Follicular Dendritic Cell-Specific Prion Protein (PrP<sub>C</sub>) Expression Alone Is Sufficient to Sustain Prion Infection in the Spleen

Laura McCulloch<sup>1</sup>, Karen L. Brown<sup>1</sup>, Barry M. Bradford<sup>1</sup>, John Hopkins<sup>1</sup>, Mick Bailey<sup>2</sup>, Klaus Rajewsky<sup>3</sup>, Jean C. Manson<sup>1</sup>, Neil A. Mabbott<sup>1*</sup>

1 The Roslin Institute & Royal (Dick) School of Veterinary Sciences, University of Edinburgh, Midlothian, United Kingdom, 2 Division of Veterinary Pathology, Infection and Immunity, School of Clinical Veterinary Science, University of Bristol, Avon, United Kingdom, 3 Program in Cellular and Molecular Medicine, Children's Hospital, and Immune Disease Institute, Harvard Medical School, Boston, Massachusetts, United States of America

Abstract

Prion diseases are characterised by the accumulation of PrP<sub>Sc</sub>, an abnormally folded isoform of the cellular prion protein (PrP<sub>C</sub>), in affected tissues. Following peripheral exposure high levels of prion-specific PrP<sub>Sc</sub> accumulate first upon follicular dendritic cells (FDC) in lymphoid tissues before spreading to the CNS. Expression of PrP<sub>C</sub> is mandatory for cells to sustain prion infection and FDC appear to express high levels. However, whether FDC actively replicate prions or simply acquire them from other infected cells is uncertain. In the attempts to-date to establish the role of FDC in prion pathogenesis it was not possible to dissociate the Prnp expression of FDC from that of the nervous system and all other non-haematopoietic lineages. This is important as FDC may simply acquire prions after synthesis by other infected cells. To establish the role of FDC in prion neuroinvasion transgenic mice were created in which PrPC expression was specifically “switched on” or “off” only on FDC. We show that PrPC-expression only on FDC is sufficient to sustain prion replication in the spleen. Furthermore, prion replication is blocked in the spleen when PrP<sub>C</sub>-expression is specifically ablated only on FDC. These data definitively demonstrate that FDC are the essential sites of prion replication in lymphoid tissues. The demonstration that Prnp-ablation only on FDC blocked splenic prion accumulation without apparent consequences for FDC status represents a novel opportunity to prevent neuroinvasion by modulation of PrP<sub>C</sub> expression on FDC.

Introduction

Prion diseases (Transmissible spongiform encephalopathies; TSE) are sub-acute neurodegenerative diseases that affect both humans and animals. Many prion diseases, including natural sheep scrapie, bovine spongiform encephalopathy, chronic wasting disease in mule deer and elk, and kuru and variant Creutzfeldt-Jakob disease in humans, are acquired by peripheral exposure (e.g. orally or via lesions in skin or mucous membranes). After peripheral exposure prions accumulate first upon follicular dendritic cells (FDC) as they make their journey from the site of infection to the CNS (a process termed, neuroinvasion) [1–7]. FDC are a unique subset of stromal cells resident within the primary B cell follicles and germinal centres of lymphoid tissues [8]. Prion accumulation upon FDC is critical for efficient disease pathogenesis as in their absence neuroinvasion are impaired [1–4]. From the lymphoid tissues prions invade the CNS via the peripheral nervous system [9].

During prion disease aggregations of PrP<sub>Sc</sub>, an abnormally folded isoform of the cellular prion protein (PrP<sub>C</sub>), accumulate in affected tissues. Prion infectivity co-purifies with PrP<sub>Sc</sub> [10] and is considered to constitute the major, if not sole, component of infectious agent [11]. Host cells must express cellular PrP<sub>C</sub> to sustain prion infection [12] and FDC appear to express high levels of PrP<sub>C</sub> on the cell membrane in uninfected mice [13,14]. Although prion neuroinvasion from peripheral sites of exposure is dependent upon the presence of FDC in lymphoid tissues, it is not known whether FDC actually replicate prions themselves. FDC characteristically trap and retain native antigen on their surfaces for long periods in the form of immune complexes, consisting of antigen-antibody and/or complement components. Prions are also considered to be acquired by FDC as complement-opsinized immune complexes [15–18]. Thus, during prion infection FDC might simply trap and retain PrP<sub>Sc</sub>-containing immune complexes on their surfaces following synthesis by other infected cells such as neurones.

Many cell types including classical DC, lymphocytes, mast cells, platelets, reticulocytes and epithelial cells secrete membrane vesicles termed exosomes that are enriched in cell-specific protein [19,20]. Although the functions of exosomes are uncertain FDC can bind them on their surfaces. These microvesicles permit FDC to passively acquire and display proteins on their surfaces that they do not express at the mRNA level [21]. Studies have shown that prions only accumulated in the spleens of mice in which the FDC-
Prion diseases are infectious neurological disorders and are considered to be caused by an abnormally folded infectious protein termed PrPSc. Soon after infection, prions accumulate first upon follicular dendritic cells (FDC) in lymphoid tissues before spreading to the brain where they cause damage to nerve cells. Cells must express the normal cellular prion protein PrPc to become infected with prions. However, whether FDC are infected with prions or simply acquire them from other infected cells is unknown. To establish the role of FDC in prion disease PrPc expression was specifically “switched on” or “off” only on FDC. We show that PrPc-expressing FDC alone are sufficient to sustain prion replication in the spleen. Furthermore, prion replication is blocked in the spleen when PrPc-expression is switched off only on FDC. These data definitively demonstrate that FDC are the essential sites of prion replication in lymphoid tissues.

Author Summary

Prion diseases are infectious neurological disorders and are considered to be caused by an abnormally folded infectious protein termed PrPSc. Soon after infection, prions accumulate first upon follicular dendritic cells (FDC) in lymphoid tissues before spreading to the brain where they cause damage to nerve cells. Cells must express the normal cellular prion protein PrPc to become infected with prions. However, whether FDC are infected with prions or simply acquire them from other infected cells is unknown. To establish the role of FDC in prion disease PrPc expression was specifically “switched on” or “off” only on FDC. We show that PrPc-expressing FDC alone are sufficient to sustain prion replication in the spleen. Furthermore, prion replication is blocked in the spleen when PrPc-expression is switched off only on FDC. These data definitively demonstrate that FDC are the essential sites of prion replication in lymphoid tissues.

containing stromal compartment expressed PrPc [13,14]. However, in each of those studies it was not possible to dissociate the Prnp expression status of the FDC from that of the nervous system and all other host-derived non-haematopoietic and stromal cell populations [13,14,22]. This is important as prion infection can occur within inflammatory PrPc-expressing stromal cells that are distinct from FDC [23]. Furthermore, as both PrPc and PrPsc can be released from cells in association with exosomes [20] FDC may passively acquire PrPc and prions after release in exosomes from other infected cells [24,25].

