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RSPO1/β-Catenin Signaling Pathway Regulates Oogonia Differentiation and Entry into Meiosis in the Mouse Fetal Ovary

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Abstract

Differentiation of germ cells into male gonocytes or female oocytes is a central event in sexual reproduction. Proliferation and differentiation of fetal germ cells depend on the sex of the embryo. In male mouse embryos, germ cell proliferation is regulated by the RNA helicase Mouse Vasa homolog gene and factors synthesized by the somatic Sertoli cells promote gonocyte differentiation. In the female, ovarian differentiation requires activation of the WNT/β-catenin signaling pathway in the somatic cells by the secreted protein RSPO1. Using mouse models, we now show that Rspo1 also activates the WNT/β-catenin signaling pathway in germ cells. In XX Rspo1⁻/⁻ gonads, germ cell proliferation, expression of the early meiotic marker Stra8, and entry into meiosis are all impaired. In these gonads, impaired entry into meiosis and germ cell sex reversal occur prior to detectable Sertoli cell differentiation, suggesting that β-catenin signaling acts within the germ cells to promote oogonial differentiation and entry into meiosis. Our results demonstrate that Rspo1/β-catenin signaling is involved in meiosis in fetal germ cells and contributes to the cellular decision of germ cells to differentiate into oocyte or sperm.


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Introduction

Germ cells have the unique capacity to ensure the propagation of genetic information between generations. Once they are sex determined, they become competent for sexual reproduction by undergoing meiosis [1,2]. During fertilization, male and female gametes join to form a diploid zygote with a mix of maternal and paternal heritable information. The sex of the resulting individual is determined by paternal transmission of either the Y chromosome or the X chromosome, which promotes the formation of an ovary [3].

Male gonad development starts with somatic cell differentiation that is initiated by expression of Sox9, an activator of the transcription factor Sry [4]. In turn, activation of SOX9 leads to differentiation of somatic cells into Sertoli cells [5]. Rpo1 and Wnt4 are required for ovarian somatic differentiation [6,7,8,9]. Loss-of-function of either Rpo1 or Wnt4 promotes sex reversal of the supporting cell lineages in XX gonads with differentiation of Sertoli cells around birth and development of oovestes. RSPO1 is synthesized and secreted by somatic cells. RSPO proteins are regulators of the canonical WNT/β-catenin signaling pathway [10] and in vitro mediate their action through LRP6, the co-receptor of this signalling pathway [11,12,13]. However, the molecular mechanism remains to be elucidated. In vivo, RSPO1 can compete with DKK1, a negative regulator of the WNT/β-catenin signalling pathway, by binding to Kremen1 and then triggering the release of LRP6 [14]. However, Kremen receptors do not seem to be crucial for Rspo1 signaling in vivo [15]. It has been shown that Rspo1 binds directly to LRP6 [11,12] suggesting that this interaction is involved in transduction of the signal. This signal promotes stabilization of β-catenin which can then interact with the transcription factors LEF/TCF to induce expression of downstream target genes [16]. In oocytes, Rspo1 activates the β-catenin signaling pathway, promoting the up-regulation of Wnt4 and differentiation of follicular cells [6].

In both XX and XY embryos, primordial germ cells migrate through the hindgut to colonize the gonad at around E10.5 [17]. In XY, but not XX fetal gonads, Mvh (Mouse vasal homolog) is required for germ cell proliferation [18] indicating that fetal germ cell proliferation is regulated by different pathways in XX and XY fetal gonads. However, the pathway(s) inducing germ cell proliferation in fetal ovaries remain to be elucidated. In fetal ovaries, germ cells proliferate, differentiate into oogonia and enter meiosis at E12.5–13.5. In the fetal testis, germ cells differentiate into gonocytes and become blocked in the G0/G1 phase of the cell cycle at around E14.5 [19]. Proliferation of these cells will only
resume after birth when spermatogenesis begins. Thus, the initial decision to become an oocyte or a gonocyte is closely coupled with differential regulation of the cell cycle of fetal germ cells [20].

In XX fetal gonads, it has been shown that the onset of meiosis requires up-regulation of the Stra8 gene in fetal oogonia [21]. Retinoic acid (RA) can induce Stra8 expression in mouse fetal germ cells in organ cultures [22,23,24]. Stra8-deficient germ cells normally proliferate, but they fail to undergo premeiotic DNA replication and meiosis in fetal ovaries and arrest as premeiotic germ cells [21]. In rats that are nutritionally deficient for vitamin A, postnatal XX germ cells have defects in upregulation of Stra8 and meiosis begins [24]. However, it has been reported that in XX Raldh2−/−/embryos, Stra8 expression is up-regulated normally in fetal oogonia in the absence of physiologically detectable RA levels in vivo [25]. Thus multiple signaling pathways may be involved in Stra8 induction and meiosis initiation in mammalian gonads. In addition, RA promotes germ cell proliferation and germ cell survival in cultured embryonic ovaries [26,27] underlying the multiple roles of RA in germ cell fate in embryonic ovaries.

In XY gonads, Cyp26b1, which is a member of a family of enzymes that degrades RA, is required to prevent Stra8 expression in mouse fetal testes in vivo and in vitro [28,29]. Tight control of RA levels is important for testicular development since RA can impair peritubular myoid cell migration and affect Sertoli cell differentiation in cultured rat embryonic testes [30]. In the developing testis, Sertoli cells, the somatic cells required to support spermatogenesis, contribute to gonocyte differentiation [31]. Consequently, defects in Sertoli cell differentiation promote male-to-female sex reversal of germ cells [5]. Indeed, XY germ cell sex reversal can be achieved by blocking the secretory pathway in cultured fetal testes [32], indicating that secreted factors, presumably originating from Sertoli cells, are required for male germ cell differentiation. Sertoli cells and Sox9 are required for Fgf9 up-regulation in the fetal testis [33,34]. FGF9 is a secreted growth factor that promotes Sertoli cell differentiation and proliferation [34] and inhibits germ cell meiosis in culture [35,36]. In addition to environmental signals, germ cell meiosis is also controlled by intrinsic factors that favor or prevent meiosis during embryogenesis [37,38]. Indeed, the translational regulator Nana2 is required to maintain germ cells in G0/G1 phase in the fetal testis and ectopic expression of this gene in XX germ cells prevents entry into meiosis [39].

It is now clear that somatic and germ cell factors are required for oogonia to enter meiosis. Here we show that Rspo1 directly activates β-catenin in XX germ cells. In turn, Rspo1/β-catenin signaling promotes XX germ cell proliferation and entry into meiosis.

