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Pleiotropic Impacts of Macrophage and Microglial Deficiency on Development in Rats with Targeted Mutation of the Csf1r Locus


We have produced Csf1r-deficient rats by homologous recombination in embryonic stem cells. Consistent with the role of Csf1r in macrophage differentiation, there was a loss of peripheral blood monocytes, microglia in the brain, epidermal Langerhans cells, splenic marginal zone macrophages, bone-associated macrophages and osteoclasts, and peritoneal macrophages. Macrophages of splenic red pulp, liver, lung, and gut were less affected. The pleiotropic impacts of the loss of macrophages on development of multiple organ systems in rats were distinct from those reported in mice. Csf1r−/− rats survived well into adulthood with postnatal growth retardation, distinct skeletal and bone marrow abnormalities, infertility, and loss of visceral adipose tissue. Gene expression analysis in spleen revealed selective loss of transcripts associated with the marginal zone and, in brain regions, the loss of known and candidate novel microglia-associated transcripts. Despite the complete absence of microglia, there was little overt phenotype in brain, aside from reduced myelination and increased expression of dopamine receptor-associated transcripts in striatum. The results highlight the redundant and nonredundant functions of CSF1R signaling and of macrophages in development, organogenesis, and homeostasis.

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Targeted mutation of Csf1r in mice depleted tissue macrophages from most organs and had pleiotropic impacts on growth and development (9). The more penetrant phenotype of the receptor mutation, compared with the natural mutation of the Csf1 ligand in the Csf1topo mouse, predicted the existence of the second ligand, IL-34 (9). Phenotypes of the Csf1 or Csf1r mutant mice include increased bone density (osteopetrosis), abnormalities of the sensory nervous system, global defects in brain development, infertility, failure of pancreatic β cell development, and severe postnatal growth retardation (reviewed in Ref. 10, 11). On the original cross-bred background, around 50% of Csf1r−/− mice survived to weaning. On an inbred background, survival beyond 3 weeks was rare (12).

Rat models have been used extensively in the study of human disease and are preferred to mice in areas of inflammation, development, physiology, neurobiology, pharmacology, and behavior (reviewed in Ref. 13). The use of rats has recently been accelerated by the expanding availability of genetic modification technologies via homologous recombination in embryonic stem cells (ESC) and, more recently, CRISPR–Cas9 and other targeted nucleases. This has led to a comeback of the rat in biomedical research (14). Rats provide many experimental advantages over mice such as larger sample volumes, higher resolution in vivo imaging, improved performance in learning/memory behavioral assays, easier surgical procedures, and physiological similarities with humans (15). They are the model of choice in areas such as diabetes, breast cancer, and chronic inflammatory and cardiovascular diseases as well as age-related illnesses (reviewed in Ref. 16). Current knowledge of CSF1R biology in rats has been based upon limited analysis of the toothless (tl/tl) mutation in the ligand CSF1, focused largely upon the osteopetrotic phenotype in bone (17). In this article, we report on the generation Csf1r−/− rats by homologous recombination in ESC and their detailed phenotypic characterization. The results highlight significant differences in CSF1R between species and suggest the rat may be a better predictive model for human macrophage biology.

Materials and Methods
Production of Csf1r−/− rats
An EGFP-PGK-Neo cassette flanked by 2.2 kb 5′ and 5 kb 3′ homology arms (Supplemental Fig. 1A) was introduced into ESC derived from a male Dark Agouti rat [clone DAK31 (18) provided by Austin Smith] by electroporation. Positive selection was provided by a PGK-Neo cassette, and negative selection was provided by a diphertheria toxin-A chain cassette. The diphertheria toxin-A chain cassette was placed upstream of the 5′ homology arm and so was not incorporated into the genome. Csf1r gene-targeted rat ESC clones were identified by 5′ PCR screening and 3′ Southern blot analysis. Of the 24 clones analyzed, 20 were positive at the 5′ end for the targeted allele by PCR, and 18 were positive at the 3′ end for the targeted allele by Southern blot (Supplemental Fig. 1D, 1E). Fifty percent of clones analyzed had normal karyotypes (data not shown), and clone DAK31-C2 was used to generate the colony (Supplemental Fig. 1B).

All experiments were carried out under the authority of a U.K. Home Office Project License under the regulations of the Animals (Scientific Procedures) Act 1986. Approval was obtained from ethics committees of The Roslin Institute and The University of Edinburgh. Procedures (Ref. 9) and 2 h, and a counterstain was not used. Epidermal sheets were obtained by guest on October 1, 2018 http://www.jimmunol.org/ Downloaded from

Western blot
Plasma was separated from EDTA whole blood by centrifugation at 2000 G for 15 min at 4°C. A 1:50 dilution was prepared using SDS-PAGE loading buffer. Proteins were separated by SDS-PAGE and transferred to nitrocellulose membranes for immunoblotting. Membranes were probed with polyclonal primary Abs: rabbit anti-CSF1 (1:500, MB551037; MyBioSource) and 1:2000 sheep anti-Transferrin (1:2000, ab9033; Abcam) in 1:2000 sheep anti-Transferrin (1:2000, ab9033; Abcam) in PBS/3% milk powder/0.1% (v/v) Tween 20 at 4°C overnight. The secondary Abs used were goat anti-rabbit HRP (1:5000, no. 7074; Cell Signaling Technologies) and rabbit anti-sheep HRP (1:10,000, ab71710; Abcam). Blots were visualized using Pierce ECL Western blotting solution (Thermo Fisher Scientific).

Histology and immunohistochemistry
Formalin fixed organs were processed into paraffin using standard procedures. For histological examination, sections were stained with H&E or LUXol fast blue. For immunohistochemical analysis, details of Ag retrieval, Ab concentrations, and detection systems used are listed in Table II. Fe- murs were decalcified in EDTA, and sections were stained using an acid phosphatase and tartrate-resistant acid phosphatase (TRAP) kit (Sigma-Aldrich) according to instructions, except incubation time was increased to 2 h, and a counterstain was not used. Epidermal sheets were obtained after incubating the dorsal face of both ears in 20 mM EDTA for 2 h (37°C), followed by fixation in 4% PFA for 20 min and washing in PBS. Tissues

Gene expression analysis
Livers and spleens were preserved in RNA later (Invitrogen). Total RNA was isolated in TRIzol (Invitrogen), followed by purification with an RNeasy Mini kit (QIAGEN) according to instructions. cDNA synthesis and quantitative real-time PCR (qRT-PCR) were performed as described in (6). The oligonucleotides used are listed in Table I.