No therapies are available to treat prions diseases. A thorough characterization of the host cells that are infected by prions is imperative for the identification of candidate molecular targets for therapeutic intervention, the development of useful pre-clinical diagnostics and to aid our understanding of the risk of transmission. To definitively determine the role of FDC in prion pathogenesis, two unique compound transgenic mouse models were created in which PrPc expression was specifically “switched on” or “switched off” only on FDC. These mice were then used to establish: i) whether FDC express PrPc or simply acquire it from other host cells; and ii) whether FDC amplify prions, or simply acquire them from other infected host cells. Our data clearly show that PrPc-expressing FDC alone are sufficient to sustain prion replication in the spleen. Furthermore, prion replication in the spleen is blocked in mice in which PrPc-expression is specifically ablated only on FDC.

Results

Mice expressing Cre recombinase specifically in FDC

To study FDC-specific gene function transgenic mice were used that expressed Cre recombinase under the control of the C2 locus (CD21-Cre mice) which directs expression in FDC and mature B cells [26,27]. First the cellular specificity of the Cre recombination was assessed by crossing the CD21-Cre mice with the ROSA26stop/geo reporter strain [28]. Histological analysis showed efficient LacZ expression indicative of Cre-mediated gene recombination in FDC and B cell follicles in the spleens, lymph nodes and Peyer’s patches of CD21-Cre ROSA26stop/geo mice (Figure 1A, B). No recombination was observed in FDC and mature B cells in the spleens of ROSA26stop/geo reporter mice that lacked Cre expression (Figure 1B). Unlike lymphocytes, FDC do not derive from bone marrow precursors [29]. As a consequence, it is possible to mix-and-match the genotype of FDC and lymphocytes by grafting bone marrow cells from donor mice into recipients of a different genetic background [13,14,22]. To restrict Cre-expression to FDC, adult CD21-Cre ROSA26stop/geo mice were lethally γ-irradiated and 24 h later reconstituted with bone marrow from Cre-deficient C57BL/6 wild-type (WT) mice (termed WT→CD21-Cre ROSA26stop/geo mice) and tissues from six mice from each group analysed 100 days after transfusion. Using this approach, in these mice all B cells lack Cre-expression as they derive from the WT donor bone marrow, whereas the FDC express Cre as they are host-derived. Analysis of the cellular sites of LacZ expression in WT→CD21-Cre ROSA26stop/geo mice confirmed that Cre-mediated recombination was associated with FDC (Figure 1B). No other cellular sites of Cre-mediated recombination were observed in the spleens of WT→CD21-Cre ROSA26stop/geo mice. Furthermore, no other cellular sites of Cre-mediated recombination were observed in a wide range of non-lymphoid peripheral tissues from CD21-Cre ROSA26stop/geo and WT→CD21-Cre ROSA26stop/geo (heart, liver, kidney, pancreas, ear, tongue, skeletal, muscle, ovary, uterus, bladder, testes, epididymis, sciatic nerve and spinal cord; data not shown). These data clearly demonstrate that CD21-Cre mice are a useful tool to study FDC-specific gene expression and function.

Expression of Cre recombinase by the Cr2 promoter is not toxic to FDC

Cre toxicity can occur in some Cre transgenic mouse lines whereby Cre recombinase causes mis-recombination, DNA damage and death of Cre-expressing cells [30]. However, immunohistochemical (IHC) analysis of spleens from CD21-Cre ROSA26stop/geo mice and WT→CD21-Cre ROSA26stop/geo mice showed no significant effect of Cre-expression on the status of FDC networks and B cell follicles when compared to spleens from WT control mice and ROSA26stop/geo mice that lacked Cre expression (Figure 1C). Furthermore, the expression of Cre recombinase under the control of the C2 locus had no observable effect on CD21/35 expression (Figure 1C).

FDC express Prnp and do not acquire PrPc from neighbouring cells

Next, mice were created in which Prnp expression (which encodes PrPc) was restricted only to FDC. To do so, CD21-Cre mice were first bred onto a Prnp-deficient (Prnpstop/-) background. The resulting CD21-Cre Prnpstop/- mice were then crossed with Prnpstop/+ mice in which a floxed βgeo stop cassette was inserted into intron 2 of the Prnp gene upstream of exon 3 [31]. In the progeny CD21-Cre Prnpstop/+ mice, PrPc is only expressed in cells expressing Cre recombinase (CD21-expressing FDC and mature B cells). To restrict the Prnp-expression to FDC, CD21-Cre Prnpstop/+ mice were lethally γ-irradiated and grafted with bone marrow from Cre-deficient Prnpstop/+ mice (Prnpstop/+ → CD21-Cre Prnpstop/+ mice). We also performed bone marrow transfers from CD21-Cre Prnpstop/+ donors into CD21-Cre Prnpstop/+ recipients (CD21-Cre Prnpstop/+ → CD21-Cre Prnpstop/+ mice). In CD21-Cre Prnpstop/+ recipients, PrPc is only expressed in cells expressing Cre recombinase (CD21-expressing FDC and mature B cells). To restrict the Prnp-expression to FDC, CD21-Cre Prnpstop/+ mice were lethally γ-irradiated and grafted with bone marrow from Cre-deficient Prnpstop/+ mice (Prnpstop/+ → CD21-Cre Prnpstop/+ mice). We also performed bone marrow transfers from CD21-Cre Prnpstop/+ donors into CD21-Cre Prnpstop/+ recipients (CD21-Cre Prnpstop/+ → CD21-Cre Prnpstop/+ mice) and Prnpstop/+ donors into Prnpstop/+ recipients (Prnpstop/+ → Prnpstop/+ mice) as controls (Figure 2A).

Spleens, tails and blood from six mice from each group were examined 100 days after bone marrow transfusion. PCR analysis of DNA isolated from the tails, blood and spleens of mice in each group was used to confirm the presence of Cre (Figure 2B, upper panel) and Cre-mediated DNA recombination (Figure 2B, lower panel) within the stromal, haematopoietic or both compartments (respectively). The detection of Cre in the tail and spleen but not
Figure 1. Cre-mediated gene recombination in FDC in the spleens, lymph nodes and Peyer’s patches of CD21-Cre ROSA26^flox/Flox mice. A) Analysis of the cellular sites of LacZ expression (blue) in the spleens, inguinal lymph nodes, Peyer’s patches and mesenteric lymph nodes of CD21-Cre ROSA26^flox/Flox mice shows Cre-mediated recombination in a focus of cells within the B cell follicles. Sections were counterstained with nuclear fast red (red). B) IHC analysis of FDC (CD35^+ cells, upper row, red) and B cells (CD45R^+ cells, lower row, red) confirmed that Cre-mediated LacZ expression (blue) was associated with FDC in the spleens of WT→CD21-Cre ROSA26^flox/Flox mice. No LacZ expression was associated with FDC in spleens from ROSA26^flox/Flox mice that lacked Cre. C) IHC analysis of the status of FDC (CD35^+ and C4-binding cells; red) and B cells expressing CD45R, CD19, and CD1d (red) in spleens from WT, CD21-Cre ROSA26^flox/Flox, WT→CD21-Cre ROSA26^flox/Flox and ROSA26^flox/Flox mice. Scale bars 100 μm. n = 6 mice/group.

