Loss of Topoisomerase I leads to R-loop-mediated transcriptional blocks during ribosomal RNA synthesis

Citation for published version:

Digital Object Identifier (DOI):
10.1101/gad.573310

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Genes & Development

Publisher Rights Statement:
Article is online at http://www.genesdev.org/cgi/doi/10.1101/gad.573310. Freely available online through the Genes & Development Open Access option.

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Loss of Topoisomerase I leads to R-loop-mediated transcriptional blocks during ribosomal RNA synthesis

Aziz El Hage,1 Sarah L. French,2 Ann L. Beyer,2 and David Tollervey1,3

1Wellcome Trust Centre for Cell Biology, University of Edinburgh, Edinburgh EH9 3JR, United Kingdom; 2Department of Microbiology, University of Virginia Health System, Charlottesville, Virginia 22908, USA

Pre-rRNA transcription by RNA Polymerase I (Pol I) is very robust on active rDNA repeats. Loss of yeast Topoisomerase I (Top1) generated truncated pre-rRNA fragments, which were stabilized in strains lacking TRAMP (Trf4/Trf5–Air1/Air2–Mtr4 polyadenylation complexes) or exosome degradation activities. Loss of both Top1 and Top2 blocked pre-rRNA synthesis, with pre-rRNAs truncated predominately in the 18S 5′ region. Positive supercoils in front of Pol I are predicted to slow elongation, while rDNA opening in its wake might cause R-loop formation. Chromatin immunoprecipitation analysis showed substantial levels of RNA/DNA hybrids in the wild type, particularly over the 18S 5′ region. The absence of RNase H1 and H2 in cells depleted of Top1 increased the accumulation of RNA/DNA hybrids and reduced pre-rRNA truncation and pre-rRNA synthesis. Hybrid accumulation over the rDNA was greatly exacerbated when Top1, Top2, and RNase H were all absent. Electron microscopy (EM) analysis revealed Pol I pileups in the wild type, particularly over the 18S. Pileups were longer and more frequent in the absence of Top1, and their frequency was exacerbated when RNase H activity was also lacking. We conclude that the loss of Top1 enhances inherent R-loop formation, particularly over the 5′ region of the rDNA, imposing persistent transcription blocks when RNase H is limiting.

[Keywords: rRNA synthesis; RNA polymerase I; transcription elongation; topoisomerase; RNase H; R-loops]

Received December 22, 2009; revised version accepted May 18, 2010.

Yeast rDNA is comprised of ~150–200 tandem repeats, about half of which are actively transcribed. The primary transcript of 35S pre-rRNA is processed into the mature 18S rRNA component of the 40S ribosome subunit and 5.8S and 25S rRNA components of the 60S subunit (see Fig. 1A; Supplemental Fig. S1). Active rDNA repeats are heavily transcribed by RNA Polymerase I (Pol I), as visualized by “Miller” chromatin spreads (French et al. 2003), and this was proposed to involve “transcription factories” where the DNA template is reeled through a polymerase array with constrained mobility [for review, see Cook 1999; Sutherland and Bickmore 2009].

Electron microscopy (EM) analyses (Osheim et al. 2004) and metabolic labeling (Kos and Tollervey 2010) demonstrate that ~50%–70% of yeast pre-rRNA transcripts are cleaved cotranscriptionally. This indicates that yeast preribosome assembly is at least partly cotranscriptional. Consistent with this, ribosome assembly is impaired in mutants with defects in Pol I elongation, indicating a coupling between these processes [Schneider et al. 2007; Zhang et al. 2009]. Surveillance systems monitor yeast ribosome synthesis, and the pre-rRNA components of aberrant preribosomes are polyadenylated by the Trf4/Trf5–Air1/Air2–Mtr4 polyadenylation complexes (TRAMP) and targeted for degradation by the exosome nuclease complex (Dez et al. 2006; Schneider et al. 2007; Wery et al. 2009).

During transcription, either the DNA or the polymerase must rotate once for every ~10 nucleotides (nt) transcribed. DNA rotation generates positive supercoils [more tightly wound DNA] ahead of the transcription bubble, whereas negative supercoils [more loosely wound DNA] are left behind the polymerase [for review, see Wang and Lynch 1993; Cook 1999]. Eukaryotic Topoisomerase I (Top1) Top2 are able to relax both positive and negative DNA supercoils [for review, see Wang 2002], and play essential roles during rDNA transcription in Saccharomyces cerevisiae [Brill et al. 1987; Schultz et al. 1992].

The accumulation of positive supercoiling ahead of the transcription bubble resists opening of the DNA, which can slow or impede transcription elongation by Pol I.
as seen following treatment with the Top1 inhibitor camptothecin (Koster et al. 2007). In contrast, negative supercoiling behind the transcription bubble can lead to opening of the DNA. When this happens, the nascent RNA may hybridize to the transcribed strand, creating RNA–DNA hybrids known as R-loops (Drolet 2006). R-loops can also impede transcription elongation, as well as exposing the nontemplate strand to cleavage and recombination (for review, see Aguilera and Gomez-Gonzalez 2008). Yeast top1Δ mutants show hyperrecombination of the rDNA array (Houseley et al. 2007), while top1Δ top2-ts double mutants undergo major excisions of the rDNA repeats due to a failure to relax negative supercoils (Trigueros and Roca 2002).

