



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

## Rapid homeostatic downregulation of LTP by extrasynaptic GluN2B receptors

### Citation for published version:

Delgado, JY, Fink, AE, Grant, SGN, O'Dell, TJ & Opazo, P 2018, 'Rapid homeostatic downregulation of LTP by extrasynaptic GluN2B receptors' *Journal of Neurophysiology*. DOI: 10.1152/jn.00421.2018

### Digital Object Identifier (DOI):

[10.1152/jn.00421.2018](https://doi.org/10.1152/jn.00421.2018)

### Link:

[Link to publication record in Edinburgh Research Explorer](#)

### Document Version:

Publisher's PDF, also known as Version of record

### Published In:

*Journal of Neurophysiology*

### General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

### Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact [openaccess@ed.ac.uk](mailto:openaccess@ed.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.



## RAPID REPORT | Cellular and Molecular Properties of Neurons

# Rapid homeostatic downregulation of LTP by extrasynaptic GluN2B receptors

Jary Y. Delgado,<sup>1</sup>  Ann E. Fink,<sup>1</sup> Seth G. N. Grant,<sup>2</sup> Thomas J. O'Dell,<sup>1</sup> and Patricio Opazo<sup>1</sup>

<sup>1</sup>Department of Physiology, University of California, Los Angeles, Los Angeles, California; and <sup>2</sup>Genes to Cognition Programme, Centre for Clinical Brain Science, Edinburgh University, Edinburgh, United Kingdom

Submitted 26 June 2018; accepted in final form 14 August 2018

**Delgado JY, Fink AE, Grant SG, O'Dell TJ, Opazo P.** Rapid homeostatic downregulation of LTP by extrasynaptic GluN2B receptors. *J Neurophysiol* 120: 2351–2357, 2018. First published August 15, 2018; doi:10.1152/jn.00421.2018.—Although the activation of extrasynaptic GluN2B-containing *N*-methyl-D-aspartate (NMDA) receptors has been implicated in neurodegenerative diseases, such as Alzheimer's and Huntington's disease, their physiological function remains unknown. In this study, we found that extrasynaptic GluN2B receptors play a homeostatic role by antagonizing long-term potentiation (LTP) induction under conditions of prolonged synaptic stimulation. In particular, we have previously found that brief theta-pulse stimulation (5 Hz for 30 s) triggers robust LTP, whereas longer stimulation times (5 Hz for 3 min) have no effect on basal synaptic transmission in the hippocampal CA1 region. Here, we show that prolonged stimulation blocked LTP by activating extrasynaptic GluN2B receptors via glutamate spillover. In addition, we found that this homeostatic mechanism was absent in slices from the SAP102 knockout, providing evidence for a functional coupling between extrasynaptic GluN2B and the SAP102 scaffold protein. In conclusion, we uncovered a rapid homeostatic mechanism that antagonizes LTP induction via the activation of extrasynaptic GluN2B-containing NMDA receptors.

**NEW & NOTEWORTHY** Although long-term potentiation (LTP) is an attractive model for memory storage, it tends to destabilize neuronal circuits because it drives synapses toward a maximum value. Unless opposed by homeostatic mechanisms operating through negative feedback rules, cumulative LTP could render synapses unable to encode additional information. In this study, we uncovered a rapid homeostatic mechanism that antagonizes LTP induction under conditions of prolonged synaptic stimulation via the activation of an extrasynaptic GluN2B-SAP102 complex.

extrasynaptic; GluN2B; homeostasis; SAP102; scaling

### INTRODUCTION

Extrasynaptic GluN2B-containing *N*-methyl-D-aspartate (NMDA) receptors have been implicated in neurodegenerative diseases, such as Alzheimer's and Huntington's disease via the activation of proapoptotic signaling cascades (Hardingham and Bading 2010; Li et al. 2011). Although consistently involved in disease, the physiological function of extrasynaptic GluN2B receptors remains unknown.

In general, GluN2B-containing NMDA receptors have been implicated in various, and sometimes opposite, forms of syn-

aptic plasticity. For example, GluN2B receptors have been implicated in the induction of long-term potentiation (LTP) and long-term depression (LTD) (Barria and Malinow 2005; Liu et al. 2004). This paradox has raised the question as to how GluN2B receptors can couple to divergent signaling pathways. Because GluN2B can interact with multiple members of the MAGUK family of scaffolding proteins, including PSD95, PSD93, and SAP102 (Grant and O'Dell 2001; Kim and Sheng 2004), and even form distinct supramolecular complexes depending on the bound MAGUK member (GluN2B receptors form 1.5-MDa supercomplexes with PSD95 and 350-kDa complexes with SAP102) (Frank and Grant 2017; Frank et al. 2016), it is plausible that different GluN2B-MAGUK supercomplexes could potentially couple to specific signaling pathways. The complexity of the GluN2B-MAGUK signaling is further increased by the preferential localization of different MAGUKs to synaptic or extrasynaptic sites. For instance, PSD95 is synaptically located at discrete nanodomains (MacGillavry et al. 2013; Nair et al. 2013), whereas SAP102 is preferentially located at extrasynaptic sites (Chen et al. 2012; Zheng et al. 2011).

Although the physiological function of extrasynaptic receptors remain unknown, it is interesting to note that their activation, not only antagonizes, but also dominates over synaptic signaling (Hardingham et al. 2002). For instance, although synaptic NMDA receptors promote ERK and cAMP response element-binding protein (CREB) phosphorylation, extrasynaptic NMDA receptors promote an ERK and CREB dephosphorylation pathway (Hardingham et al. 2002). In this context, it is possible that extrasynaptic NMDA receptors homeostatically prevent the overactivation of synaptic signaling in conditions of strong synaptic stimulation that lead to glutamate spillover.

We have previously observed that LTP in the hippocampal CA1 region can be induced by brief period of theta-pulse stimulation (TPS) (5 Hz for 15–30 s). Surprisingly, prolonging the time of stimulation (5 Hz for 1–3 min) results in no significant potentiation of synaptic transmission relative to baseline recordings (Thomas et al. 1996). In this study, we explore whether extrasynaptic GluN2B receptors might homeostatically antagonize the signaling pathways underlying the induction of LTP during periods of prolonged synaptic stimulation as previously hypothesized (Rusakov et al. 2004). Here, we show that prolonged synaptic stimulation promotes the spillover of glutamate to extrasynaptic sites, the activation of a GluN2B-SAP102 functional complex, and the homeostatic downregulation of LTP.

Address for reprint requests and other correspondence: P. Opazo, Clem Jones Centre for Ageing Dementia Research, Queensland Brain Inst., Univ. of Queensland, St. Lucia, Queensland, Australia (e-mail: p.opazo@uq.edu.au).