doi:10.1371/journal.ppat.1002402.g001
Figure 2. FDC-restricted PrP<sup>C</sup> expression in the spleens of Prnp<sup>stop</sup>→CD21-Cre Prnp<sup>stop</sup> mice. A) The anticipated distribution of PrP<sup>C</sup> expression on FDC and B cells in tissues from each mouse group. B) The detection of Cre in the tail and spleen but not blood of the Prnp<sup>stop</sup>→CD21-Cre Prnp<sup>stop</sup> mice confirmed the restriction of the Cre-expression to the stromal but not haematopoietic compartments of these mice (upper panel). Efficient Cre-mediated recombination of Prnp<sup>stop</sup> was restricted to the FDC-containing stromal compartment of the spleens of Prnp<sup>stop</sup>→CD21-Cre Prnp<sup>stop</sup> mice when compared to control mice. Cre-mediated recombination by CD21-expressing lymphocytes was efficiently prevented in these mice by the irradiation and transfer of Prnp<sup>stop</sup> bone marrow as demonstrated by the lack of a Prnp<sup>stop</sup> band in DNA extracted.
blood of the Prnp<sup>stop/-</sup>→CD21-Cre Prnp<sup>stop/-</sup> mice confirmed the restriction of the Cε-expression to the stromal but not hematopoietic compartments of these mice. In addition, PCR analysis also confirmed that in these mice efficient Cre-mediated recombination of the Prnp<sup>stop</sup> allele was restricted to the FDC-containing stromal compartment of the spleen (Figure 2B). In Prnp<sup>stop/+</sup>→CD21-Cre Prnp<sup>stop/+</sup> mice the recombined Prnp<sup>stop</sup> allele (Prnp<sup>stop/B+</sup>) was detected in the spleen, but not blood and tail. Thus, these data indicate that in the spleens of Prnp<sup>stop/+</sup>→CD21-Cre Prnp<sup>stop/+</sup> mice Cre-mediated recombination is restricted to FDC and not B cells. As anticipated, in the spleens of Prnp<sup>+/+</sup>→Prnp<sup>+/+</sup> control mice high levels of PrP<sub>C</sub> expression were observed upon FDC and tyrosine hydroxylase (TH)-positive sympathetic nerves (Figure 2C). In contrast, in the spleens of Prnp<sup>stop/+</sup>→CD21-Cre Prnp<sup>stop/+</sup> mice PrP<sub>C</sub> was only expressed on FDC (Figure 2C). In the absence of Cre-recombinase expression by FDC and peripheral nerves in CD21-Cre Prnp<sup>stop/+</sup>→Prnp<sup>stop/-</sup> mice, PrP<sub>C</sub> expression was not expressed by either cell population (Figure 2C).

Morphometric analysis confirmed that the amount of the PrP<sub>C</sub> expression co-localized upon the surfaces of FDC in the spleens of Prnp<sup>stop/+</sup>→CD21-Cre Prnp<sup>stop/+</sup> mice was not significantly different from that observed upon FDC in spleens from Prnp<sup>+/+</sup>→Prnp<sup>+/+</sup> control mice (P<0.69, n = 48 FDC networks/group). In contrast, in the absence of Cre-recombinase expression by FDC in CD21-Cre Prnp<sup>stop/+</sup>→Prnp<sup>stop/-</sup> mice, PrP<sub>C</sub> expression was substantially lower than that observed upon FDC in spleens from Prnp<sup>+/+</sup>→Prnp<sup>+/+</sup> control mice (P<1×10<sup>-25</sup>, n = 48 FDC networks/group). E) Morphometric analysis confirmed that PrP<sub>C</sub> expression upon the surfaces of sympathetic nerves in the spleens of Prnp<sup>stop/+</sup>→CD21-Cre Prnp<sup>stop/+</sup> mice was similar to that observed upon FDC in spleens from Prnp<sup>+/+</sup>→Prnp<sup>+/+</sup> control mice (P<0.969, n = 48 FDC networks/group). In contrast, in the absence of Cre-recombinase expression by FDC in CD21-Cre Prnp<sup>stop/+</sup>→Prnp<sup>stop/-</sup> mice, PrP<sub>C</sub> expression was substantially lower than that observed upon FDC in spleens from Prnp<sup>+/+</sup>→Prnp<sup>+/+</sup> control mice (P<1×10<sup>-25</sup>, n = 48 FDC networks/group). For all panels n = 6 mice/group.

doi:10.1371/journal.ppat.1002402.g002
spleen. Furthermore, consistent with data above, our analysis showed that PrP expression only of FDC was sufficient to sustain high levels of prion infectivity within the spleen (Figure S2). These data demonstrate that PrP expression only on FDC is sufficient to sustain prion replication in the spleen. In the absence of PrP expression on FDC, the prions appeared to be scavenged by tingible body macrophages resident within the B cell follicles.

FDC-specific Prnp-ablation

Next, mice were created in which Prnp expression was specifically ablated in FDC. To do so, CD21-Cre Prnp−/− mice were crossed with mice carrying a “floxed” Prnp gene (Prnp flox/flox mice; [31]). In the progeny CD21-Cre Prnp flox/−, mice, Prnp expression is conditionally ablated in cells expressing Cre recombinase (CD21-expressing FDC and mature B cells). To restrict the Prnp-ablation to FDC, CD21-Cre Prnp flox/− mice were lethally γ-irradiated and grafted with bone marrow from Cre-deficient Prnp flox/−/− mice (Prnp flox/−→CD21-Cre Prnp flox/−−/− mice). We also performed bone marrow transfers from CD21-Cre Prnp flox/−−/− donors into CD21-Cre Prnp flox/− recipients (CD21-Cre Prnp flox/−−/− mice, CD21-Cre Prnp flox/−−/− mice into Cre-deficient Prnp flox/−−/− mice (CD21-Cre Prnp flox/−−/− mice, and Prnp−/−−/− donors into Prnp−/−−/− recipients (Prnp−/−−/−→Prnp−/−−/− mice) as controls (Figure 5A). Spleens, tails and blood from 6 mice from each group were examined 100 days after bone marrow transplantation. PCR analysis of DNA isolated from the spleens, blood and tails of each mouse were collected 70 days after exposure. As anticipated, heavy PrPd accumulations consistent with localisation upon FDC were detected in spleens from Prnp−/−−/− and CD21-Cre Prnp−/−−/− mice (Figure S2). These data indicate that in the spleens of Prnp−/−−/− and CD21-Cre Prnp−/−−/− mice and Prnp−/−−/− control mice (n = 32 FDC networks/group). D and E, Comparison of the sympathetic innervation in spleens from each mouse group. D) IHC detection of TH-positive sympathetic nerves (green) and FDCs (CD35+ cells; red). Scale bar, 50 µm. E) Quantitative analysis of the relative positioning of the FDC and sympathetic nerves showed there were no significant differences in average distance between these cell populations in spleens from each mouse group (P = 0.932, n = 48 FDC networks/group). For all panels n = 6 mice/group.

doi:10.1371/journal.ppat.1002402.g003

Effect of FDC-specific Prnp-ablation on FDC status and splenic microarchitecture

Data in the current study definitively demonstrate that FDC express high levels of PrP but the role PrP plays in FDC function and homeostasis is not known. IHC analysis showed that the microarchitecture of the FDC networks from Prnp−/−−/− CD21-Cre Prnp−/−−/− mice were normal when compared to control mice (Figure 6A). Furthermore, no significant difference was observed in the size (P = 0.750, n = 32) and number (P = 0.713, n = 32) of the FDC networks in spleens from each mouse group (Figure 6B & C, respectively). The relative positioning of the FDC and sympathetic nerves was likewise similar in spleens from each mouse group (Figure 6D & E; P<0.765, n = 48).

