The Chromosomal Passenger Complex Activates Polo Kinase at Centromeres

Mar Carmena1,*, Xavier Pinson2, Melpi Platani1, Zeina Salloum2, Zhenjie Xu1, Anthony Clark3, Fiona MacIsaac1, Hiromi Ogawa1, Ulrike Eggert3, David M. Glover4, Vincent Archambault2,5,*, William C. Earnshaw1

1 Wellcome Trust Centre for Cell Biology, University of Edinburgh, Edinburgh, Scotland, United Kingdom, 2 Institut de Recherche en Immunologie et en Cancérologie, Université de Montréal, Montréal, Québec, Canada, 3 Dana-Farber Cancer Institute and Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, Boston, Massachusetts, United States, 4 Cancer Research UK, Cell Cycle Genetics Research Group, Department of Genetics, University of Cambridge, Cambridge, United Kingdom, 5 Département de Biochimie, Université de Montréal, Montréal, Québec, Canada

Abstract

The coordinated activities at centromeres of two key cell cycle kinases, Polo and Aurora B, are critical for ensuring that the two sister kinetochores of each chromosome are attached to microtubules from opposite spindle poles prior to chromosome segregation at anaphase. Initial attachments of chromosomes to the spindle involve random interactions between kinetochores and dynamic microtubules, and errors occur frequently during early stages of the process. The balance between microtubule binding and error correction (e.g., release of bound microtubules) requires the activities of Polo and Aurora B kinases, with Polo promoting stable attachments and Aurora B promoting detachment. Our study concerns the coordination of the activities of these two kinases in vivo. We show that INCENP, a key scaffolding subunit of the chromosomal passenger complex (CPC), which consists of Aurora B kinase, INCENP, Survivin, and Borealin/Dasra B, also interacts with Polo kinase in Drosophila cells. It was known that Aurora A/Bora activates Polo at centrosomes during late G2. However, the kinase that activates Polo on chromosomes for its critical functions at kinetochores was not known. We show here that Aurora B kinase phosphorylates Polo on its activation loop at the centromere in early mitosis. This phosphorylation requires both INCENP and Aurora B activity (but not Aurora A activity) and is critical for Polo function at kinetochores. Our results demonstrate clearly that Polo kinase is regulated differently at centrosomes and centromeres and suggest that INCENP acts as a platform for kinase crosstalk at the centromere. This crosstalk may enable Polo and Aurora B to achieve a balance wherein microtubule mis-attachments are corrected, but proper attachments are stabilized allowing proper chromosome segregation.

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Abbreviations: CPC, chromosomal passenger complex; KMN, KNL-1/Mis12/Ndc80; KT-MT, kinetochore-microtubule; NEB, nuclear envelope breakdown; OA, okadaic acid; PLA, proximity ligation assay
* E-mail: mar.carmena@ed.ac.uk (MC); vincent.archambault.1@umontreal.ca (VA)

Introduction

Executive decisions concerning when cells enter and exit mitosis are made by Cdk1 with cyclins A and B as cofactors. Once cells have entered mitosis, Plk1 and the Aurora kinases direct spindle formation, regulate chromosome attachments to spindle microtubules, ensure the operation of the spindle checkpoint, and enable daughter cells to complete cytokinesis [reviewed in [1–4]]. Plk1 and Aurora A also function in the regulation of mitotic entry (reviewed in [5]).

In higher eukaryotes, Plk1 and Aurora B have potentially antagonistic activities during the early stages of chromosome attachment and alignment on the mitotic spindle. Plk1 phosphorylation of kinetochore components and microtubule plus-end-associated proteins is required for the establishment of stable kinetochore-microtubule (KT-MT) interactions. Electron micrographs of human cells treated with the Plk1 inhibitor BI2536 show fewer microtubule connections per kinetochore [6]. Tension-sensitive phosphorylation of BubR1 by Plk1 regulates the initial stability of KT-MT interactions [7], as do phosphorylation of CLIP-170 [8] and NudC [9]. Plk1 also phosphorylates components of the Ska and KNL-1/Mis12/Ndc80 (KMN) kinetochore complexes as well as centromere proteins CENP-B, CENP-C, CENP-E, and CENP-F. However, the function of these phosphorylations is not known [10].

The chromosomal passenger complex (CPC), consisting of Aurora B kinase, INCENP, Survivin, and Borealin [11], has a role in the correction of kinetochore-microtubule attachment errors by promoting the release of kinetochore-microtubule attachments [11–15]. The localization of the CPC relative to kinetochores is...
critical for regulation of kinetochore-microtubule attachments [16], CPC targeting to inner centromeres occurs as a result of Survivin binding to histone H3 phosphorylated on Thr3 by Haspin kinase [17–19] and is helped by an Aurora B-dependent positive feedback loop [20].

The temporal and spatial regulation of Plk1 activation is complex. While Polo activity is not required for mitotic entry in unperturbed cell cycles, its activation by T-loop phosphorylation is needed for its functions during mitosis. At centromeres, Aurora A kinase-Bora phosphorylates Plk1 on Thr210, thereby activating Plk1 at the G2-M transition in human cells [21,22]. Subsequently, Plk1 triggers the degradation of both Bora [23,24] and Aurora A [25]. The regulation of Plk1 activity at kinetochores is a critical and largely unstudied question.