Results

Rspo1 promotes XX germ cell proliferation

Although the precise mechanisms controlling germ cell proliferation in XY fetal gonads remain to be clarified, they involve the RNA helicase protein MVH [18]. In XX gonads, MVH is expressed in the germ cells but is not required for oogonial proliferation indicating that the regulation of the proliferation of female fetal germ cells involves distinct molecular pathways. Rspo1 has been shown to regulate proliferation [40]. This signaling protein was found bound to the cellular membrane of the germ cells and somatic cells of the ovaries [10,41,42] suggesting that Rspo1 plays a role not only in somatic but also in germ cell proliferation. In addition, Rspo1 becomes strongly up-regulated in the somatic cells of the XX gonad from E11.5 onwards [6,7,43] when germ cells are highly proliferative. To address whether this secreted protein might promote germ cell proliferation in XX gonads, we carried out bromodeoxyuridine (BrdU) labeling experiments at E12.5 in XX Rspo1 mutant and control gonads (Fig. 1A). In XX Rspo1−/− gonads and in XY and XX controls, BrdU positive cells were detected, implying that germ cells were proliferating. In XX Rspo1−/− gonads, the percentage of germ cells that incorporated BrdU was significantly reduced compared to XX controls (31% in XX Rspo1−/− gonads versus 48% in XX controls), suggesting a decrease in germ cell proliferation. However, some reduction of BrdU incorporation could also be due to pre-meiotic S phase failure in germ cells within the mutant gonads. As premeiotic S phase is not followed by cell division, we were able to assess the relationship of this reduction in BrdU incorporation to germ cell proliferation, by counting the number of germ cells at E14.5 in XX controls and XX Rspo1−/− gonads. Mutant mice showed a small, but statistically significant decrease of germ cells at this stage (Fig. 1B).

Rspo1 is involved in Wnt4 up-regulation in XX gonads. Indeed, Wnt4 is expressed in XX and XY gonads at a basal level [33] and Rspo1 is required for the up-regulation of Wnt4 in the XX gonad. Previous results showed that 90% of germ cells undergo apoptosis in XX Wnt4−/− gonads [8]. In addition, analysis of another designed Rspo1 mutant showed an increase of cell apoptosis potentially germ cells in XX Rspo1 mutant gonads at E16.5 [43]. However, statistical analysis showed no significant increase in germ cell apoptosis (TUNEL experiments) takes place at E12.5, E14.5 or at E16.5 in XX Rspo1−/− gonads when compared to XX controls (Fig. 1). This suggests that some remaining Wnt4 expression can still promote survival of germ cells in these Rspo1 mutants [6]. The increase of apoptotic cells observed in another XX Rspo1−/− gonads could be due either to a different genetic background or a different mutant allele [43]. Thus depletion in germ cell number in XX Rspo1−/− mutants is not due to apoptosis, but must be caused by defects in germ cell proliferation.

Loss of Rspo1 impairs XX germ cell meiosis

After proliferation, XX fetal germ cells enter meiosis at around E13.5–E14.5 whereas male germ cells arrest in G0/G1 phase of the cell cycle. At E14.5, when female germ cells enter meiosis, the expression of Oct4 is down-regulated in ovaries whereas it is maintained in XY gonads [44,45,46]. In situ hybridization studies for Oct4 at E14.5 revealed robust expression in XX Rspa1−/− gonads and XY gonads, but not in XX Rspa1−/− (Fig. 2) suggesting that Rspo1 is required for Oct4 down-regulation in XX germ cells. We therefore asked whether entry into meiosis was occurring normally in XX Rspa1−/− gonads. Immunostaining using two meiotic markers, SCP3 and γH2AX, demonstrated a dramatically reduced number of meiotic cells in XX Rspa1−/− gonads to XX Rspa1−/−/2 mutants is not due to apoptosis, but must be caused by defects in germ cell proliferation.

Rspo1 contributes to Stra8 expression

Entry into meiosis is sensitive to the genetic background [37]. Indeed, a function of Dazl in the initiation of meiosis in fetal
ovaries could only be unravelled on a pure C57BL6 background. Stra8 (Stimulated by Retinoic Acid 8) is essential for meiosis induction (Baltus et al. 2006) and previously, we observed a small but not significant down-regulation of Stra8 expression (Fig. 3 in [6]). To know the relevance of this down-regulation, Stra8 expression was studied in XX Rspo12/2 gonads with a higher content of C57BL6/J. It has been shown that RA induces Stra8 expression in germ cells in XX gonads [22,23]. However, a recent report that Stra8 expression is induced normally in XX gonads from XX Raldh22/2 embryos that lack any detectable RA in their urogenital ridges, has led to the suggestion that Stra8 expression can be induced independently of RA in this developmental context [25]. This suggests that other signaling pathways might also be able to induce Stra8 expression in fetal female germ cells. To address whether Rspo1 could be involved in Stra8 expression, we analysed Stra8 expression levels in XX Rspo1+/+ and Rspo1−/− gonads at E12.5. Measurements of Cyp26b1 expression levels at E12.5 showed these remained higher in XX Rspo1−/− gonads when compared to XX controls. Although at E12.5, Cyp26b1 expression was 40 fold lower than in XY gonads, we cannot exclude that CYP26B1 is involved in Stra8 repression.

In the embryonic testis, meiosis inhibition and Stra8 repression are at least partially regulated via CYP26B1, an enzyme that metabolizes RA [25,36]. Measurements of Cyp26b1 expression levels at E12.5 showed these remained higher in XX Rspo1−/− gonads when compared to XX controls. Although at E12.5, Cyp26b1 expression was 40 fold lower than in XY gonads, we cannot exclude that CYP26B1 is involved in Stra8 repression.

Figure 1. Rspo1 promotes XX germ cells proliferation. A- Reduction of germ cell proliferation in XX Rspo1 mutant gonads. Immunodetection of the proliferating germ cells with BrdU and MVH (upper panel), and the apoptotic germ cells with TUNEL and MVH (middle panel), in XY and XX Rspo1+/+, and XX Rspo1−/− gonads at E12.5 gonads. DAPI (blue): nuclei. Inset middle panel: positive control for TUNEL. Histograms: Percentage of proliferating germ cells or apoptotic germ cells in XY, XX Rspo1+/+ and Rspo1−/− gonads at E12.5. Bars represent mean±1 SEM, n = 24 sections of each genotype. B- Ablation of Rspo1 does not trigger germ cell apoptosis. Quantification of germ cell number and germ cell apoptosis in XX Rspo1+/+ and Rspo1−/− gonads at E14.5 and E16.5. Bars represent mean±1 SEM, n = 24 sections of each genotype.