FDC characteristically trap and retain native antigen on their surfaces in the form of immune complexes, consisting of antigen-antibody and/or complement components. Antigens trapped on the surface of FDC are considered to promote immunoglobulin-isotype class switching, affinity maturation of naive IgM B cells and the maintenance of immunological memory [38–42]. Indeed, prions are also considered to be acquired by FDC as complement-opsonized immune complexes [15–18]. To determine whether antigen retention by Prnp-ablated FDC was affected six mice from each group were passively immunized with preformed PAP immune complexes, and 24 h later, the presence of FDC-antigen retention by Prnp-ablated FDC was affected six mice from each group.
with data above (Figure 4), in spleens of mice with PrPC-deficient FDC PrP accumulations were only occasionally observed within tingible body macrophages (Figure 8A and B, arrowheads; Figure S1). We also analysed prion infectivity levels in spleens from Prnp<sup>flox/−</sup>→CD21-Cre Prnp<sup>lox/−</sup> mice in which cellular PrP<sup>C</sup> expression was ablated only on FDC (Figure S2; n=3). Consistent with data above this analysis showed that in the absence of PrP expression only on FDC the accumulation of high levels of prion infectivity in the spleen was blocked (Figure S2). Taken together, these data show that in the specific absence of PrP<sup>C</sup> expression FDC are unable to sustain prion replication upon their surfaces and as a consequence the agent is scavenged by tingible body macrophages.

FDC-restricted PrP<sup>C</sup>-ablation does not influence prion disease and susceptibility when infection is established directly within the CNS

When mice with PrPC-ablated FDC (Prnp<sup>lox/−</sup>→CD21-Cre Prnp<sup>lox/−</sup> mice) were injected intracerebrally (i.c.) with the ME7 scrapie agent strain directly into the CNS all mice succumbed to clinical signs of scrapie approximately 300 days after exposure with incubation periods indistinguishable from those of Prnp<sup>+/+</sup>-control mice [43] (Prnp<sup>lox/−</sup>→CD21-Cre Prnp<sup>lox/+</sup>, 297±4 days, n=4; Prnp<sup>−/−</sup>, 290±4 days, n=5; P=0.386). Histopathological analysis showed that brains from all clinically-affected mice from each group displayed the characteristic spongiform pathology, astrogliosis, microgliosis and PrP<sup>D</sup> accumulation typically associated with terminal infection with the ME7 scrapie agent (Figure S3, third and fourth columns). In contrast, none of the mice with PrPC-ablated FDC (Prnp<sup>lox/−</sup>→CD21-Cre Prnp<sup>lox/−</sup> mice, n=0/6; CD21-Cre Prnp<sup>lox/−</sup>→CD21-Cre Prnp<sup>lox/−</sup> mice, n=0/7) succumbed to clinical prion disease during their life-spans (Table S1). Although we cannot exclude the possibility that if the clinically-negative mice with PrPC-ablated FDC mice had lived longer some may have succumbed to clinical prion disease after substantially extended incubation periods, no PrP<sup>F</sup> or other characteristic histopathological hallmarks of prion disease were detected in their brains (Figure S3, first two columns). Together, these data suggest that in the specific absence of PrP<sup>F</sup> expression on FDC neuroinvasion following peripheral exposure is impaired.

**Discussion**

These data definitively demonstrate that FDC are essential sites of prion replication in lymphoid tissues. In order to precisely establish the role of FDC in prion pathogenesis two unique compound transgenic mouse models were created in which PrP<sup>F</sup> expression was specifically “switched on” or “off” only on FDC. Our data confirm that FDC express high levels of PrP<sup>F</sup> and do not simply acquire it from other host cells. Furthermore, we show that following peripheral exposure PrP<sup>F</sup>-expressing FDC alone are sufficient to sustain high levels of prion replication in the spleen. Accordingly, when PrP<sup>F</sup>-expression was specifically ablated only on FDC prion replication in the spleen was blocked. These data likewise demonstrate that FDC do not simply acquire prions after their release from other infected host cells. Our analysis showed that the effects of Prnp-ablation on prion replication in the spleen were specific to FDC and had no effect on prion neuropathogenesis when the infection was established directly within the CNS. In the absence of PrP<sup>F</sup> expression on FDC the PrP<sup>S</sup> from the initial inoculum appeared to be scavenged by tingible body macrophages residing within the B cell follicles. Together, these data definitively demonstrate that FDC are the critical early sites of prion replication in lymphoid tissues. This study is the first to demonstrate that the specific ablation of a cellular protein only on FDC, without apparent consequences for FDC status and function, blocks the replication of an important pathogen in the spleen.

FDC reside in the primary B cell follicles and germinat centres of lymphoid tissues and are a completely distinct cell lineage from bone-marrow-derived classical dendritic cells [47–49]. FDC possess many slender and convoluted dendritic processes which provide the FDC with an extremely large surface area. This helps the FDC to efficiently trap and retain large amounts of native antigen in the form of immune complexes, consisting of antigen-
Figure 5. FDC-restricted PrP\textsuperscript{C}-ablation in the spleens of Prnp\textsuperscript{flox/-}–CD21-Cre Prnp\textsuperscript{flox/-} mice. A) The anticipated distribution of PrP\textsuperscript{C} expression on FDC and B cells in tissues from each mouse group. B) PCR analysis of DNA isolated from the spleens, blood and tails of Prnp\textsuperscript{flox/-}–CD21-Cre Prnp\textsuperscript{flox/-} mice confirmed that efficient Cre-mediated DNA recombination and Prnp-ablation (Prnp\textsuperscript{deflox}) was restricted to the FDC-containing stromal compartment of the spleen. Cre-mediated recombination of CD21-expressing lymphocytes was efficiently prevented in these mice by the irradiation and transfer of Prnp\textsuperscript{flox/-} bone marrow as demonstrated by the lack of a Prnp\textsuperscript{deflox} band in DNA extracted from blood (lower panel). B, blood; S, spleen; T, tail; M, DNA size markers; a, b, c, d control DNA samples for each transgene combination tested which were (a) Prnp\textsuperscript{flox/flox}, (b) Prnp\textsuperscript{flox/-}, (c) Prnp\textsuperscript{-/-} and (d) Prnp\textsuperscript{flox/-} with complete recombination of the floxed exon 3. C) IHC analysis of PrP\textsuperscript{C} expression (blue) by FDC (CD35\textsuperscript{+} cells; Role of Follicular Dendritic Cells in Prion Pathogenesis