Here, we have examined the mechanism of Polo kinase activation at Drosophila and human kinetochores. We show that Aurora B is required for Polo T-loop phosphorylation at the centromere, but not at centrosomes. Our studies identify a new regulatory link between the Aurora B and Polo kinases mediated by INCENP. Furthermore, we demonstrate that this mechanism of regulation of Polo kinase at the kinetochore by the CPC is conserved in human cells. These results support our previous hypothesis that INCENP acts as a platform coordinating the activities of these kinases on chromosomes during early mitosis [26].

Results

Polo Kinase Localizes to the Centromere/Kinetochore Region Before the CPC Does

As a starting point to examine the relationship between Polo and the CPC in Drosophila, we compared their localization in space and time. INCENP and the CPC are diffuse during early prophase in Drosophila cultured cells (Figure 1A2). In contrast, Polo kinase in these early stages is localized at centromeres

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This confirmed a recent study that found positive PLA signals between various members of the CPC in all phases of mitosis in human cells [33].

In Drosophila, the positive PLA signals overlapped with Aurora B-GFP, confirming that epitopes on INCENP and Aurora B are in close proximity in the inner centromere (Figure 1G4). A parallel assay using antibodies against INCENP and GFP confirmed this close association of INCENP with exogenous GFP-tagged Aurora B (unpublished data). In two negative controls, we failed to observe PLA signals using antibodies to INCENP and γ-tubulin and between Polo and the centromere histone CENP-A/CID (Figure 1J,1K).

We next used PLA to define where in cells interactions occur between INCENP and Polo. Cells co-stained for INCENP and Polo showed PLA signals on chromosomes in early mitosis (Figure 1H1). These signals colocalized with Aurora B-GFP, confirming that the interaction occurs at inner centromeres (Figure 1H4). PLA also detected a close association between INCENP and Polo\(^{T182ph}\), the activated form of Polo kinase, using an anti-phospho-epitope-specific antibody (Figure 1H1, see validation of the antibody below). These positive PLA signals were also present in inner centromeres (Figures 1I).

We conclude that INCENP and Polo physically interact and are in close proximity at inner centromeres during early mitosis in Drosophila.

INCENP Is Required for Polo Kinase Activation at the Inner Centromere

The association between INCENP and Polo described above suggested that INCENP and other CPC components might have a role in Polo activation by T-loop phosphorylation. To test this hypothesis, we asked whether reducing INCENP protein levels by RNAi affected the localization and activation of Polo kinase, as detected by monitoring Polo T-loop phosphorylation. Phosphorylation on the highly conserved T-loop residue Thr-210 (Thr-182 in Drosophila Polo, Figure 2A) is crucial for kinase activation [34]. T182 of Drosophila Polo is a major phosphorylation site detected by mass spectrometry, and a phosphomimetic mutation at that site (Polo\(^{T182D}\)) increases Polo kinase activity in vitro (VA, unpublished observations).

To examine Polo activation, we used a phospho-epitope-specific antibody raised against human Polo\(^{T182ph}\) that also recognizes Drosophila Polo\(^{T182ph}\) (Figure 2B). In asynchronous cultures, Polo\(^{T182ph}\) was barely detectable by Western blotting. However, we could readily observe endogenous Polo\(^{T182ph}\) using this antibody after treatment with the phosphatase inhibitor okadaic acid (OA; Figure 2B). This signal disappeared following Polo depletion by RNAi, confirming that it comes from the Polo protein (see below). Moreover, Polo-Myc gives a signal at the expected higher molecular weight, while Polo\(^{T182A}\)-Myc and Polo\(^{T182D}\)-Myc are not recognized by this antibody (Figure 2B). In these experiments, Polo was detected as a doublet by Western blotting. This mobility shift is not caused by T182 phosphorylation and may be caused by an-as-yet uncharacterized modification.

We used this antibody to examine the distribution of Polo\(^{T182ph}\) by immunofluorescence in cycling DMel-2 cells that had not been treated with okadaic acid. The antibody detected Polo\(^{T182ph}\) at centromeres/kinetochores, centrosomes, the cleavage furrow, and midbody. This staining was largely abolished following Polo RNAi-depletion (Figure S3).

Although most Polo accumulates at prometaphase kinetochores (Figure 2C3; Figure S2B,C; Figure S4A-C linescans), a minor fraction of the kinase localizes to inner centromeres (Figure 2C3 arrows; Figure 2C linescan). Indeed, the active kinase (detected with anti-Polo\(^{T182ph}\)) is clearly detectable at inner centromeres (Figure 2C4+inset, arrows), where it colocalizes with INCENP, as predicted by the PLA results (Figure 2C4; linescan). We first detect this inner-centromeric pool of active Polo in late prophase cells (Figure S4A-C). Polo\(^{T182D}\) is no longer detected at the inner centromeres of chromosomes aligned at the metaphase plate. Instead it accumulates at kinetochores in metaphase cells (Figure 2D4, arrows; linescan).

Depletion of INCENP by RNAi substantially reduced levels of activated Polo\(^{T182ph}\) at kinetochores (Figure 2E4,F4; linescans). In contrast, total Polo localized normally to kinetochores following INCENP knockdown (Figure 2E3,F3; linescans). This is consistent with the observation that Polo localization to this region precedes that of INCENP (Figure 1A). Importantly, we could still readily detect active Polo\(^{T182ph}\) at centromeres of cells following INCENP knockdown (Figure 2E4,F4, asterisks).