## MATERIAL AND METHODS

Standard techniques were used to prepare slices (400  $\mu\text{m}$  thick) from the hippocampus of halothane-anesthetized 5- to 8-wk-old male C57BL/6 mice or from 4- to 7-mo-old male SAP102 knockout (KO) mice and their wild-type (WT) littermates (MF1 strain) as before (Cuthbert et al. 2007). All techniques were approved by the UCLA Institutional Animal Care and Use Committee and done in accordance with UK Home Office Guidelines.

Slices were maintained in interface-type chambers (Fine Science Tools, Foster City, CA) that were continuously perfused (at 2–3 ml/min) with a warm (30°C–31°C), oxygenated (95%  $\text{O}_2$ -5%  $\text{CO}_2$ ) artificial cerebrospinal fluid (aCSF) containing the following: 124.0 mM NaCl, 4.4 mM KCl, 25.0 mM  $\text{Na}_2\text{HCO}_3$ , 1.0 mM  $\text{NaH}_2\text{PO}_4$ , 1.2 mM  $\text{MgSO}_4$ , 2.0 mM  $\text{CaCl}_2$ , and 10.0 mM glucose. In our initial experiments, field excitatory postsynaptic potentials (fEPSPs) were recorded in slices maintained in an interface-type recording chamber, whereas in later experiments extracellular recordings were done using slices completely submerged in aCSF. The type of recording configuration, which is noted in the figure legends, had no apparent effect on our observations.

Standard extracellular recording techniques were used to record fEPSPs in the CA1 region of hippocampal slices (Babiec et al. 2017). Presynaptic fiber stimulation pulses were delivered once every 50 s (0.02 Hz, 0.01- to 0.02-ms-duration pulses using tungsten wire bipolar stimulation electrodes placed in stratum radiatum of the CA1 region) using a stimulation strength that evoked fEPSPs that were approximately half of the maximal fEPSP amplitude that could be evoked by strong-intensity stimulation. LTP was induced using either short-duration trains of TPS (150 pulses of presynaptic stimulation delivered at 5 Hz) or long-duration trains of TPS (900 pulses) either in the absence (controls) or presence of inhibitors in a randomized manner to minimize bias when allocating slices to treatment. LTD and depotentiation were induced using a classical protocol at 1-Hz stimulation for 15 min (900 pulses). The average of fEPSP slopes recorded between 40 and 45 min after TPS was used for statistical comparisons. In all experiments, individual slices were considered the experimental and analytical units ( $n$ ). The number of animals used in each experiment is stated in the corresponding figure legend (e.g.,  $n = 6$  from 4 animals).

Statistical significance of results was assessed using ANOVAs (followed by Bonferroni  $t$ -tests) or paired and unpaired  $t$ -tests (2-tailed). When used in reporting statistical results, asterisks denote the following significance level: \*\*\* $P < 0.001$ . Nonstatistical significance ( $P > 0.05$ ) is denoted by ns.

## RESULTS

We have previously found that the duration of TPS (5 Hz) has an important impact on the direction of synaptic plasticity (Thomas et al. 1996). In particular, brief trains of stimulation trigger LTP (5 Hz for 30 s), whereas longer stimulation times (5 Hz for 3 min) have no effect on basal synaptic transmission. Because prolonged stimulation might promote spillover to extrasynaptic sites, we reasoned that 5 Hz for 3 min might recruit a population of extrasynaptic receptors that antagonize the induction of LTP. Because NMDA receptors containing the GluN2B are enriched at extrasynaptic sites (Stocca and Vicini 1998; Tovar and Westbrook 1999), it is possible that they are recruited upon prolonged stimulation. To test this hypothesis, we stimulated at 5 Hz for 3 min in the absence and presence of 5  $\mu\text{M}$  ifenprodil, a specific GluN2B inhibitor (Williams 1993). As previously shown (Thomas et al. 1996), we found that 3 min of 5-Hz stimulation has little lasting effect on synaptic strength (Fig. 1, *A* and *B*, fEPSPs were  $115 \pm 4\%$  of baseline

45 min after 5-Hz stimulation,  $n = 14$ ). However, when the same stimulation protocol was applied in the presence of ifenprodil (5  $\mu\text{M}$ ), a robust LTP was revealed (Fig. 1, *A* and *B*, fEPSPs were  $182 \pm 16\%$  of baseline 45 min after 5-Hz stimulation,  $n = 9$ ,  $P < 0.001$ ). Another group of receptors enriched at peri-/extrasynaptic sites consists of metabotropic glutamate receptors (mGluR) (Baude et al. 1993). To examine whether mGluR activation might also antagonize LTP during prolonged stimulation, we stimulated at 5 Hz for 3 min in the presence of the mGluR antagonist LY341495 (10  $\mu\text{M}$ ). As shown in Fig. 1, *B* and *C*, LY341495 had no effect on the outcome of 5-Hz 3-min stimulation (fEPSPs were  $113.0 \pm 11.0\%$  and  $115.1 \pm 13.0\%$  of baseline values at 45 min after 5-Hz stimulation in the absence or presence of the mGluR antagonist, respectively,  $P = 0.90$ ). These findings suggest that the activation of extrasynaptic GluN2B receptors but not mGluRs antagonizes the induction of LTP under prolonged theta-frequency stimulation. Although controversial, it has previously been shown that activation of GluN2B receptors can trigger LTD under certain conditions (Liu et al. 2004; but see Morishita et al. 2007). It is thus possible that 5 Hz for 3 min might trigger LTD through GluN2B receptors and thus mask the expression of LTP. To explore this alternative, we tested the contribution of GluN2B receptor signaling to a classical LTD protocol (1 Hz for 15 min). As shown in Fig. 1*D*, ifenprodil 5  $\mu\text{M}$  had no effect on the induction of LTD, suggesting that extrasynaptic GluN2B seems to antagonize LTP rather than simply mask LTP via LTD induction (fEPSPs were 53.1% and 46.7% of baseline values at 60 min after LTD induction in the absence or presence of the ifenprodil 5  $\mu\text{M}$ , respectively;  $P = 0.73$ ). As for LTD and consistent with previous studies (Massey et al. 2004), we found that GluN2B receptors do not mediate depotentiation of synaptic transmission (Fig. 1*E*; 1 Hz for 15 min delivered 2 min after a high-frequency stimulation LTP protocol, 2 times 100 Hz for 1 s; fEPSPs were 109.5% and 108.3% of baseline values at 60 min after depotentiation in the absence or presence of the ifenprodil 5  $\mu\text{M}$ , respectively;  $P = 0.9$ ).