<table>
<thead>
<tr>
<th>Gp</th>
<th>Mouse model</th>
<th>Cells expressing PrP\textsuperscript{C}</th>
<th>Nerves, other stromal cells, etc.</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Prnp\textsuperscript{flox/-}–CD21-Cre Prnp\textsuperscript{flox/-}</td>
<td>no</td>
<td>yes</td>
</tr>
<tr>
<td>2</td>
<td>CD21-Cre Prnp\textsuperscript{flox/-}–CD21-Cre Prnp\textsuperscript{flox/-}</td>
<td>no</td>
<td>no</td>
</tr>
<tr>
<td>3</td>
<td>CD21-Cre Prnp\textsuperscript{flox/-}–Prnp\textsuperscript{flox/-}</td>
<td>yes</td>
<td>no</td>
</tr>
<tr>
<td>4</td>
<td>Prnp\textsuperscript{-/-}</td>
<td>yes</td>
<td>yes</td>
</tr>
</tbody>
</table>
antibody and/or complement components. The longevity of FDC ensures that antigen is retained upon their surfaces for long periods [50,51]. Antigens trapped on the surface of FDC are considered to promote immunoglobulin-isotype class switching, affinity maturation of naive IgM B cells and the maintenance of immunological memory [38–42]. FDC are also considered to aid the clearance of apoptotic B lymphocytes [52], and play a role in infection with human immunodeficiency virus [53] and the pathogenesis of chronic inflammatory and autoimmune diseases [54] and peripherally-acquired prion infections.

A number of studies have addressed the role of FDC in prion pathogenesis. They show that prion replication in the spleen and subsequent neuroinvasion are both impaired in immunodeficient mice that lack FDC [4,44,45], or following their temporary deletion [1,33,46]. Although the precise identity of FDC: precursor cells is unknown, other studies have exploited their non-haematopoietic origins to address their role in prion pathogenesis. In these bone marrow chimaera studies, mismatches were created in Prnp expression between the FDC-containing stromal and haematopoietic compartments by grafting bone marrow cells from PrP-deficient (Prnp-/-) mice into PrP-expressing wild-type mice, and vice versa [13,14]. Using this approach FDC and all other stromal cells were derived from the recipient, whereas lymphocytes and other haematopoietic lineages were derived from the donor cells. Following peripheral exposure, prion accumulation upon FDC was only detected in the spleens of mice with a Prnp-expressing stromal compartment.

While the above studies clearly show that the presence of FDC is important for prion replication in the spleen, it was not possible to dissociate the Prnp expression status of FDC from that of the nervous system and all other non-haematopoietic host-cell populations and therefore precisely characterise the role of FDC in prion neuroinvasion [13,14]. This is important for a number of reasons. Firstly, prion infection can occur within inflammatory PrP+-expressing stromal cells that are distinct from FDC [23]. Secondly, the FDC's ability to bind exosomes may have lead to the wrong interpretation to be made in earlier studies describing their ontology [55]. Most evidence indicates that FDC do not derive from haematopoietic precursors [29,49]. However, the detection of donor bone marrow derived MHC class-I molecules, and other donor-derived antigens, on the surface of FDC in recipient mice was considered evidence of FDC precursor cells within bone marrow [55]. With hindsight these observations are most likely due to the FDC's capacity to acquire exosome-associated antigens from other cell types [21]. Both PrPSc and PrPSc can be released from cells in association with exosomes [20]. The possibility, therefore, cannot be excluded that FDC passively acquire prions after their release in exosomes from other infected non-haematopoietic cell populations. Finally, FDC characteristically trap and retain immune complexes on their surfaces. FDC express negligible levels of complement component C4 at the mRNA level but the detection of abundant activated C4 on their surfaces by IHC using mAb FDC-M2 (as used in this study) is indicative of the capture and retention of immune complexes by FDC [32].

Opossum subcomponent components and cellular CR are likewise considered to play an important role in the retention of prions by FDC [15,16,18]. Thus FDC may simply act as concentrating depots for prion-containing complement-opsonized immune complexes.

The practical hurdles that are encountered when attempting to isolate highly purified FDC from lymphoid tissues have made detailed analysis of their pathobiological functions extremely difficult. The main issues include: contamination with other cell types such as B cells and tingle body macrophages which express MFGE8 (FDC-M1), a common marker used to identify FDC [52,56], low yield, and their dependence on constitutive lymphotoxin β receptor-stimulation to maintain their differentiated state [57]. FDC and mature B cells express prion receptors [58] which encode the complement receptors CR2/CR1 (CD21/35) [18,27]. A previous study used CD21-cre mice to study FDC-specific gene function [27]. In the current study, our data confirm that Cre/loxP-mediated DNA recombination was specific to FDC and mature B cells in CD21-cre mice, and could be restricted to FDC by transfusing the mice with Cre-deficient bone marrow. In some Cre transgenic mouse lines Cre-toxicity is encountered whereby Cre recombinase causes mis-recombination, DNA damage and death of Cre-expressing cells [30]. However, our analysis suggested no significant effect of Cre-expression on the number, size and status of FDC networks and B cell follicles. CD21-cre mice are therefore a powerful in vivo tool in which to study FDC-specific gene expression and function.

Expression of PrPSc is mandatory for host cells to sustain prion infection [43]. In the current study to establish whether FDC actively amplify prions a compound transgenic mouse model was created using the CD21-cre mouse line to specifically "switch on" PrPSc expression only on FDC (Prnpfloxed→CD21-Cre Prnpfloxed/mice). As a consequence, only FDC in these mice had the potential to be actively infected with and replicate prions. Our analysis showed that expression of PrPSc only on FDC was sufficient to sustain high levels of PrPSc accumulation upon FDC in the spleen after peripheral prion exposure. These data definitively demonstrate that FDC are the critical sites of prion replication in lymphoid tissues. Ultrastructural analysis of the cellular compartments within which PrPSc localizes upon/within FDC has failed to show any intracellular accumulation. Instead the PrPSc appears to be restricted to the plasmalemma of their dendritic processes [58]. This implies that early de novo PrPSc conversion occurs upon the surface of FDC.

A second compound transgenic mouse model was created in which PrPSc expression was specifically "switched off" only on FDC (Prnpfloxed→CD21-Cre Prnpfloxed/mice). If, as shown above, FDC do actively amplify prions, then one would also expect the specific ablation of PrPSc expression only on FDC to block prion replication in the spleen. Our data confirmed this to be the case. As PrPSc expression in all other host cells (eg neurons) in these mice was unaffected, these data clearly show that FDC do not simply acquire prions following release from other infected host cells, even in mice with clinical prion disease in the brain. IHC
analysis implied that in the spleens of mice with PrP-deficient FDC the prions appeared to be scavenged by tingible body macrophages resident within the B cell follicles. The lack of detection of PrPSc within tingible body macrophages in the spleens of clinically-affected mice with PrP-deficient FDC (Figure 9) clearly demonstrates that these cells are not alternative sites of replication of ME7 scrapie prions. High levels of prions rapidly accumulate within the spleen and other lymphoid tissues within weeks of peripheral exposure. The magnitude of the prion accumulation within the spleen rapidly reaches a plateau level which is maintained for the duration of the disease [13,44]. The maintenance of this plateau may be the consequence of a competitive state whereby FDC act to amplify prions above the threshold required to achieve neuroinvasion, whereas phagocytic cells such as macrophages act to destroy them [59,60]. Indeed increased numbers of PrPSc-containing tingible body macrophages are found within the B cell follicles of TSE-affected animals [58]. Thus, our data suggest that in the specific absence of PrPC expression by FDC the initial inoculum is phagocytosed and gradually degraded by mononuclear phagocytes such as tingible body macrophages [59,60]. These data are congruent with data from our earlier study which likewise occasionally detected trace levels of prions from the initial inoculum within tingible body macrophages in the spleens of mice with a PrPSc-deficient FDC-containing stromal compartment [13].