R-loops can be removed by cleavage by RNase H1 and RNase H2, which have overlapping functions and similar catalytic mechanisms (Arudchandran et al. 2000). Yeast RNase H1 is a single protein, whereas RNase H2 is a complex of three proteins: Rnh2A [Rnh201], Ydr279 [Rnh202], and Ylr154 (Rnh203) [Jeong et al. 2004]. Seventy percent of RNase H activity is lost upon deletion of RNH201 (Jeong et al. 2004). However, rnh1Δ rnh201Δ double-mutant
strains are viable, indicating that they retain some RNase H activity. Moreover, a residual activity was still able to degrade poly[4A]:poly[dT] substrates in double-mutant cell extracts (Arudchandran et al. 2000), confirming that an additional RNase H activity exists. Yeast RNase H is reported to alleviate transcriptional blocks in THO complex mutants (Huertas and Aguilera 2003), and to destroy RNA–DNA hybrids formed by telomeric TERRA transcripts that accumulate in the absence of Rat1 5′–3′ exonuclease activity (Luke et al. 2008). In eukaryotes, overexpression of RNase H can suppress the genomic instability otherwise induced by R-loop formation following depletion of the pre-mRNA splicing factor ASF/SF2 (Li and Manley 2005) or Top1 (Tuduri et al. 2009).

**Results**

**Aberrant pre-rRNA fragments are generated in top1Δ mutants, and are targeted for degradation by the exosome and TRAMP complexes**

To investigate the effects of loss of Top1 on rRNA synthesis, total RNA extracted from wild-type and top1Δ strains was analyzed by Northern. Several truncated species were identified (labeled 1–9 in Fig. 1), which were mapped by comparison of hybridization patterns using multiple probes across the 35S pre-rRNA (see Fig. 1A).

Two major truncated pre-rRNAs, species 3 and 4, were detected with probes 130 and 033, which hybridize to different regions of the 5′-ETS [Fig. 1B [lane 3], F [lanes 2, 7]], but not with probe 004, which hybridizes between sites D and A2 [Fig. 1C, lane 3]. The RNAs were separated on agarose gels, which have limited resolution, but species 3 and 4 have estimated sizes of ~1.1 kb and ~2.1 kb, respectively. This indicates that species 3 extends from the 5′ end of the pre-rRNA to a position ~0.4 kb into the 18S rRNA region, while species 4 extends ~1.4 kb into 18S. Two less abundant RNAs, species 5 (~4 kb) and 6 (~6 kb), are predicted to extend from the 5′ end of the pre-rRNA to sites within the 25S rRNA [Fig. 1F, lanes 2, 7]. Species 7 and 8 have 5′ ends at the cotranscriptional cleavage site A2, since they were detected with probe 003 (which lies 3′ to site A2) but not with 004 [5′ to A2] [Fig. 1D [lane 3], G [lane 1], Supplemental Fig. S3B, lane 2]. Species 7 was ~0.4–0.5 kb in size, corresponding to 3′ ends within ITS2, whereas species 8 was ~1.2 kb, corresponding to a 3′ end ~0.4 kb into the 25S rRNA region.

The presence of these truncated pre-rRNA species strongly suggested that the absence of Top1 impedes Pol I transcription, leading to cleavage or release of nascent pre-rRNAs.

In other mutants, aberrant preribosomes are targeted cotranscriptionally by the TRAMP nuclear surveillance complex and degraded by the exosome (Dez et al. 2006; Schneider et al. 2007; Wery et al. 2009). We therefore deleted the gene encoding the nonessential, nuclear exosome component Rrp6 in top1Δ strains. The top1Δ rrp6Δ double-mutant strain showed strongly increased accumulation of RNA species 3, 4 [Fig. 1B [lane 4], F [lane 4]], 7, and 8 [Fig. 1D, lane 4, Supplemental Fig. S3B, lane 4] relative to the top1Δ single mutant. Additional pre-rRNA species were also detected, probe 130, which lies close to the 5′ end of the pre-rRNA primary transcript, detected a small mRNA of ~240 nt, designated species 1 [Fig. 1F, lane 4, Supplemental Fig. S3A, lane 4], while 5′-ETS probes 130 and 033 detected an RNA with a size <1 kb, designated species 2 [Fig. 1B [lane 4], F [lane 4]].

Surveillance of aberrant pre-rRNAs by the TRAMP complex is associated with RNA polyadenylation. Purification of poly[A]⁺ RNAs from the top1Δ rrp6Δ strain showed that species 2, 3, 4 (Fig. 1B, lane 8), and 8 (Fig. 1D, lane 8) are indeed polyadenylated. In contrast, species 7 was not detectably polyadenylated (Fig. 1D, lane 8), although it remains possible that short oligo[A] tails were present and escaped our purification method. Deletion of TRF4 in a top1Δ background led to stabilization of species 1, 2 [probe 130] [Fig. 1F [lane 8], G [top panel, lane 2]], 7, and 8 [probe 003] [Fig. 1G, lane 2].

Pre-rRNA fragments extending from the 5′ end of the rDNA into the 5′ region of the 18S were detected with probes 130 and 033 in both single rrp6Δ and double trf4Δ rrp6Δ mutants (TOP1⁺) [species labeled 2 and 3] [Fig. 1F, lanes 3, 6]. Thus, Pol I transcription through the 5′ region of the 18S rDNA might be a limiting step even in otherwise wild-type cells, as proposed previously (Schneider et al. 2007).

In conclusion, the absence of Top1 leads to the accumulation of two major groups of RNAs: species 1–4 extend from the transcription start site to sites within the 5′-ETS and 18S rDNA, whereas species 7 and 8 extend from the A2 site of cotranscriptional cleavage to positions in ITS2 and the 5′ region of the 25S rDNA (see Fig. 1A). These aberrant pre-rRNAs are polyadenylated by TRAMP and degraded by the exosome.