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down-regulation in XX Rspo1 mutants. However, in situ hybridization analysis showed that Cyp26b1 was weakly or not expressed in XX Rspo1−/− gonads at E14.5 or at E16.5 (Fig. 3B & data not shown), when compared to XY gonads. At these stages, Stra8 and other meiotic markers (Fig. 3A and Fig. 2) were strongly down-regulated suggesting that Cyp26b1 is not the main factor repressing Stra8 expression in XX Rspo1−/− gonads at E14.5.

Stra8 expression can be induced by RA treatments [47], RA acts by binding to the Retinoic Acid Receptors (RAR), which in turns bind to retinoic acid response elements (RAREs) in the regulatory regions of direct target genes [48]. One of these direct targets is RARβ, and thus RARβ expression is up-regulated by RA signaling [49]. We next addressed whether the RA signaling pathway is affected in XX Rspo1−/− gonads by quantification of the level of expression of RARA and RARβ using Q-PCR experiments. At E12.5, RARA and RARβ levels were not changed between the different genotypes (Fig. 3A), but the variability in RAR expression levels do not allow us to completely exclude small differences in RAR expression levels in XX Rspo1−/− gonads". However, these results suggest that Stra8 induction by Rspo1 is not dependent on RA signaling.
Sex reversal of germ cells in XX Rspo1<sup>−/−</sup> gonads

To investigate how germ cells differentiate in XX Rspo1<sup>−/−</sup> mutants, we analysed the expression of known gonocyte makers. In wild type mice, Nanos2 is expressed specifically in male gonocytes. Moreover, transgenic analysis showed that Nanos2 is sufficient to repress Stra8 expression, prevent meiosis, and induce XX germ cells to adopt a male fate [39]. In situ hybridization and Q-PCR analyses demonstrated strong expression of Nanos2 in XX germ cells of Rspo1<sup>−/−</sup> gonads compared to XX controls at E14.5 whereas Nanos2 expression was not detectable at E12.5 ([Fig. 4A] and data not shown). The localization of Nanos2 expression was variable and not always at the anterior pole in the XX Rspo1<sup>−/−</sup> animals (data not shown), a pattern likely associated with the variability of the genetic background. The upregulation of Nanos2 in XX Rspo1<sup>−/−</sup> gonads was around 10-fold lower than XY control gonads which is consistent with in situ hybridization data showing that only some germ cells in XX Rspo1<sup>−/−</sup> gonads express high levels of Nanos2. This ectopic activation of Nanos2...
provides a possible explanation for the partial repression of Stra8 expression and meiosis inhibition observed in XX *Rspo1*<sup>−/−</sup> gonads at E14.5. Moreover, we quantified the level of expression of other gonocyte markers such as *Dmalt2* and *Tdh35s*. Q-PCR analyses showed that their expression was significantly up-regulated in the XX *Rspo1*<sup>−/−</sup> gonads compared to XX control gonads at E14.5 (Fig. 4A). However, their level of expression was 100 fold lower in XX *Rspo1* mutant gonads versus XY gonads suggesting that only some of the germ cells in the mutant gonads followed a male differentiation pathway or that progression along the male differentiation pathway is impaired or delayed, in comparison to SSCs in XY gonads. These cells, representing about 50% of the germ cells in the mutant gonads, may stay in an undifferentiated/pluripotent state as suggested by the maintenance of Oct4 expression observed in *in situ* hybridization experiments at E14.5 (Fig. 2).

These results were illustrated by histological analyses of gonads at E14.5 (Fig. 4B), when all the female germ cells have normally entered meiosis. Indeed, germ cell chimaerism was observed in XX *Rspo1*<sup>−/−</sup> gonads with germ cells that have initiated meiosis as expected from oocytes (Fig. 4B - arrows) and others which were comparable to quiescent gonocytes in XY gonads (Fig. 4B - arrowheads) further supporting that a certain proportion of germ cells were sex reversed in XX *Rspo1*<sup>−/−</sup> gonads. At this stage, these XX gonocytes were single, isolated cells and not enclosed by sex cords in XX mutant gonads.

**Germ cell proliferation and meiosis defects are independent of Sertoli cells in XX *Rspo1*<sup>−/−</sup> gonads**

The germ cell proliferation defect and meiotic blockage that we observed in XX *Rspo1*<sup>−/−</sup> gonads could be either a consequence of supporting cell sex reversal in these gonads, or could reflect a more direct role for *Rspo1* in promoting proliferation and meiosis in fetal germ cells. In order to check whether the supporting cells sex reversed and differentiated into Sertoli cells before sex reversal of the germ cells in XX *Rspo1*<sup>−/−</sup> gonads, we performed Q-PCR analysis for the Sertoli cell markers *Sox9* and *Pds*. These markers were slightly increased in XX *Rspo1*<sup>−/−</sup> gonads compared to the XX *Rspo1*<sup>+/+</sup> gonads at E12.5 (data not shown) and E14.5, but this increase was much lower than the high expression levels of these genes observed in XY gonads (Fig. 5A, right panels). To check whether Sertoli cells were present in XX *Rspo1*<sup>−/−</sup> gonads at E14.5, we performed *in situ* analysis using the Sertoli cell markers *Sox9*, *Pdgfs*, *SDMG1* and *Sxda* (Fig. 5A & data not shown). While these genes were highly expressed in XY gonads, no significant expression could be detected in XX *Rspo1*<sup>−/−</sup> mutants by *in situ* hybridization. *Sox9* expression and Sertoli cell differentiation did eventually occur in XX *Rspo1*<sup>−/−</sup> gonads by E19.5 ([6] & data not shown), but the prevention of meiosis and changes in germ cell gene expression seen by E14.5 in XX *Rspo1*<sup>−/−</sup> gonads preceded detectable Sertoli cell differentiation.

**Meiosis defects are independent of *Fgf9* expression in XX *Rspo1*<sup>−/−</sup> gonads**

Deletion of *Fgf9* induces massive germ cell loss in fetal XY gonads and about half of the remaining germ cells enter meiosis ([50]. Furthermore, FGF9 inhibits Stra8 up-regulation and entry into meiosis ([35,36]. Although *Fgf9* is up-regulated in Sertoli cells of XY gonads, it is initially expressed in both, XX and XY undifferentiated gonads ([33]. We therefore asked whether ablation of *Rspo1* would allow gonads to express high levels of *Fgf9* that in turn could inhibit meiosis. Previous Q-PCR analysis showed an increase of *Fgf9* expression at E14.5 ([6]. However, new Q-PCR analyses at E11.5, E12.5 and E14.5 did not show significant increase in *Fgf9* levels in XX control versus XX *Rspo1*<sup>−/−</sup> gonads. *Fgf9* is expressed in mesonephric tubules which are closely linked to the developing gonads and we cannot exclude that some *Fgf9* signals come from these mesonephric cells. To check these results, *in situ* hybridization experiments were carried out and showed no differences in staining between XX control and XX *Rspo1*<sup>−/−</sup> gonads at E12.5 and E14.5 (Fig. 3B).