Radiographs and micro–computed tomography analysis of bone architecture
Radiographs were produced at the Hospital for Small Animals located at the Royal (Dick) School of Veterinary Studies (Easter Bush, U.K). Micro–computed tomography (µCT) analysis was performed on formalin fixed tibias and skulls using a SkyScan 1172 (Bruker) at a resolution of 17.22 µm with a 0.5-mm aluminum filter and four-frame averaging to improve the signal to noise ratio of the images. The tibias were scanned using a source voltage of 71 kV and a source current of 139 µA. To accommodate the larger dimensions of the skulls, a double width acquisition mode was used with a source voltage of 81 kV and a source current of 122 µA. An exposure time of 1180 ms was used for all scans. The images were reconstructed using NRecon v1.6.9 (Bruker) and analyzed using CTAn v1.13.5 (Bruker) software.

Complete blood count analysis
Whole EDTA blood was analyzed as described in (20).

Serum biochemistry analysis
Serum was analyzed using the ILab 650 automated biochemistry analyzer (Instrumentation Laboratory).

Flow cytometry
Blood was collected into EDTA tubes via cardiac bleeds and prepared using Dako Uti-Lyse erythrocyte lysing solution according to instructions. Blood was stained with AF647-labeled CSF1-Fc (6) and the following Abs: CD32a/P72.5 (Clone REA256, 1:40 and CD161/67PE (Clone REA227, 1:40), from Miltenyi Biotec, B220PE (Clone His24, 1:400; eBioscience), CD45TTC (Clone W3/25, 1:10; Bio-Rad Laboratories), CD3α4488 (Clone IPF4, 1:200), and SIRPα7/8 (Clone OX-41, 1:400) from BioLegend. Myelin-depleted single-cell suspensions of brain were prepared as described in (21), except rats were perfused with physiological saline and heparin and then stained with CD4555555 (Clone OX1, 1:200; ebioscience) and CD11b/6447 (Clone OX-42, 1:500; BioLegend). Peritoneal cavity cells were isolated as described in (22) and stained with SIRPa/β and CD11b/6447 as described above. Isotype controls were used for all flow cytometry data. Cells were analyzed by flow cytometry on a FACSCalibur, LSRFortessa X-20 or LSRFortessa (BD Biosciences). Analysis was performed with FlowJo software (FlowJo).

KNOCKOUT RATS

3′ Southern blot analysis of ESC clones
Genomic DNA (7.5 μg) from ESC clones were digested with XbaI and used in Southern blot analysis as per standard procedures. The probe was 503 bp and spanned the third exon of rat Csf1r (Rat Genome Sequencing Consortium 6.0/rn6, chr18:56,457,991–56,458,493).

Csf1r KNOCKOUT RATS
were blocked for 1 h (room temperature [RT]) in PBS/0.1% BSA/5% goat serum, then stained with anti-rat MHC class II (MHC-II) (clone OX-6, 1:100, Abcam). Secondary Ab was F(ab')2-Goat anti-Mouse IgG (H+L)AF647 (1:500, A-21053; Invitrogen). Epidermal sheets were washed in HBSS (Thermo Fisher Scientific), and nuclei were stained for 5 min (RT) with Hoechst 33258 (1:1000, 861405; Sigma-Aldrich). Stained epidermal sheets were mounted with ProLong Gold (Life Technologies) and cured for 24 h (RT) before imaging.

Microarrays
Total RNA was prepared from snap frozen brain regions (hippocampus, striatum, olfactory bulbs, and pituitary gland) and spleens as described above from nonperfused rats (two female rats per genotype). Library preparation and hybridization to the Affymetrix Gene Rat Gene 2.1 ST array was performed by Edinburgh Genomics, The University of Edinburgh. CEL files were normalized (RNA [23]) and annotated in R/Bioconductor. The data from the microarrays are available at Gene Expression Omnibus, National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/geo/), under accession number GSE100696. To identify spleen-specific genes downregulated by the loss of Csf1r, the normalized transcriptional data were loaded into Graphia Pro (Kajeka, U.K.). Using a Pearson correlation threshold cutoff of R = 0.98, a graph was obtained comprising 20,515 nodes (individual probe sets). Clustering of the graph using the Markov clustering algorithm (MCL) was used with an MCL inflation value of 1.5 to determine the granularity of clusters.

Evaluation of blood–brain barrier permeability
At 48 h at 55˚C, Evans blue stain was measured by spectroscopy. After 48 h at 55˚C, Evans blue stain was measured by spectroscopy. The organs were dried overnight in a 55˚C oven, and Evans blue was extracted by addition of formamide (16 μl/mg spleen and 8 μl/mg brain). After 48 h at 55˚C, Evans blue stain was measured by spectrophotometer at 620 nm and quantified according to a standard curve. The results are presented as microgram of Evans blue per milligram of dried tissue.

Statistical analysis
Analysis was performed using GraphPad Prism version 6.04, and statistical significance was assessed using Student two-tailed t test. Resulting values were considered statistically significant at p < 0.05.

Results
Generation and gross phenotype of Csf1r+/- rats
Csf1r+/- rats were produced using ESC in which the first coding exon of rat Csf1r was disrupted with a drug selection cassette by homologous recombination (Supplemental Fig. 1A). qRT-PCR demonstrated partial (~50%) and complete loss of Csf1r mRNA in spleens from Csf1r+/- and Csf1r-/- rats, respectively (Fig. 1A), indicating that there was no dosage compensation in the heterozygote. The loss of functional CSF1R was demonstrated using a labeled CSF1R ligand [porcine CSF1-Fc (6)]. There was reduced (>60%) binding activity on monocytes (SIRPα, SSclo cells) from Csf1r+/- heterozygous rats. There was no CSF1 binding activity on any blood cells from Csf1r-/- rats (Fig. 1B, 1C). The loss of CSF1R activity was associated with an increase in the ligand in the circulation, as observed in Csf1r-/- mice (9), confirmed by Western blot analysis (Fig. 1D). By contrast, Csf1r-/- rats displayed a significant reduction in Csf1 mRNA in the spleen (Fig. 1E). This likely reflects the depletion of CSF1-dependent macrophages, which in rats (unlike mice, see data on www.biogps.org or www.immgen.org) express abundant CSF1 mRNA (see below).