We next examined whether brief stimulation at 5 Hz, which we have previously shown to induce robust LTP, might be sufficient to activate the extrasynaptic pool of GluN2B receptors. We therefore stimulated slices at 5 Hz for 30 s in the presence of ifenprodil (5  $\mu\text{M}$ ). As shown in Fig. 2, *A* and *D*, we found that 5 Hz for 30 s induces LTP of similar magnitude in the absence or presence of ifenprodil (fEPSPs were  $143.7 \pm 6.0\%$  and  $146.7 \pm 11.0\%$  of baseline 45 min after 5-Hz stimulation with or without ifenprodil, respectively,  $P = 0.930$ ). This indicates that extrasynaptic GluN2B populations are preferentially recruited after prolonged period of stimulation and that 30 s of 5-Hz stimulation does not induce sufficient spillover to activate these receptors in a way that causes synaptic plasticity. Because glutamate uptake is the main mechanism to prevent spillover (Huang and Bergles 2004), we examined whether blocking glutamate transporters might facilitate the activation of GluN2B receptors even during 5 Hz for 30 s, as it has been previously observed with 100-Hz stimulation (Scimemi et al. 2009; Varga et al. 2015). To test this hypothesis, we stimulated at 5 Hz for 30 s in the presence and absence of the general glutamate transporter inhibitor DL-threo-beta-benzyloxyaspartic acid (TBOA). As shown in Fig. 2, *B* and *D*, LTP induced by stimulation at 5 Hz for 30 s was significantly

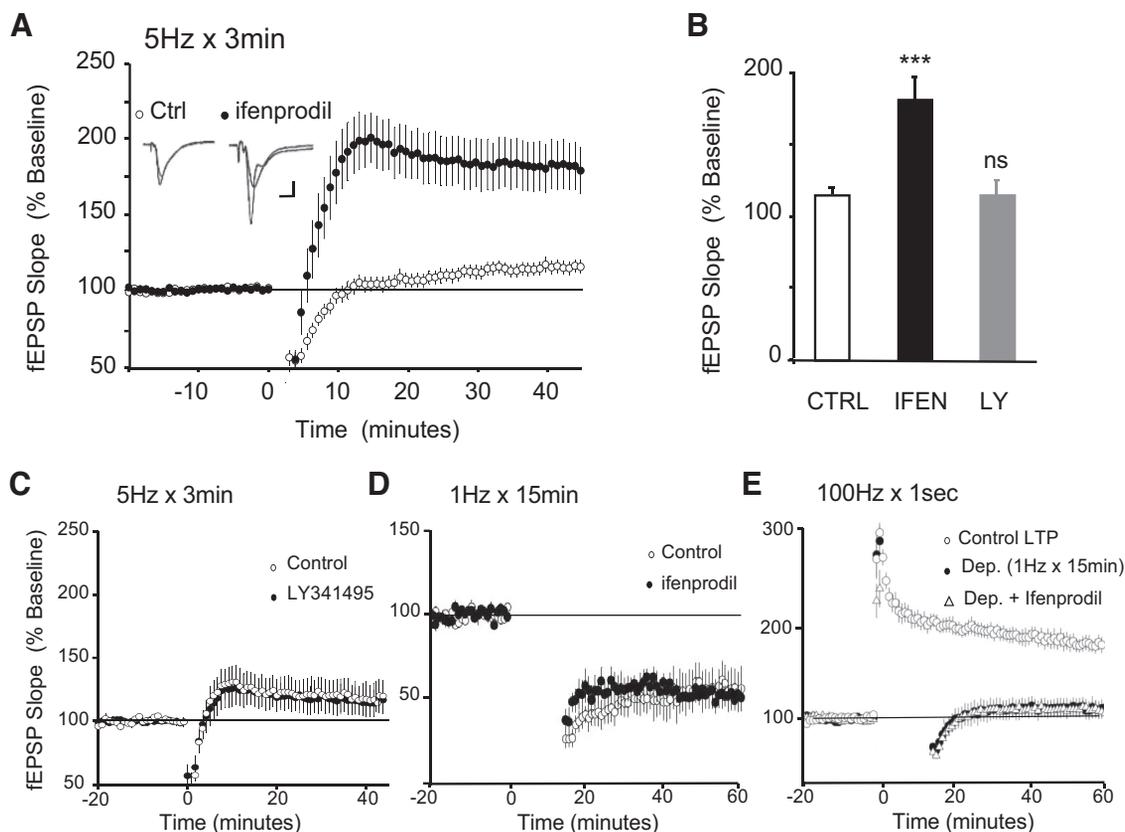


Fig. 1. Extrasyaptic GluN2B-containing *N*-methyl-D-aspartate (NMDA) receptors homeostatically antagonize long-term potentiation (LTP). **A**: in control experiments, 3 min of 5-Hz stimulation (delivered at time = 0,  $\circ$ ,  $n = 14$  from 12 animals) produced a small enhancement of synaptic transmission [field excitatory postsynaptic potentials (fEPSPs) were  $115 \pm 4\%$  of baseline 45 min after 5-Hz stimulation]. However, 5 Hz for 3 min delivered in the presence of the GluN2B blocker ifenprodil ( $5 \mu\text{M}$ ) induced robust LTP ( $\bullet$ , fEPSPs were  $182 \pm 16\%$  of baseline 45 min after 5-Hz stimulation,  $n = 9$  from 8 animals,  $P < 0.001$  compared with controls). *Inset*: fEPSPs recorded during baseline and 45 min after 5-Hz stimulation in control (*left*) and ifenprodil (*right*). Calibration: 2 mV, 2.5 ms. **B**: summary of the data presented in **A** and **C**. Bar graph indicating the mean fEPSP averaged from the last 5 min of experiment in each condition [ $***P < 0.001$  and nonstatistical significance (ns),  $P > 0.05$  compared with untreated controls]. IFEN, ifenprodil; LY, LY341495. **C**: extrasynaptic metabotropic glutamate receptors do not contribute to the homeostatic pathway. A 10-min bath application of  $10 \mu\text{M}$  mGluR antagonist LY341495 does not rescue LTP. fEPSPs were  $113.0\%$  of baseline for control experiments ( $n = 6$  from 5 animals) and  $115.1\%$  of baseline for the experiment carried out in the presence of LY341495 ( $n = 5$  from 4 animals);  $P = 0.9$ . **D**: GluN2B-containing NMDA receptors are not required for long-term depression (LTD). fEPSPs were  $53.1\%$  ( $n = 6$  from 5 animals) and  $46.7\%$  ( $n = 5$  from 4 animals) of baseline values at 60 min after LTD induction in the absence or presence of the ifenprodil  $5 \mu\text{M}$ , respectively;  $P = 0.73$ . **E**: GluN2B-containing NMDA receptors are not required for depotentiation (1 Hz for 15 min delivered 2 min after a high-frequency LTP protocol; 2 times 100 Hz for 1 s). fEPSPs were  $109.5\%$  and  $108.3\%$  of baseline values at 60 min after depotentiation in the absence ( $n = 6$  from 4 animals) or presence ( $n = 6$  from 4 animals) of the ifenprodil  $5 \mu\text{M}$ , respectively;  $P = 0.9$ .