It has been shown previously that *Rspo1* is genetically up-stream of *Wnt4* during ovarian development and *Rspo1* is required for *Wnt4* up-regulation in the XX gonad. Since *Wnt4* expression is antagonistic of *Fgf9* expression ([33,51], it could be expected that Rspo1 would indirectly prevent *Fgf9* expression. However, *Rspo1* deletion did not trigger robust *Fgf9* expression suggesting that the remaining expression of *Wnt4* in XX *Rspo1*<sup>−/−</sup> gonads is sufficient to prevent *Fgf9* expression. In addition, it is not known whether other pathways, independent of the *Wnt4* pathway and potentially involved in *Fgf9* regulation, are activated by Rspo1. In summary, the failure to enter meiosis does not seem to be caused by an increase of *Fgf9* expression in XX *Rspo1*<sup>−/−</sup> gonads but rather suggests a direct effect of Rspo1 promoting germ cells to differentiate along the female pathway.

**Rspo1 regulates the transcriptional activity of β-catenin in XX germ cells**

*R-spondins* are activators of the canonical WNT/β-catenin signaling and *Rspo1* activates this pathway to promote ovarian differentiation ([6,11,12,13]. Recent data show that the secreted Rspo1 protein localizes at the plasma membrane of not only somatic, but also germ cells ([41,42] suggesting that Rspo1 may be able to activate β-catenin directly in germ cells.

To investigate whether the WNT/β-catenin signaling pathway is active in fetal germ cells, we performed immunostaining at E14.5. Stabilized β-catenin was mainly found in the nuclei of XX somatic and germ cells whereas in XY gonads it predominantly localized to the cellular membranes ([Fig. 6A](#fig6){ref} see also Fig. 6 in [6]). Since nuclear staining was weaker or absent in both XY and in XX *Rspo1*<sup>−/−</sup> germ cell nuclei ([Fig. 6A](#fig6){ref}), we conclude that Rspo1 is required for the specific activation of β-catenin in XX germ cells at this stage.

The nuclear form of β-catenin has a transcriptional function and together with TCF/LEF transcription factors induces expression of target genes like *Axin2* and *Lef1* ([52,53]. To confirm a sex specific activation of β-catenin signalling, we next investigated whether effectors of this signaling pathway were expressed in fetal germ cells. For this, we purified germ cells using FACs sorting from E13.5 *Oct4-GFP* transgenic gonads ([54], a transgene that is specifically expressed in germ cells but not in somatic cells of the gonad. Q-PCR experiments were used to quantify the expression levels of known WNT/β-catenin partners and shown that *Axin2*, *Lef1* and *LRP6* (the Rspo1 receptor) ([11] were strongly expressed in XX somatic cells. This is in agreement with our previous study showing that Rspo1 is involved in β-catenin activation in ovarian somatic cells [6]. In addition, *Axin2*, *Lef1* and *LRP6* were also expressed in XX germ cells but at a lower level then in somatic tissues ([Fig. 6B](#fig6){ref}). Importantly, levels of the markers of β-catenin signaling, *Axin2*, *Lef1* and *LRP6*, were significantly higher in XX germ cells compared to XY germ cells. Analyses of the expression levels of *Meh* (germ cell marker), *Sf1* (highly expressed in male somatic cells at E13.5), *Bmp2*, *Fst* and *Rspo1* (female somatic cell markers) confirmed that there was no significant contamination of germ cells by somatic cells ([Fig. S1](#figs1){ref}). Activation of β-catenin was expected in XY somatic cells from the urogenital ridges as evidenced by expression of *Axin2* ([Fig.S2](#figs2){ref}) since
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A- XY Rspe1+/c  XX Rspe1+/c  XX Rspe1 -/c

Nanos2

E14.5

p<0.0001

XY Rspe1+/c  XX Rspe1+/c  XX Rspe1 -/c

Dnmt3L

Tdrd5

p<0.0001  p<0.001

B- XY Rspe1+/c  XX Rspe1+/c  XX Rspe1 -/c

H&E

E16.5

25 µm

10 µm
the WNT/β-catenin signalling pathway has been shown to be active in the Mullerian duct [55]. Taken together, these data show that members of the canonical β-catenin signalling pathway are expressed in XX germ cells, suggesting that this pathway is also functional within germ cells.

We had previously investigated the transcriptional function of β-catenin in fetal gonads at E14.5 using the Axin2/CAT reporter in gonads. To confirm that the β-catenin In situ hybridization at E14.5 showed strong β-catenin expression in XX fetal gonads but not in XY gonads. Using Hprt as the normalization control. Bars represent mean ± SEM, n = 8 individual embryos. β- Haematoxylin & Eosin staining of gonadic sections of XY, XX Rspo1+/− and XX Rspo1+/+ gonads at E16.5. Meiotic oogonia are indicated by arrows GO/G1 quiescent gonocytes are indicated by arrowheads.

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Figure 4. Up-regulation of β-catenin in XX Rspo1 mutant gonads. A- In situ hybridizations at E14.5 in XY and XX control and XX Rspo1−/− gonads for Nanos2. Histograms: Quantitative RT-PCR analysis of Nanos2, Dmrt3L and Tard5 expression in E14.5 XY control, XX control and Rspo1−/− gonads, using Hprt as the normalization control. Bars represent mean ± SEM, n = 8 individual embryos. B- Haematoxylin & Eosin staining of gonadic sections of XY, XX Rspo1+/− and XX Rspo1+/+ gonads at E16.5. Meiotic oogonia are indicated by arrows GO/G1 quiescent gonocytes are indicated by arrowheads.

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RSPO1 Promotes Germ Cell Sexual Differentiation

The WNT/β-catenin signalling pathway has been shown to be active in the Mullerian duct [55]. Taken together, these data show that the canonical β-catenin signalling pathway is expressed in XX germ cells, suggesting that this pathway is also functional within germ cells.