**Pre-rRNA fragments are increased in top1Δ mutants at low growth temperatures**

If positive supercoiling before, and/or negative supercoiling behind, transcribing Pol I restrict elongation in top1Δ strains, this might be exacerbated by a more rigid DNA structure at lower growth temperatures (Drolet 2006). To assess this, top1Δ strains were grown at 18°C,
25°C, or 30°C. Northern analyses showed that accumulation of truncated RNA species 3–6 (probe 130) [Fig. 2A] and 7–9 (probe 003) [Fig. 2B,C] increased progressively from 30°C to 25°C to 18°C. The ratio between species 3 and 4 increased as growth temperature decreased [Fig. 2A, cf. lanes 6,4,2]. Comparison of the levels of truncated pre-rRNA species with those of 35S pre-rRNA [Fig. 2A–C] showed that species 3 and 4 are more abundant than species 7, 8, and 9, arguing against the possibility that all of these fragments might be generated by cleavage of a single truncated pre-rRNA species.

At 18°C, synthesis of 27S pre-rRNA was reduced, whereas levels of 20S pre-rRNA were affected little [Fig. 2D,E, probes 004 and 003; see Supplemental Fig. S1 for the rRNA maturation pathway]. Fast tritium-labeling 2D,E, probes 004 and 003; see Supplemental Fig. S1 for where levels of 20S pre-rRNA were affected little (Fig. 2A–C) increased progressively at lower temperatures, increased transcriptional blockages, in particular in the 5’ region of 18S, would affect the synthesis of all pre-rRNA species. However, blockages downstream from site A2 [Supplemental Fig. S1] would affect only 27S and 35S synthesis, potentially leading to the observed increase in 20S to 27S ratios.

These data indicate that rRNA synthesis in top1Δ strains is more affected at low growth temperatures, where excess supercoils may be more frequent and/or stable [Baaklini et al. 2008]. The predominance of species 3 at lower temperatures suggests increased transcription blockage in the 5’ region of 18S rDNA, with reduced synthesis of downstream RNA species. Increased transcription elongation defects may also produce the elevated 20S to 27S pre-rRNA ratio seen at low temperatures.

**rRNA transcription stalls in the 5’ region of the 18S rDNA in the absence of both Top1 and Top2**

Although pre-rRNAs were truncated and degraded in the absence of Top1, synthesis of full-length pre-rRNA was only partially inhibited. Top1 is partially redundant with Top2, and rRNA synthesis is fully inhibited in double top1Δ top2-ts strains at nonpermissive temperatures [Brill et al. 1987, Schultz et al. 1992]. To analyze rRNA synthesis and pre-rRNA fragment accumulation in the absence of Top2 or both Top1 and Top2, we constructed single P_{GAL-TOP2} and double P_{GAL-TOP2} top1Δ strains. As expected, both mutant strains showed growth inhibition in glucose medium, since Top2 is essential for viability in yeast. Northern analyses detected no major rRNA synthesis defects in the single P_{GAL-TOP2} strain even after several hours of depletion [Fig. 3B–H, lanes 1–6; Supplemental Fig. S3A–F, lanes 9–14]. In double P_{GAL-TOP2} top1Δ strains, depletion of Top2 reduced formation of truncated species 4 relative to the top1Δ single mutant [Fig. 3B, cf. lanes 7 and 8–11]. Formation of the normal 35S, 20S, 27S [Fig. 3B–D, lanes 7–11], and 7S [Supplemental Fig. S3C, lanes 15–19] pre-rRNAs was also inhibited. However, this was not the case for truncated species 1 and 3 [Fig. 3B, lanes 7–11; Supplemental Fig. S3A, lanes 15–19]. In addition, multiple heterogeneous species were detected in the double mutant, with predicted 3’ ends at sites encompassing the region from site A0 (+600 nt in 5’-ETS) to ~500 nt into the 18S rDNA region [Fig. 3A,B, Supplemental Fig. S3A, species labeled by stars].

The accumulation of these truncated fragments, together with the loss of fragment 4 [see Fig. 3A] and all normal pre-rRNAs, strongly indicates that Pol I elongation is arrested predominately within the 5’ region of the 18S rDNA in strains lacking both Top1 and Top2.

Northern analyses on RNA from a top1Δ top2-ts strain gave similar results, showing the preferential loss of truncated species 4 relative to 1 and 3 at the restrictive temperature of 37°C, as well as significant reductions in the 20S and 27S pre-rRNAs (Supplemental Fig. S4A–C, lanes 1,2).
Together, the data indicate that, in the absence of Top1 alone, Pol I transcription elongation is affected mainly over the 18S sequences. In the absence of Top1 at lower growth temperatures, or when Top1 is lacking and Top2 is limiting, transcriptional blocks become predominant in the 5' region of 18S, leading to the inhibition of rRNA synthesis.

The absence of both Top1 and RNase H inhibits rRNA synthesis and increases non-protein-coding RNA transcript (ncRNA) expression from intergenic spacers (IGSs)

If the absence of Top1, or both Top1 and Top2, results in rDNA opening behind Pol I, the associated, nascent pre-rRNA transcripts may form R-loops with the DNA template. R-loops are predicted targets for RNase H activity, which might therefore generate the observed pre-rRNA fragments.