The density of sympathetic nerves can significantly influence the amount of prion accumulation in the spleen [33]. In the current study the distribution of TH-positive sympathetic nerves in the spleens of the FDC-specific gene targeted mouse lines was not adversely affected. Furthermore, when prions were injected directly to the brain, FDC-specific Prnp ablation had no influence on the onset of clinical disease or the neuropathology. These data provide strong evidence that the effects of Cre-mediated Prnp ablation on prion replication in the spleen were specific to FDC and not due to unregulated ablation of PrPSc expression within the nervous system. In the current study PrPSc accumulation upon PrPSc-ablated FDC (Prnpflox/−→CD21-Cre Prnpflox/− mice) was blocked even in spleens from i.c. injected clinically-scrapie affected mice. These data contrast those reported by Crozet and colleagues [61] which used Tg(OvPrP4) mice that express the ovine Prnp gene under the control of the neuron-specific enolase promoter on neurones it is plausible that greater prion replication occurred within the peripheral nervous system, which may have been subsequently trapped on the surface of the FDC and scavenged by macrophages as the prion burden increased. Similarly, hyper-immunisation of the spleen likewise leads to increase prion burden in this tissue [33].

In conclusion, our data demonstrate that PrPSc-expressing FDC are the essential sites of prion replication in lymphoid tissues. Indeed, PrPSc-expression on FDC alone was sufficient to sustain high levels of prion replication. In contrast, the specific ablation of PrPC expression on FDC blocked prion replication. Although FDC have the capacity to bind exosomes and immune complexes which may contain PrPSc, this finding clearly demonstrates that FDC do not simply passively acquire prions from other infected cell populations such as neurons. Previous data show treatments which impair the status or immune complex-trapping function of FDC reduce prion susceptibility after peripheral exposure [1,16,35,46,62]. The demonstration that Prnp-ablaction only on FDC blocked splenic prion replication without apparent consequences for FDC status represents a novel opportunity to prevent neuroinvasion by modulation of PrPSc expression on FDC.

Materials and Methods

Ethics statement

All studies using experimental mice and regulatory licences were approved by both The Roslin Institute’s and University of Edinburgh’s Protocols and Ethics Committees. All animal experiments were carried out under the authority of a UK Home Office Project Licence within the terms and conditions of the strict regulations of the UK Home Office ‘Animals (scientific procedures) Act 1986’. Where necessary, anaesthesia appropriate for the procedure was administered, and all efforts were made to minimize harm and suffering. Mice were humanely culled using a UK Home Office Schedule One method.

Mice

The CD21-Cre [26], ROSA26lox/lox reporter strain [28], PrPflox/+ [12] mice and tga20 mice over-expressing PrP [63] were generated as described previously. Prnpflox/− mice have loxP sites flanking exon 3 of the Prnp gene [31]. Prnpflox/+ mice have a floxed β-gro cassette inserted into intron 2 of the Prnp gene upstream of exon 3 [31]. Mice were maintained under SPF conditions.

Genotype confirmation by PCR analysis

Prior to their use in experiments, the genotype of each mouse was confirmed by PCR analysis. DNA was prepared from tails, blood and spleens using the DNaseasy blood and tissue kit (Qiagen, Crawley, UK) according to the manufacturer’s instructions. Where indicated DNA samples were analysed for presence of Cre, LacZ, Prnpflox/+ , Prnpflox/− , Prnpflox, recombined Prnpflox (Prnpflox/+ ), Prnpflox and recombined Prnpflox (Prnpflox/−) using the primers listed in Table 1. PCR products were resolved by electrophoresis through a 1.0% agarose gel containing 0.002% GelRed (Biotium, Cambridge Biosciences Ltd, Cambridge, UK).
Figure 7. Effect of FDC-restricted PrP<sup>C</sup>-ablation on immune complex trapping. A) Mice were passively immunized with preformed PAP immune complexes, and 24 h later, the presence of immune complexes (red) upon FDC (CD35<sup>+</sup> cells, green) assessed by IHC. Scale bar, 100 μm. B) Morphometric analysis confirmed that the magnitude of the immune complex-trapping co-localized upon the surfaces of FDC from Prnp<sup>flox/null</sup> mice was not significantly different from that observed in spleens from control mice. This analysis also confirmed that the immune complexes were preferentially associated with FDC in these tissues and significantly greater than the null hypothesis that the pixels were randomly distributed. *, P<1 x 10<sup>-21</sup>; **, P<1 x 10<sup>-32</sup>; ***, P<9 x 10<sup>-28</sup>; n = 40 FDC networks/group. For all panels n = 6 mice/group.

doi:10.1371/journal.ppat.1002402.g007
Bone-marrow from the femurs and tibias of donor mice was prepared as single-cell suspensions (3 × 10^7–4 × 10^7 viable cells/ml) in HBSS (Invitrogen, Paisley, UK). Recipient adult (6–8 weeks old) mice were γ-irradiated (950 rad) and 24 h later reconstituted with 100 µl bone-marrow by injection into the tail vein. Recipient mice were used in subsequent experiments as described 100 days after bone marrow reconstitution to allow sufficient time for removal of PrPSc.
Figure 9. Effect of FDC-restricted PrP<sup>C</sup>-ablation on PrP<sup>Sc</sup> accumulation in the brains and spleens of scrapie-affected mice. Control mice (Prnp<sup>+</sup> mice) and Prnp<sup>flx/-</sup>→CD21-Cre Prnp<sup>flx/-</sup> mice that lacked PrP<sup>C</sup>-expressing FDC were injected i.c. with the scrapie agent directly into the CNS. Brains and spleens were collected from clinically scrapie-affected mice to compare the neuropathology and cellular sites of PrP<sup>Sc</sup> accumulation.

A) High levels of spongiform pathology (H&E, upper row), heavy accumulations of PrP<sup>d</sup> (brown, second row), reactive astrocytes expressing GFAP (brown, third row) and active microglia expressing Iba-1 (brown, bottom row) were detected in the hippocampi of the brains of all clinically scrapie-affected mice. Scale bars, 500 μm.