We had previously investigated the transcriptional function of β-catenin in fetal gonads at E14.5 using the Axin2/CAT reporter in gonads. To confirm that the β-catenin In situ hybridization at E14.5 showed strong β-catenin expression in XX fetal gonads but not in XY gonads. Using Hprt as the normalization control. Bars represent mean ± SEM, n = 8 individual embryos. β- Haematoxylin & Eosin staining of gonadic sections of XY, XX Rspo1+/− and XX Rspo1+/+ gonads at E16.5. Meiotic oogonia are indicated by arrows GO/G1 quiescent gonocytes are indicated by arrowheads.

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Figure 4. Up-regulation of β-catenin in XX Rspo1 mutant gonads. A- In situ hybridizations at E14.5 in XY and XX control and XX Rspo1−/− gonads for Nanos2. Histograms: Quantitative RT-PCR analysis of Nanos2, Dmrt3L and Tard5 expression in E14.5 XY control, XX control and Rspo1−/− gonads, using Hprt as the normalization control. Bars represent mean ± SEM, n = 8 individual embryos. B- Haematoxylin & Eosin staining of gonadic sections of XY, XX Rspo1+/− and XX Rspo1+/+ gonads at E16.5. Meiotic oogonia are indicated by arrows GO/G1 quiescent gonocytes are indicated by arrowheads.

Figure 4. Up-regulation of β-catenin in XX Rspo1 mutant gonads. A- In situ hybridizations at E14.5 in XY and XX control and XX Rspo1−/− gonads for Nanos2. Histograms: Quantitative RT-PCR analysis of Nanos2, Dmrt3L and Tard5 expression in E14.5 XY control, XX control and Rspo1−/− gonads, using Hprt as the normalization control. Bars represent mean ± SEM, n = 8 individual embryos. B- Haematoxylin & Eosin staining of gonadic sections of XY, XX Rspo1+/− and XX Rspo1+/+ gonads at E16.5. Meiotic oogonia are indicated by arrows GO/G1 quiescent gonocytes are indicated by arrowheads.

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A- | XY Rspo1+/− | XX Rspo1+/− | XX Rspo1−/−
---|---|---|---
Sox9 | ![Image](image1.png) | ![Image](image2.png) | ![Image](image3.png)
*Sox9* **p<0.001**

**E14.5**

PgdS | ![Image](image4.png) | ![Image](image5.png) | ![Image](image6.png)
*PgdS* **p<0.001**

**B-**

E11.5 | E12.5 | E14.5
---|---|---
Fgf9 | ![Image](image7.png) | ![Image](image8.png) | ![Image](image9.png)
*Fgf9/Dep* **p<0.001**

E12.5 | ![Image](image10.png) | ![Image](image11.png) | ![Image](image12.png)
Fgf9

E14.5 | ![Image](image13.png) | ![Image](image14.png) | ![Image](image15.png)
Fgf9

**Legend:**
- XY Rspo1+/−
- XX Rspo1+/−
- XX Rspo1−/−
- NS
- ***p<0.001***
- **NS**
Figure 5. Germ cell sex reversal is not caused by Sertoli cell factors in XX Rspos1 mutant gonads. A- Lack of Sertoli cell differentiation at E14.5 in XX Rspos1 mutant gonads. In situ hybridizations of Sox9 and Pgds in E14.5 XY and XX control and XX Rspos1−/− gonads. Right panels: Quantitative RT-PCR analysis of Sox9 and Pgds, using Hprt as the normalization control. Bars represent mean±1 SEM, n = 8 individual embryos. B- Fgf9 is not significantly expressed at E11.5 and E12.5 in XX Rspos1 mutant gonads. Upper panels: Quantitative RT-PCR analysis of Fgf9 at E11.5, E12.5 and E14.5 using Hprt as the normalization control. NS: Not significant. Bars represent mean±1 SEM, n = 8 individual embryos. Lower panels: In situ hybridizations of Fgf9 (E12.5 and E14.5) in XY and XX control and XX Rspos1−/− gonads.

Figure 6. Rspos1 activates β-catenin signaling pathway in XX germ cells. A- Stabilization of β-catenin in XX germ cells is mediated by Rspos1. Active β-catenin (red) and MVH immunostaining (germ cells, green) in XY, XX Rspos1+/+ and Rspos1−/− gonads at E14.5. DAPI (blue): nuclei. Arrowheads: nuclear β-catenin staining. B- Effectors of the WNT/β-catenin signaling pathway are expressed in XX germ cells. Quantitative RT-PCR analysis of Axin2, Lef1, LRP6 expression in E13.5 germ cells (XY and XY Oct4-positive cells) and somatic cells (XX and XY Oct4-negative cells), using respectively Hprt as the normalization control. Bars represent mean±1 SEM, n = 3 individual experiments. C- Expression of Axin2, a target of β-catenin in XX germ cells. X-Gal staining (AXIN2) (blue) and MVH immunostaining (germ cells, red) in XY Axin2+/+LacZ, XX Axin2+/+LacZ and XX Axin2−/−LacZ; Rspos1−/− gonads at E12.5. Arrowheads: germ cells. G: gonad. X-Gal staining (AXIN2) (blue) and MVH immunostaining (germ cells, green) on isolated germ cells from XY Axin2+/+LacZ and XX Axin2−/−LacZ gonads at E12.5. Arrowheads: germ cells, arrows: somatic cells.
Statistical analyses showed that the percentage of proliferating germ cells was similar in XX Catnb^{b/flox}; Sf1:Cre^{gr} gonads when compared to XX control gonads (Fig. 8A), suggesting that depletion of β-catenin activation in Sf1-positive cells does not impair germ cell proliferation. Moreover, histological analysis demonstrated that while XY gonocytes were arrested in G0/G1 phase, oogonia in both wild type XX and XX Catnb^{b/flox}; Sf1:Cre^{gr} gonads initiated meiosis before becoming apoptotic (Fig. 8B). These data and others [63] showed that depletion of β-catenin activation in Sf1-positive somatic cells impairs ovarian somatic differentiation but does not affect germ cell proliferation and meiosis.

