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**Report**

**Fission Yeast Apc15 Stabilizes MCC-Cdc20-APC/C Complexes, Ensuring Efficient Cdc20 Ubiquitination and Checkpoint Arrest**

**Highlights**

- Two small fission yeast APC/C subunits regulate MCC binding
- Apc14 is required for checkpoint silencing and Apc15 for checkpoint arrest
- Apc15 enhances Cdc20 ubiquitination and turnover
- Two Cdc20 molecules are found in MCC-APC/C and free MCC-Cdc20Δ accumulates in apc15Δ

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**In Brief**

May et al. dissect the roles of fission yeast Apc14 and Apc15 in spindle checkpoint regulation of APC/C activity. Apc15 is needed for efficient Cdc20 turnover and checkpoint arrest in fission yeast. Apc15 and the C terminus of Mad3 are shown to co-ordinate the interactions of two molecules of Cdc20 within inhibited APC/C complexes.
Fission Yeast Apc15 Stabilizes MCC-Cdc20-APC/C Complexes, Ensuring Efficient Cdc20 Ubiquitination and Checkpoint Arrest

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SUMMARY

During mitosis, cells must segregate the replicated copies of their genome to their daughter cells with extremely high fidelity. Segregation errors lead to an abnormal chromosome number (aneuploidy), which typically results in disease or cell death [1]. Chromosome segregation and anaphase onset are initiated through the action of the multi-subunit E3 ubiquitin ligase known as the anaphase-promoting complex or cyclosome (APC/C [2]). The APC/C is inhibited by the spindle checkpoint in the presence of kinetochore attachment defects [3, 4]. Here we demonstrate that two non-essential APC/C subunits (Apc14 and Apc15) regulate association of spindle checkpoint proteins, in the form of the mitotic checkpoint complex (MCC), with the APC/C. apc14Δ mutants display increased MCC association with the APC/C and are unable to silence the checkpoint efficiently. Conversely, apc15Δ mutants display reduced association between the MCC and APC/C, are defective in poly-ubiquitination of Cdc20, and are checkpoint defective. In vitro reconstitution studies have shown that human MCC-APC/C can contain two molecules of Cdc20 [5–7]. Using a yeast strain expressing two Cdc20 genes with different epitope tags, we show by co-immunoprecipitation that this is true in vivo. MCC binding to the second molecule of Cdc20 is mediated via the C-terminal KEN box in Mad3. Somewhat surprisingly, complexes containing both molecules of Cdc20 accumulate in apc15Δ cells, and the implications of this observation are discussed.

RESULTS AND DISCUSSION

Cdc20Sip1 (Sip1 is the fission yeast homolog of Cdc20) was identified several years ago as the key effector of the spindle checkpoint [8, 9], and the mitotic checkpoint complex (Cdc20-Mad3-Mad2-Bub3) was found to be the most potent anaphase-promoting complex or cyclosome (APC/C) inhibitor [10]. The crystal structure of fission yeast mitotic checkpoint complex (MCC) has been solved [11], and recently cryoelectron microscopy (cryo-EM) structures of human MCC in complex with APC/C were obtained from recombinant complexes produced in baculovirus-infected insect cells [6, 7]. These models provide an excellent structural framework within which spindle checkpoint inhibition takes place, yet there are several areas requiring further insight. The Gould laboratory previously demonstrated that deletion of either the Apc14 or Apc15 subunit of fission yeast APC/C did not affect its ubiquitin ligase activity [12]. Apc14 is not well conserved, but in human and budding yeast cells it has been shown that there are spindle checkpoint silencing defects in the absence of Apc15 [13–15]. Therefore, we analyzed the fission yeast apc14Δ and apc15Δ strains for defects in checkpoint establishment, maintenance, and/or silencing.

apc15Δ Mutants Display Spindle Checkpoint Defects

First, we employed the cold-sensitive beta-tubulin mutant nda3-KM311 to analyze the ability of cells to arrest in mitosis in the absence of spindle microtubules [16]. Figure 1A demonstrates that whereas apc14Δ cells arrest like wild-type, apc15Δ cells display severe defects in this checkpoint assay. We also analyzed the ability of these apc mutants to arrest in response to Mad2 and Mps1Mph1 kinase (Mph1 is the fission yeast homolog of Mps1 kinase) overexpression [17, 18], and once again found that apc15Δ strains were significantly defective in arrest (Figure 1B). The inability to respond to Mad2 overexpression places Apc15 function downstream in the spindle checkpoint pathway, as the only other checkpoint protein required for the Mad2 overexpression arrest is Mad3, the other key component of fission yeast MCC [19]. This suggested that Apc15 might have a role in MCC assembly and/or its association with the APC/C. To test this, we analyzed MCC assembly in an unperturbed mitosis, having synchronized cells at G2/M using the cdc25-ts allele. Figure 1C shows that there is little effect on MCC assembly in the absence of either Apc14 or Apc15. However, we note that in the apc15Δ mutant, the levels of Cdc20Sip1 and the MCC complex are 2- to 3-fold higher and stay high for longer, even though they display no delay in anaphase (see Figure S1).

