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The \textit{INT6} Cancer Gene and MEK Signaling Pathways Converge during Zebrafish Development

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**Background.** \textit{Int-6} (integration site 6) was identified as an oncogene in a screen of tumorigenic mouse mammary tumor virus (MMTV) insertions. \textit{INT6} expression is altered in human cancers, but the precise role of disrupted \textit{INT6} in tumorigenesis remains unclear, and an animal model to study \textit{Int-6} physiological function has been lacking. **Principal Findings.** Here, we create an \textit{in vivo} model of \textit{Int6} function in zebrafish, and through genetic and chemical-genetic approaches implicate \textit{Int6} as a tissue-specific modulator of MEK-ERK signaling. We find that \textit{Int6} is required for normal expression of MEK1 protein in human cells, and for Erk signaling in zebrafish embryos. Loss of either \textit{Int6} or \textit{Mek} signaling causes defects in craniofacial development, and \textit{Int6} and Erk-signaling have overlapping domains of tissue expression. **Significance.** Our results provide new insight into the physiological role of vertebrate \textit{Int6}, and have implications for the treatment of human tumors displaying altered \textit{INT6} expression.


**INTRODUCTION**

Embryonic development and tumour development often share underlying molecular mechanisms—a concept illustrated by the identification of genes disrupted by the mouse mammary tumor virus (MMTV) in mammary cancers [1]. An important example, the \textit{Int-1} gene which is a common integration site for MMTV in mammary tumours, encodes the homologue of the \textit{Drosophila wingless} gene [2,3] and was subsequently named \textit{Wnt1} (wingless/Int) in recognition of this conserved function. Wnt signaling is now known to be disrupted in many human tumor types, especially colon cancer [4]. Other \textit{Int} genes, such as \textit{Int-2} and 4 (Fgf3, 4), and \textit{Int-3} (Notch4), encode mitogens and regulators of development that are also misactivated in many cancers [1,5].

In the majority of cases, MMTV activates \textit{Int} gene expression as a result of proviral integration upstream of the promoter region. Remarkably, all three MMTV insertions found in \textit{Int-6}, which encodes a component of the eukaryotic translation initiation factor 3 (eIF3), were found to lie within introns, and in the opposite transcriptional orientation to the \textit{Int-6} gene, creating a truncated \textit{Int-6} mRNA [1,6]. Ectopic expression of equivalently truncated \textit{Int-6} can transform cell cultures [7,8], and promote persistent mammary alveolar hyperplasia and tumorigenesis in transgenic mice [9].

Despite important evidence in favor of a role for \textit{INT6} in human tumourigenesis [10–12], the molecular basis for \textit{INT6} in cancer development remains unresolved. Highly conserved in eukaryotes, \textit{INT6} contains a PCI domain, found in proteins of the 19S regulatory lid of the proteasome, the COP9 signalosome (CSN), and the eIF3 translation initiation complex; all three complexes share overall structural similarity, and \textit{INT6} has been found associated with each [13]. When over-expressed in yeast, \textit{Int6} induces multi-drug resistance by activating an AP-1 transcription factor [14,15], and in human cells, the range of \textit{INT6} function includes orderly progression through mitosis [16], regulation of the proteasome-dependent stability of MCM7 [17] and HIF2z [18], and nonsense mediated mRNA decay [19].

With no animal model for \textit{Int6} loss-of-function available, we reasoned that an understanding of \textit{INT6} during development would provide novel insight into \textit{INT6} function in normal vertebrate cells, thereby providing a new perspective on \textit{INT6} function in cancer formation. Here, using zebrafish and mammalian cells, we describe the first \textit{Int6} loss-of-function phenotype in an animal, and link \textit{Int6} with a signaling pathway, that like those effected by other \textit{Int} genes, is critical for both development and cancer.