A recent analysis reported that overexpression of the Rnh201 subunit of RNase H2 increased degradation of RNA–DNA hybrids over telomeric regions in the 5'–3' exonuclease rat1-1 mutant (Luke et al. 2008). However, overexpression of Rnh201 in a top1D mutant did not alter the abundance or distribution of pre-rRNA fragments (Supplemental Fig. S2). This was also the case for overexpression of Rnh1 or Rnh201 in top1D top2-ts mutants at 25°C and 37°C (Supplemental Fig. S4).

To test whether pre-rRNA fragments accumulated in top1D mutants are generated by RNase H cleavage, we initially attempted to delete both RNH1 and RNH201 in top1D strains. Deletion of both RNH1 and RNH201 in TOP1 strains had no detectable effect on growth (Arudchandran et al. 2000; data not shown) or rRNA synthesis (Fig. 4B–H, cf. lanes 1 and 9). However, we could not delete both RNH1 and RNH201 in top1D strains, suggesting that top1D, rnh1D, and rnh201D are lethal in combination. We therefore placed TOP1 expression under the control of a conditional PGAL promoter and deleted both RNH1 and RNH201 in this strain. Northern analyses were performed using total RNA extracted from single PGAL-TOP1 and triple PGAL-TOP1 rnh1D rnh201D mutants, which were shifted from permissive galactose-containing medium [0 h] to nonpermissive glucose-containing medium for 2–14 h at 30°C.

Growth of the single PGAL-TOP1 mutant strain was only slightly inhibited after the shift to glucose, consistent with the viability of top1Δ mutants (data not shown). Growth of the PGAL-TOP1 rnh1Δ rnh201Δ triple-mutant strain slowed after 10 h of Top1 depletion in glucose-containing medium, and completely ceased after ~14–16 h (data not shown).

Northern hybridization showed a marked reduction in accumulation of truncated pre-rRNA fragments 3, 4, 8, and 9 in the triple mutant relative to the single mutant during Top1 depletion [Fig. 4D,E [lanes 1–8], I,J], as was observed in top1Δ strains [see Fig. 2D,E]. In contrast, both 20S and 27S accumulation decreased in the triple mutant [Fig. 4D,E [lanes 9–16], I,J].

**Figure 3.** Pre-rRNAs are truncated predominately within the 5' region of 18S rRNA when both Top1 and Top2 are absent. (A) Diagram representing the 5'-ETS and 18S rDNA sequences. Heterogeneous, truncated pre-rRNA fragments that accumulate in the absence of both Top1 and Top2 are depicted by dotted arrows followed by a star. (B–H) Strains P_GAL-TOP2 and P_GAL-TOP2 top1Δ were shifted from galactose-containing medium [0 h] to glucose-containing medium at 30°C [2–10 h]. Total RNA was analyzed by Northern hybridization. The membrane was hybridized successively with probes 130 (B), 004 (C), 003 (D), 008 (F), and 007 (G). (E) Random primed probe IGS2 was used to detect ncRNAs transcribed by RNA Pol II from the intergenic rDNA spacers IGS1 and IGS2, located between the 35S rDNA units. (H) Cytoplasmic SCR1 RNA was used as a loading control. Intact pre-rRNAs and truncated pre-rRNA fragments are labeled. Heterogeneous truncated pre-rRNA fragments are labeled by stars. “(18S)” and “(25S)” indicate the position of migration of 18S and 25S rRNAs. Probe names are bracketed [see Fig. 1A for location of probes].
At early times of Top1 depletion (6 h), the triple mutant showed reductions in both pre-rRNA truncation and synthesis of 20S and 27S pre-rRNA (Fig. 4B–D, lane 12). In the absence of both Top1 and RNase H1 and H201, stable RNA–DNA hybrids may impose transcriptional blocks along the rDNA, further reducing pre-rRNA transcription, as shown in vitro [Tous and Aguilera 2007]. This is supported by the almost complete disappearance of pre-rRNA fragments in the triple mutant after 14 h of Top1 depletion [Fig. 4B,C, lane 16]. The reduced level of pre-rRNA fragments still seen in the triple PGAL-TOP1 rnh1D rnh201D mutant after 6 h of Top1 depletion could arise from the residual RNase H activity observed previously in extracts from strains lacking both RNase H1 and H201 [Arudchandran et al. 2000], or from other RNA cleavage activities (see the Discussion).

RNA Pol II generates ncRNA from the rDNA IGSs [IGS1 and IGS2]. Northern hybridization showed that ncRNAs IGS1-F and IGS2-R accumulated in mutants lacking Top1 [Fig. 3E, lane 7; Fig. 4G, lanes 7,8], as reported previously [Houseley et al. 2007]. Accumulation of these transcripts increased strongly in the triple mutant [Fig. 4G, lanes 9–16], but this was not the case in the P<sub>cal</sub>-TOP2 top1D strain depleted of Top2 [Fig. 3E, lanes 7–11]. Transcription of IGS1-F and IGS2-R is normally limited by a repressive chromatin structure that requires the histone deacetylase Sir2 [Li et al. 2006]. IGS transcription in mutants lacking Top1 is possibly due to disruption of the chromatin structure caused by accumulation of supercoils in the IGS1 and IGS2 regions. The increase in ncRNA levels seen in the triple-mutant strain indicates a specific role for RNase H in the synthesis or degradation of IGS transcripts.

RNA–DNA hybrids accumulate over the Pol I promoter, the 5′-ETS, and the IGS regions of the rDNA when both Top1 and RNase H are absent

The accumulation of pre-rRNA fragments with 3′ ends within the 5′ region of the 18S rRNA, in the absence of Top1 alone or both Top1 and Top2, strongly suggested that Pol I might have to overcome a particular problem that requires topoisomerase activity during transcription of that region.