These experiments reveal that INCENP is required for T182 phosphorylation and activation of Polo kinase at inner centromeres in early mitosis. Activation of centrosomal Polo does not require INCENP.

Polo T-Loop Phosphorylation Is Required for Mitotic Progression But Not For Mitotic Entry

In order to investigate the function(s) of Polo T182 phosphorylation in mitosis, we established stable cell lines allowing inducible expression of Polo\(^{WT}\)-GFP or Polo\(^{T182A}\)-GFP. Endogenous Polo could be depleted in those cells by RNAi against the 3'UTR of the native transcript (Figure 3A). Expression of Polo\(^{WT}\)-GFP rescued the viability and proliferation of cells depleted of endogenous Polo. However, expression of Polo\(^{T182A}\)-GFP did not, and cells died (unpublished data). Thus, Polo T-loop phosphorylation is essential for viability.

Polo-depleted cells accumulated in mitosis, exhibiting phenotypes similar to those observed for the first polo mutants [33]. Expression of Polo\(^{WT}\)-GFP restored mitotic progression in cells depleted of endogenous Polo (Figure 3B), but expression of Polo\(^{T182A}\)-GFP did not. Cells expressing only Polo\(^{T182A}\)-GFP accumulated in prometaphase/metaphase (Figure 3C), often with...
Figure 2. INCENP is required for the activation of Polo kinase at the inner centromere. (A) Sequence alignment showing the conservation of Thr182 across species. (B) Immunoblot of cell lines stably expressing wild-type, T182A, or T182D Polo in the absence or presence of okadaic acid. The phosphoepitope antibody anti-Pik1T210Ph recognizes Drosophila PoloT182Ph, both Myc-tagged and endogenous (e). Both endogenous and Myc-tagged PoloT182Ph were detected as a doublet, suggesting that they can be modified at another site. The asterisk indicates a non-specific band that does not disappear after Polo RNAi. This band increases following okadaic treatment, and therefore could correspond to a non-specific phosphoepitope. (C–F) Control or RNAi-treated DMel-2 cells stably expressing Polo-GFP showing colocalization of INCENP and Polo/PoloT182Ph. Arrows point to chromosomes shown in zoomed images. Linescans show fluorescence intensity across the kinetochores (dashed lines). (C) Control prometaphase. PoloT182Ph is visibly enriched at the inner centromere compared to Polo (arrows). Linescans show both Polo and PoloT182Ph are present at the inner centromere (double-ended arrows show difference in intensity with respect to background levels; green, Polo; blue, PoloT182Ph). (D) Control metaphase. Asterisks point to centrosomes, and PoloT182Ph is virtually undetectable in the inner centromere (linescan; note that intensity drops to background level). (E,F) INCENP RNAi-treated cells. Asterisks point to PoloT182Ph on centrosomes (note absence from kinetochores, also shown in linescans). Zoomed images show localization at the centromere/kinetochore of the indicated proteins. doi:10.1371/journal.pbio.1001250.g002
unaligned chromosomes (Figure 3D–F). Interestingly, while the loss of Polo led to an increase in monopolar spindles, substitution of endogenous Polo with PoloT182A-GFP did not (Figure 3D–F). This suggests that T-loop phosphorylation of Polo may be dispensable for its role in bipolar spindle assembly.

The observation that INCENP-dependent activation of Polo by phosphorylation at T182 at centromeres/kinetochores is required for chromosome alignment in prometaphase is consistent with the known role of Polo in regulating kinetochore function. **Aurora B Activity is Required for Polo Kinase Activation at Centromeres**

Because the best known role of INCENP is to activate Aurora B kinase in the CPC, we next asked whether Aurora B has a role in Polo T-loop phosphorylation at centromeres. Drosophila Polo T182 (corresponding to human Pkl1 T210) is preceded by a conserved stretch of basic residues resembling the consensus site for Aurora kinases (Figure 2A) [31,36,37]. Indeed, Drosophila Aurora B complexed with a fragment of INCENP can directly phosphorylate Polo in vitro (Figure 4A). A T182A mutation in the Polo used as a substrate reproducibly reduced its phosphorylation by about one half. Thus, Polo T182 is a major phosphorylation site for Aurora B (Figure 4A). Similar results were obtained using human Aurora B on GST-PoloWT or GST-PoloT182D (unpublished data).

Kinase inhibition studies suggest that Aurora B is responsible for PoloT182 phosphorylation in vivo. Binuclein 2 is the only specific published data). PoloT182 phosphorylation at centromeres. Drosophila Polo T182 phosphorylation in vivo, Binuclein 2 is the only specific published data). When DMel-2 cells were treated with 20 μM Binuclein 2 for 2 h, H3S10ph was undetectable in mitotic cells (unpublished data); [39] and INCENP and Aurora B were dispersed in clumps on the chromosomes ([38,39]; Figure S5B,C, compare with Figure S5A). Both of these phenotypes are characteristic of the loss of Aurora B function [40,41].

