologous recombination repair deficiency; HR, homologous recombination repair; LT, long term; ST, short term; WT, wild type

<table>
<thead>
<tr>
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<th>N=24</th>
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<tr>
<td>HRD score (≥42)</td>
<td>21 (88%)</td>
<td>9 (69%)</td>
</tr>
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<td>25 (96%)</td>
<td>13 (76%)</td>
</tr>
<tr>
<td>HR non-deficient</td>
<td>1 (4%)</td>
<td>4 (24%)</td>
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</table>
FIGURE LEGENDS

Figure 1. Location of the BRCA1/2 mutation on the BRCA1/2 gene in patients on olaparib – Study 19

Somatic deletion exon 17 (not shown) found in BRCA1 in LT responder group. Large insertion (somatic, now shown) and deletion (not shown) found in BRCA2 in ST and LT responder group, respectively. VUS and SNP variants not shown

Figure 2. Mutations and other gene alterations in LT and ST responders to olaparib – Study 19

Only events that occurred in at least two patients are shown. Columns represent patients. Dark grey columns are samples that failed sequencing or analysis. A total of 51 patients were analyzed (32 LT responders, 19 ST responders), but seven failed sequencing; therefore, the molecular profiles of 44 patients are shown. Novel rearrangements of unknown significance, short variants of unknown significance, and non-focal lower-level (copy number [CN] ≤8) amplifications of genes known to be recurrently amplified in cancer are excluded

Figure 3. Study 41 data showing LT and ST responders to olaparib and placebo, with mutations and other gene alterations given for LT and ST responders

Novel rearrangements of unknown significance, short variants of unknown significance, and non-focal lower-level (CN ≤8) amplifications of genes known to be recurrently amplified in cancer are excluded
FIGURES

Figure 1. Location of the *BRCA1/2* mutation on the *BRCA1/2* gene in patients on olaparib

Somatic deletion exon 17 (not shown) found in *BRCA1* in LT responder group. Large insertion (somatic, now shown) and deletion (not shown) found in *BRCA2* in ST and LT responder group, respectively. VUS and SNP variants not shown.
Figure 2. Mutations and other gene alterations in LT and ST responders to olaparib

Only events that occurred in at least two patients are shown. Columns represent patients. Dark grey columns are samples that failed sequencing or analysis. A total of 51 patients were analyzed (32 LT responders, 19 ST responders), but seven failed sequencing; therefore, the molecular profiles of 44 patients are shown. Novel rearrangements of unknown significance, short variants of unknown significance, and non-focal lower-level (copy number [CN] ≤8) amplifications of genes known to be recurrently amplified in cancer are excluded.
Figure 3. Study 41 data showing LT and ST responders to olaparib and placebo, with mutations and other gene alterations given for LT and ST responders

Novel rearrangements of unknown significance, short variants of unknown significance, and non-focal lower-level (CN ≤8) amplifications of genes known to be recurrently amplified in cancer are excluded.
Figure 1

**BRCA1 (17q21)**
- **RING**
- **NLS**
- **NLS**
- **DNA binding**
- **CHK2**
- **ATM**
- **PALB2**
- **CCDC98 Ctip BRIP1**
- **BRCT1**
- **BRCT2**
- **M1T**
- **E23fs***
- **E418X**
- **S956fs***
- **R1203X**
- **R1443X**
- **S1655fs***
- **Q1756fs***

**BRCA2 (13q12.3)**
- **BRC**
- **BRC**
- **BRC**
- **BRC**
- **BRC**
- **BRC**
- **DNA binding**
- **CHK2**
- **PALB2**
- **RAD51**
- **E23fs***
- **V1283fs***
- **E1493fs***
- **E1646fs***
- **S1830fs***
- **D2515fs***
- **T2515fs***
- **A938fs***
- **W1692fs***
- **S1982fs*(1)**
- **D1321fs***
- **S1830fs***
- **D2005fs***
- **F2254fs***
- **L2587fs***
- **K2939X R3128X**

**Germline**
- **Tumor**
- **Germline**
- **Somatic mutation**
- **Founder mutation**

**Binding domain**
**DNA damage repair**
- BRCA1
- BRCA2
- NBN
- RAD52
- FANCE
- XRCC3
- BRIP1

**Cell cycle regulation**
- CCNE1
- RB1
- CCND2
- CDKN1B
- CCND3
- MDM4

**MAPK signaling**
- MYC
- FGF8
- MAP3K13
- KRAS
- NF1

**Akt/PI3K signaling**
- Pik3C2G
- PTEN
- TSC2
- RPTOR
- AKT1

**Apoptosis**
- MCL1
- DAXX

**Others**
- RUNX1T1
- KDMSA
- BCL6
- ARID1A
- KHL6
- ASXL1
- MLL2
- ARFRP1
- PRDM1
- WISP3
- GNAS
- ZNF217
- NF2

**Legend**
- V – VUS
- S – SNP
- HRR deficient, defined as BRCA1/2 mutation and/or HRD score ≥42
- Known somatic or likely somatic short variants
- High-level (CN >8) and focal (CN >5) amplifications
- Homozygous deletions in cancer
- Observed copy number gains in other genes
- Likely functional rearrangements (e.g., gene fusions)
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<th>Genes seen in at least 2 patients</th>
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S – SNP
- Known somatic or likely somatic short variants
- High-level (CN >8) and focal (CN >5) amplifications
- Homozygous deletions in cancer
- Observed copy number gains in other genes