impaired in the presence of TBOA  $10 \mu\text{M}$  (fEPSPs were  $166.5 \pm 11.0\%$  of baseline 45 min after 5-Hz stimulation in the absence and  $119.2 \pm 5.0\%$  in the presence of TBOA,  $P < 0.001$ ). To confirm that LTP impairment was due to activation of GluN2B-containing NMDA receptors, we applied TBOA in the presence of the GluN2B blocker ifenprodil  $5 \mu\text{M}$ . As shown in Fig. 2, **C** and **D**, ifenprodil completely rescued the LTP impairment caused by TBOA  $10 \mu\text{M}$  (fEPSPs were  $160.4 \pm 5.0\%$  and  $161.3 \pm 6\%$  of baseline at 45 min after 5-Hz stimulation for control and in the presence of TBOA and ifenprodil, respectively,  $P = 0.91$ ). Taken together, these findings support the hypothesis that extrasynaptic GluN2B receptors antagonize LTP in conditions that promote glutamate spillover, such as prolonged stimulation or glutamate transporter block.

The involvement of GluN2B receptors in various, and sometimes opposite, forms of synaptic plasticity raises the question of how GluN2B can activate divergent signaling cascades. Numerous studies have stressed the central role of MAGUK family scaffolding proteins in coupling NMDA receptors to

specific signaling pathways (Grant and O'Dell 2001; Kim and Sheng 2004). Because GluN2B preferentially interacts with the extrasynaptically enriched SAP102 scaffolding protein (Chen et al. 2011; Murata and Constantine-Paton 2013; Sans et al. 2000; Zhang and Diamond 2009), it is likely that a GluN2B-SAP102 complex mediates signaling at extrasynaptic sites. To test this hypothesis, we stimulated hippocampal slices obtained from SAP102 KO mice with short and long trains of 5-Hz stimulation. Given the absence of SAP102, we hypothesized that activation of extrasynaptic GluN2B might be uncoupled from the signaling cascades that antagonize LTP. Consistent with this idea and our previous work (Cuthbert et al. 2007), we found that prolonged stimulation (5 Hz for 3 min) triggered an LTP as robust as that induced with brief stimulation (5 Hz for 30 s) (Fig. 3A; fEPSPs were  $177\%$  and  $184\%$  of baseline for 5 Hz for 30 s and 5 Hz for 3 min, respectively;  $P = 0.12$ ). To make sure that this finding was not simply attributable to a deficit in synaptic transmission and subsequent failure to activate extrasynaptic GluN2B receptors, we promoted glutamate spillover using the glutamate transporter inhibitor TBOA. Al-