To gain more insight into the roles of topoisomerase and RNase H activities during Pol I transcription elongation, we analyzed by chromatin immunoprecipitation (ChIP) the distribution of both Pol I [Rpa190] and RNA/DNA hybrids along the rDNA in mutants depleted of Top1, or Top1 and Top2, in the presence or absence of RNase H1 and H201 [Fig. 5]. Wild-type, PGAL-TOP1, and PGAL-TOP1 rnh1D rnh201D strains were harvested after 6 h of Top1 depletion in glucose medium at 30°C, at which time growth was affected little in the triple mutant.
In the wild type (Fig. 5A, blue), the Pol I (Rpa190) signal was maximal over the 5'-ETS (Fig. 5A, probe c), dropped across the 18S, and declined to background levels over the terminator region (Fig. 5A, probe m). ChIP with antibodies directed against the smallest subunit of Pol I (Rpa34-13MYC) gave similar profiles (data not shown). In the single and triple mutants (Fig. 5A, red and green), Pol I signal over the 5'-ETS was shifted toward the promoter region (Fig. 5A, probes a, b), and the overall Pol I signal was decreased in the single and decreased more in the triple mutant.

Together, these data suggest that Pol I occupancy is highest over the 5' region of the rDNA. We propose that R-loops and accumulation of positive supercoils in the 5' region of the 18S rDNA slow transcription elongation. Stalling or pausing of any polymerase results in other polymerases becoming backed up over the upstream regions, and this is particularly frequent in the absence of Top1.

To assess the occurrence of R-loops along the rDNA, we performed ChIP using the S9.6 antibody, which specifically recognizes RNA–DNA hybrids [see Supplemental...
EM reveals polymerase pileups over the 5′-ETS and 5′ region of 18S rDNA in the absence of Top1, RNase H1, and H201

To directly test the model that stable R-loops impose transcriptional blocks along the rDNA, we analyzed Pol I transcription by EM analysis of chromatin spreads. These were prepared from wild-type, P_{CAG-TOP1} [single mutant], and P_{CAG-TOP1} mh1Δ mh201Δ [triple mutant] strains 6 h after transfer to glucose medium at 30°C.

Inspection of Pol I distribution revealed the presence of “pileups,” which were defined as arrays of five or more polymerases with no visible intervening DNA [examples are bracketed in Fig. 6A]. We attribute these to transcriptional pausing or stalling of the leading polymerase (situated at the right end of a bracket in Fig. 6A; see Supplemental Fig. S6A for schematic), hindering the progression of subsequent polymerases. In wild-type cells, analysis of all visualized, active rDNA genes [data not shown] revealed pileups on 37% of genes [the number of genes analyzed, N = 105], showing that transient transcriptional pausing is surprisingly frequent. The fraction of genes showing pileups increased to 55% in the P_{CAG-TOP1} single mutant [N = 180], and to 81% in the triple mutant [N = 209].

To determine whether polymerases were preferentially paused at particular sites, we divided the 35S gene into 20 equal segments and analyzed all genes that could be unambiguously traced from the 5′ to 3′ ends. We mapped the position of the leading polymerase along a schematic 35S gene, in all pileups [five or more polymerases] [Fig. 6B] or in only longer pileups, which presumably reflect more persistent pauses [20 or more polymerases] [Supplemental Fig. S6B]. Notably, the distribution of inferred pause sites along the rDNA was similar in all strains [Fig. 6B; Supplemental Fig. S6B], indicating that pausing is a normal feature of Pol I transcription, which is exacerbated in the mutant strains. In wild type, the major pause sites fell within the 18S rDNA, whereas lower levels of pausing were observed at sites in the 25S rDNA [Fig. 6B, blue]. As Top1 was depleted, pausing strongly increased along the 18S rDNA [Fig. 6B], and this was exacerbated when RNase H was also limiting [Fig. 6B, green]. The increase in pausing was more prominent in the 5′ region of the rDNA when both Top1 and RNase H were deficient, with stalled polymerases at the 5′ region of 18S backed up into the 5′-ETS. The proportion of 35S genes with pileups increased from wild-type to single- to triple-mutant strains [Fig. 6D]. Both mutants showed similar numbers of polymerases per pileup, but this was greater than the wild type [Fig. 6E].
Figure 6. Polymerase pileups over the 5’-ETS and 18S increase when Top1 and RNase H1 and H201 are absent. (A) Representative rRNA genes from Miller spreads of wild-type, \( P_{GAL-TOP1} \), and \( P_{GAL-TOP1\,mh1\Delta\,mh201\Delta} \) strains after 6 h of Top1 depletion in glucose at 30°C. Brackets indicate sites of polymerase pileups. These were defined as at least five tightly packed polymerases. The leading polymerase corresponds to the polymerase situated at the right end of a bracket [Supplemental Fig. S6A]. (B) Sites of Pol I pausing across the 35S rDNA gene. The gene was divided into 20 equal segments (~337 bp each), and the position of the leading polymerase in each pileup was plotted onto the segment in which it occurred. The Y-axis shows the percentage of all rDNA genes for each strain with a pileup starting at the indicated position along the gene [X-axis]. All rDNA genes that could be visualized from the 5’ to 3’ ends were included in the analysis, and their lengths were normalized. (C) EM analysis of Pol I occupancy over the rDNA unit. For each of the three strains, polymerase positions were measured along 77 rDNA genes, yielding the position of 15,115 polymerases. Each gene was divided into 20 equal segments, and the number of polymerases in each segment was determined. Data were plotted using the midpoints of the 20 gene segments for positioning on the X-axis using smoothed lines. (D) Total frequencies of pileup occurrence for wild-type and mutant strains. (E) Plot of pileup lengths [number of polymerases per pileup] for wild-type and mutant strains. The same sample of genes was used in B.