When we analyzed the ability of the MCC complex to bind to the APC/C, we saw striking effects in both the apc14Δ and apc15Δ strains (Figure 2A). Cells were synchronized at G2/M and then released into mitosis. The levels of MCC bound to the APC/C in apc15Δ strains were reduced to a level similar to those in the absence of Mps1Mph1 kinase activity [20]. Conversely, the
MCC levels bound in apc14Δ strains were 2- to 3-fold higher than in wild-type cells (see Figure S2 for quantitation). The simplest interpretation of these data is that Apc15 is required for stable MCC binding to the APC/C, which is consistent with the checkpoint defects observed in apc15Δ strains (Figure 1), whereas Apc14 function is required for efficient MCC release and checkpoint silencing.

apc14Δ Mutants Display Checkpoint Silencing Defects

To analyze the apc14Δ phenotype in more detail, we employed a checkpoint silencing assay that we previously developed during studies of protein phosphatase 1 [21]. Cells are arrested without microtubules using the nda3 mutation [16], and then Aurora activity is inhibited using the ark1-as3 allele [22]. Because Ark1 activity is necessary to maintain spindle checkpoint arrests [21, 23], these cells rapidly degrade cyclin B. Figure 2B shows that there is a clear delay in cyclin B degradation in apc14Δ cells, consistent with the hypothesis that Apc14 has a role in checkpoint silencing. This Apc14 silencing function is the subject of ongoing work, but was not analyzed further in this study.

Apc15 Is Cell-Cycle Regulated

Transcript data in PomBase [24] show that Apc15 expression is tightly cell-cycle regulated, with highest expression at G2/M. Therefore, we analyzed the cell-cycle levels of the Apc15 protein. Apc15 is a rather small protein, and we were concerned that epitope tags might perturb its function. Therefore, we generated specific, polyclonal antibodies to Apc15. Figure S2B shows that the levels of Apc15 increase ~2-fold during mitosis, and Figure 2C demonstrates that Apc15 levels increase further during prolonged checkpoint arrest. As is the case for many APC/C subunits, Apc15-GFP accumulates in the nuclei of mitotic cells [25]. We carried out several experiments to test whether the Apc15 newly synthesized at G2/M is important for checkpoint function and whether Apc15 is itself subject to ubiquitin-mediated degradation. As yet, we have found no clear evidence that this regulation is physiologically important for mitosis (see Figure S2).

At this point, we had two simple hypotheses for Apc15 function: (1) Apc15p could act as a loading factor, by first binding to the MCC and then helping it associate with the APC/C core particle, or (2) Apc15 might form an important part of the MCC docking site on the APC/C. To distinguish between these modes of action, we analyzed the association of Apc15 with the APC/C and with MCC proteins in yeast lysates. To test whether there...
was a free pool of Apc15 and whether this could interact with the MCC, we carried out immunodepletion of Apc15-GFP from mitotic extracts. As expected, Apc15 co-immunoprecipitates with Apc1-GFP (Figure 2D). No Apc15 remains associated with Apc1-GFP in an extract after three rounds of Apc1-GFP immunodepletion, yet there is still a significant pool of free Apc15.
Importantly, when we immunoprecipitated this free pool of Apc15 it was not associated with Mad3, Mad2, or Cdc20 (Figure 2D). This experiment indicates that although there was a significant non-APC/C-bound pool of Apc15, it was not associated with the MCC complex or proteins therein. This argues against the model where Apc15 acts as an MCC loading factor, as it would be expected to bind MCC components independent of APC/C binding. The role of the free Apc15 pool remains unclear, but its existence most likely explains why there was no significant phenotype upon transcriptional depletion of Apc15 (Figures S2D–S2F).