B) High levels of PrP<sup>d</sup> (red) were detected in association with FDC in spleens from clinically scrapie-affected control mice that contained PrP<sup>C</sup>-expressing FDC. In contrast, no PrP<sup>d</sup> or PrP<sup>Sc</sup> was detected in spleens of Prnp<sup>flx/-</sup>→CD21-Cre Prnp<sup>flx/-</sup> that lacked PrP<sup>C</sup>-expressing FDC. Scale bars = 500 μm.

doi:10.1371/journal.ppat.1002402.g009
mice were injected i.c. with 20 μl of each homogenate. The scrapie titre in each sample was determined from the mean incubation period in the indicator mice, by reference to a dose/incubation period response curve for ME7 scrapie-infected spleen tissue serially titrated in tga20 mice using the relationship: \( y = 4.5333 - 0.0595x \) \( (\gamma = \log \text{ID}_{50} \ \text{U}/20 \ \mu\text{l of homogenate}; \ x = \text{incubation period}; R^2 = 0.9562) \). As the expression level of cellular PrP controls the prion disease incubation period, tga20 mice overexpressing PrP\(^e\) are extremely useful as indicator mice in prion infectivity bioassays as they succumb to disease with much shorter incubation times than conventional mouse strains [63].

IHC and immunofluorescent analyses

Spleens were removed and snap-frozen at the temperature of liquid nitrogen. Frozen serial sections (10 μm in thickness) were cut on a cryostat and immunostained with the following antibodies: FDCs were visualized by staining with mAb 7G6 to detect CR2/CR1 (CD21/CD35; BD Biosciences PharMingen), mAb FDC-M2 to detect C4 (AMS Biotechnology, Oxon, UK) or mAb 8C12 to detect CR1 (CD35; BD Biosciences PharMingen). Cellular PrP\(^d\) was detected using PrP-specific polyclonal antibody (pAb) B13 [66]. B cells were detected using mAb B220 to detect CD45R (Caltag, Towcester, UK), or anti-CD19 (BD biosciences PharMingen). Marginal zone B cells were detected using mAb B1 to detect CD1d (BD Biosciences PharMingen). Sympathetic nerves were detected using tyrosine hydroxylase (TH)-specific pAb (Chemicon Europe).

For the detection of disease-specific PrP (PrP\(^d\)) in spleens and brains, tissues were fixed in periodate-lysine-paraformaldehyde fixative and embedded in paraffin wax. Sections (thickness, 6 μm) were deparaffinised, and pre-treated to enhance the detection of PrP\(^d\) by hydrated autoclaving (15 min, 121°C, hydration) and subsequent immersion formic acid (98%) for 5 min [67]. Sections were then immunostained with B3 PrP-specific pAb. For the detection of EGF-like module-containing mucin-like hormone receptor-like 1 (EMR1)-expressing macrophages, paraffin-embedded spleen sections were micro-waved in citric acid buffer (pH 6.0) for 10 min. Endogenous peroxidase activity was blocked using 1% hydrogen peroxidase in methanol, and macrophages detected using rat mAb F4/80 to detect EMR1 (clone CI:A3-1, AbD Serotec). For the detection of astrocytes, brain sections were immunostained with anti-glia fibrillary acidic protein (GFAP; Dako, Ely, UK). For the detection of microglia, deparaffinised brain sections were first pre-treated with Target Retrieval Solution (DAKO) and subsequently immunostained with anti-oxidized calcium-binding adaptor molecule 1 (Iba1; Wako Chemicals GmbH, Neuss, Germany). Immunolabelling was revealed using HRP-conjugated to the avidin-biotin complex (Novared kit, Vector laboratories, Peterborough, UK).

Table 1. PCR primers used to confirm the genotypes of mice used in this study.

<table>
<thead>
<tr>
<th>Allele</th>
<th>Details</th>
<th>Primer sequence</th>
<th>Product size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cre</td>
<td>Fwd</td>
<td>CGAGTGTAGGAGGTCGCAAGACCT</td>
<td>786</td>
</tr>
<tr>
<td></td>
<td>Rev</td>
<td>GCTAGTTGCTCTCTACATCTGC</td>
<td>300</td>
</tr>
<tr>
<td>LacZ</td>
<td>Fwd</td>
<td>TACCAACAGCGAGTGTTCGGG</td>
<td>300</td>
</tr>
<tr>
<td></td>
<td>Rev</td>
<td>GTGTGGTGTATGCGGCAATGCG</td>
<td>300</td>
</tr>
<tr>
<td>Prnp(^{fox})</td>
<td>1</td>
<td>AATGGTTAACTTCCATAGGATTG</td>
<td>Prnp(^{fox}) 344</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>GCCGACATGCTAGCATA</td>
<td>Prnp(^{fox}) 210</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>GGTTGACGCATGACCTTC</td>
<td>Prnp(^{WT}) 167</td>
</tr>
<tr>
<td>Prnp(^{ad})</td>
<td>Fwd</td>
<td>GCCATCACAGAGATTCCATGATT</td>
<td>1,200</td>
</tr>
<tr>
<td></td>
<td>Rev</td>
<td>ATCCCCAGATCGAAAGATG</td>
<td>1,200</td>
</tr>
<tr>
<td>Prnp(^{WT})</td>
<td>Fwd</td>
<td>TCAATCCACAGATCGGAAAGATG</td>
<td>600</td>
</tr>
<tr>
<td></td>
<td>Rev</td>
<td>ATGGGGAACCCTGGGTACCTGGCCTG</td>
<td>600</td>
</tr>
<tr>
<td>Prnp(^{flo})</td>
<td>1</td>
<td>ACAATGGTTAGGATGCTAGGATTG</td>
<td>Prnp(^{flo}) 1,000</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>ATGATGATTGCAACAGATGATGAGT</td>
<td>Prnp(^{flo}) 840</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>TACACAGAAGCTGCGGATAG</td>
<td>Prnp(^{WT}) 750</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>GGCAGAGGCTAAGGACAACA</td>
<td>Prnp(^{WT}) 750</td>
</tr>
</tbody>
</table>

Fwd, forward primer; Rev, reverse primer; (R), Cre-mediated DNA recombinated allele.

doi:10.1371/journal.ppat.1002402.t001

long-lived B lymphocyte populations and their replacement from the donor bone marrow.

Histological assessment of LacZ expression

Tissues were first immersed in LacZ fixative (PBS [pH 7.4] containing 2% paraformaldehyde, 0.2% glutaraldehyde, 0.02% Nonidet P40, 0.01% sodium deoxycholate, 5 mM EGTA, 2 mM MgCl\(_2\)) and washed in LacZ wash buffer (PBS [pH 7.4] containing 0.02% Nonidet P40, 0.01% sodium deoxycholate, 2 mM EGTA, 2 mM MgCl\(_2\)). Tissues were subsequently incubated in 15% (wt/vol) sucrose in PBS overnight followed by a further overnight incubation in 30% (wt/vol) sucrose in PBS and embedded in Tissue-Tek O.C.T. compound (Bayer PLC, Newbury, UK). Serial sections (thickness, 6 μm) were cut on a cryostat and stained with LacZ, using 1B3 PrP-specific pAb. For the construction of lesion profiles, sections were immunostained with the following antibodies:

- Marginal zone B cells were detected using mAb 1B1 to detect CD1d (BD Biosciences PharMingen).
- Sympathetic nerves were detected using tyrosine hydroxylase (TH)-specific pAb (Chemicon Europe).