Csf1r-/- rats were indistinguishable from their littermates at birth but were identified from P9–11 by the absence of tooth eruption, smaller body size, and shorter snout (Fig. 2A), similar to the reported phenotype of the Csf1 mutant (tl/tl) rats (24). The frequencies of wild type, heterozygous, and homozygous genotypes at birth were 27, 52, and 21% (n = 266), respectively, not significantly different from the expected 1:2:1 ratio expected under Mendelian inheritance (p = 0.3). All female Csf1r-/- rats survived beyond weaning (n = 22), but 23% of male Csf1r-/- rats were lost postnatally (n = 8) without obvious clinical signs. Csf1r-/- rats weighed less than their littermates at weaning, and their body weight plateaued after 10wk. As in Csf1r-/- mice, the differential increase in growth/body size normally seen in wild type males was abolished in the Csf1r-/- mutant animals, most likely reflecting a lack of testosterone production (9) (Fig. 2B). Tlh/tlh rats lack circulating insulin-like growth factor 1 (IGF1; responsible for body growth) in the postnatal period, implying a link between CSF1R and growth hormone receptor (GHR) signaling (25). Mouse macrophages grown in CSF1 express high levels of IGF1 mRNA and are likely to be the major extrahepatic source (25). We have confirmed this finding in rats (see below). In the Csf1r-/- rats, the mutation also impacted upon the major source of IGF1 in the postpubertal growth surge. Both Ghr and total IGF1 mRNA levels were consistently reduced by around 50% in the liver of mutant rats compared with age- and sex-matched littermate controls (Fig. 2C).

Csf1r-/- rats were osteoporotic, with reduced bone marrow (BM) cavities and disrupted growth plates in the tibia (Fig. 2D). Although Csf1r-/- rats lack any visibly erupted teeth, μCT analysis of the skull clearly showed molars and rudimentary incisors as well as a domed skull (Fig. 2E). By 6mo of age, a severe curvature of the spine was apparent (Fig. 2F). Ageing Csf1r-/- rats developed further phenotypes, including bulging eyes

<table>
<thead>
<tr>
<th>Table I. Oligonucleotides</th>
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<tr>
<td>Gene</td>
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<tr>
<td>Adgre4</td>
</tr>
<tr>
<td>Cd209b</td>
</tr>
<tr>
<td>Csf1r (genotyping)</td>
</tr>
<tr>
<td>Csf1</td>
</tr>
<tr>
<td>Csf1r</td>
</tr>
<tr>
<td>Fmod</td>
</tr>
<tr>
<td>Gal3s52</td>
</tr>
<tr>
<td>Gapdh</td>
</tr>
<tr>
<td>Ghr</td>
</tr>
<tr>
<td>Hgf</td>
</tr>
<tr>
<td>Il22ra2</td>
</tr>
<tr>
<td>Mpeg1</td>
</tr>
<tr>
<td>Nr1h3 (Lrxa)</td>
</tr>
</tbody>
</table>

*Spans local and circulating transcripts.
R, reverse.

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that secreted porphyrin. The secretions were likely a consequence of the inability to blink properly, causing corneal dryness and increased porphyrin production. Postmortem analysis revealed that both male and female Csf1r<sup>2/2</sup> rats had an almost complete absence of visceral white adipose tissue at 4.5 wk (Fig. 2H) yet still developed interscapular brown adipose tissue (Supplemental Fig. 2A). White adipose tissue was detected in older Csf1r<sup>2/2</sup> rats but was still greatly reduced when compared with littermate controls (Fig. 2I). As the rats aged, they developed increased respiratory rates, most likely a result of cranial bone defects limiting the space available for soft tissues such as the soft palate and tongue, similar to brachycephalic syndrome in certain dog breeds (26). Indeed, the tongues of Csf1r<sup>2/2</sup> rats were the same size as littermate controls, despite their much smaller body size (Supplemental Fig. 2B). A subset of males exhibited a protruding penis, without any adverse effects on behavior or urination, with osteopetrosis of the penile bone being the most probable cause.

Aside from the obvious difference in viability between mutant rats and mice, many pleiotropic impacts of macrophage deficiency described in Csf1 and Csf1r mutant mice (9, 27, 28) were not observed in the Csf1r<sup>2/2</sup> rats. For example, they showed no evidence of sensory defects (sight, hearing, and smell were apparently normal), produced insulin in histologically normal pancreatic islets (Supplemental Fig. 2C), and retained Paneth cells in the intestines, which also showed normal villous architecture (Supplemental Fig. 2D).

Analysis of blood from Csf1r<sup>2/2</sup> rats

Csf1r<sup>2/2</sup> rats had a significant decrease in PBMCs, whereas the percentages of granulocytes and erythroblasts were increased (Fig. 3A, 3B). By contrast with the CSF1 ligand–deficient Csf1<sup>0/l0</sup> rat (29), there was no evidence of thrombocytopenia, nor any change in total RBC or WBC count (Fig. 4A). SIRPα<sup>+</sup> monocytes were unaffected in the heterozygote but were almost undetectable in the homozygous mutant (Fig. 3C, 3D). Conversely, neutrophils

![Figure 1](http://www.jimmunol.org/)

**Figure 1.** Expression of Csf1r and Csf1 in rats. (A) cDNA was prepared from total splenic RNA and Csf1r expression analyzed by qRT-PCR (n = 7<sup>+/+</sup>, 7<sup>+/−</sup>, and 3<sup>−/−</sup>). (B) Whole EDTA blood was used to analyze SIRPα<sup>+</sup> expression and binding of CSF1R to its ligand (CSF1-Fc) by flow cytometry. Regions R1–R3 highlighted in Csf1r<sup>−/−</sup> rat blood were back-gated onto a forward versus side scatter (FSC/SSC) dot plot to highlight the three main cell populations in blood. (C) Combined mean fluorescent intensity of CSF1-Fc binding in SIRPα<sup>+</sup> monocytes (n = 6<sup>+/+</sup>, 8<sup>+/−</sup>, and 5<sup>−/−</sup>). (D) Protein lysates were prepared from EDTA plasma (female adults) and assessed for CSF1 expression by Western blot. An anti-transferrin (TRF) Ab was used as the loading control. (E) cDNA was prepared from total spleen RNA and Csf1 expression analyzed by qRT-PCR (n = 7<sup>+/+</sup>, 7<sup>+/−</sup>, and 3<sup>−/−</sup>). Graphs show the mean ± SEM. Significance compared with wild type is indicated by *p = 0.015 and ****p < 0.0001 using a t test.