**Fission Yeast Apc15 Is Required for Efficient Cdc20 Ubiquitination**

Deletion of Apc15 had no effect on the ability of cells to assemble the MCC, but we noted that the levels of Cdc20 and MCC remained high for longer in apc15Δ than in wild-type cells (Figure 1C). This suggested that Apc15 might regulate Cdc20 protein levels and that it could be required for efficient Cdc20 ubiquitination and/or degradation. To determine whether this was the case, we arrested cells in metaphase using mitotic fission yeast cells. Our findings are confirmed in the accompanying fission yeast manuscript [30], where two forms of Cdc20 (one tagged and one untagged) are shown to interact with itself or on a larger platform. In this model, all members of the MCC complex would contain two Cdc20 molecules [6, 7]. Importantly, our Cdc20-Cdc20 co-immunoprecipitation experiment is carried out in whole-cell extracts made from mitotic fission yeast cells. Our findings are confirmed in the accompanying fission yeast manuscript [30], where two forms of Cdc20 (one tagged and one untagged) are shown to interact in mitosis. One way to form these Cdc20-Cdc20 complexes would be for the MCC as a whole to form dimers, either simply with itself or on a larger platform. In this model, all members of the MCC complex would contain two molecules, rather than just Cdc20. To test this, we engineered fission yeast strains expressing normal Mad3 and Mad3-GFP, and asked whether the two forms of Mad3 can be co-immunoprecipitated in mitotically arrested cells. Figure S3B shows that this was not the case, ruling out MCC dimerization.

The model proposed in the human studies [5] was that the MCC (Cdc20M-Mad3-Mad2) binds and inhibits an active Cdc20M-APC/C complex to form Cdc20M-Mad3-Mad2-Cdc20M-APC/C (containing two Cdc20 molecules). Our apc15 mutant allowed us to test this model in fission yeast. If this were the case, one would predict that the apc15 mutant, which impairs the interaction between the MCC and APC/C, will reduce or abolish the interaction between the two Cdc20 molecules. Figure 3B shows that this is not the case, as efficient co-immunoprecipitation of

**Fission Yeast MCC Can Contain Two Cdc20 Molecules, and Binding of the Second Molecule Is Mad3-KEN2 but Not Apc15 Dependent**

Musacchio and co-workers proposed a few years ago that a second molecule of Cdc20 in MCC complexes would help explain the need for two conserved KEN boxes in Mad3/BubR1 and aspects of Cdc20 turnover [2, 29]. It was then argued from in vitro studies that human MCC can bind a second molecule of Cdc20, enabling it to inhibit active Cdc20-APC/C [5]. Recent cryo-EM studies have demonstrated that when human MCC-APC/C is reconstituted in insect cells there are indeed MCC complexes that contain two Cdc20 molecules [6, 7]. To distinguish the two Cdc20 molecules, we will refer to them as the MCC-bound form (Cdc20M) and the APC/C-bound or activator form (Cdc20A).

To provide formal proof of the second Cdc20 molecule in a living system, we generated a fission yeast strain containing a second copy of the Cdc20M^KEN1 gene. This copy has an internal 3×FLAG tag [31], enabling it to be readily distinguished from the first copy, which has a C-terminal 3×HA (hemagglutinin) tag. Both of these forms of Cdc20M^KEN1 are functional and bind the APC/C (see below). If two copies of Cdc20 are present in the MCC, then the two proteins would be co-immunoprecipitated from mitotic extracts. Figure 3B shows that this is the case, that their interaction is dependent on Mad3p, and importantly that their interaction is dependent on both of the conserved Mad3-KEN boxes. The first KEN box (KEN1) is known to be critical for direct Cdc20 binding and MCC formation [11], but the function of the second KEN box (KEN2) is less well understood [32]. KEN2 is widely recognized as being necessary for checkpoint function but not for MCC formation. In BubR1, it can compete with substrates for APC/C interaction in vitro [32]. Our finding that Mad3-KEN2 is needed for interaction with the second molecule of Cdc20 is in agreement with the human study where the D box and the second KEN box in BubR1 were both found to be necessary [5] and with the recent cryo-EM structures [6, 7]. Importantly, our Cdc20-Cdc20 co-immunoprecipitation experiment is carried out in whole-cell extracts made from mitotic fission yeast cells. Our findings are confirmed in the accompanying fission yeast manuscript [30], where two forms of Cdc20 (one tagged and one untagged) are shown to interact in mitosis.
Cdc20-FLAG and Cdc20-HA was still observed in apc15Δ cells; indeed, these complexes accumulated in the mutant (see also Figure S3A). Figure 2A demonstrates that there was a significant reduction in the level of MCC bound to the APC/C in apc15Δ mutants, yet Figure 3B reveals complexes containing multiple Cdc20 molecules. To confirm that these Cdc20 complexes were not associated with the APC/C, we depleted the APC/C from apc15Δ extracts via four rounds of Cut9-GFP immunodepletion and asked whether the free pool of MCC still contained two molecules of Cdc20 (Cdc20-HA–Mad3–Mad2–Cdc20-FLAG). Figure 4A shows that this is the case, and that there is significantly more of this complex in apc15Δ mutant extracts compared to wild-type. We note that this observation is not necessarily in line with the proposed human model of Cdc20-APC/C inhibition [5], and suggest two models to explain our fission yeast observations:

1. Apc15 forms an important part of the MCC binding site on the APC/C. As a consequence, in apc15Δ cells the MCC (Cdc20ΔΔ–Mad3–Mad2) might preferentially bind to free Cdc20 rather than to Cdc20ΔΔ–APC/C complexes. This will sequester Cdc20 and form a free pool of Cdc20ΔΔ–Mad3-Mad2-Cdc20ΔΔ. Such sequestration may well help inhibit Cdc20 action, and the accompanying manuscript [30] shows this to be the case when Cdc20 levels are reduced. However, we note that the Cdc20ΔΔ in these

Figure 3. apc15Δ Mutants Have Significant Defects in the Processivity of Cdc20ΔΔ Ubiquitination, and Fission Yeast MCC-Cdc20-APC/C Contains Two Molecules of Cdc20

(A) Cdc20 ubiquitination experiments. The indicated strains all contain the mts3-1 proteasome mutation, to block cells in mitosis independent of the spindle checkpoint and to enrich for polyubiquitinated forms of cellular proteins. Cultures were shifted to 36°C 3 hr prior to harvesting. Whole-cell lysates were made in the presence of Dub inhibitors, and Cdc20-FLAG was immunoprecipitated and then immunoblotted for Cdc20-FLAG. Long exposure reveals a ladder of slow-migrating bands, which are reduced in mad3 and apc15 mutants. The indicated lysates were treated with recombinant hsUSP2 (de-ubiquitinase) or lambda phosphatase prior to running the gel. Lambda phosphatase has no effect but USP2 abolishes the ladder, confirming that this is due to modification with ubiquitin. Different modified forms of Cdc20 accumulate in the mts3 and apc15Δmts3 mutants, with shorter chains in the absence of Apc15. These are indicated with green and red markings by the relevant anti-ubiquitin blots. This experiment was repeated three times.

(B) The indicated lanes from (A) are expanded to highlight the different modified forms of Cdc20 that accumulate in the mts3, apc15Δmts3, and apc14Δmts3 mutants. Most notably, there are shorter chains in the absence of Apc15 (boxed in red).

(C) Cdc20-FLAG and Cdc20-HA co-immunoprecipitate. Cells containing both Cdc20 forms were synchronized in mitosis (60 min after cdc25 block and release), lysates were prepared, and Cdc20-FLAG was immunoprecipitated. The immunoprecipitates were then immunoblotted and analyzed for associated Cdc20-HA, Mad3-GFP, Mad2, and Apc15. The asterisk indicates Cdc20-Cdc20 co-immunoprecipitation. This experiment was repeated three times. The % of metaphase cells is indicated below the blot for the six strains.

See also Figure S3.
completed for APC/C complexes through four rounds of cut9apc6.  

Released with short ubiquitin chains in the form of Cdc20 M-Mad3-Mad2-degradation. Apc14 function is required for efficient release of the MCC.

(iia) In Mad3 (KEN2 and associated ABBA motifs) is critical for this stable interaction and in fission yeast this interaction is stabilized by Apc15. The C terminus of apc15

Apc14 function is required for efficient release of the MCC.

(ii) In wild-type cells Cdc20A is efficiently poly-ubiquitinated, leading to its degradation. Apc14 function is required for efficient release of the MCC.

(2) The MCC binds to Cdc20A-APC/C, but their interaction is weakened in the absence of Apc15 and the MCC-Cdc20A-APC/C complex is rather short-lived. A consequence of this brief interaction with the APC/C is that it would lead to a reduced processivity of Cdc20A ubiquitination in apc15Δ cells. This is consistent with the short Ub chains we have observed on Cdc20 in apc15Δ (Figure 3B).

Importantly, when the MCC complex dissociates it takes the Cdc20A activator with it to form free Cdc20A-Mad3-Mad2-Cdc20A.

Note that these models are not mutually exclusive. It is important to include the free Cdc20A-Mad3-Mad2-Cdc20A complex in overviews of MCC action and Cdc20 inhibition (see Figure 4B).