**RESULTS**

\textit{Int6} is essential for zebrafish embryogenesis

We chose to study the physiological role of zebrafish \textit{Int6} during development, using morpholino oligonucleotides (MOs) to reduce \textit{Int6} protein, as well as an \textit{int6} knock-out insertion mutant line (kindly provided by N. Hopkins, A. Amsterdam and S. Farrington, M.I.T.). Zebrafish \textit{Int6} is over 90% identical in its amino acid sequence to human \textit{INT6} (\textit{Ensembl} ENSDARG00000002549) and using an \textit{Int6} antibody raised against the N-terminus of the human

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INT6 [20] we determined that the int6 MO resulted in loss of Int6 (Figure 1A). As INT6 has been implicated in G2/M-phase cell cycle control, we first performed whole-mount immunohistochemistry with the late G2/M phase marker, phospho-histone H3, and found only slightly reduced numbers of cells in late G2/M phase in the int6 morphant compared to the control (Figure S1). Importantly, we found that embryos injected with int6 MO had specific developmental defects (Figure 1B–N), most notably reduced melanisation 2 days post-fertilization (dpf: int-6 MO n = 51/53; con MO n = 0/35; int-6 5MM n = 3/31); misplaced pigment cells in the tail 3 dpf (int-6 MO n = 46/49; con MO n = 3/30); and abnormal jaw morphogenesis, with cartilage elements reduced or malformed at 4 and 5 dpf (int-6 MO n = 81/85 4 dpf, n = 76/83 5 dpf; con MO 1/67 4 dpf, n = 1/61 5 dpf). The craniofacial and pigment cell defects observed in the int-6 morphant and hi2470 mutant suggest that int-6 might contribute to development of neural crest-cell (NCC) derivatives. We used multiple markers of NCCs and their derivatives to assess when these phenotypes arise, and found Int6 did not appear to be required for the specification or organization of premigratory and migrating cartilage precursors (Figure S2). In contrast, alcian blue cartilage staining revealed a specific loss of the five ceratobranchial cartilage elements in the int6 morphants, whereas Meckel’s, palatoquadrate, and hyoid cartilage were all present, albeit misshapen (5.5 dpf int6 MO n = 45/53; con MO n = 1/34; Figure 1I–L, Figure S3). Expression of int6 mRNA restored normal craniofacial elements to the int6 morphant (data not shown); and the int6hi2470 mutant had an almost identical craniofacial phenotype (Figure 1M, N) indicating a genuine requirement for Int6 in craniofacial development.

Loss of Int6 alters MEK protein and Erk signaling
Biochemical evidence in fission yeast suggests that Int6 is part of a specialized eIF3 translation initiation complex that may target specific mRNAs for translation [21]. Given the involvement of INT6 in cell proliferation [16], western blots using a panel of antibodies against proteins involved in the cell cycle and associated signalling pathways were performed using lysates from control and INT6 siRNA transfected MDA-MB-231 cells. Of 16 proteins investigated in this way, only MEK1 levels were altered by INT6 siRNA transfection (Figure 2). As previously reported [20], we found that Int6 siRNA cell lysates had reduced levels of INT6 protein compared with the untransfected and reverse INT6 siRNA sequence. We also found a dramatic reduction of MEK1 protein

Figure 1. Int6 is essential for zebrafish embryonic development. (A) Western blot analysis of Int6 (*) in zebrafish embryos injected with a control (con) MO or int6 MO. (B) int6 morphant melanocytes are less darkly pigmented. (C–F) Int6 is required for pigment cell placement in the tail, as int6 morphants and int6hi2470 mutants have misplaced pigment cells in the tail fin (D, arrow). Ambient light illuminates the iridophore ‘star-light’ pattern seen in the int6hi2470 embryos (F, arrow). (G–H) By 5 dpf, embryos injected with a con MO have clearly visible ceratobranchial arches, while int6 morphants do not have visible ceratobranchial arches, in addition to other abnormalities, including unconsumed yolk sac, heart and eye development. (I–N): Alcian blue staining of 5 dpf embryos shows loss of ceratobranchial arches 1 through 6. M, Meckel’s; PQ, palatoquadrate; CH, ceratohyal; CB, ceratobranchial. doi:10.1371/journal.pone.0000959.g001