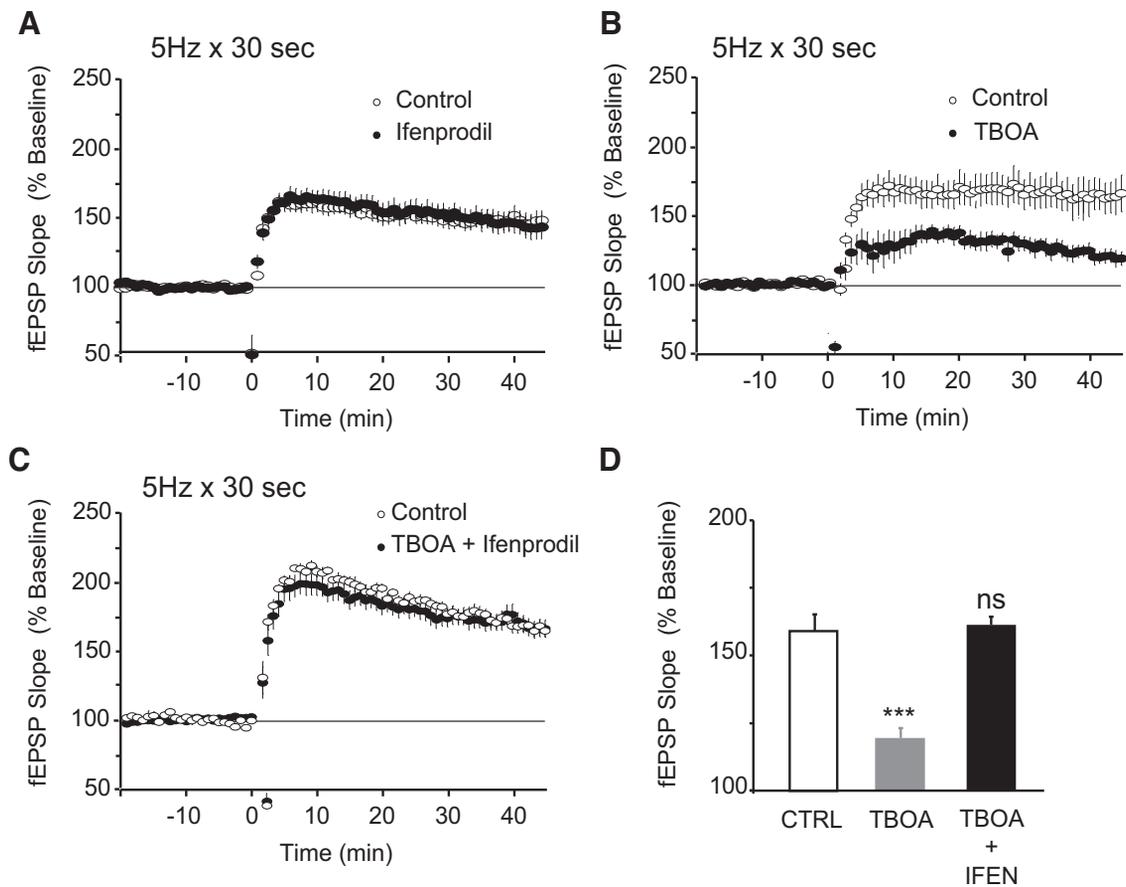


Fig. 2. Extrasyaptic GluN2B homeostasis is recruited after glutamate spillover. **A:** 5-Hz stimulation for 30 s delivered at time = 0 induced indistinguishable robust long-term potentiation (LTP) in vehicle control experiments and in the presence of ifenprodil 5  $\mu$ M. Field excitatory postsynaptic potentials (fEPSPs) were 146.7% of baseline 45 min after 5-Hz stimulation for control ( $\circ$ ,  $n = 8$  from 6 animals) and 143.7% of baseline in the presence of ifenprodil ( $\bullet$ ,  $n = 5$  from 5 animals);  $P = 0.93$ . **B:** 5-Hz, 30-s stimulation delivered at time = 0 induced robust LTP (fEPSPs were 166.5% of baseline in vehicle control experiments;  $\circ$ ,  $n = 6$  from 6 animals) but was significantly impaired in the presence of the glutamate transporter blocker DL-threo-beta-benzyloxyaspartic acid (TBOA) 10  $\mu$ M (fEPSPs were 119.2% of baseline;  $\bullet$ ,  $n = 6$  from 5 animals;  $P < 0.001$  compared with controls). **C:** 5-Hz stimulation for 30 s induced indistinguishable robust LTP in vehicle control experiments ( $\circ$ ,  $n = 11$  from 10 animals, fEPSPs were 160.4% of baseline) and when delivered in the presence of TBOA 10  $\mu$ M and ifenprodil 5  $\mu$ M ( $\bullet$ ,  $n = 8$  from 7 animals; fEPSPs were 161.3% of baseline);  $P = 0.91$ . **D:** summary of the data presented in A–C. Bar graph indicates the average fEPSP obtained from the last 5 min of recording for each condition (\*\* $P < 0.001$  compared with untreated controls). IFEN, ifenprodil.

though TBOA blocked LTP induced by stimulation at 5 Hz for 30 s in WT littermates (Fig. 3B), it caused no significant impairment in SAP102 mutant mice (Fig. 3C). In WT, fEPSPs were  $151.5 \pm 4.9\%$  and  $116.7 \pm 4.7\%$  of baseline levels in the absence or presence of TBOA, respectively ( $P < 0.001$ ). In SAP102 KO, fEPSPs were  $149.2 \pm 11.0\%$  and  $141.7 \pm 6.0\%$  of baseline levels in the absence or presence of TBOA, respectively ( $P = 0.279$ ). Taken together, these findings suggest that activation of an extrasyaptic GluN2B-SAP102 functional complex during prolonged theta-pulse stimulation activates signaling cascades that antagonize the induction of LTP.

## DISCUSSION

Although LTP is an attractive model for memory storage, it tends to destabilize neuronal circuits because it drives synapses toward a maximum value (Abbott and Nelson 2000). Unless opposed by homeostatic mechanisms operating through negative feedback rules, cumulative LTP could render synapses unable to encode additional information. In this study, we describe a novel homeostatic mechanism that rapidly antagonizes LTP induction when the synapse is faced with prolonged durations of synaptic stimulation. We have previously found

that brief periods of theta-pulse stimulation (5 Hz for 15–30 s) trigger LTP in the CA1 region of the hippocampus, whereas increasing the period of stimulation (5 Hz for 1–3 min) has no lasting effects on basal synaptic transmission (Thomas et al. 1996). In this study, we found that prolonging stimulation times promotes glutamate spillover to extrasyaptic sites and recruitment of a GluN2B-SAP102 functional complex that antagonizes LTP induction.

A number of additional homeostatic mechanisms might compensate for activity-dependent increases in excitatory synaptic drive, the most studied being synaptic scaling (Turrigiano et al. 1998). To compensate for increases in activity, synaptic scaling operates through negative feedback rules by globally scaling down the strength of all synapses in a neuron (Turrigiano et al. 1998). Although synaptic scaling seems an ideal mechanism to compensate for the increased excitatory drive caused by LTP, it has the disadvantage of being slow (requiring hours and days of altered activity to take place) and nonspecific (it modifies all synapses in a neuron). Because LTP can be induced very rapidly (standard LTP protocols take a few seconds), synapses can reach a maximum value very quickly, rendering the synapse unable to encode additional information.