Aurora B kinase activity is required for Polo activation at kinetochores, and levels of kinetochore-associated PoloT182ph were greatly reduced in Binuclein 2-treated mitotic cells (Figure 4B–D4; Figure 4E). In contrast, we observed no obvious difference in the localization of bulk Polo kinase in those cells (Figure 4B3–D3; Figure 4E). Importantly, as in the case of INCENP RNAi, we could still detect activated Polo kinase at centromeres in the same cells (Figure 4B4–D4 asterisks).

As independent confirmation of the inhibitor studies, RNAi-mediated depletion of Aurora B also led to disappearance of the PoloT182ph signal observed in Western blots after OA treatment of cells, while total Polo levels remained unchanged (Figure 4F). In striking contrast, the PoloT182ph signal actually increased after partial Aurora A depletion (Figure 4Fa), perhaps because cells accumulated in mitosis.

The above results suggested that Aurora B rather than Aurora A plays a major role to promote PoloT182 phosphorylation at centromeres in Drosophila cells. In order to exclude that our failure to detect PoloT182ph by Western blotting following Aurora B depletion was due to a cell cycle block outside mitosis caused by OA treatment, we examined the effects of RNAi depletion of Aurora A, Aurora B, and INCENP on the PoloT182ph signal at centromeres in individual mitotic cells without okadaic acid treatment. Brief (3 d) dsRNA treatments were used to avoid an accumulation of binucleate cells caused by failure in CPC function in cytokinesis.

Depletion of Aurora B or INCENP led to a significant reduction of the PoloT182ph signal at centromeres (Figure 4G). This effect was specific to centromeres, and PoloT182ph levels at centromeres were unaffected following depletion of Aurora B or INCENP (Figure S6A). In contrast, Aurora A depletion had no effect on levels of PoloT182ph at centromeres, but led to a modest reduction in PoloT182ph levels at centrosomes.

Together, these results confirm that Aurora B and INCENP are required for Polo activation at the centromere/kinetochore in early mitosis and strongly implicate Aurora B as the kinase responsible.

**The CPC Is Required for Polo Activation at Kinetochores in Larval Neuroblasts**

The CPC is required for Polo kinase activation at centromeres in live animals, and not only in aneuploid cultured cells. To demonstrate this, we examined flies homozygous for the hypomorphic female-sterile allele incenpQA26, a point mutation in the highly conserved IN-box domain [42].

We observed a strong signal of PoloT182ph concentrated at kinetochores in wild-type mitotic neuroblasts (Figure 5A). In third instar larval neuroblasts from the hypomorphic female-sterile allele incenpQA26, a point mutation in the highly conserved IN-box domain [42].

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Figure 4. Aurora B activity is required for the activation of Polo kinase at the inner centromere. (A) Aurora B phosphorylates Polo kinase in vitro. Bacterially expressed HIS-Polo or HIS-PoloT182A (which is catalytically inactive and therefore unable to autophosphorylate) were incubated with (or without) Drosophila Aurora B in complex with a fragment of INCENP (residues 654–755) in presence of 32P-g-ATP, in triplicate. Reaction products were resolved by SDS-PAGE transferred to nitrocellulose and analyzed by autoradiography (AR) and anti-Polo Western blot (WB). Quantitative measurements of signals were obtained (see Materials and Methods), and the ratios were calculated for each reaction (AR/WB, A.U.: arbitrary units). Right, average values for the relative phosphorylatin of PoloWT and PoloT182A by Aurora B. Error bars, SEM. (B–D) DMel-2 cells stably expressing Polo-GFP treated with (B) DMSO or (C–D) Binucleine-2, immunostained for INCENP, Polo, and PoloT182Ph (insets: zoomed images of kinetochores). In (C–D) asterisks indicate centrosomes. Merged images show INCENP/Polo/DNA. Zoomed images in (C–D) insets show examples of
kinetochores in prometaphase cells were measured at individual kinetochores using Image J, subtracting background. (E) Dot plot showing the quantification of INCENP/Polo/PoloT182Ph signal intensity at the kinetochores (t test: *** p < 0.0001; n.s., not significant; p = 0.4028). Signal intensities for individual kinetochores were measured using the SoftWorx Data Inspector tool; average background was subtracted; data was plotted using KaleidaGraph software. (F) RNAi depletion of Aurora B, but not Aurora A, strongly reduces PoloT182Ph levels in DMel-2 cells treated with okadaic acid. Cells were transfected with the indicated dsRNAs for 4 d, and 100 nM okadaic acid added for 4 h before immunoblotting to improve visualization of phosphorylated Polo. A dsRNA against the Kanamycin resistance bacterial gene was used as a negative control. Asterisks: non-specific bands. Both bulk Polo and PoloT182ph appear as doublets. (G) RNAi depletion of Aurora B, but not Aurora A, reduces PoloT182Ph levels at centromeres/kinetochores. Cycling cells were treated with the indicated dsRNAs for 3 d (immunoblots are shown in Figure S6B) and PoloT182Ph was detected by immunofluorescence. Levels of PoloT182Ph at centromeres/kinetochores in prometaphase and metaphase cells were measured at individual kinetochores using Image J, subtracting background (K-C, K-D). Asterisks indicate centromeres. Error bars = S.E.M.