Figure 6B shows the locations of the leading polymerase in each pileup on active rDNA genes, but we also quantified the distribution of all polymerases [Fig. 6C]. Pol I occupancy in the wild type was predominant over the 5’-ETS and the 5’ region of 18S [Fig. 6C, blue], consistent with Pol I ChIP data [cf. blue graphs in Figs. 5A and 6C]. In both mutants, Pol I occupancy increased from the promoter to approximately +4000 nt downstream [Fig. 6C, red and green], with more polymerases across the 5’-ETS and 5’ region of 18S in the triple mutant. The increase in pileup frequency and Pol I occupancy [Fig. 6B,C], together with the increased accumulation of RNA/DNA hybrids [Fig. 5B], across the 5’-ETS and 5’ region of the 18S in the triple mutant suggested that stable R-loops exacerbate Pol I pausing in these regions. Persistent polymerase pileups are predicted to impose transcriptional blocks that would lead to reduced rRNA synthesis [Tous and Aguilera 2007; Klumpp and Hwa 2008], consistent with RNA analyses in the triple mutant [Fig. 4].

Gaps [DNA free of Pol I] were generally observed in front of pileups, but there was no clear correlation between the length of the pileup and gap length [Supplemental Fig. S7]. This may simply reflect the stochastic nature of transcription initiation, but might also indicate that paused polymerases impact on the elongation of neighboring polymerases. However, a complication is that the observed pileups presumably represent a mix of those in the process of formation, in which new polymerases are joining, and those in the process of resolution, in which previously stalled polymerases are moving away from the pause site.

In EM analyses, the Pol I distribution observed in spreads of wild-type cells was in good agreement with the occupancy inferred from the ChIP data. The match was less close for \( P_{GAL-TOP1} \) and \( P_{GAL-TOP1\,mh1\Delta} \)
R-loops associated with stalled polymerases were not visible in the chromatin spreads, suggesting that they may be limited in size by the subsequent polymerases. It is, however, unclear how R-loops would appear in chromatin spreads, or whether regions of rRNA genes harboring R-loops would be amenable to dispersal, since they may be subject to strong topological constraints.

We conclude that RNA Pol I frequently pauses or stalls during rDNA transcription even in wild-type strains, in particular over the 18S region. This could be caused by the accumulation of positive supercoils in front of the polymerase and/or R-loop formation in its wake. Pausing was greatly increased by loss of Top1, and was augmented further when combined with reduced RNase H activity.

Discussion

We report that pre-rRNA transcription is affected by the absence of Top1, particularly over the 5′ region of 18S rDNA. Pre-rRNA fragments that accumulated in the absence of Top1 were consistent with cleavage of nascent transcripts. The truncated pre-rRNA fragments had predicted 5′ ends at the transcription initiation site or at cotranscriptional cleavage site A2 within ITS1 [Fig. 1A]. The most abundant fragments extended from the initiation site into the 5′ region of the 18S rRNA. Topoisomerase activity is essential for transcription by Pol I, since rRNA synthesis was abolished in top2-ts top1Δ mutants at 37°C (Brill et al. 1987; Schultz et al. 1992). We showed that, in strains lacking both Top1 and Top2, transcription blockage occurs predominantly within the 18S 5′ region [Fig. 3; Supplemental Fig. S4], again indicating the stringent requirement for topoisomerase activity in this region.

Negative torsion behind transcribing Pol I can lead to unpairing of the DNA helix followed by hybridization of the nascent pre-rRNA to the template strand, forming an R-loop (Drolet 2006; Aguilera and Gomez-Gonzalez 2008). R-loops are targets for RNase H, suggesting that this activity might generate pre-rRNA truncations in the absence of Top1 [see model in Fig. 7]. Consistent with this, decreased RNase H activity reduced pre-rRNA truncation in strains lacking Top1 [Fig. 4]. rRNA synthesis was also reduced in these strains, supporting the conclusion from in vitro data (Tous and Aguilera 2007) that stable R-loops block transcription elongation. Pre-rRNA accumulation was not abolished completely in the absence of Top1 and RNase H1 and H201. This might reflect the residual RNase H activity in these strains.

Figure 7. Model for the role of R-loops in blocking pre-rRNA transcription. [A] Polymerase movement during transcription forces the rDNA to rotate, building up positive torsion (+) in front and negative torsion (−) in its rear (Cook 1999). Torsion ahead of Pol I causes positive supercoiling, which resists strand opening. This slows or pauses transcription elongation, generating polymerase pileups, in particular over the 5′ region of the 18S rDNA. Torsion behind the polymerase leads to negative supercoiling, which facilitates DNA strand opening and stimulates formation of R-loops with the nascent pre-rRNA (Roy et al. 2010). These structures also slow transcription elongation (Tous and Aguilera 2007) and trigger pileup formation. In wild-type strains, pileups are normally transient, with Top1 resolving both negative and positive supercoiling and facilitating transcription. rDNA rotation and direction of transcription are depicted by a bent arrow and a straight arrow, respectively. [B] In strains lacking Top1, which provides the major topoisomerase activity during rDNA transcription, more torsion is accumulated (− − − and ++++) and R-loops occur more frequently, leading to an increase in pileup formation. RNase H1 and H2 cleave the RNA–DNA hybrids, releasing truncated pre-rRNA fragments that are targeted and degraded by the TRAMP and exosome complexes. Top2 resolves positive and negative supercoiling. Both activities should lead to the release of transcriptional blocks, but Top2 is not predicted to resolve strand separation induced by negative torsion (Lavelle 2008; SL French, ML Sikes, RD Hontz, YN Osheim, TE Lambert, A El Hage, MM Smith, D Tollervey, JS Smith, and AL Beyer, in prep.). (C) In the absence of both Top1 and RNase H1 and H201, persistent R-loops block rotation of the rDNA and cause severe polymerase arrests and pileups.
Pol I transcription termination (Kawauchi et al. 2008).