### Table II. Immunohistochemistry conditions

<table>
<thead>
<tr>
<th>Target (Clone)</th>
<th>Supplier</th>
<th>Ag Retrieval</th>
<th>Dilution</th>
<th>Detection</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD68 (ED1)</td>
<td>Bio-Rad</td>
<td>Proteinase K 20 min, RT</td>
<td>1:500</td>
<td>ImmPRESS anti-mouse 15 min</td>
</tr>
<tr>
<td>CD163 (ED2)</td>
<td>Thermo Fisher Scientific</td>
<td>Proteinase K 20 min, RT</td>
<td>45 min</td>
<td>DAB, hematoxylin counterstain</td>
</tr>
<tr>
<td>IBA1 rabbit polyclonal</td>
<td>Wako Pure Chemical Corp.</td>
<td>0.01 M citrate buffer, pH 6.0</td>
<td>1:100</td>
<td>ImmPRESS anti-mouse 10 min</td>
</tr>
<tr>
<td>MRC1 rabbit polyclonal</td>
<td>Abcam</td>
<td>1 mM EDTA, 0.1% Tween 20, pH 8 16 h 60°C</td>
<td>60 min</td>
<td>DAB, hematoxylin counterstain</td>
</tr>
<tr>
<td>SIGLEC1 (ED3)</td>
<td>Bio-Rad</td>
<td>0.01 M citrate buffer, pH 6 110°C 20 min</td>
<td>1:200</td>
<td>ImmPRESS anti-mouse 30 min</td>
</tr>
</tbody>
</table>

DAB, 3,3-diaminobenzidine.
were increased and lymphocytes were decreased (Fig. 3E) as reported previously in the Csf1r<sup>2/2</sup> mice (9). There was no change in relative abundance of B, T, or NK cells within the lymphoid compartment (Fig. 4B, 4C). Blood albumin and electrolyte levels were unaffected by the mutation (Fig. 4D, 4E), with the exception of calcium and inorganic phosphate, which were both significantly reduced, most likely associated with deficient bone resorption.

Fertility of Csf1r<sup>2/2</sup> rats

Csf1r<sup>2/2</sup> male rats reach sexual maturity and were reported to be fertile (17). To date, male Csf1r<sup>2/2</sup> rats have produced no pregnancies when placed with wild type females. Histological analysis revealed a severe reduction in the number of mature sperm cells in the lumen of the epididymis and a large number of atypical residual bodies, suggesting impaired maturation of spermatids (30) (Fig. 5A). The mAb ED1 recognizes rat CD68 and is commonly used to identify rat macrophages (31). In the testis of Csf1r<sup>2/2</sup> males, there was a decrease in CD68<sup>+</sup> interstitial macrophages (Fig. 5B), similar to that shown in mice treated with anti-CSF1R (32). The size difference between Csf1r<sup>2/2</sup> females and adult male wild type rats precluded test mating on welfare grounds, but the females are also likely to be infertile. Their ovaries were greatly reduced in size, and corpora lutea were either absent or greatly reduced (Fig. 5C). The few corpora lutea that were observed lacked CD68<sup>+</sup> macrophages (Fig. 5D), which are known to regulate corpus luteum development in mice (33).

Analysis of tissue macrophages in Csf1r<sup>2/2</sup> rats

There was a reduction of CD68<sup>+</sup> macrophages in both the red pulp and the periarteriolar lymphoid sheath in spleen compared with wild type and heterozygous rats (Fig. 6A, 6B). This decrease was partly due to extensive infiltration of the red pulp by small mononuclear cells. There were no nucleated RBCs and no obvious increase in the number of megakaryocytes (Fig. 6A). Hence, there was no evidence of the extramedullary hematopoiesis that occurs in Csf1r<sup>2/2</sup> mutant mice in the absence of a BM cavity (9). To evaluate the impact of the mutation on spleen cell populations, we...
The selective loss of the marginal zone macrophages may explain the disrupted splenic architecture and the migration of lymphocytes into the red pulp.

In the liver, CD68+ Kupffer cells were unaffected in the heterozygotes but partially depleted in Csf1r−/− rats. The residual CD68+ cells appeared smaller and less ramified, but there was otherwise no gross change in liver size (relative to body weight) or architecture (Fig. 7A, 7B). Liver enzymes, alanine aminotransferase, and alkaline phosphatase were marginally increased in the circulation, but bile acids and bilirubin were unchanged and there was no histological evidence of injury (Fig. 4D, 4E). As in the Csf1r−/− mice (9), peritoneal macrophages were absent in the Csf1r−/− rats (Fig. 7C).

To examine effects on bone architecture, femurs were stained for TRAP (also known as ACP5) and CD68. As noted also in Fig. 2D, despite the cortical thickening and increased trabecular bone, the Csf1r−/− rats retained a substantial BM compartment. This likely explains the lack of extramedullary hematopoiesis in the spleen. Osteoclasts were entirely deficient in Csf1r−/− rats (Fig. 7F), as reported in Csf1r−/−/− rats (17), whereas in Csf1r−/− mice TRAP+ cells are retained, albeit at a lower frequency (9). The surface of bone in mice and humans contains a population of specialized osteal macrophages, intercalated among the osteoblasts (36). Abundant osteal macrophages were detected with CD68 staining in rat bone, and they were completely absent in bones from the Csf1r−/− animals (Fig. 7G).

In mice, IL-34 is also expressed in the epidermis and is required for the development of the MHC-II–positive epidermal Langerhans cells (37). In the Csf1r−/− rats, MHC-II+ Langerhans cells,
visualized in epidermal sheets, were almost completely depleted (Fig. 7H, 7I); the few remaining cells were only weakly MHC-II+.

Dermal macrophages were also absent in Csf1r\(^{-/-}\) rats as detected by CD206 (mannose receptor, MRC1) expression (Fig. 7J).

Analysis of Csf1r\(^{-/-}\) rat brains

The brains of Csf1r\(^{-/-}\) rats at 11 wk of age revealed few abnormalities apart from some enlargement of the lateral ventricles (Fig. 8A). Whereas Csf1r\(^{-/-}\) mice had hollow and fragile olfactory bulbs (27), these were intact and apparently normal in the rat (Fig. 8B). Minor loss of myelin was apparent at the level of the corpus callosum (Fig. 8C). Csf1r-deficient mice had a gross deficiency of microglia, detected with the microglial marker, IBA1 (27). IBA1+ cells with ramified morphology were similarly not detectable in any region of the brain (Fig. 8D) or in the plexiform layers of the retina (Fig. 8F, 8G) in adult Csf1r\(^{-/-}\) rats. To confirm that this deficit was due to a loss of microglia, rather than loss of the marker, cells extracted from total brain tissue were analyzed (21). CD11b/c+CD45low microglia were entirely absent in Csf1r\(^{-/-}\) rats (Fig. 9A), and CD11b/c+CD45hi macrophages (normally a minor component in brain digests) were decreased. The population of CD11b/c+CD45hi cells was increased in Csf1r\(^{-/-}\) rats. We attribute this to blood contamination as a result of incomplete perfusion (discussed below).