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T182 phosphorylation at kinetochores in mitotic Drosophila neuroblasts. As independent confirmation that the CPC contributes to Polo regulation in vivo, we conducted a genetic experiment in a sensitized background. Tubulin-Gal4-driven overexpression of Polo mutated in a conserved destruction box (PoloΔdb—Figure S7) is semi-lethal at the pupal stage, suggesting that excessive Polo activity is detrimental to development. Interestingly, a decrease in the levels of functional INCENP rescued the semi-lethality of PoloΔdb-expressing flies. This was observed when one wild-type allele of incenp was replaced with either of the alleles incenpΔC774 or incenpΔQ476 (Figure S7). A heterozygous deletion removing Aurora B did not cause a significant rescue in the same assay, suggesting that INCENP may be the limiting CPC component for Polo regulation.

Together, these results confirm that the CPC contributes to the regulation of Polo function at kinetochores in vivo.

The CPC Is Required for Plk1 Activation at Kinetochores in Human Cells

Importantly, the regulation of Polo T-loop phosphorylation described above for Drosophila is conserved in human cells. We used siRNAs to deplete either Aurora B (Figure 6A) or INCENP (Figure 6C) in HeLa cells and subsequently measured the levels of total Plk1 and Plk1T210P at kinetochores. Both INCENP and Aurora B depletion caused a dramatic reduction in Plk1T210P levels in early mitosis (Figure 6A,B,D,Plk1T182ph). Levels of Plk1 were also slightly reduced, confirming that the CPC is at least partly required for the stable localization of Plk1 to mammalian kinetochores [27]. Comparable results were obtained when we treated cells with the Aurora B kinase inhibitor ZM447439 (Figure S8), confirming that Aurora B activity is indeed required for the presence of Plk1T210P at kinetochores in human cells.

Our results thus indicate that the CPC activates Polo kinase by T-loop phosphorylation at centromeres in both flies and humans.

Discussion

Coordination of Polo and Aurora B activity at kinetochores is critical in early mitosis, as the two kinases play potentially antagonistic but complementary roles in regulating kinetochore-microtubule interactions. Aurora B is essential for the correction of aberrant attachments [13–15], and indeed, tethering Aurora B too close to kinetochores interferes with the formation of stable attachments [16]. In contrast, Plk1 activity is required for initial stabilisation of microtubule attachments to kinetochores [7–9]. We suggest that interactions with INCENP may provide a mechanism to coordinate the activities of these two essential kinases during early mitosis.

Recent studies suggest that Plk1 is activated at centromeres when its T-loop (T210) is phosphorylated by Aurora A kinase–Bora, and that this promotes the G2/M transition upstream of Cdk1 [21,22], although Polo activity is not required for mitotic entry ([3]; this paper–Figure 3). How Plk1 is activated at kinetochores remained an important unsolved question. Our present results show that Aurora B and INCENP, which are concentrated at inner centromeres [43,44], function there to activate Polo by phosphorylating its T-loop.

Plk1 recruitment to centromeres in late G2 has been variously proposed to be mediated by Bub1 [45], INCENP [27], and BubR1 [7]. Another report implicated the self-primed interaction of Plk1 with PBIP1/CENP-U [46]. This could potentially explain why Plk1 activity is reportedly required for its localisation to kinetochores in human cells [47].

Our RNAi studies confirmed that Plk1 is partially dependent on the CPC for its centromeric localization in human cells. However, this appears not to be the case in Drosophila, where Polo is present at centromeres before NEB, at a time when INCENP is not yet concentrated at inner centromeres and before PoloT182ph, the active form of the kinase, is detected there. Indeed, we observed no significant decrease in kinetochore-associated Polo levels after INCENP RNAi in Drosophila cells.

Although Polo targeting to kinetochores is independent of the CPC in Drosophila, its activation there does require the CPC with active Aurora B. Our data suggest that INCENP binding to Polo facilitates its subsequent activation by Aurora B kinase (Figure 7B,C). Indeed, INCENP and Polo interact physically in vitro and co-immunoprecipitate in mitotic cell extracts. Although most centromeric Polo kinase is concentrated in the outer kinetochore in prophase and prometaphase, active Polo (PoloT182ph) is also found in inner centromeres, where it overlaps with INCENP as confirmed by a proximity ligation assay (PLA).

A range of evidence presented here suggests that Aurora B is the upstream kinase responsible for Polo kinase activation at centromeres. Firstly, Aurora B phosphorylates Polo at Thr182 in vitro. Secondly, RNAi depletion of INCENP or Aurora B, but not Aurora A, reduces levels of active PoloT182ph at kinetochores. Thirdly, tissue culture cells and third larval instar neuroblasts treated with a specific inhibitor of Drosophila Aurora B kinase show decreased levels of PoloT182ph at kinetochores. In all of the preceding experiments, PoloT182ph levels are affected at kinetochores but not at centromosomes, where Polo is presumably activated by Aurora A [21,22]. Importantly, this involvement of Aurora B in Polo activation at centromeres discovered in Drosophila is conserved for Plk1 in human cells.