Altogether, these data suggest that defects in germ cell proliferation and meiosis in XX Rspo1^{-/-} gonads are not an indirect consequence of ablating β-catenin signaling in the somatic cells, but are rather due to a direct action of RSPO1 to activate β-catenin signaling within germ cells to promote oogenesis in the fetal ovary.
RSPO1 Promotes Germ Cell Sexual Differentiation

A-

| XY; βcat fl/+ | XX; βcat fl/+ | XX; βcat fl/fl, Sf1:CreTr |

MVH  
BrdU  

E12.5

% GC BrdU+

XY βcat fl/+  
XX βcat fl/+  
XX βcat fl/fl, Sf1:CreTr

B-

H&E  

E17.5

% apoptotic GC

p<0.001

C-

SOX9  

E18.5

100µm
Figure 8. Somatic ablation of β-catenin does not affect germ cell fate. A- Proliferation is not impaired in XX Catnb flox/flox; Sf1Cre + /- gonads. Immunodetection of the proliferating germ cells with BrdU (green) and MVH (red) in XY, XX control and XX Catnb flox/flox; Sf1Cre + /- gonads at E12.5. DAPI (blue); nuclei. Histograms: Percentage of BrdU-positive germ cells in XY, XX control and XX Catnb flox/flox; Sf1Cre + /- gonads at E12.5. Bars represent mean±1 SEM, n = 24 sections of each genotype. B- Meiosis in not hampered in XX Catnb flox/flox; Sf1Cre + /- gonads. Haematoxylin & Eosin staining of sections from E17.5 XY, XX control and XX Catnb flox/flox; Sf1Cre + /- gonads. Arrowheads: gonocytes, arrows: meiotic germ cells, dotted arrows: apoptotic germ cells. Histograms: Percentage of apoptotic germ cells versus total germ cells after TUNEL staining in XY, XX control gonads and XX Catnb flox/flox; Sf1Cre + /- gonads. Bars represent mean±1 SEM, n = 24 sections of each genotype. C- Immunodetection of SOX9 (red) in XY, XX control gonads and XX Catnb flox/flox; Sf1Cre + /- gonads at E18.5. DAPI (blue) was used to detect nuclei.

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Discussion

In mice, germ cell differentiation is influenced by differentiation of the somatic environment. Although XX and XY germ cells go through similar processes of maturation (proliferation and meiosis), they are regulated in a sex-specific manner. Rspo1 encodes an activator of the WNT/β-catenin signaling pathway that is secreted by gonadal somatic cells and is required for ovarian differentiation [6,40,43]. In ovariies, Rspo1 is up-regulated from E11.5 onwards [7]. In previous studies, we have shown that loss-of-function of Rspo1 induces not only somatic sex reversal but also germ cell masculinization [6]. In XX Rspo1 -/- gonads, cell-cell adhesions involving germ cells show a similar organization as in XY gonads. These adherens junctions are lost when XX germ cells enter meiosis in ovariies [64]. This seems to be linked to the down-regulation of Wnt4 expression since this phenomenon has also been described in XX Wnt4 -/- gonads [65]. In addition, germ cell sex reversal was observed in XX Rspo1 -/- gonads [6]. We now have shown that germ cell defects in XX Rspo1 mutant gonads are independent of Sertoli cell differentiation. The first testis-like structures that appear in XX Rspo1 -/- gonads are the coelomic vessels and steroidogenic lineages expressing Cyp11a1-P450scc [6]. Although there is no evidence suggesting that endothermal or steroidogenic cells are involved in germ cell proliferation and meiotic defects in XX Rspo1 -/- gonads, we cannot exclude completely such a hypothesis.

In the embryonic ovary, germ cells are firstly dividing before entering meiosis. Primordial germ cell proliferation during their migration towards the gonads in both sexes [66]. Once within the gonads, the germ cell intrinsic factor RNA helicase MVH is only required for XY germ cell proliferation while it is expressed in germ cells of both sexes [18]. Indeed, XX germ cells in a testicular environment do not proliferate as efficiently as XY germ cells in the testis or XX germ cells in the ovary [67]. This suggests that sex specific signalling pathways are not only required for sexual differentiation of germ cells [68] but are also involved in germ cell proliferation once they have reached the gonads.

Rspo1 is produced by ovarian somatic cells and induces functional β-catenin activation in germ cells in a paracrine manner. In XX Rspo1 -/- gonads, down-regulation of β-catenin in germ cells is associated with defects of their proliferation, inhibition of meiosis and partial sex-reversal of germ cells. Since Rspo1 is not expressed in the environment of XY germ cells [7] and β-catenin is not activated in male fetal germ cells, this molecular pathway appears to be female specific. Thus male and female fetal germ cells seem to have acquired independent mechanisms to regulate their proliferation and differentiation, with Rspo1 and WNT/β-catenin signaling holding a key position within the female lineage.

However, when germ cell proliferation resumes in the postnatal testis prior to entry into meiosis, it is associated with activation of WNT/β-catenin signaling but not with Rspo1 expression in spermatogonia [69] and our results (Fig.S2). Indeed, Axin 2+/LacZ reporter expression can be detected in the postnatal XX testes at P0 when germ cells resume mitosis and a robust Axin 2+/LacZ reporter expression is present in XY postnatal testes at 12 days after birth (Fig.S1). Immunostaining with markers for germ cells or Sertoli cells whose cytoplasm surrounds the germ cells at this stage (MVH and SDMG1 respectively) identify the Axin 2+/LacZ-positive germ cells exclusively as germ cells. As parsimony predicts that the same function of β-catenin is conserved in germ cell differentiation in both sexes, WNT/β-catenin signaling may be involved in cell cycle progression in pre-meiotic proliferation in both sexes. Whereas the function of β-catenin remains to be elucidated in spermatogonia, the extracellular signals regulating germ cell proliferation in these tissues are likely to be different.

In addition to proliferation, Rspo1 is also involved in oogonial differentiation and entry into meiosis. Ectopic activation of β-catenin in XY fetal germ cell increases germ cell proliferation, and prevents germ cells from becoming quiescent, but does not allow them to enter meiosis. Since these mutant germ cells are surrounded by Sertoli cells in a developing testis, Sertoli cell-derived factors such as FGF9 may prevent them from entering meiosis. These conflicting messages acting on the germ cells with β-catenin as a meiosis stimulating factor and Sertoli cell-derived meiosis preventing factors may lead the germ cells to become apoptotic.

In XX gonads, entry into meiosis has been linked to RA signaling, which induces the expression of Stra8 [22,23]. However, it has been reported that Stra8 remains expressed in XX gonads devoid of RA synthesis and signalling and that Stra8 expression and entry into meiosis may be stimulated by an alternative mesonephros-derived substrate of CYP26b1 [25]. At E12.5, the remaining expression of Cyp26b1 in XX Rspo1 -/- gonads may participate in Stra8 repression by metabolizing this mesonephric substrate. Our data suggest that the Rspo1/β-catenin signaling pathway is also involved in entry into meiosis. It would be interesting to investigate whether expression of the Rspo1/β-catenin signaling pathway in the female gonad is induced by the putative Raidl-independent meiosis inducer that has been proposed to originate in the mesonephros [25].