RNA–DNA helicases like Sen1, which plays a role in
Schneider et al. 2009), or dissociation of the duplex by
endonuclease activity of the exosome sub-
unit Rrp44 [Lebreton et al. 2008; Schaeffer et al. 2009,
Schneider et al. 2009], or dissociation of the duplex by
RNA–DNA helicases like Sen1, which plays a role in
Pol I transcription termination [Kawauchi et al. 2008].

Truncated pre-rRNA fragments were stabilized in
top1Δ strains defective in TRAMP [trf4Δ] or the exosome
{rtp6Δ}, indicating that these complexes degrade the 5′
pre-rRNA fragments released by RNase H cleavage. The
fate of the 3′ fragments resulting from RNase H cleavage
is unclear, but rapid degradation by the nuclear 5′-to-3′
exonuclease Rat1 would be predicted. Rat1 functions in
transcription termination at the 3′ end of the rDNA [El
Hage et al. 2008, Kawauchi et al. 2008], and might also
play a role in the release of stalled polymerases associated
with transcripts that are engaged in R-loops. Consistent
with this model, Rat1 has been shown to function in the
termination of RNA Pol II molecules engaged in the
production of uncapped mRNA [Jimeno-Gonzalez et al.
2010].

Strains carrying rpa49Δ and rpa34Δ are defective in Pol
I elongation [Kuhn et al. 2007], and also accumulated pre-
rRNA fragments that terminated in the 5′ region of the
18S rDNA when combined with the exosome mutation
rtp6Δ [data not shown], as seen previously in
rpa135(D7784C) rtp6Δ double mutants [Schneider et al.
2007]. Thus, transcription through the 5′ region of the
18S rDNA appears to pose a particular challenge to
elongation-compromised Pol I. Moreover, pre-rRNA frag-
ments with 3′ ends that extended to the same region were
detected in TRAMP and exosome mutants with intact
topoisomerase and RNA polymerase activity, indicating
that some level of pre-rRNA truncation normally occurs
here in wild-type strains.

In EM analyses, pileups of five or more polymerases
were seen on 37% of active wild-type rDNA transcription
units, principally over the 18S rDNA. We interpret these
as resulting from the collision of multiple transcribing
polymerases with a single stalled or paused Pol I. In
strains lacking Top1, the proportion of rDNA units with
stalled Pol I was increased to 53%, and this rose to 84% in
strains lacking both Top1 and RNase H. Notably, the
distribution of the leading stalled polymerase was not
clearly altered in the different strains, suggesting that the
loss of these activities exacerbates an underlying pause
that is present in the wild type. In the absence of Top1,
both the frequency of pileups and the numbers of contiguous
polymerases increased, presumably reflecting in-
creased duration of stalling of the leading polymerase. In
strains lacking both Top1 and RNase H activity, pileup
frequency increased further. The presence of persistent
R-loops might slow down local rotation of the rDNA,
reducing the speed of elongation of Pol I in the same
pileup, which would increase the residency times of
pileups and impede their resolution [see model in Fig.
7]. The density of R-loops was increased dramatically in
strains lacking both Top1 and RNase H activity, partic-
ularly over the promoter and 5′-ETS region [Fig. 5]. This
may reflect both the increased time available for R-loop
formation due to the stalled polymerases, and R-loop
stabilization due to reduced RNase H activity.

The similarities in the distribution of stalled Pol I,
R-loops, and pre-rRNA cleavage sites suggest that un-
derlying features of transcription in the 5′ region of the
18S rDNA are distinct from other segments of the 6.7-kb
pre-rRNA transcript. This region lies ~1 kb, or ~100
turns of the rDNA, from the start of transcription, and it
is feasible that, simply due to physical features of tran-
scription, Top1 activity first becomes crucial for Pol I
transcription over this region. However, the existence of
a checkpoint for ribosome assembly, which would slow
Pol I elongation until the pre-rRNA is folded and/or
bound by r-proteins and pre-40S assembly factors, was
suggested previously [Moss et al. 2007; Schneider et al.
2007]. In yeast, the U3 snoRNA binds to the 5′-ETS and to
the extreme 5′ end of the 18S rRNA sequence, where it
may play a major role in the mechanism or timing of
formation of the central pseudoknot in 18S rRNA [for
review, see Henras et al. 2008]. It could be envisaged that
binding and/or release of U3 and other early-acting
ribosome synthesis factors is synchronized with the
transcription elongation rate to promote productive pre-
rRNA folding and reduce the tendency of exposed, un-
folded RNA sequences to generate R-loops. Such surveil-
ance of preribosome assembly might occur preferentially
at a location where Pol I is slowed by physical processes.