The brain, like the spleen, contains several macrophage populations in addition to microglia (38). To further analyze the impact of Csf1r mutation on brain myeloid populations and on other brain cells, we profiled gene expression in the striatum, hippocampus, olfactory bulbs, and pituitary gland. The full dataset is provided in Supplemental Table I. There was no global change in expression of generic neuron-associated genes in Csf1r\(^{-/-}\) rats shared by the four brain regions examined, but there was some evidence of region-specific impacts, discussed below.

The set of 131 transcripts with expression ratios from 0.03 (Ctss, Cx3cr1, and Csf1r) to 0.5 (Supplemental Table I) contains many microglia-enriched transcripts identified in mice (Fig. 9B) (39–41) and rats (42). By contrast to the loss of microglia-specific markers, many macrophage-associated marker genes remained readily detectable in Csf1r\(^{-/-}\) brains (e.g., Cd14 and Fcgr1a encoding CD64).

We attempted to locate residual macrophages with known markers. As in the Csf1r-deficient mouse (27), there were no detectable CD68+ cells in Csf1r\(^{-/-}\) rat brains via immunohistochemistry (e.g., Cd14 and Fcgr1a encoding CD64). We attempted to locate residual macrophages with known markers. As in the Csf1r-deficient mouse (27), there were no detectable CD68+ cells in Csf1r\(^{-/-}\) rat brains via immunohistochemistry (data not shown). Csf1r\(^{-/-}\) rats retained IBA1+ macrophages in the choroid plexus (Fig. 9C) but lacked detectable CD163+ perivascular macrophages (43) and MRC1+ meningeal macrophages (38) (Fig. 9D, 9E).

The blood–brain barrier was intact in Csf1r\(^{-/-}\) rats, as evident from Evans blue exclusion. Interestingly, there was an obvious
reduction in Evans blue staining in the periphery of Csf1r2/2 rats, suggesting a subtle vascular phenotype. Evans blue staining in the spleen was also reduced (Fig. 9A), despite no differences in levels of the carrier albumin (Fig. 4E), potentially highlighting the known role of macrophages in angiogenesis (reviewed in Ref. 44).

Network analysis of gene expression in the spleen of Csf1r-deficient rats

In previous studies of the effect of CSF1 treatment on macrophage numbers and gene expression profiles in the liver, BMDM were used as an external reference to identify genes that were enriched in macrophages relative to the tissue (45). These genes provided a signature of CSF1-dependent expansion of the macrophage population detectable in total mRNA from the liver (45). To provide a similar reference for the rat, BMDM were generated by cultivation in a medium containing recombinant human CSF1 for a period of 7 d as described for mouse (45), mRNA was isolated, and transcript expression was profiled in parallel with tissue mRNA from adult rats.

To identify the set of genes that were spleen enriched, based upon comparison with BMDM grown in CSF1, the wild type, heterozygous, and homozygous Csf1r mutant expression profiles were combined with BMDM and clustered using Graphia Pro. Surprisingly, one of the two heterozygous mutant animals had much lower expression of Csf1r, albeit higher than in the knockout. (Supplemental Table I).

The network graph divided into two major aggregates, essentially comprising clusters of macrophage-enriched and spleen-enriched
transcripts. Fig. 10A shows the clustered nodes for the main element in the layout graph. Five clusters of specific interest are among those listed in Supplemental Table I, together with their average expression profile. The largest cluster, cluster 1 (2171 genes; green nodes), was strongly enriched in the spleen compared with BMDM and not sensitive to \textit{Csf1r} mutation. The second largest cluster (cluster 2, 1986 genes; purple nodes) was strongly enriched in the BMDM and also was not affected by the loss of \textit{Csf1r}.

This list is a complex mixture of genes associated with phagocyte biology and include \textit{Adgre1} (F4/80), \textit{Itgam} (CD11b), components of the vacuolar ATPase (\textit{Atp6v} family), \textit{Fcgr}, and \textit{Clec} families. The existence of this cluster indicates that there is a substantial residual macrophage population in the spleen of \textit{Csf1r} \textsuperscript{2/2} rats, and they share with BMDM the majority of the generic macrophage transcriptome. That conclusion is consistent with the residual CD68 staining in Fig. 6A and 6B.

Within cluster 2, there were more general anabolic/metabolism-associated genes involved in mitochondria (e.g., \textit{Ndaf}, \textit{Cox}, \textit{Mrpl} families) and RNA/protein synthesis. The enrichment of these genes in cluster 2 reflects the difference between a pure growing macrophage population and the splenic lymphoid population, in which most of the cells are relatively metabolically inactive.

\textit{Csf1r}-dependent clusters varied in their level of spleen enrichment. The expression profile of cluster 6 (238 genes) was much higher in spleen than in BMDM and downregulated in the knockout and in the low \textit{Csf1r}-expressing heterozygote. Cluster 30 (24 genes) had a similar expression profile to cluster 6, but with higher relative expression in BMDM. The spleen-specific cluster 6 contained the signature gene for the marginal zone macrophages (\textit{Siglec1}) which was among the most downregulated with the loss of \textit{Csf1r} (Supplemental Table I). The IL-22–binding protein, encoded by \textit{Il22ra2}, has recently been studied in detail in Peyer
patch, where it is highly expressed by myeloid cells underlying follicle-associated epithelium, regulates IL-22 signaling, and indirectly affects bacterial uptake (46). The loss of this transcript was confirmed by qRT-PCR (Fig. 6F).

The gene expression profile in the spleen of Csf1r-deficient rats is consistent with almost complete loss of the marginal zone macrophage populations. These cells, and the related Ag capture cells in the subcapsular sinus of lymph nodes, were also depleted in op/op mice (47). The data suggest that relatively few Csf1r-dependent genes are absolutely restricted to the marginal zone.

Genes that are less spleen specific but are nevertheless Csf1r dependent (e.g., those in cluster 13, including Sirpa) may be shared by macrophages of the marginal zone and red pulp, or specifically associated with red pulp macrophages. Overall, the main impact of Csf1r-deletion is to compromise spleen-specific macrophage differentiation.