For the detection of disease-specific PrP (PrP\(^d\)) in spleens and brains, tissues were fixed in periodate-lysine-paraformaldehyde, fixative and embedded in paraffin wax. Sections (thickness, 6 μm) were deparaffinised, and pre-treated to enhance the detection of PrP\(^d\) by hydrated autoclaving (15 min, 121°C, hydration) and subsequent immersion formic acid (98%) for 5 min [67]. Sections were then immunostained with B3 PrP-specific pAb. For the detection of EGF-like module-containing mucin-like hormone receptor-like 1 (EMR1)-expressing macrophages, paraffin-embedded spleen sections were micro-waved in citric acid buffer (pH 6.0) for 10 min. Endogenous peroxidase activity was blocked using 1% hydrogen peroxidase in methanol, and macrophages detected using rat mAb F4/80 to detect EMR1 (clone CI:A3-1, AbD Serotec). For the detection of astrocytes, brain sections were immunostained with anti-glia fibrillary acidic protein (GFAP; Dako, Ely, UK). For the detection of microglia, deparaffinised brain sections were first pre-treated with Target Retrieval Solution (DAKO) and subsequently immunostained with anti-oxidized calcium-binding adaptor molecule 1 (Iba1; Wako Chemicals GmbH, Neuss, Germany). Immunolabelling was revealed using HRP-conjugated to the avidin-biotin complex (Novared kit, Vector laboratories, Peterborough, UK)
IHC. immune complexes (Sigma). Spleens were removed 24 h later and after i.p. infection. Due to the absence of PrP C-expressing FDC was ablated on FDC and B cells, lower row) were collected 70 days after i.p. injection of ME7 scrapie prions. Prion infectivity titres were determined by transmission of tissue homogenates into groups of 4 indicator tga20 mice. Each point represents data derived from an individual spleen. Data below the horizontal line indicate disease incidence in the recipient mice <100% and considered to contain trace levels of prion infectivity. High levels of prion infectivity were detected in spleens of control mice and those in which cellular PrP C was expressed only on FDC (left-hand and middle panels, respectively). However, this accumulation was blocked in spleens with FDC-restricted PrP C ablation as only trace levels of infectivity were detected (right-hand panel).

Figure S2 Follicular dendritic cell-specific PrP C expression alone is sufficient to sustain high levels of prion infectivity in the spleen. Prion infectivity levels were assayed spleens from control mice (Prnp flox/-→Prnp flox/- mice), Prnp flox/-→Prnp flox/-→CD21-Cre Prnp flox/- mice in which cellular PrP C was expressed only on FDC and Prnp flox/-→Prnp flox/-→CD21-Cre Prnp flox/- mice with FDC-restricted PrP C ablation (n = 3/group) collected 70 days after i.p. infection with ME7 scrapie prions. Prion infectivity titres were determined by transmission of tissue homogenates into groups of 4 indicator tga20 mice. Each point represents data derived from an individual spleen. Data below the horizontal line indicate disease incidence in the recipient mice <100% and considered to contain trace levels of prion infectivity. High levels of prion infectivity were detected in spleens of control mice and those in which cellular PrP C was expressed only on FDC (left-hand and middle panels, respectively). However, this accumulation was blocked in spleens with FDC-restricted PrP C ablation as only trace levels of infectivity were detected (right-hand panel).

Figure S3 Effect of FDC-restricted PrP C-ablation on disease pathogenesis within the brain after i.p. prion exposure. Mice were injected i.p. with ME7 scrapie prions. Brains were collected from clinically scrapie-affected mice and mice which were free of the clinical signs of prion disease at the time of cull and the neuropathology within each brain compared. High levels of spongiform pathology (H&E, upper row), heavy accumulations of PrP C (brown, second row), reactive astrocytes expressing GFAP (brown, third row) and active microglia expressing Iba-1 (brown, bottom row) were detected in the hippocampi of the brains of all clinically scrapie-affected control mice (right-hand column, n = 5) and mice in which PrP C expression was ablated in B cells only (CD21-Cre Prnp flox/-→Prnp flox/- mice, third column, n = 3). In contrast, none of the mice with PrP C-ablated FDC (FDC-restricted, Prnp flox/-→CD21-Cre Prnp flox/- mice, first column, n = 6; FDC and B cells, CD21-Cre Prnp flox/-→CD21-Cre Prnp flox/- mice, second column, n = 7) developed clinical signs of prion disease during their life-spans or histopathological signs of prion disease in their brains. Scale bar, = 500 μm. Clin., presence of clinical signs of scrapie at the time of cull; Path., histopathological detection of spongiform pathology in the brain; dpi, days post i.p. prion infection.

Supporting Information

Figure S1 In the absence of PrP C expression by follicular dendritic cells prions are scavenged by tingible body macrophages in the spleen. Mice were injected i.p. with ME7 scrapie prions. Spleens from CD21-Cre Prnp flox/-→Prnp flox/- mice (in which cellular PrP C was expressed only on B cells, upper row), Prnp flox/-→CD21-Cre Prnp flox/- mice (with FDC-restricted PrP C ablation, middle row) and CD21-Cre Prnp flox/-→CD21-Cre Prnp flox/- mice (in which PrP C expression was ablated on FDC and B cells, lower row) were collected 70 days after i.p. infection. Due to the absence of PrP C-expressing FDC prion replication in these tissues was blocked. However, in the spleens of some of these mice, low levels of PrP C (left-hand column, red) were occasionally localised within cells with characteristics typical of tingible body macrophages. These cells contained the remnants of many phagocytosed apoptotic lymphocytes (tingible bodies, arrowheads) and expressed the tissue macrophage marker EGF-like module-containing mucin-like hormone receptor-like 1 (EMR1) detected by mAb F4/80 (right-hand column, brown). Data are representative of spleens from at least 4 mice from each group. Sections are counterstained with haematoxylin (blue). Scale bar, 20 μm.

Statistical analyses

Data are presented as mean ± SE. Unless indicated otherwise, significant differences between samples in different groups were sought by one-way ANOVA. Values of P<0.05 were accepted as significant.

Passive immunization

To assess antigen trapping by FDC in vivo, mice were passively immunized by intravenous injection with 100 μl preformed PAP immune complexes (Sigma). Spleens were removed 24 h later and the presence of FDC-associated immune complexes identified by IHC.

Acknowledgments

We thank Bob Fleming, Nadia Tuzi, Irene McConnell, Fraser Laing, Simon Cumming and the Pathology Services Group (University of Edinburgh, UK) for helpful discussion and excellent technical support; Nathalie Uyttersprot (current address, Artemis Pharmaceuticals GmbH, Germany) and Ari Waisman (Johannes Gutenberg University of Mainz, Germany) for supply of the CD21-cre mice; and Christine Farquhar (University of Edinburgh, UK) for provision of pAb IB3.

Author Contributions

Conceived and designed the experiments: NAM LM BMB KLB JCM KR JH MB. Performed the experiments: LM KLB NAM. Analyzed the data: LM NAM MB JH. Contributed reagents/materials/analysis tools: MB JCM KR. Wrote the paper: NAM LM BMB KLB JCM KR MB JH.
References