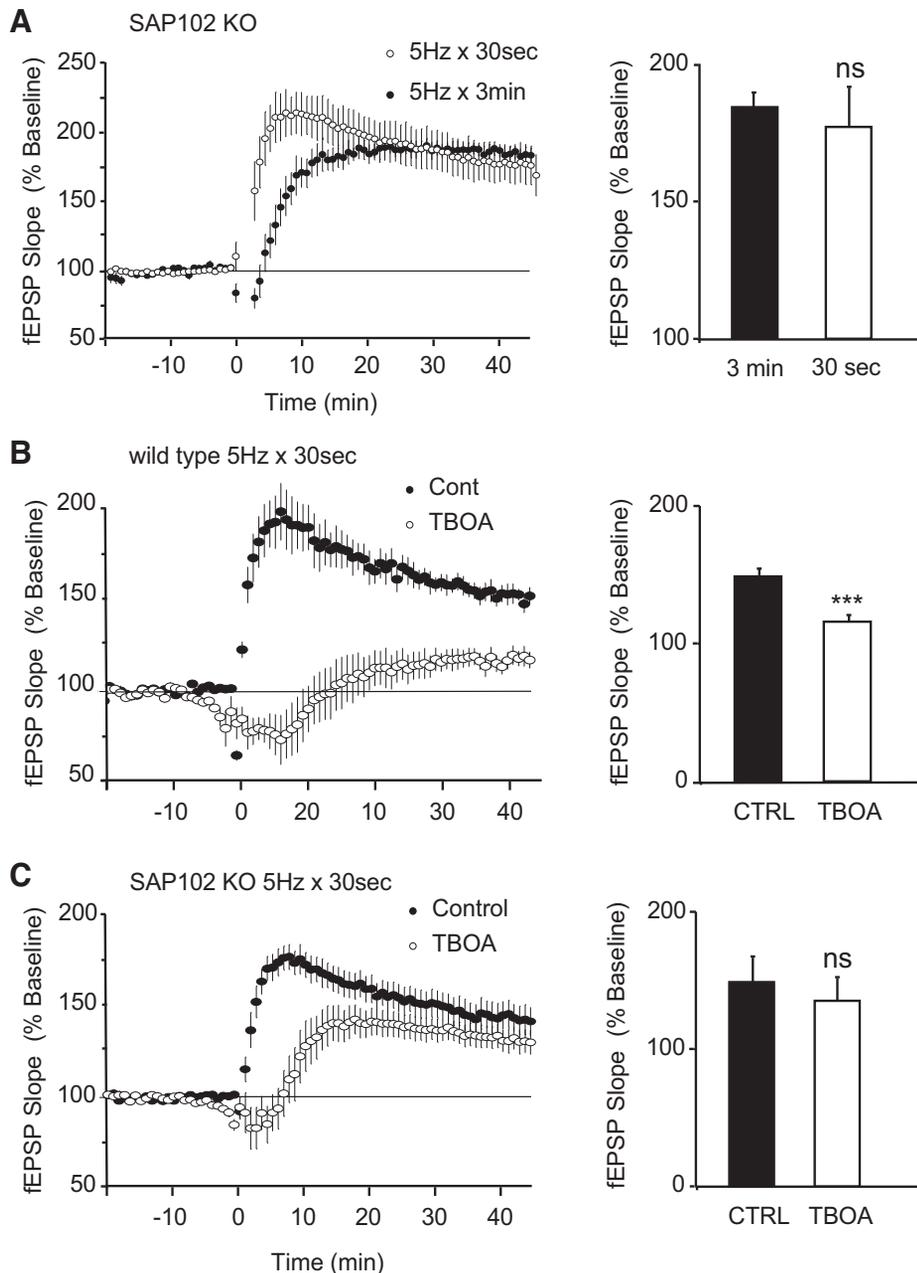


Fig. 3. Extrasynaptic GluN2B homeostasis is uncoupled in the SAP102 knockout (KO) mice. *A, left*: indistinguishable amount of long-term potentiation (LTP) is induced by both short (30 s) and long (3 min) periods of 5-Hz stimulation in the SAP102 KO mice. Field excitatory post-synaptic potentials (fEPSPs) were 177% ( $n = 11$  from 5 animals) and 184% ( $n = 10$  from 6 animals) of baseline for 30 s and 3 min, respectively. *Right*: bar graph indicating the average fEPSP obtained from the last 10 min of recording for each condition ( $P = 0.12$ ). *B, left*: 5-Hz, 30-s stimulation delivered at time = 0 induced robust LTP in wild-type littermates. fEPSPs were 159.5% of baseline in vehicle control experiments (●,  $n = 6$  from 4 animals) but were significantly impaired in the presence of the glutamate transporter blocker DL-threo-beta-benzyloxyaspartic acid (TBOA) (fEPSPs were 116.7% of baseline; ○,  $n = 6$  from 4 animals). *Right*: bar graph indicating the average fEPSP obtained from the last 10 min of recording for each condition ( $***P < 0.001$ ). *C, left*: 5-Hz stimulation for 30 s caused a strong potentiation in the SAP102 KO mice either in the presence or absence of TBOA. fEPSPs were 149.6% ( $n = 6$  from 4 animals) and 141.7% ( $n = 8$  from 4 animals) of baseline in the absence or presence of TBOA, respectively. *Right*: bar graph indicating the average fEPSP obtained from the last 5 min of recording for each condition.  $P = 0.279$ .

The disparity in the time courses of homeostatic scaling and LTP outlines a need for a fast downregulating homeostatic mechanism (Keck et al. 2017; Zenke et al. 2013), such as the extrasynaptic GluN2B-mediated mechanism described in this study.

This study also emphasizes the critical role of glutamate transporters in regulating synaptic transmission and plasticity through activation of extrasynaptic GluN2B receptors. We found that even brief trains of stimulation can activate these receptors under pharmacological blockade of glutamate transporters, either directly via glutamate spillover or indirectly via some other mechanism. These results are consistent with the LTP deficits found in mice lacking either the glial (GLT-1) or neuronal glutamate transporter (EAAC1) (Katagiri et al. 2001; Scimemi et al. 2009). Interestingly, the LTP deficits observed in the EAAC1<sup>-/-</sup> mice can be rescued with the GluN2B antagonist ifenprodil (Scimemi et al. 2009).

Although most GluN2B-containing NMDA receptors are located extrasynaptically, they can also be found at synaptic sites (Thomas et al. 2006). Because of this, we cannot completely rule out the possibility that synaptic GluN2B-containing NMDA receptors also contribute to the homeostatic process. Nevertheless, this possibility is unlikely because the GluN2B blocker ifenprodil does not further increase the amount of LTP obtained with brief trains of stimulation (5 Hz for 30 s). It is also unlikely that ifenprodil failure to enhance LTP is attributable to saturation of synaptic potentiation because we have previously shown that high-frequency stimulation (100 Hz for 1 s) can increase the amount of LTP obtained with this pattern of stimulation (5 Hz for 30 s) (Moody et al. 1999). Furthermore, the fact that even a brief period of stimulation can recruit the GluN2B-mediated homeostatic pathway when delivered in the presence of the glutamate transporter blocker strongly suggests that only extrasynaptic GluN2B can antagonize LTP.

Given that the GluN2B-mediated homeostatic mechanism was absent in SAP102 KO mice, our study suggests that extrasynaptic GluN2B acts in a complex with the scaffolding protein SAP102 to antagonize LTP induction. Although the present finding could be explained by a lack of GluN2B in the SAP102 KO, we have previously shown that GluN2B location and function are intact in this mutant (Cuthbert et al. 2007). The functional coupling between extrasynaptic GluN2B and SAP102 is consistent with their preferential binding (Chen et al. 2011; Murata and Constantine-Paton 2013; Sans et al. 2000) and extrasynaptic location (Petralia 2012; Zheng et al. 2011).

The potential relevance of this novel extrasynaptic GluN2B/SAP102-mediated homeostatic mechanism in vivo has been recently highlighted in a study showing that loss of the extrasynaptic GluN2B-SAP102 complex is associated with seizure susceptibility (Okuda et al. 2017). On the other hand, it has been shown that overactivation of extrasynaptic GluN2B attributable to alterations in glutamate uptake underlies the LTP deficits found in Alzheimer's disease models (Li et al. 2011; Varga et al. 2015). In fact, it has been shown that promoting glutamate spillover with TBOA mimics and occludes the LTP deficits mediated by oligomeric A $\beta$  (Huang et al. 2018; Li et al. 2009, 2011; Varga et al. 2015). In light of our present findings, it is plausible that the aberrant activation of the extrasynaptic GluN2B-SAP102 homeostatic mechanism described here might contribute to the LTP deficits found in Alzheimer's disease models.

In conclusion, we have uncovered a novel homeostatic mechanism that opposes LTP induction under conditions of prolonged theta-pulse stimulation at excitatory synapses in the hippocampal CA1 region. Because this feedback loop is built into the synapse and its surrounding extrasynaptic site, it is fast and presumably input specific and thus complements other homeostatic mechanisms operating at slower time scales such as scaling (Zenke et al. 2013).

