Structural evidence for the covalent modification of FabH by 4,5-dichloro-1,2-dithiol-3-one (HR45)

Citation for published version:
Ekström, AG, Kelly, V, Marles-wright, J, Cockroft, SL & Campopiano, DJ 2017, 'Structural evidence for the covalent modification of FabH by 4,5-dichloro-1,2-dithiol-3-one (HR45)' Organic & Biomolecular chemistry, vol. 15, no. 30, pp. 6310-6313. DOI: 10.1039/C7OB01396E

Digital Object Identifier (DOI):
10.1039/C7OB01396E

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Peer reviewed version

Published in:
Organic & Biomolecular chemistry

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Download date: 22. Apr. 2019
Structural evidence for the covalent modification of FabH by 4,5-dichloro-1,2-dithiol-3-one (HR45)

Alexander G. Ekström¹, Van Kelly², Jon Marles-Wright³, Scott L. Cockcroft², and Dominic J. Campopiano³

We use mass spectrometry analysis and molecular modelling to show the established antimicrobial inhibitor 4,5-dichloro-1,2-dithiol-3-one (HR45) acts by forming a covalent adduct with the target β-ketoacyl-ACP synthase III (FabH). The 5-chloro substituent directs attack of the essential active site thiol (C112) via a Michael-type addition elimination reaction mechanism.

Fatty acid biosynthesis is essential for cell viability and growth, deriving fatty acids from the fundamental building block acetyl-CoA. In eukaryotes, this process is facilitated by type I fatty acid synthase (FAS I); a large, multi-domain enzyme containing all required functionalities. In contrast, most bacteria utilise a complex of proteins termed type II fatty acid synthase (FAS II), whereby each functionality is carried out by a discreet enzyme, between which the growing acyl chain is transported by the acyl carrier protein (ACP). Substantial differences in the architecture and chemistry carried out by FAS I and II systems has led to significant interest in the bacterial pathway as a target for new antibacterial compounds.

β-Ketoacyl-ACP synthase III (FabH) is the first condensing enzyme in the FAS II pathway, and has attracted significant attention as a target for novel antibiotic design. This ubiquitous, highly conserved enzyme catalyses the cysteine-mediated, Claisen-like condensation between malonyl-ACP and short chain acyl-CoAs. Despite the suggestion that Gram-positive bacteria can absorb and utilise exogenous fatty acids, FabH is widely believed to be essential for cell viability. A few natural product inhibitors of FAS II exist in the literature, but despite research efforts there are no FabH-specific inhibitors approved for clinical use.

Of the natural products, the thiolactone antibiotic thiolactomycin (TLM, Fig. 1) has selectivity for FAS II condensing enzymes by mimicking malonyl-ACP binding. He et al. used TLM as a starting point to develop potent FabH inhibitors by searching the National Cancer Institute (NCI) database for structurally similar molecules. The most potent hit was 4,5-dichloro-1,2-dithiol-3-one (also referred to in the literature as HR45 and DDCP, Fig. 1). Subsequent structure-activity relationship studies showed that the chlorine in the 5-position was found to be essential for irreversible inhibition of FabH isoforms from both Staphylococcus aureus and Escherichia coli (ecFabH) with reported IC₅₀ values of 156 nM and 2.0 μM, respectively. The reported mode of inhibition was inconclusive, and despite postulating covalent modification via a Michael-type mechanism they were unable to obtain data to support this hypothesis. This commercially available 1,2-dithiol-3-one (CAS 1192-52-5) has since been frequently reported as a positive control for FabH inhibition, and also has FDA approval as a slimicide additive in the paper industry for food packaging.

A less efficacious hit from the same study was HR12 (also known as RWJ-3981, Fig. 1), an analogue of HR45 with a phenyl group replacing the chlorine in the 4-position, with reported IC₅₀ values of 0.98 μM and 5.7 μM for soFabH and ecFabH respectively. Interestingly, in a separate study HR12 was identified as a hit against Escherichia coli uridine diphosphate-β-acetyl glucosamine enolpyruvyl transferase (ecMurA), a cysteine-dependent enzyme involved in the first committed step of peptidoglycan cell wall biosynthesis.

![Figure 1](https://example.com/figure1.png)

Fig. 1 HR45 and HR12, hits against soFabH and ecFabH from an NCI database screen by He et al. based on Thiolactomycin. HR12 identified as a hit against ecMurA in a separate study.

¹ EasChEM School of Chemistry, University of Edinburgh, Joseph Black Building, David Brewster Road, Edinburgh, EH9 3FJ, UK.
² School of Biological Science, Newcastle University, Devonshire Building, Newcastle upon Tyne, NE1 7RU, UK.

DOI: 10.1039/x0xx00000x

This journal is © The Royal Society of Chemistry 20xx

J. Name., 2013, 00, 1-3 | 1
The mode of inhibition was speculated to be covalent modification of the catalytic ecMurA C115 residue. Due to the lack of homology between soFabH and ecMurA, it is unlikely that the HR45 mechanism is restricted to these two enzymes. The authors were unable to obtain supporting mass spectrometry (MS) data and suggested that the modification did not survive the MS conditions. A key point to note was that the inhibition could be reversed through reduction of the disulfide by dithiothreitol (DTT).