Our results suggest a model for interactions between Polo kinase and the CPC at centromeres (Figure 7). In Drosophila cells, Polo targets to centromeres before the CPC is recruited by Survivin binding to histone H3T3ph [17–19]. At inner centromeres of chromosomes whose kinetochores are not under tension, Polo now binds to INCENP. This promotes Polo kinase activation, as Aurora B phosphorylates PoloT182. We suggest that interactions with INCENP link the two complementary kinase activities, thereby potentially creating a microtubule attachment/detach-
Figure 5. The centromeric activation of Polo in mitosis depends on INCENP and Aurora B in vivo. (A–E) Immunostaining analysis of the phosphorylation of Polo^T182^ in third instar neuroblasts of (A) wild type larvae (Canton-S), (B) *incenp^QA26^* mutant larvae, and (C–D) wild type larvae treated with the Aurora B-specific inhibitor Binucleine 2. INCENP (2, green), Polo^T182^ (3, red). Arrows point to INCENP blocks characteristic of the *incenp^QA26^* and Binucleine-2 treatment phenotype. Asterisks indicate centrosomes. (E) Dot plot showing the quantification of INCENP/Polo^T182^ signal intensity at the kinetochore (*t* test: ***p < 0.0001; **p = 0.003). Signal intensities for individual kinetochores were measured using the SoftWorx Data Inspector tool; average background was subtracted; data was plotted using KaleidaGraph software. (F–I) Levels of Polo kinase are not affected
by defects in Incenp or Aurora B function (F) wild type larvae (Canton-S), (G) incenp<sup>QA26</sup> mutant larvae, and (H) wild type larvae treated with the Aurora B-specific inhibitor Binucleine 2. INCENP (5, red), Polo (6, green). (I) Dot plot showing the quantification of INCENP/Polo signal intensity at the kinetochore (t test: *** p<0.0001). Signal intensities for individual kinetochores were measured using the SoftWorx Data Inspector tool; average background was subtracted; data were plotted using KaleidaGraph software.

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At metaphase, when chromosomes are bi-oriented and under tension, the CPC and Polo kinase exhibit only a partial overlap. A weakening of the INCENP/Polo PLA signals in metaphase suggests that Polo may be released from INCENP after its activation— weakening of the INCENP/Polo PLA signals in metaphase suggests that Polo may be released from INCENP after its activation—

In summary, our results reveal that INCENP and Aurora B are responsible for Polo kinase activation at centromeres but not at centromeres during mitosis. These experiments support the hypothesis that INCENP acts as a scaffold integrating the cross-talk between these two important mitotic kinases [26].

**Materials and Methods**

**Drosophila Strains**

Fly strains were grown at 25°C in standard Drosophila medium. The following stocks were used: Canton-S; incenp<sup>QA26</sup>/SM6a, incenp<sup>/E3747</sup>/SM6a, Tubulin Gal-4/TM3. UAS<sup>p</sup>POLO<sup>Δ6DB</sup>-MYC transgenic flies were generated by BestGene Inc. Immunostaining of testes and larval neuroblasts was performed as described previously [49].

**Antibodies**

Primary antibodies and dilutions for immunofluorescence analysis were as follows: mouse monoclonal B512 anti-α-Tubulin (SIGMA, 1:2,000); Rabbit Polyclonal Rhb-801, Rhb-803 [41], 1:500; mouse monoclonal anti-Phospho-T210 Plk1 (Abcam ab39068, 1:100); mouse monoclonal anti-Polo Mab294 (kind gift of A. Tavares and David Glover, 1:100); rabbit polyclonal anti-Incenp (Abcam, 1:100); Rat monoclonal anti-Incenp (kind gift of Kim McKim, 1:300)/ Rabbit polyclonal anti-Aurora B 963 [41,1:500]; Rabbit polyclonal anti-Incenp Rb801 [41], 1:500/mouse monoclonal anti-Phospho-T210 Plk1 (Abcam, 1:300); Rat monoclonal anti-Incenp (kind gift of Kim McKim, 1:300)/ Rabbit polyclonal anti-Aurora B 963 [41,1:500]; Rabbit polyclonal anti-Incenp Rb801 [41], 1:500/mouse monoclonal anti-γTubulin (Sigma 1:50); and Rabbit polyclonal anti-CID (a gift from S. Henikoff, 1:500/mouse monoclonal anti-α-Polo-Mab294 (1:100). In each experiment a negative control using only one antibody of each pair was included.

For each antibody pair, exponentially growing DMel-2 cells were seeded on Con-A treated coverslips and fixed for immunostaining as described previously [41]. After overnight incubation with primary antibody at 4°C, half of the samples were processed following the normal immunostaining protocol [41] and the other half was used for the PLA assay.

Imaging was performed using Olympus IX-71 microscope controlled by Delta Vision SoftWorx (Applied Precision, Issaquah, WA, USA). Image stacks were deconvolved, quick-projected, and saved as tiff images to be processed using Adobe Photoshop. Linescans were generated using Image-Pro software.

**Plasmids and Mutagenesis**

**POLO-T182A, T182D, and POLO<sup>Δ6DB</sup>** (R309A, L312A) were generated in the pDONR221 (Invitrogen) using QuickChange (Stratagene). The expression vectors pAC5-POLO-MYC, pAC5-POLO-T182A-MYC, pAC5-POLO-T182D-MYC, and pUASP-POLO<sup>Δ6DB</sup>-MYC were generated by Gateway recombination (Invitrogen) of pDONR-based entry clones into pDEST-AC5-Clam-MYC and pDEST-UASp-Clam-MYC, respectively. POLO<sup>-WT</sup> and POLO-T182A were cloned into pETDuet for expression as N-terminal fusions with a HIS tag at the MCS1 position. Aurora B was cloned into pDONR221, which was then recombined into pDEST-AC5-Clam-GFP to generate pAC5-AURORA B-GFP.