The penetrance of the meiotic blockage in XX Rspo1 -/- gonads is not complete indicating that β-catenin is probably not the only germ cell intrinsic factor involved in the regulation of Stra8 expression, as germ cells in XX Rspo1 -/- gonads enter meiosis. At E14.5, Nanos2 is expressed in XX Rspo1 -/- gonads suggesting that this meiosis preventing factor [39] is involved in Stra8 down-regulation at this stage. However, Nanos2 is not expressed at E12.5 in XX Rspo1 mutant gonads (data not shown), thus it cannot be involved in the initial repression of Stra8 and subsequent meiosis blockage nor in proliferation defects.

Rspo1 is an activator of β-catenin both in germ cells and somatic cells and thus it is difficult to uncouple the germinal and somatic effect of Rspo1. However, depletion of β-catenin signaling in the SfiI positive somatic cell lineage did not affect either germ cell proliferation or meiosis in XX Catnb flox/flox; Sfi1:Cre + /- ovaries (our data and [63]). This suggests that the germ cell defects observed in XX Rspo1 -/- gonads are rather due to impairment of β-catenin signaling within germ cells. We cannot exclude that the germ cell proliferation defect, impaired entry into
meiosis and germ cell sex reversal in XX Rspo1 

mutants are an indirect consequence of loss of Rspo1-mediated b-catenin signalling in a Sf1-negative expressing cell type in the gonad. However, any indirect effect of Rspo1 deficiency on XX germ cells will have to occur in the absence of any detectable Sertoli cell differentiation. Given that b-catenin signalling is active in female germ cells in a sex-specific and Rspo1-dependent manner, and that these germ cell phenotypes are observed prior to detectable Sertoli cell differentiation, and do not occur in XX Catnbex3/flox; Sf1:CreTR gonads, we favour the interpretation that loss of Rspo1-mediated b-catenin signalling in the germ cells themselves is responsible for the germ cell proliferation defect, impaired entry into meiosis and germ cell sex reversal in XX Rspo1 

mutants. At present, how b-catenin promotes Stra8 expression and represses Nanos2 in XX germ cells remains to be elucidated (See Model in Fig. 9).

Whereas germ cell sex reversal is explained by regulation of key genes like Stra8 and Nanos2 in XX germ cells, it is surprising that Sertoli cell differentiation occurs so late, i.e. after sex reversal of germ cells in XX Rspo1 mutant gonads. One possible explanation is that the differentiation of gonocytes in XX Rspo1 

gonads influences masculinisation of the surrounding somatic cells promoting them to differentiate into Sertoli cells. In support of this hypothesis, germ cells can have a masculinizing effect on gonad development in some mouse models for sex-reversal [56,70]. It would be interesting to determine whether signalling molecules such as prostaglandin D2 play a role in masculinisation of the somatic cells in XX Rspo1 

gonads [56].

It is now clear that somatic sex determination is regulated by a tight balance between two different pathways [33], the SRY/ SOX9/FGF9 pathway for testis development and Rspo1/

WNT/b-catenin for ovarian differentiation [71]. Our data suggest that a similar balance exists for the sexual differentiation of germ cells and we have shown that Rspo1/b-catenin signalling has a key function in oogonial differentiation by regulating the proliferation and meiosis initiation of the XX germ cells.

**Materials and Methods**

Mouse strains and genotyping

The experiments here described were carried out in compliance with the relevant institutional and French animal welfare laws, guidelines and policies. These experiments have been approved by the French ethics committee Comité Institutionnel d’Ethique Pour l’Animal de Laboratoire (number: NCE/2011-12). All mouse lines were kept on a mixed 129/C57BL6/J background (4 to 5 backcrosses on C57BL6/J background, a purer background results in unhealthy animals). Rspo1 

and Axin2/LacZ transgenic mice were previously described in [6,52]. TnAP:CreTR mice [39] were mated with mice carrying the b-catenin exon3 floxed allele (Catalb3/+) [58] to obtain Catalb3/++; TnAP:CreTR embryos.

Sf1:CreTR mice (kind gift from Keith Parker) [62] were mated with mice carrying the b-catenin conditional knock out allele (Catalbex3/+; Catnbex3/flox) [58] to obtain Catalbex3/++; TnAP:CreTR embryos. Oct4-GFP embryos were generated by breeding Oct4-GFP homozygous transgenic male mice (a kind gift from Jenny Nichols, CSCR, Cambridge, UK) with CD1 female mice. Embryonic samples were collected from timed matings (day of vaginal plug = E0.5). Genotyping was performed using DNA extracted from tail tip of embryos or ear biopsies of mice with published

![Figure 9. Opposing signals regulate germ cell sexual differentiation.](http://www.plosone.org/doi/10.1371/journal.pone.0025641.g009)
primers related to the strains. The presence of the Y chromosome (Sry PCR) was determined as described previously [5]. Pax6 primer set 5′ GCAACAGGAAAGGGGAGA 3′; 5′ CGTTCTTC- CAGAGCTCATTGT 3′ was included in each PCR reaction as an internal control.

Germ cell isolation

For FACS sorting. Urogenital ridges (gonad and mesonephros) were dissected from E13.5 Oct4-GFP embryos, sorted into testes or ovaries, and trypsinised to single-cell suspensions. The cells were then resuspended in ice-cold PBS and sorted for GFP fluorescence using a BD FACSaria II cell sorter. GFP-positive cells (germ cells) and GFP-negative (somatic cells) were isolated for each sex.

For XGal/immunostaining. Urogenital ridges were dissected from XY and XX E12.5 Axin2+/+/-/lacZ gonads and germ cells were isolated according to [56]. Germ cells were allowed to adhere on a 12 mm cover glass pre-coated with a 0.1 mg/mL solution of poly-L-Ornithine (Sigma cat P4957) for 2 hours at RT. They were then processed for XGal staining according to [73]. After 12 hours of staining at 37°C, they were washed in PBS, fixed in 4% PFA for 10 minutes and then processed for DDX4/MVH immunostaining (see below).

Histological analysis

This technique was performed as described previously [6]. Urogenital organs were dissected, fixed in Bouin’s solution overnight, and processed to obtain 5 mm or 3 mm thickness paraffin sections. For each genotype, 5 sections of 3 different embryos were processed for Haematoxylin and Eosin staining and quiescent germ cells were analysed with a light microscope, using a 100× objective. The quiescent state was identified by a uniform size of the gonocytes and the absence of any heterochromatin in the nuclei whereas the proliferative state was characterized by the variation in nuclear size, perinuclear heterochromatin (late S phase and G2 phase), and appearance of chromosome condensation and chromosomal threads (mitosis). In the present study, in the counts it was determined whether or not the cells were in one of the phases of a mitotic division. All the sections obtained from 2 XY wild type gonads and 6 XY Cebd+/+/-; TNAxis/Cit+/+ gonads were used for the quantification. Pictures were taken with an AxioCam mrm camera (Zeiss) and processed with Adobe Photoshop.