**Conclusions**

At first glance, fission yeast Apc15 appears to have a simple role to play in helping the MCC stably bind to the APC/C core particle, and this is consistent with its position in recent high-resolution cryo-EM structures of the APC/C. Budding yeast and in vitro human studies have argued that Apc15 is needed for efficient Cdc20 ubiquitination and subsequent MCC release [13, 15]. Based on the cryo-EM models, it was proposed that human Apc15 undergoes a conformational change to shift the MCC into a suitable orientation for Cdc20 auto-ubiquitination [6, 7]. Our study is consistent with this, as Cdc20Δplp displays reduced ubiquitination and is stabilized in fission yeast apc15Δ cells (Figure 3A). However, fission yeast Apc15 is also needed for stable MCC binding and thus for spindle checkpoint arrest, which is not the case in human and budding yeast cells.

Importantly, we also provide in vivo evidence for complexes containing two Cdc20 molecules in fission yeast. Interaction with the second molecule of Cdc20 is mediated by the C terminus of Mad3, including its second KEN box and the nearby ABBA motifs [32–34]. Our work and the accompanying manuscript [30] both describe in vivo fission yeast studies that provide an important confirmation of models proposed from in vitro reconstitution studies of human MCC proteins and their interactions with the APC/C. New findings are presented here where, paradoxically, although apc15Δ mutants display reduced levels of MCC bound to the APC/C (Figure 2A), they also accumulate Cdc20-Cdc20 complexes (Figure 3B). We have suggested two models for generating the free pool of Cdc20A-Mad3-Mad2-Cdc20A that we observe in apc15Δ mutants (see Figure 4B).

In fission yeast, we find that deletion of Apc14 leads to checkpoint silencing defects. In its absence there are higher than normal levels of the MCC bound to the APC/C, and experiments are ongoing to understand its mode of action. It will be interesting to see whether in the absence of Apc14 the MCC simply binds more tightly to the APC/C, or whether Apc14 has an active role similar to p31comet (not conserved in S. pombe), TRIP13, or the CCT chaperone in disrupting MCC-APC/C complexes [35, 36].

(iii) In wild-type cells Cdc20A is efficiently poly-ubiquitinated, leading to its degradation. Apc14 function is required for efficient release of the MCC.

(iv) The checkpoint is off; APC/C is now free to be bound by the Cdc20A activator, which can catalyze the poly-ubiquitination of securin and cyclin, leading to anaphase onset.

See also Figure S4.

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**Figure 4. A Free Pool of Cdc20A-Mad3-Mad2-Cdc20A Accumulates in apc15Δ Mutants**

(A) The MCC (Cdc20A-Mad3-Mad2)-Cdc20A complex can be found in the APC/C-depleted supernatant, and the complex accumulates in apc15Δ. Mitotic lysates were prepared from cdc25-22 cdc20-HA cdc20-FLAG Apc6Cut9-GFP (60 min after cdc25 block and release) and then immunodepleted for APC/C complexes through four rounds of cut9apc6 immunodepletion. Cdc20-FLAG was then immunoprecipitated from the resulting supernatant and immunoblotted to look for associated Cdc20-HA and checkpoint proteins. The MCC (Cdc20A-Mad3-Mad2)-Cdc20A is immuno-precipitated without Apc6Cut9-GFP or Apc15. This experiment was repeated twice.

(B) Models of MCC binding and Cdc20 ubiquitination, in wild-type cells and apc15 mutants. (i) When the checkpoint is on, the MCC binds to Cdc20-APC/C and in fission yeast this interaction is stabilized by Apc15. The C terminus of Mad3 (KEN2 and associated ABBA motifs) is critical for this stable interaction with the second molecule of Cdc20 (Cdc20A). (ii) In apc15Δ cells, the MCC could preferentially bind free Cdc20A. (iii) In the absence of Apc15, the MCC complex is weakly bound and Cdc20A is inefficiently ubiquitinated. It is released with short ubiquitin chains in the form of Cdc20A-Mad3-Mad2-Cdc20A. Note that both molecules of Cdc20 are released from the APC/C. (iv) In wild-type cells Cdc20A is efficiently poly-ubiquitinated, leading to its degradation. Apc14 function is required for efficient release of the MCC.
EXPERIMENTAL PROCEDURES

See the Supplemental Experimental Procedures.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures and four figures and can be found with this article online at http://dx.doi.org/10.1016/j.cub.2017.03.013.

AUTHOR CONTRIBUTIONS

K.M.M. conceived and designed the experiments, acquired data, performed analysis and interpretation of the data, and drafted the figures. F.P. generated Slp1 constructs and yeast strains and performed certain Cdc20 co-immuno-precipitations. K.G.H. conceived the project, helped with analysis and interpretation of the data and figure construction, and wrote the manuscript. All authors reviewed the manuscript.

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REFERENCES