In the absence of Top1, levels of Pol II ncRNA tran-
scripts derived from the rDNA IGS regions and rDNA
recombination are both increased relative to wild-type
strains [Bryk et al. 1997; Houseley et al. 2007]. We spec-
ulate that underwound DNA in the IGS regions dis-
rupts transcriptional silencing and forms R-loops with
cncRNAs, which might be poorly packaged. This leaves
the complementary strand single-stranded and prone
to cleavage and recombination [Aguilera and Gomez-
Gonzalez 2008]. In strains lacking Top1 and RNase H,
rRNA–DNA hybrids accumulated over IGS1 and IGS2,
and ncRNA levels were increased relative to top1Δ single
mutants, suggesting that RNase H-mediated cleavage
is a major pathway of ncRNA degradation in the top1Δ
strains.

In yeast, cleavage of the nascent transcripts generated
by RNA Pol II is essential to avoid transcriptional
roadblocks and for cell viability [Sigurdsson et al. 2010].
Similarly, cleavage of the pre-rRNA in top1Δ and Pol I
elongation mutants might be essential to avoid transcrip-
tional blocks, in particular over the 5′ region of 18S.
Persistent accumulation of RNA polymerase pileups
along the rDNA was also predicted to reduce rRNA
transcription in modeling data from bacteria [Klumpp
and Hwa 2008]. R-loop formation also occurs in Top1
mutants in Escherichia coli, and leads to growth defects,
impaired transcription elongation on the rDNA, and
extensive RNA degradation by RNase H [Hraikey et al.
2000; Drolet 2006; Baaklini et al. 2008]. This supports
the conclusion [Li and Manley 2006] that the deleterious
effects that R-loops can have on transcription have been
conserved over a long evolutionary distance.
Materials and methods

Strains, plasmids, and growth conditions
All mutants and tagged yeast strains were constructed using strain BY4741 (derived from S288C). The strains and plasmids used in this study are listed in Supplemental Table S1. Regulated $P_{\text{GAL}}$ expression and deletions were constructed by one-step PCR using template plasmids pFA6a-kanMX6-PGAL1-3HA and pFA6a-kanMX6/NatMX6/HphMX6, respectively. Growth and handling of S. cerevisiae were by standard techniques. Cells were grown in YPD medium at 18°C, 25°C, or 30°C. For RNA and ChIP analyses, strains where TOP1 or TOP2 was replaced under $P_{\text{GAL1}}$ promoter were grown at 30°C to OD$_{600}$ = 0.3–0.4 in complete synthetic medium containing 2% galactose and 2% sucrose, then transferred to the same prewarmed medium containing 2% galactose. For EM analysis, wild-type, $P_{\text{GAL1}}$:TOP1, and $P_{\text{GAL1}}$:TOP1 rnh1A rnh201A strains were grown at 30°C in YP-Gal, then shifted to YPD for 6 h.

RNA analyses

RNA extraction, Northern hybridization, and quantifications were as described (Supplemental Material; El Hage et al. 2008). Poly(A) RNA extraction was as described (Supplemental Material; Dez et al. 2006). Primers for RNA analysis are listed in Supplemental Table S2.

ChIP

ChIP experiments were performed essentially as described (Supplemental Material; El Hage et al. 2008). RNA–DNA hybrids were detected using Ab S9.6, kindly provided by Dr. S. Leppla, or purified by Eurogentec from supernatants of a mouse hybridoma cell line purchased from American Type Culture Collection. Pol I subunit Rpa190 was detected with rabbit polyclonal antibody (kind gift from David Schneider). Pol I subunit Rpa34-13MYC was detected using mouse monoclonal antibody e-Myc 9E11 (Santa Cruz Biotechnology). Primers used for quantitative PCRs are listed in Supplemental Table S3.

For detection of both RNA–DNA hybrids and Rpa190 in Figure 5, sonicated DNA from the same strain was split into three equal parts that served for no antibody control, S9.6, and anti-Rpa190. Values for ChIPs were determined using the formula $Ct = 2^{\frac{1}{dCT}}$, where $dCT$ is the cycle number for immunoprecipitate, and “Ct Background” is the cycle number for control without antibody. To normalize Pol I ChIP values for Figure 5, A and D, each set of Rpa190–ΔCt values was normalized to 2SS = average of ΔCt values of primers j and k in the same data set. The means of three to five independent experiments (replicates) were calculated with standard errors. This shows the relative distribution of Pol I along the length of the rDNA in each strain. However, because the values are normalized internally, differences in overall Pol I ChIP signals between different strains will be lost. To assess this, we divided the average of the Rpa190–ΔCt values from each mutant to the average of the Rpa190–ΔCt values from the wild type. The normalized ChIP values obtained for each data set for the mutants were then multiplied by this correction factor. A different correction was applied in Figure 5, B and E, to account for differences in ChIP efficiencies and Pol I loading. The ΔCt values for RNA/DNA hybrids were each normalized to the average of Rpa190–ΔCts within the same strain and replicate. The means of three to five independent experiments were calculated with standard errors.

EM

Miller chromatin spreads and EM analyses were performed as described (Supplemental Material; Osheim et al. 2009).

Acknowledgments

We thank Steven Leplla, David Schneider, Andrés Aguilera, and Brian Luke for providing reagents. We thank Jean Beggs, Steve Innocente, Laura Milligan, Olivier Cordin, and HannahMischofor fruitful discussions; Lea Harrington and Laura Guardano for helping with the production of S9.6 antibodies; and Alexandra Moreira and Steve West for critical reading of the manuscript. This work was supported by the Wellcome Trust and by U.S. Public Health Service grant GM-63952 to A.L.B.

References