X-gal staining and immunological analyses

These techniques were performed as described previously [6]. Embryonic samples were fixed with 4% paraformaldehyde overnight and then processed for paraffin embedding. Embryonic samples for cryosection were successively fixed 2 hours in 4% paraformaldehyde, washed in cold PBS, equilibrated in 10% sucrose during 3 hours, then in 30% sucrose overnight at 4°C, embedded in Cryomount (Histolab) and stocked at −80°C before cryosection. For each genotype, 5 cryostat or microtome sections of 8 μm thickness of 2 to 3 different embryos were processed for XGal staining or/and immunostaining. The following dilutions of primary antibodies were used: DDX4/MVH (cat 13840, Abcam), 1:200, γH2AX (cat 16-193, Upstate) 1:300, SCP3 (ab15091, Abcam) 1:50, SOX9 (kindly provided by Michael Wegner) 1:1500, active β-catenin (anti-ABC, clone 8E7, cat 05-665, Millipore) 1:200, FOXL2 (ab5096, Abcam), 1:250 and SDMG1 (generated in [32]) 1:2000. DAPI (blue) was used to detect nuclei. For histology, 5 μm thickness sections of 2 embryos of each genotype were stained with Hematoxylin & Eosin. Fluorescent studies were performed with a motorized Axio Imager Z1 microscope (Zeiss), and pictures were taken with an AxioCam mrm camera (Zeiss) and processed with Adobe Photoshop.

Counting of proliferating, apoptotic or meiotic germ cells

Urogenital organs were dissected and fixed in 4% paraformaldehyde, and processed to obtain 5 μm paraffin sections as described above.

Counting of meiotic germ cells. For each genotype, 5 sections of 3 different embryos were processed for immunohistological experiments with DDX4/MVH antibody, and SCP3 and γH2AX at respectively E14.5 and E16.5. Then the percentage of meiotic germ cells versus total germ cells (MVH positive) was determined. Both γH2AX and MVH-positive germ cells were quantified on the same sections after a co-immunostaining, whereas SCP3-positive and MVH-positive germ cells were independently quantified on consecutive sections since a co-immunostaining was not technically possible.

Counting of proliferating and apoptotic germ cells. For each genotype, 8 sections of 3 embryos (24 pictures per genotype) were used for immunohistological experiments with DDX4/ MVH antibody. Then proliferation analysis was performed on the same sections by way of 5-Bromo-2’-deoxy-Uridine labelling and detection using an appropriate kit (cat 11 296 736 001, Roche) at E12.5. TUNEL analysis was performed with the In Situ Cell Death Detection kit, TMR red (cat 11 684 795 910, Roche) at E14.5 and E16.5. Total germ cells, proliferating (both BrdU- and MVH-positive cells) and apoptotic (both TUNEL- and MVH-positive cells) were quantified on the entire section using ImageJ software with a macro designed by Cédric Matthews (platform of microscopy, IBDC Nice). For each picture, the number of BrdU-positive/TUNEL-positive germ cells and the number of total germ cells (MVH positive) were counted. Then the percentage of BrdU-positive/TUNEL-positive germ cells versus total germ cells (MVH positive) was determined. For each genotype (~24 pictures), the mean and mean±1 SEM of these percentages were calculated and reported on a graph after statistical analysis (for details, see paragraph Statistical Analysis above).

In situ hybridization

In situ hybridization was carried out as described previously [6]. Stra8, Cyp26b1, Oct4 and Fgf9 riboprobes were a kind gift of David Page, Peter Koopman, and Jennifer Colvin respectively. Digoxigenin riboprobe for Nanos2 was generated by amplifying a cDNA fragment by RT-PCR from Nanos2 (NM_194064, bases 126-696) and inserting it into TA cloning vector pCR2.1- TOPO (Invitrogen). The plasmid was then linearized by BamHI and transcribed with T7 RNA polymerase in the presence of Dig-labeling mix (Roche).

Quantitative PCR analysis

Individual gonads without mesonephros were dissected in PBS from E12.5 and E14.5 embryos and immediately frozen at −80°C. RNA was extracted using the RNAeasy Qiagen kit, and reverse transcribed using the RNA RT-PCR kit (Stratagene). Primers and probes were designed by the Roche Assay design center (http://www.rocheappliedscience.com/sis/rtPCR/upl/adc.jsp). Primers: Hprt1: 5’-ctcctccgacgacgttt-3’ and 5’-cctggttcat- cgcataac-3’ (probe 95), Mvh: 5’-ccagcatgacaggcaatg-3’ and 5’-ctggttagttcacgttc-3’ (probe 77), Nanos2: 5’-agctccctcaca-3’ and 5’-cagccacagagaggctg-3’ (probe 98), Dmrt3L: 5’-aaccggacaggtgaaa-3’ and 5’-cgctctgagtgaggt-3’ (probe 43), Tsid5: 5’- agcagcagcaagaggg-3’ and 5’-taacccctgtttc- catta-3’ (probe 102). RARs: 5’-aagcagcagacgggtg-3’ and 5’-cctccgacgacgac-3’.
**Supporting Information**

**Figure S1** Control experiments for the purification of FACS sorted cells. Quantitative RT-PCR analysis of *Meh*, *Bmp2*, *Fst* and *Rspo1* expression in E15.5 germ cells (XX and XY Oct4-positive cells) and somatic cells (XX and XY Oct4-negative cells), using *Hpo1* as the normalization control. Bars represent mean±1 SEM, n = 3 individual experiments. (TIF)

**Figure S2** β-catenin signalling pathway is activated postnatally in XY proliferating germ cells. Upper panel: X-Gal staining ([AXIN2] and immunostaining with MVH (germ cells) or SDM1 (Sertoli cells) in XY and XX *Axin2*Δ/Δ gonads) at P0 and P12. Black arrowheads: germ cells. Lower panel: *Rspo1 in situ* hybridization at P0 in XY gonads and epididymis (positive control, inset). (TIF)

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**Author Contributions**

Conceived and designed the experiments: AAC IRA MCC. Performed the experiments: AAC EPG RL. Analyzed the data: AAC DGR IRA MCC. Contributed reagents/materials/analysis tools: MT IRA. Wrote the paper: AAC DGR IRA MCC.

**References**