#### ACKNOWLEDGMENTS

Present address for P. Opazo: Clem Jones Centre for Aging Dementia Research, Queensland Brain Institute, Univ. of Queensland, St. Lucia, Queensland, Australia.

Present address for J. Y. Delgado: Dept. of Neurobiology, Univ. of Chicago, Chicago, IL.

#### GRANTS

This work was supported by HHS, NIH, National Institute of Mental Health (NIMH) (T. O'Dell), Wellcome Trust Grant 202932 (S. Grant), and the European Research Council under the European Union Horizon 2020 research and innovation program (Grant Agreement 695568) (S. Grant).

#### DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

#### AUTHOR CONTRIBUTIONS

J.Y.D., T.J.O., and P.O. conceived and designed research; J.Y.D., A.E.F., and P.O. performed experiments; J.Y.D., A.E.F., and P.O. analyzed data; J.Y.D., A.E.F., and P.O. interpreted results of experiments; P.O. prepared figures; P.O. drafted manuscript; J.Y.D., A.E.F., S.G.G., T.J.O., and P.O. edited and revised manuscript; J.Y.D., A.E.F., S.G.G., T.J.O., and P.O. approved final version of manuscript.

#### REFERENCES

- Abbott LF, Nelson SB. Synaptic plasticity: taming the beast. *Nat Neurosci* 3, Suppl: 1178–1183, 2000. doi:10.1038/81453.
- Babiec WE, Jami SA, Guglietta R, Chen PB, O'Dell TJ. Differential regulation of NMDA receptor-mediated transmission by SK channels underlies dorsal-ventral differences in dynamics of Schaffer collateral synaptic function. *J Neurosci* 37: 1950–1964, 2017. doi:10.1523/JNEUROSCI.3196-16.2017.
- Barria A, Malinow R. NMDA receptor subunit composition controls synaptic plasticity by regulating binding to CaMKII. *Neuron* 48: 289–301, 2005. doi:10.1016/j.neuron.2005.08.034.
- Baude A, Nusser Z, Roberts JD, Mulvihill E, McIlhinney RAJ, Somogyi P. The metabotropic glutamate receptor (mGluR1 alpha) is concentrated at perisynaptic membrane of neuronal subpopulations as detected by immunogold reaction. *Neuron* 11: 771–787, 1993. doi:10.1016/0896-6273(93)90086-7.
- Chen BS, Thomas EV, Sanz-Clemente A, Roche KW. NMDA receptor-dependent regulation of dendritic spine morphology by SAP102 splice variants. *J Neurosci* 31: 89–96, 2011. doi:10.1523/JNEUROSCI.1034-10.2011.
- Chen BS, Gray JA, Sanz-Clemente A, Wei Z, Thomas EV, Nicoll RA, Roche KW. SAP102 mediates synaptic clearance of NMDA receptors. *Cell Reports* 2: 1120–1128, 2012. doi:10.1016/j.celrep.2012.09.024.
- Cuthbert PC, Stanford LE, Coba MP, Ainge JA, Fink AE, Opazo P, Delgado JY, Komiyama NH, O'Dell TJ, Grant SG. Synapse-associated protein 102/dlgh3 couples the NMDA receptor to specific plasticity pathways and learning strategies. *J Neurosci* 27: 2673–2682, 2007. doi:10.1523/JNEUROSCI.4457-06.2007.
- Frank RA, Grant SG. Supramolecular organization of NMDA receptors and the postsynaptic density. *Curr Opin Neurobiol* 45: 139–147, 2017. doi:10.1016/j.conb.2017.05.019.
- Frank RA, Komiyama NH, Ryan TJ, Zhu F, O'Dell TJ, Grant SG. NMDA receptors are selectively partitioned into complexes and supercomplexes during synapse maturation. *Nat Commun* 7: 11264, 2016. doi:10.1038/ncomms11264.
- Grant SG, O'Dell TJ. Multiprotein complex signaling and the plasticity problem. *Curr Opin Neurobiol* 11: 363–368, 2001. doi:10.1016/S0959-4388(00)00220-8.
- Hardingham GE, Bading H. Synaptic versus extrasynaptic NMDA receptor signalling: implications for neurodegenerative disorders. *Nat Rev Neurosci* 11: 682–696, 2010. doi:10.1038/nrn2911.
- Hardingham GE, Fukunaga Y, Bading H. Extrasynaptic NMDARs oppose synaptic NMDARs by triggering CREB shut-off and cell death pathways. *Nat Neurosci* 5: 405–414, 2002. doi:10.1038/nrn835.
- Huang YH, Bergles DE. Glutamate transporters bring competition to the synapse. *Curr Opin Neurobiol* 14: 346–352, 2004. doi:10.1016/j.conb.2004.05.007.
- Huang S, Tong H, Lei M, Zhou M, Guo W, Li G, Tang X, Li Z, Mo M, Zhang X, Chen X, Cen L, Wei L, Xiao Y, Li K, Huang Q, Yang X, Liu W, Zhang L, Qu S, Li S, Xu P. Astrocytic glutamatergic transporters are involved in A $\beta$ -induced synaptic dysfunction. *Brain Res* 1678: 129–137, 2018. doi:10.1016/j.brainres.2017.10.011.
- Katagiri H, Tanaka K, Manabe T. Requirement of appropriate glutamate concentrations in the synaptic cleft for hippocampal LTP induction. *Eur J Neurosci* 14: 547–553, 2001. doi:10.1046/j.0953-816x.2001.01664.x.
- Keck T, Toyozumi T, Chen L, Doiron B, Feldman DE, Fox K, Gerstner W, Haydon PG, Hübener M, Lee HK, Lisman JE, Rose T, Sengpiel F, Stellwagen D, Stryker MP, Turrigiano GG, van Rossum MC. Integrating Hebbian and homeostatic plasticity: the current state of the field and future research directions. *Philos Trans R Soc Lond B Biol Sci* 372: 20160158, 2017. doi:10.1098/rstb.2016.0158.
- Kim E, Sheng M. PDZ domain proteins of synapses. *Nat Rev Neurosci* 5: 771–781, 2004. doi:10.1038/nrn1517.
- Li S, Hong S, Shepardson NE, Walsh DM, Shankar GM, Selkoe D. Soluble oligomers of amyloid beta protein facilitate hippocampal long-term depression by disrupting neuronal glutamate uptake. *Neuron* 62: 788–801, 2009. doi:10.1016/j.neuron.2009.05.012.
- Li S, Jin M, Koeglsperger T, Shepardson NE, Shankar GM, Selkoe DJ. Soluble A $\beta$  oligomers inhibit long-term potentiation through a mechanism involving excessive activation of extrasynaptic NR2B-containing NMDA receptors. *J Neurosci* 31: 6627–6638, 2011. doi:10.1523/JNEUROSCI.0203-11.2011.