Network analysis of gene expression in the brain of Csf1r-deficient rats

As with the spleen, the first pass analysis of gene expression data for two brain regions was to determine the expression ratios (ratio of Csf1r<sup>−/−</sup>/littermates) for each of the transcripts on the arrays in each of the tissues. There was no global change in expression of generic neuron-associated genes in Csf1r<sup>−/−</sup> rats shared by the brain regions examined, but there was some evidence of region-specific impacts (Supplemental Table I). There were several overlaps between the Csf1r-dependent genes in the brain regions and spleen, including Gpr31, Gpr34, Cd33, Tmem119, Tmem176a, P2ry13, Pld4, Clec4a1, and C1q genes, but
most were not shared, consistent with the tissue-specific adaptation of Csf1r-dependent macrophages in the two locations. Fig. 10B shows the network graph for the combined analysis of the four brain regions; gene lists are provided in Supplemental Table I. The largest cluster was cluster 1 (4283 genes), containing olfactory bulb–associated genes. Other region-specific clusters were cluster 3 (pituitary gland; 878 genes), cluster 5 (hippocampus; 513 genes), and cluster 7 (striatum; 384 genes). The second largest cluster (cluster 2; 1253 genes) contained genes that were more highly expressed in both hippocampus and striatum. Two other main clusters also shared highly expressed genes between tissues: cluster 4 (pituitary gland, striatum and hippocampus; 757 genes) and cluster 6 (pituitary gland and olfactory bulb; 471 genes). None of these clusters showed any evidence of genotype association. Cluster 10 (107 genes) contained genes that were downregulated in the Csf1r−/− mice in all brain regions, including Csf1r itself (Fig. 10B). This set contains most known microglia-specific transcripts and is presumably attributed to the loss of microglia.

Discussion

The Csf1r knockout rats described in this article join a small set of rat knockout lines that have been produced via homologous recombination in ESC ([48] and reviewed in Ref. 13). The comparison between rat and mouse mutant phenotypes supports the view that the rat, in many cases, provides a superior model for studying human genetic disease. For example, p53–deficient rats exhibited a more human-like tumor spectrum than mice (49), and dystrophin mutant rats displayed heart defects which were more similar to the human condition Duchenne muscular dystrophy (50, 51). The phenotype of Csf1r−/− rats is clearly very different from mice, and because of the improved viability, we have assayed phenotypes that could not be accessed in mice. In any insertional mutagenesis, or deletion, there is always the possibility of cis-acting impacts on neighboring genes. The two genes neighboring Csf1r, namely Hmgxb3 and Pdgfrb, are both robustly expressed in all three tissues examined by gene expression profiling, and there was no significant change in their expression in response to insertional mutagenesis of Csf1r.

A common trait between Csf1r−/− rats and mice is increased bone density (osteopetrosis) attributed to either a reduction (mice) or complete absence (rats) of bone-resorbing osteoclasts. In humans, genetic susceptibility to disordered bone turnover in Paget disease has been linked to variation at the Csf1 locus (52). In Csf1ltdo rats, there is a severe loss of osteoclasts in addition to the loss of osteoblasts (53). Unlike Csf1ltdo rats (54), the Csf1r−/− rats developed a BM cavity, albeit reduced, and showed no evidence of extramedullary hematopoiesis in the spleen. Nevertheless, only monocytes were greatly reduced in the blood. We speculate that the loss of macrophages in liver and spleen increases the half-life of red cells, platelets, and neutrophils, compensating for reduced production by the BM. The receptor mutants also do not display the early onset deafness seen in Csf1ltdo rats, attributed to auditory ossicle abnormalities (55). The loss of osteoclasts in Csf1r−/− rats is probably balanced in part by deficient calcification by osteoblasts. Osteoblasts on the bone surface interact intimately with a population of osteal macrophages, which are essential for intramembranous and endochondral ossification (36, 56). In the Csf1r−/− rat, there were no CD68+ osteal macrophages detectable on the bone surfaces (Fig. 7F). The difference between ligand

![Image](https://www.jimmunol.org/)

FIGURE 8. Analysis of brains from Csf1r−/− rats. All brains were formalin fixed and paraffin embedded for histology and immunohistochemistry. (A) Brains from 11-wk-old rats were stained with H&E. Images are representative of seven rats per genotype. Scale bar, 5 mm. (B) Adult heads were fixed in formalin and stained with H&E following EDTA decalcification. Image shows olfactory bulbs in situ. Scale bar, 1 mm. Image representative of seven rats per genotype. EPL, external plexiform layer; GL, glomerular layer; IGC, internal granular cell layer of the olfactory bulbs; MCL, mitral cell layer. (C) Formalin-fixed brains from 3-wk-old rats were stained with Luxol fast blue. Arrows point to myelin preservation and myelin pallor in wild type and mutant rats, respectively. Image of three rats per genotype. (D) Adult brains (8–14 wk) were stained with an Ab against IBA1. Scale bar, 1 mm (left) or 100 μm (right). (E) For each rat, 10 20× images of the cortex were analyzed for the percentage of IBA1+ staining using ImageJ (n = 7 per genotype). Graphs show mean ± SEM. Significance compared with wild type is indicated by ****p < 0.0001 using a t test. (F) and (G) Formalin-fixed eyes were stained with an Ab against IBA1. Arrows point to IBA1+ microglia in the inner plexiform layer of the retina. Images are representative of two rats per genotype, two repeat experiments. Scale bar, 250 μm (F) and 50 μm (G).
and receptor mutations presumably reflects the role of the second ligand, IL-34. Both CSF1 and IL-34 (57) are produced by osteoblasts and contribute to osteoclastogenesis. It appears likely that IL-34 is essential for rat osteocytic macrophage development. Conversely, there is a lack of sperm development and fertility in male Csf1r Knockout rats, whereas Csf1r knockout male animals are fertile. This suggests that the alternative ligand, IL-34, has a specific function in testis development in the rat. Consistent with this hypothesis, Il34 mRNA is highly expressed in rat testis and downregulated by candidate male contraceptives (58).

Other phenotypes shared between Csf1r Knockout rats and mice include postnatal growth retardation, lack of tooth eruption, reduced circulating monocyte numbers, defects in fertility, reduction in liver and peritoneal macrophages, and loss of Langerhans cells. The most obvious difference between Csf1r Knockout rats and mice is the greatly increased viability, relative lack of a severe brain phenotype, and absence of effects on the sensory nervous system, especially the latter two, probably contribute to the improved postweaning survival of the rat knockout compared with the mouse. The gut phenotype in mice has been attributed to the consequent loss of circulating IGF1, as noted previously in the liver of Csf1r Knockout rats (45). A previous study also noted the loss of GHR expression in the bone of Csf1r Knockout mice, and was associated with an increase in goblet cells (32). More recently, we demonstrated that Csf1r mRNA is not expressed in mouse Paneth cells, but macrophages intimately associated with the crypt control the differentiation of these cells and Lgr5 stem cells. Whereas lysozyme expression was Csf1r dependent, other Paneth cell markers, including defensins, were not. In addition to increased goblet cell formation, anti-CSF1R treatment produced a complete loss of Ag-sampling M cells (61). In the rat Csf1r knockout, there was no effect on the cellularity of the lamina propria, no loss of Paneth cells, no increase in goblet cells, and no discernible change in villus architecture (Supplemental Fig. 2). In the rat Csf1r mutant, there was no apparent impact on the intestine.