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Figure 2. INT6 is required for MEK protein levels and Erk-signaling. A. Osteosarcoma U2-OS cells untransfected (U), or transfected with siRNA targeted against the INT6 mRNA (si) or the reverse sequence (R), show reduced levels of INT6 and MEK1 protein specifically after transfection with INT6-siRNA, but no reduction in BAX, tubulin or actin protein levels. (B) Semi-quantitative-PCR shows MEK1 mRNA is unaffected in reverse sequence and INT6-siRNA treated cells, coupled with the expected reduced levels of the INT6 message in the INT6-siRNA transfected cells. c, PCR control without DNA. (C) Phospho-Erk levels are reduced in int6 morphants, while ponceau stain detects equal loading of protein on the gel. doi:10.1371/journal.pone.0000959.g002
levels that correlated with loss of INT6, while BAX, tubulin and actin protein levels appeared unaffected in the INT6-siRNA transfected cells (Figure 2A). The loss of MEK1 was specifically at the protein level, as semi-quantitative-PCR showed normal levels of MEK1 mRNA in INT6-siRNA treated cells, as well as the expected reduced levels of the INT6 message in the INT6-siRNA transfected cells (Figure 2B). The possibility that INT6 may affect MAPK signaling through control of MEK protein levels prompted us to examine the phosphorylation state of Erk1/2, downstream targets of the MeK kinases, in int6 morphant zebrafish embryos. Compared with control MO embryos, int6 morphant embryo lysates had reduced phospho-Erk levels (Figure 2C). These data suggest a novel function for Int6 in the control of MAPK signaling in the developing embryo, possibly by direct control of MEK1 protein levels.

**Int6 and Mek pathways converge during development**

If Int6 controls Mek activity in the developing embryo, we theorized that specific developing tissues might have overlapping expression domains of Int6 protein and phospho-Erk activity. Indeed, immunohistochemistry with antibodies directed against Int6 and phospho-Erk revealed overlapping domains of expression in the developing craniofacial region in 3 and 4 dpf embryos (Figure 3A–F). Strong Int6 tissue-specific expression was also detected in the developing intestine and lens, regions that had little or no phospho-Erk expression (Figure S4). Given the observed phospho-Erk and Int6 expression in the craniofacial region, we hypothesized that some of the Int6 phenotypes, such as the jaw formation defect, might be phenocopied by repression of Erk signaling. As interpretation of MO phenotypes has recently been complicated by the identification of MO-induced p53-dependent craniofacial defects [22], we used an alternative approach – the highly selective, clinically active MEK inhibitor CI-1040 [23] – to reduce Mek signaling in zebrafish. We added the drug at 4 hpf at a concentration of 0.25, 0.5, and 1.0 μM, and confirmed loss of phospho-Erk expression by Western blot analysis (data not shown). Notably, the addition of CI-1040 caused a dose-dependent loss of the posterior structures of the embryo, such that 1.0 μM CI-1040 caused a severe anterior-posterior (AP) axis defect (Figure 4A–C), consistent with a role for FGF signaling in the development of the AP axis [24]. CI-1040 also caused loss of ceratobranchial cartilage elements, while the anterior elements – Meckel’s, palatoquadrate and byoid cartilages – were present but misshapen (Figure 3G–J), similar to the effects seen in int6 morphants and mutants.

To further elucidate the biological relevance of Int6 and Mek signaling, we took advantage of the ease with which signaling pathways can be altered pharmacologically in specific genetic contexts in the zebrafish system. We reasoned that if Int6 contributes to activation of Mek signaling, then embryos with reduced Int6 should be hypersensitive to low doses of the MEK inhibitor CI-1040. In control embryos treated with 0.25 μM CI-1040, no changes in the anterior-posterior axis were detected (Figure 4B). In addition, int6 morphants generated by low doses of MO (0.25 ng) did not have an altered AP axis (Figure 4D). In contrast, in combination with low doses of CI-1040, the low dose int6 morphant showed a severely enhanced AP axis phenotype (Figure 4E). Taken together, these data provide further evidence that int6 may play a role in modulating MEK signaling in vivo.