- Liu L, Wong TP, Pozza MF, Lingenhoehl K, Wang Y, Sheng M, Auberson YP, Wang YT. Role of NMDA receptor subtypes in governing the direction of hippocampal synaptic plasticity. *Science* 304: 1021–1024, 2004. doi:10.1126/science.1096615.
- MacGillavry HD, Song Y, Raghavachari S, Blanpied TA. Nanoscale scaffolding domains within the postsynaptic density concentrate synaptic AMPA receptors. *Neuron* 78: 615–622, 2013. doi:10.1016/j.neuron.2013.03.009.
- Massey PV, Johnson BE, Moulton PR, Auberson YP, Brown MW, Molnar E, Collingridge GL, Bashir ZI. Differential roles of NR2A and NR2B-containing NMDA receptors in cortical long-term potentiation and long-term depression. *J Neurosci* 24: 7821–7828, 2004. doi:10.1523/JNEUROSCI.1697-04.2004.
- Moody TD, Carlisle HJ, O'Dell TJ. A nitric oxide-independent and beta-adrenergic receptor-sensitive form of metaplasticity limits theta-frequency stimulation-induced LTP in the hippocampal CA1 region. *Learn Mem* 6: 619–633, 1999. doi:10.1101/lm.6.6.619.
- Morishita W, Lu W, Smith GB, Nicoll RA, Bear MF, Malenka RC. Activation of NR2B-containing NMDA receptors is not required for NMDA receptor-dependent long-term depression. *Neuropharmacology* 52: 71–76, 2007. doi:10.1016/j.neuropharm.2006.07.005.
- Murata Y, Constantine-Paton M. Postsynaptic density scaffold SAP102 regulates cortical synapse development through EphB and PAK signaling pathway. *J Neurosci* 33: 5040–5052, 2013. doi:10.1523/JNEUROSCI.2896-12.2013.
- Nair D, Hosy E, Petersen JD, Constals A, Giannone G, Choquet D, Sibarita JB. Super-resolution imaging reveals that AMPA receptors inside synapses are dynamically organized in nanodomains regulated by PSD95. *J Neurosci* 33: 13204–13224, 2013. doi:10.1523/JNEUROSCI.2381-12.2013.
- Okuda K, Kobayashi S, Fukaya M, Watanabe A, Murakami T, Hagiwara M, Sato T, Ueno H, Ogonuki N, Komano-Inoue S, Manabe H, Yamaguchi M, Ogura A, Asahara H, Sakagami H, Mizuguchi M, Manabe T, Tanaka T. CDKL5 controls postsynaptic localization of GluN2B-containing NMDA receptors in the hippocampus and regulates seizure susceptibility. *Neurobiol Dis* 106: 158–170, 2017. doi:10.1016/j.nbd.2017.07.002.
- Petralia RS. Distribution of extrasynaptic NMDA receptors on neurons. *Sci World J* 2012: 267120, 2012. doi:10.1100/2012/267120.
- Rusakov DA, Scimemi A, Walker MC, Kullmann DM. Comment on “Role of NMDA receptor subtypes in governing the direction of hippocampal synaptic plasticity” [author reply]. *Science* 305: 1912, 2004. doi:10.1126/science.1101128.
- Sans N, Petralia RS, Wang YX, Blahos J II, Hell JW, Wenthold RJ. A developmental change in NMDA receptor-associated proteins at hippocampal synapses. *J Neurosci* 20: 1260–1271, 2000. doi:10.1523/JNEUROSCI.20-03-01260.2000.
- Scimemi A, Tian H, Diamond JS. Neuronal transporters regulate glutamate clearance, NMDA receptor activation, and synaptic plasticity in the hippocampus. *J Neurosci* 29: 14581–14595, 2009. doi:10.1523/JNEUROSCI.4845-09.2009.
- Stocca G, Vicini S. Increased contribution of NR2A subunit to synaptic NMDA receptors in developing rat cortical neurons. *J Physiol* 507: 13–24, 1998. doi:10.1111/j.1469-7793.1998.013bu.x.
- Thomas MJ, Moody TD, Makhinson M, O'Dell TJ. Activity-dependent beta-adrenergic modulation of low frequency stimulation induced LTP in the hippocampal CA1 region. *Neuron* 17: 475–482, 1996. doi:10.1016/S0896-6273(00)80179-8.
- Thomas CG, Miller AJ, Westbrook GL. Synaptic and extrasynaptic NMDA receptor NR2 subunits in cultured hippocampal neurons. *J Neurophysiol* 95: 1727–1734, 2006. doi:10.1152/jn.00771.2005.
- Tovar KR, Westbrook GL. The incorporation of NMDA receptors with a distinct subunit composition at nascent hippocampal synapses in vitro. *J Neurosci* 19: 4180–4188, 1999. doi:10.1523/JNEUROSCI.19-10-04180.1999.
- Turrigiano GG, Leslie KR, Desai NS, Rutherford LC, Nelson SB. Activity-dependent scaling of quantal amplitude in neocortical neurons. *Nature* 391: 892–896, 1998. doi:10.1038/36103.
- Varga E, Juhász G, Bozsó Z, Penke B, Fülöp L, Szegedi V. Amyloid- $\beta$ 1-42 disrupts synaptic plasticity by altering glutamate recycling at the synapse. *J Alzheimers Dis* 45: 449–456, 2015. doi:10.3233/JAD-142367.
- Williams K. Ifenprodil discriminates subtypes of the N-methyl-D-aspartate receptor: selectivity and mechanisms at recombinant heteromeric receptors. *Mol Pharmacol* 44: 851–859, 1993.
- Zenke F, Hennequin G, Gerstner W. Synaptic plasticity in neural networks needs homeostasis with a fast rate detector. *PLOS Comput Biol* 9: e1003330, 2013. doi:10.1371/journal.pcbi.1003330.
- Zhang J, Diamond JS. Subunit- and pathway-specific localization of NMDA receptors and scaffolding proteins at ganglion cell synapses in rat retina. *J Neurosci* 29: 4274–4286, 2009. doi:10.1523/JNEUROSCI.5602-08.2009.
- Zheng CY, Wang YX, Kachar B, Petralia RS. Differential localization of SAP102 and PSD-95 is revealed in hippocampal spines using super-resolution light microscopy. *Commun Integr Biol* 4: 104–105, 2011. doi:10.4161/cib.14172.