**In Vitro Binding Assays**

GST tagged full-length Drosophila Incenp was expressed in bacteria (BL21) and purified on Glutathione sepharose beads as

described previously [42]. Polo, Aurora B, and Luciferase were in vitro translated using a coupled transcription/translation reticulocyte lysate system (Promega’s TNT system). Binding buffer—50 mM Tris pH 7.5, 10 mM MgCl2, 1 mM EGTA, 1 mM DTT, 0.1% Triton X-100, 0.5 mM PMSF, and 1 mg/ml CLAP.

Immunoprecipitation

Mouse anti-GFP (Roche) and mouse IgG (Abcam)—negative control—antibodies were crosslinked to protein G Dynabeads (Invitrogen) at 0.5 μg of antibody/1 μl of beads. Exponentially growing D-Mel2 cells were lysed on ice in lysis buffer (for Polo-GFP cell line: 40 mM Tris-Cl [pH 7.5], 100 mM NaCl, 1 mM PMSF, 1 mM DTT, 10 mM EGTA, 1% Triton-X-100, and 0.1% NP40, 0.5% deoxycholate, and protease inhibitor cocktail - Roche). Cell lysates were separately incubated with either mAb anti-GFP or mouse IgG bound to Dynabeads protein G for 1 h at 4°C. Samples were spun down, then washed first with lysis buffer, and then twice with wash buffer (40 mM Tris-Cl [pH 7.5], 100 mM NaCl, 1 mM PMSF, 10 mM EGTA, 0.1% Triton-X-100, protease inhibitor cocktail, Roche, UK). Finally the beads were boiled in Laemmli sample buffer. All samples were subjected to SDS-PAGE and analyzed by immunoblotting as described before.

Kinase Assay

HIS-Polo and HIS-Polo-T182A were expressed in BL21 bacteria from pETDuet-based constructions (see above). Protein purification was done using Talon resin (Clontech) and purified proteins were stored on the resin at −80°C.

For the kinase assay, HIS-Polo and HIS-Polo-T182A on the resin were incubated with Drosophila Aurora B in complex with INCENP645–755 [39] for 15 min at 30°C in 20 mM K-HEPES pH 7.5, 2 mM MgCl2, 1 mM EDTA, 1% NP40, 0.5% deoxycholate, and protease inhibitor cocktail - Roche). Cell lysates were separately incubated with either mAb anti-GFP or mouse IgG bound to Dynabeads protein G for 1 h at 4°C. Samples were spun down, then washed first with lysis buffer, and then twice with wash buffer (40 mM Tris-Cl [pH 7.5], 100 mM NaCl, 1 mM PMSF, 10 mM EGTA, 0.1% Triton-X-100, protease inhibitor cocktail, Roche, UK). Finally the beads were boiled in Laemmli sample buffer. All samples were subjected to SDS-PAGE and analyzed by immunoblotting as described before.

siRNAi, Drug Treatments, Immunofluorescence, and Imaging in Human Cells

HeLa Kyoto were grown in Dulbecco’s modified Eagle’s medium, supplemented with 10% foetal calf serum, 0.2 mM l-Glutamine, 100 U/ml penicillin, and 100 μg/ml streptomycin. RNAi experiments were performed using annealed siRNA oligos (Qiagen) diluted in serum free OptiMem and transfected
using HiPerFect reagent (Qiagen) according to the manufacturer’s protocol. HeLa cells were seeded on coverslips at a density of 1 x 10⁶ cells/ml and diluted siRNA was added to cells so that the final concentration of siRNA was 40 nM. Coverslips were fixed at 48 h. For control transfections non-silencing random scramble siRNA oligos were used at the same concentration. The full sequences of siRNA oligos used are as follows: for Aurora B siRNA, 5'-AAGCGGGAACACTCAATTTGA-3'; for INCENP siRNA 5'-AGATGACAGCCATGATACTA-3'. For drug treatments, ZM447439 (Tocris Bioscience) or DMSO as control were added to the cells at the concentration of 5 μM for 1 h.

All fixation, permeabilisation and immunostaining were performed at room temperature, as previously described [53]. Anti-Aurora B rabbit polyclonal at 1:100 (Abcam, ab2245), anti-INCENP rabbit polyclonal at 1:100 (Upstate), anti-Plk1 1:100 mouse monoclonal (Abcam), anti-P-Pol (T210) 1:100 mouse monoclonal (Abcam), and anti-phospho-Histone H3 (Ser10) rabbit polyclonal (Upstate). All affinity purified donkey secondary antibodies (labelled either with FITC, TRITC, or CY5) were purchased from Jackson ImmunoResearch.

Quantification of Plk1 and Plk1T210Ph on approximately 1,000 centromeres per condition was carried out as follows: Deconvolved images were imported into OMERO [54] and segmentation of centromere foci (ACA, Cy5, reference channel) performed using Otsu segmentation implemented in Matlab. Masks stored in OMERO were then used to calculate intensities, and output into comma-separated value file for plotting in Excel.