In contrast, the almost complete loss of visceral fat in Csf1r Knockout rats at an early age is a novel phenotype that has not previously been noted in mice. The Csf1r Knockout mouse is not deficient in adipose tissue but does have a reduction of macrophages in this tissue (63). Adipose tissue growth is related to growth hormone and IGF1 production (64). Hence, the loss of adipose tissue is most likely linked to the reduced GHR and IGF1 we observed in the liver and the consequent loss of circulating IGF1, as noted previously in Csf1r Knockout rats (45). A previous study also noted the loss of GHR expression in the bone of Csf1r Knockout rats (65).

Injection of Evans blue stain highlighted a deficiency in development of the peripheral vasculature in Csf1r Knockout rats. Notably,
FIGURE 10. Network analysis of gene expression in the spleen and brains of Csf1r deficient rats. RMA-normalized microarray data from Supplemental Table 1 was analyzed with Graphia Pro. Edges have been removed for ease of visualization. Nodes allocated to the same cluster are the same color. Histograms show the averaged expression patterns of all genes in the cluster. Boxed clusters refer to genes affected by loss of Csf1r. Unboxed clusters refer to genes that are tissue specific (*). (A) Key clusters from spleen. All known genes in which no sample reached an intensity of 20 were excluded. Analysis was performed at a Pearson correlation coefficient ≥0.95 (12,305 nodes making 1,746,925 edges). Clustering was performed at an inflation of 2.0 with a minimum cluster size of 10. (B) Key clusters from brain. All known genes in which no sample reached an intensity of 20 were excluded. Analysis was performed at a Pearson correlation coefficient ≥0.85 (11,833 nodes making 3,617,804 edges). Clustering was performed at an inflation of 2.0 with a minimum cluster size of 10. Three clusters (circled numbers) shared gene expression with multiple brain regions: cluster 2 (striatum and hippocampus), cluster 4 (pituitary gland, striatum, and hippocampus), and cluster 6 (olfactory bulb and pituitary gland). Histograms for these clusters are shown in Supplemental Table 1.
the apparent leakiness in the spleen (Supplemental Fig. 3B) appears to be associated with the marginal zone and may be an indirect consequence of the loss of the marginal zone macrophages. The \( \beta / \beta \) CSF1 mutant rats display impaired capillary proliferation in the femur, which can be rescued with CSF1 treatment (66). However, the peripheral blood vessels have not been examined in these rats.

The gene expression analysis in Fig. 6C and 6D allowed us to identify spleen-specific genes that were downregulated with the loss of CSF1r. The large majority of genes enriched in BMMDM relative to spleen were not affected. Transcripts associated with classical dendritic cell differentiation, such as \( \text{Ilgax} \), \( \text{Znhr46} \), \( \text{Lyz57} \), or \( \text{Clec9a} \), were also unaffected, suggesting that these cells are not CSF1r dependent. Those genes that were both spleen enriched and CSF1r dependent provide a surrogate indicator of the loss of specific cell populations. The marginal zone macrophages of the mouse have resisted isolation and characterization (41). The commonly studied marker genes \( \text{Siglec1} \) (marginal zone macrophages) and \( \text{CD209b} \) (marginal metallophil) (35) were almost ablated in \( \text{CSF1r}^{-/-} \) rats, and coregulated genes are implicated, by association, in marginal zone macrophage differentiation and function (Supplemental Table I). Very few genes show the same level of CSF1r dependence. One example is the IL-22-binding protein, encoded by \( \text{Il22ra2} \). This transcript is highly expressed by myeloid cells underlying mouse follicle-associated epithelium, regulates IL-22 signaling, and indirectly affects bacterial uptake (46).

The marginal zone macrophage population is also absent from \( \text{CSF1r}^{+/+} \) mice (47). Marginal zone macrophages in mice are distinguished from red pulp macrophages by expression of the widely used macrophage marker, \( \text{F4/80} \), encoded by \( \text{Adgre1} \) (previously \( \text{Emr1} \)) (67). \( \text{Adgre1} \) is also highly expressed by rat macrophages (Supplemental Table I), and the relative loss of expression in the knockout spleen (∼70%) is consistent with expression in the red pulp macrophages. The related gene, \( \text{Adgre4} \) (previously \( \text{Emr4} \)), was strongly spleen enriched and highly CSF1r dependent (Fig. 6F). ADGRE4 was shown to mediate binding to expression in the red pulp macrophages. The related gene, \( \text{Csf1r} \), is associated with differentiation of plasmacytoid dendritic cells (78). Plasmacytoid dendritic cells express CSF1r and were found to be 70% reduced in the spleen of \( \text{op/op} \) (CSF1-deficient) mice (79). The alteration of the gene expression profile in the spleen of CSF1r-deficient rats is consistent with the immunohistochemistry (IHC) data and indicates almost complete loss of the marginal zone macrophage populations. These cells, and the related Ag capture cells in the subcapsular sinus of lymph nodes, were also depleted in \( \text{op/op} \) mice (47). Genes that were less spleen specific, but are nevertheless CSF1r dependent (e.g., those in cluster 13 and 30), may be shared by macrophages of the marginal zone and red pulp or specifically associated with red pulp macrophages.

\( \text{CSF1r}^{-/-} \) rats had no microglia detected by staining for IBA1 in the brain and retina or in flow cytometry profiles of brain digests (Figs. 8D–G, 9A). This observation was strongly supported by the network analysis of gene expression in the four brain regions shown in Fig. 10B. Cluster 10 (Supplemental Table I) provides a list of 107 rat microglia-enriched genes that have been previously published as expressed in microglia from mouse (21, 39, 40, 80, 81) and human (81, 82). Cluster 10 also includes 18 of the top 35 genes downmodulated in the brains of mice treated with a CSF1R kinase inhibitor to deplete microglia (83). As expected, CSF1r was almost undetectable in the homoygotes and was the only transcript that was also reproducibly reduced, by 40–50%, in the heterozygous mutants, in all brain regions. Expression of \( \text{Cx3cr1} \), as well as cathepsin \( \text{S} \) (\( \text{Ctsb} \)), the enzyme required for CX3CL1 cleavage (84), was also almost completely ablated, to the same extent as \( \text{CSF1r} \). Microglia-associated transcripts are of particular interest given the emerging consensus that these cells are central players in the pathologic condition of Alzheimer disease. Genes including \( \text{CSF1r} \) and others within cluster 10, including \( \text{Trem2} \), \( \text{C1q} \), \( \text{Tyrbp} \), \( \text{Abi3} \), and \( \text{Spil} \), are associated with disease susceptibility (85–87).