The structure of HR45 suggests that the Michael-type acceptor nature of the cyclic α,β-unsaturated ketone could define it as a pan assay interference compound (PAIN) with non-specific activity towards nucleophilic amino acid residues. Whilst PAINs are often removed from traditional small molecule screens, it has been suggested that this should not be the case when screening for antimicrobial activity. Encouraged by this observation, the reaction of HR45 with the irreversible, cysteine-specific label N-ethylmaleimide (NEM), which indicated C12 as the most electrophilic centre, was quanitatively removed by incubation with HR45 with the loss of HCl. This data supports our proposed mechanism, involving attack of the nucleophilic cysteine residue followed by elimination of the ipso-chlorine (Fig. S2). The HR45 modification was quantitatively removed by incubation with excess DTT (2 mM) for 15 mins, resulting in the native protein mass. We also treated the native soFabH with the irreversibly alkylating agent N-ethylmaleimide (NEM), which resulted in a mass shift of +125 Da, signifying quantitative labelling of a single cysteine residue (Fig. 2c). The NEM-modified sample was then incubated with HR45 for 3 h; resulting in no further mass change (Fig. 2d). This indicates that alkylation of C112 prevents the reaction of soFabH with HR45 (Fig. 2d).

To further confirm modification of C112, HR45-modified soFabH was digested with trypsin to generate peptide fragments. As well as the absence of the ion corresponding to the C112-containing tryptic peptide (103-VASMDQLAACSGFMYSMITAK-123, predicted monoisotopic mass 2224.0036 Da, observed 1113.0042 Da [M+2H]+) in the control digest, we also failed to detect a peptide corresponding to the expected mass of the C112-HR45 adduct. This observation was consistent with previous studies that attempted to detect HR12 adducts. To discover the fate of the C112-containing peptide, it was purified from a large-scale trypsin digest of soFabH with activated thiol sepharose. After elution with DTT, the peptide was desalted and resuspended in either ammonium bicarbonate (pH 8.0) or ammonium acetate (pH 7.0) buffer and incubated in the presence or absence of HR45 to investigate the stability of the peptide and the HR45 adduct. In ammonium acetate, both the unmodified and HR45-modified forms were observed at the predicted masses (Fig. 3a and c), consistent with observations on the intact protein (Fig. 2b). In contrast, in ammonium bicarbonate (Fig. 3b), the unmodified peptide was found to exist predominantly as a [M+4H]+ disulfide dimer. Perplexingly, upon incubation of the HR45-modified form in ammonium bicarbonate (Fig. 3d) a new signal appeared corresponding to a loss of 34 Da from the unmodified peptide (predicted 2189.9837 Da, observed 1095.9991 Da [M+2H]+). Using CID tandem MS, we obtained a fragmentation spectrum which we interpreted as the conversion of C112 to dehydroalanine (Dha) (Fig. S7). A small amount of this Dha-containing peptide also formed from the disulfide dimer at pH 8.0 (Fig. 3b).

This C112 to Dha conversion under typical tryptic digest conditions may be the reason for previous failures to observe...
The covalent modification of saFabH and ecMurA by HR45 and similar inhibitors. The alkaline hydrolysis pathway of 1,2-dithiol-3-ones is a highly complex one. Although we do not yet have sufficient data to speculate on the mechanism of the C112-HR45 to Dha conversion, it is likely that one of the hydrolysis products facilitates the transformation, possibly by a similar mechanism to other Dha formation chemistry.

The instability of the adduct under standard trypsin digest conditions led us to seek an alternative method. We digested HR45-modified saFabH at pH 2.0 with pepsin, and identified two peptic peptides in the unmodified control, AACSGF and AACSGFM^Ox, where C^ denotes a cysteine modification of +150 Da, corresponding to addition of HR45 with the loss of HCl. Clearly a chlorine atom is incorporated in the modified peptides due to the distinctive isotope pattern. CID tandem MS was used to fragment AAC*SGFM^Ox, confirming that the modification was indeed on the cysteine residue. When similar pH-dependence experiments were conducted on the intact saFabH-HR45 adduct, we observed no evidence of dehydroalanine formation. In the intact protein, C112 is located at the N-terminus of a long α-helix which lowers the pKa of C112 from 8.8 to 7.2. What impact this environment has on the stability of the HR45 adduct requires further study.

Despite quantitative modification, we were unable to obtain crystallographic data of HR45-modified saFabH. We modelled the adduct by docking the recent crystal structure of HR45 into the active site of saFabH (PDB: 3IL7) guided by our proposed mechanism (Fig. 5 and S4). Although there is ample space for HR45 in the active site, catalytically essential residue...
C112 has little freedom of movement, limiting the likely positions of the ligand and suggesting possible hydrogen bonding between the C-5 oxygen of HR45 and the side chain of catalytically essential N268. This model could be a starting point to develop highly specific inhibitors targeting FabH.

Conclusions

We have shown that the 1,2-dithiol-3-one HR45 covalently modifies sFabH through a Michael-type addition elimination at the catalytic residue C112. We suggest that this is also the mode of inhibition of the molecule on other FabH isoforms, and potentially other systems including the inhibition of ecMurA by HR12. We have also shown that the reason this covalent modification has previously escaped detection is that under normal trypptic digest conditions the alkaline hydrolysis pathway of HR45 facilitates the conversion of the C112 to Dha, the mechanism of which remains to be determined. Describing the precise mode of action of molecules isolated from screening campaigns that failed to make the clinic may have impact on the design and discovery of new antimicrobial leads. Our detailed mechanistic study of HR45 provides insight for the design of new antimicrobials, and this approach will no doubt be useful for other systems.

We thank SULSA for providing PhD studentship funding (AE). We also thank the BBSRC and EPSRC for funding (BB/M003493/1, BB/N005570/1 and EP/K039717/1).

Notes and references