**Supporting Information**

**Figure S1** Localization of Polo, INCENP, and tubulin in mitosis in Drosophila cultured cells. (A) Prophase, Polo on centrosomes and kinetochores. No Incenp at centromeres, (B) Early prometaphase, (C) Late prometaphase, (D) Metaphase. Scale bar = 10 μm. (TIF)

**Figure S2** Relative localizations of Polo and Aurora kinases during mitosis in Drosophila cultured cells. (A–B) Aurora A (red) and Polo (green) colocalize at centrosomes throughout mitosis (arrow-heads), (C–D) Aurora B (red) and Polo (green) both localize at the centromere/kinetochores region in early mitosis. Scale bar = 5 μm. (TIF)

**Figure S3** Characterization of the anti-Polo T182Ph antibody by immunofluorescence. (A–D) Distribution of Polo kinase (green) and the active form of the kinase (Polo T182Ph, red) during mitosis. (A) Polo POLO T182Ph are present at centrosomes at a time in which Polo— but not Polo T182Ph — accumulates at kinetochores. (B) Polo POLO T182Ph colocalize at kinetochores and centrosomes in metaphase and (C–D) also at the central spindle at anaphase and telophase. (E) Specificity of the antibody against Polo T182Ph in immunofluorescence. D-Mel cells were treated with polo dsRNA or not for 60 h, fixed, and stained for pT182-Polo and alphatubulin or CENP-A (centromere). The pT182-Polo stainings at centromeres/KTs and centrosomes are largely abolished, pT182Polo stainings of the centrosomes and the midbody in cytokinesis were strongly diminished, but never completely abolished, probably because cells that could complete mitosis were those for which Polo depletion was only partial. In addition, we always observed a non-specific staining of unknown nature at or near the DNA, which remained visible during mitosis in Polo-depleted cells more than in control cells. (TIF)

**Figure S4** Colocalization of INCENP/Polo/Polo T182Ph changes through mitosis. High magnification images of kinetochores in (A–C) late prophase/early prometaphase and (D–F) metaphase in cultured cells. INCENP, red; Polo, green; and Polo T182Ph, blue. Linescans show signal intensity across a kinetochore/inner centromere/kinetochore line. The graph profile shows specific accumulation of Polo T182Ph at the inner centromere at the earlier stages of mitosis; at later stages the Polo T182Ph graph resolves in two clear peaks closer to the kinetochore. (TIF)

**Figure S5** CPC localization is similar in Binucleine-2 treated cells and incep mutants in prometaphase. DMel-2 cells treated with (A) DMSO or (B–C) Binucleine-2 and stained for INCENP (green) and Aurora B (red). (B) Prometaphase. (C) Binucleate cell. (D) Wild type and (E) incep H4T10Ph mutant neuroblasts stained for INCENP (red) and Histone3S10Ph. (TIF)

**Figure S6** RNAi depletion of Aurora B or INCENP does not reduce Polo T182Ph levels at centromeres. (A) Cells were treated with the indicated dsRNAs for 3 d and Polo T182Ph was detected by immunofluorescence. Levels of Polo T182Ph at individual centromeres in prometaphase and metaphase cells were measured using Image J as in Figure 4G. (B) Immunoblots showing levels of protein depletion after dsRNA treatments. (TIF)

**Figure S7** A decrease in INCENP activity partially rescues the lethality induced by a gain of Polo function. (A) A conserved destruction box in Polo was mutated in Polo∆db. (B) Female flies heterozygous for a UAS-Polo∆db-Myc transgene and strongly hypomorphic incep alleles were crossed to males heterozygous for the Tubulin-Gal4 driver. (C) Expression of this UAS-Polo∆db-Myc transgene driven by Tubulin-Gal4, is semi-lethal. Progeny flies combining the incep allele, UAS-Polo∆db-Myc transgene, and the Tubulin-Gal4 driver were identified by the absence of phenotypic markers from balancer chromosomes. The number of flies obtained relative to the expected numbers (one-fourth of the total progeny) is shown for each incep genotype. X, total numbers of progeny obtained and scored from three vials for each cross. The only definitely null aberrations that we could obtain were large deficiencies that could not be tested because they interacted with balancer chromosomes in our crosses; therefore, it remains formally possible, although unlikely, that both incep alleles tested here have antimorphic effects. (TIF)

**Figure S8** Aurora B activity is required for activation of Plk1 at centromere/kinetochores in human cells. (A) HeLa cells treated with DMSO or ZM44739 immunoostained for Histone H3 P-Ser10 (green), Plk1 (red), or Plk1 T210Ph (red) and DNA (blue). Scale bar = 10 μm. (B) Quantification graph of Plk1 and Plk1 T210Ph levels at centromeres in Control and ZM44739 treated cells. Fluorescent intensities are in Arbitrary Units (A.U.). t test: *** p<0.0001. (TIF)

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**Author Contributions**

The author(s) have made the following declarations about their contributions: Conceived and designed the experiments: MC XP VA.
Performed the experiments: MC XP ZS AC ZX VA. Contributed reagents/materials/analysis tools: MC XP UE DMG VA. Wrote the paper: MC VA WCE. Provided technical assistance: HO.

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