Many known macrophage-associated transcripts were less affected by \( \text{CSF1r} \) mutation, including \( \text{Cd14} \), \( \text{Csf2ra} \), \( \text{Merk} \), \( \text{Fcg2a} \) (CD32), \( \text{Stab1} \), \( \text{Mrsl} \), \( \text{Marc0} \), \( \text{Gpr84} \), \( \text{Icam1} \), \( \text{Clec7a} \), and several TLR (\( \text{Tlr2,4,8,9,11} \)) (average 25% less expression; Supplemental Table I). Among macrophage-expressed transcription factors, \( \text{Spil} \) (\( \text{PU.1} \)) was reduced by around 75%, but \( \text{Ifnr9} \), \( \text{Cebpa} \), \( \text{Cebph} \), \( \text{Runx1} \), \( \text{Tfrc} \), and \( \text{Stat5A} \), all of which were readily detectable and are expressed by microglia in the mouse (39, 41), were reduced by <37%. One interpretation is that these transcripts are more highly expressed in the nonmicroglial macrophage population, which is also less \( \text{CSF1r} \) dependent. The \( \text{CD45}^{hi} \) brain macrophage population in rats was isolated and characterized by Ford et al. (88). They were larger than microglia, with a lower nucleus/cytoplasm ratio. A subset expressed MHC-II genes (RT1-D, RT1-A, RT1-DM, RT1-Db1, RT1-Db2, RT1-Doh) and were found to be 70% reduced in the spleen of \( \text{op/op} \) (CSF1-deficient) mice (79).

Few impacts on overall brain architecture are shared with the \( \text{CSF1r} \) knockout mouse (27). There was little overt phenotype and specifically no impact on the olfactory bulb, where the mouse mutation had the greatest effect (Fig. 8B). A recent study described a specific microglial population associated with myelination in the brain in the postnatal period (89). These cells were proposed to be the major source of \( \text{IGF1} \) required for myelination. The loss of these cells could explain the minor deficiency of myelination and myelination-associated transcripts (Figs. 8C, 9B).
observed in the mutant rats. However, in the gene expression profiles of rat brain regions, there was no deficiency in Igf1 mRNA associated with the Csf1r mutation. Similarly, microglia have been considered an important source of brain-derived neurotrophic factor (BDNF), although conditional microglial depletion did not reduce the total level of Bdnf mRNA in the mouse cortex or hippocampus (90). Bdnf mRNA was not reduced in the Csf1r mutant rat brain. Presumably, the other cellular sources of these trophic factors can compensate for the absence of microglia. It remains to be seen whether the loss of microglia produces the alterations in synaptic proteins and synaptic plasticity observed in mouse microglia depletion studies (90, 91). Microglia in mice have been associated with the outgrowth of dopaminergic neurons, and their depletion produced an imbalance in dopaminergic innervation of the striatum (92). In the striatum of Csf1r-deficient rats (Supplemental Table 1), we observed a 30–40% increase in mRNA encoding dopamine receptors (Drd1, 2, and 3), the dopamine transporter Slc6a3, and other genes correlated with dopamine receptor expression across brain regions, Gnal, Dgkb, Pcp4l1, Kcnq1, and Tac1 (93). More subtle brain phenotypes may emerge from further investigation. The advantage of the rat model is that these phenotypes can be studied in adult animals in the absence of gross changes in brain architecture. The flow cytometry analysis and gene expression profiling indicate that there is no compensatory monocyte recruitment or inflammatory cytokine production in the Csf1r-/- brain, consistent with the absence of circulating monocytes. Monocyte recruitment is the hallmark of brain pathologic conditions (38, 94) and has been observed in other models of microglial deficiency (40). We suggest that the impact of the loss of the neuroprotective functions of microglia in the Csf1r-/- rat may be mitigated by the absence of monocytes.

The human disease ALSP (formerly known as hereditary diffuse leukoencephalopathy with spheroids (95)) is associated with mutations affecting conserved amino acids in the kinase domain of CSF1R (96), which are likely to have a dominant-negative impact on signaling by forming inactive heterodimers with the wild type receptor (8). Fifty-eight different mutations have been described in ALSP patients (7), and within the human exome database (exac.broadinstitute.org), there are a further 38 variants, each identified only as a singleton, that affect amino acids conserved in humans to chickens. One report claimed that the disease could be associated with haploinsufficiency arising from a frame-shift mutation (7), but their data indicate >50% loss of CSF1R protein in the brains of these patients. Chitu and colleagues (97) have characterized Csf1r-/- mice as a model for ALSP. Like these authors, we found that there is no dosage compensation for loss of one Csf1r allele, which is surprising because CSF1 signaling regulates Csf1r mRNA levels (98). However, despite the lack of dosage compensation, we have found no significant impact of the Csf1r heterozygous mutation on gene expression in the adult brain (Supplemental Table 1). We have maintained heterozygous mutant rats for more than 12 months and observed no overt phenotype. Accordingly, the impact of dominant-acting mutations in patients is likely to depend in part upon the level of expression of the other allele and on the activity of the compensatory mechanisms that permit relatively normal brain function when microglial differentiation is compromised.

In overview, analysis of Csf1r-/- rats highlights selective impacts of the mutation on macrophage populations and clear differences in the cell populations affected and the pleiotropic consequences compared with mutant mice. The effect of Csf1 mutation in mice on macrophages and osteoclast populations is gradually corrected with age. The macrophage phenotype can be corrected with exogenous GM-CSF (CSF2), but endogenous CSF2 is not required (99). Conversely, mutation of Flt1 prevents the age-dependent development of osteoclasts (100). As noted above, one major difference between rats and mice is that rat macrophages themselves express high levels of Csf1 mRNA. This explains the loss of Csf1 mRNA in the spleen of Csf1r mutant rats. In the brain of rats, Csf1 and Il34 were both highly expressed and not notably region specific. The Csf1r mutation did not reduce the level of expression, suggesting that microglia are not the major source. We suggest that differences between the rodent species derive from differences in expression or function of alternative growth factors that can compensate for the lack of Csf1r. Although female Csf1r mutant rats appear infertile, hormone-dependent expression of CSF2 and other factors by the uterus (101) might also mitigate the impact of the Csf1r mutation on macrophage numbers and function in females relative to males.

The increased longevity and plethora of phenotypes found in Csf1r-/- rats provide a unique model for studying the role of CSF1R during development and adulthood, the functions of microglia, and other tissue macrophage populations and the molecular basis for adult onset disease associated with human CSF1R mutations. Many of the macrophage phenotypes are shared between mice and rats, so the differences mainly lie in the relative redundancy of macrophages in developmental processes in the two species.

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