**DISCUSSION**

Activated in most cancers, the MAPK signaling pathway is among the most attractive targets for novel anti-cancer therapies [23]. Like MAPK signaling pathways, most of the Int pathways - Wnt,
Fgf, and Notch - are conserved regulators of development that are frequently activated to promote oncogenesis. We provide evidence that, like other Int gene products, Int6 is required for vertebrate development (Figure 1), in part by providing a novel layer of MAPK signal transduction regulation (Figure 2). With the wide range of cellular activities attributed to INT6, the mechanistic detail of this control remains to be understood; our early investigations indicate reduction of MEK1 in INT6-siRNA treated mammalian cells is not dependent on the proteasome ([M.G. & C.J.N., unpublished data], making direct MEK1 regulation by INT6-dependent translation a possibility.

Recently, importance of RAS, RAF and MEK in human disease has been extended beyond cancer by the discovery that in MMTV induced mammary tumors, the truncated Int-6 protein may act as an oncogene by altering MEK-ERK signaling. We propose that the diverse cellular locations of Int6, combined with the temporal expression and localization of Mek1/2 and Erk1/2, may result in fine-tuning of Mek-Erk signaling pathways in specific tissues during development, and may have important implications for the role of INT6 in tumorigenesis.

METHODS

Zebrafish husbandry and morpholino studies

Zebrafish (Danio rerio) lines AB, AB*, and AB*-TPL were raised and staged as described [32,33]. MOs (Table 1) were designed by and purchased from Gene Tools, LLC (USA), and 1 ng injected into one-cell stage embryos.

Phenotype analysis

Phenotype analysis were performed as described: cell cycle studies [34]; alcian blue staining [35]; probe synthesis and whole-mount in-situ hybridizations [36]. cDNA probes for neural crest markers [34]; alcian blue staining [35]; probe synthesis and whole-mount in-situ hybridizations [36].
were the kind gift of David Raible (University of Washington, USA). Polyadenylated int6 mRNA was generated using Ambion mMessage mMachine (#1340).

Cell culture and RT-PCR analysis
MDA-MB-231 cells were grown and transfected as described [19] using Lipofectamine (Invitrogen) with si-oligouracilode (Table 1; Eurogentec) at a final concentration of 100 nM in Optimum (Gibco). Forty-eight hours after transfection total RNA was isolated (RNaseasy Mini Kit; Qiagen) and one-step RT-PCR reactions (Qiagen) accomplished using specific primers (Table 1).

Immunoblotting
Whole-cell lysates and zebrafish extracts were generated [31,19] and immunohistochemistry was performed as described [36]. Antibodies used as in Table 2.

SUPPORTING INFORMATION

Figure S1 Cell cycle analysis of int6 morphants. Whole-mount immunohistochemistry with the late G2/M phase marker, phoso-histone H3 shows only slightly reduced numbers of cells in late G2/M phase in the int6 morphant compared to the control. Similarly, DNA content as measured by flow cytometry reveals only a slight reduction of cells in G2/M phase in the int6 morphant. Thus, we find that loss of Int6 in normal vertebrate cells (as well as in additional human cancer cell lines, M.G. & C.J.N, unpublished data) does not appear result in an accumulation of cells in G2/M progression.

Figure S2 Lateral views of in situ hybridization of neural crest markers in control and int6 morphants, revealing no change in cell number or migration as indicated by the apparently normal expression of dinkel (stages 6–36 hpf, examined at two hour intervals), nor of early markers of NCC and melanocytes, such as sox10, cresin, snail and mitfa (24 hpf) in int6 morphants. These observations were extended by examination of a transgenic sot10-GFP line [1] revealing unaltered GFP-expressing NC-derived cells in int6 morphants within the first 48 hpf, but a loss of GFP expressing differentiated pharyngeal arches 3–7 by 3 dpf (data not shown). Found at: doi:10.1371/journal.pone.0000959.s002 (40.41 MB TIF)

Figure S3 Development of the pharyngeal arches in the developing control (A–C) and (D–F) int6 morphant animals. Note the loss of pharyngeal arches (A, bracket) in the int6 morphants (bracket). Sections were stained with methylene blue. Anterior to the left.

Figure S4 Immunohistochemistry of Int6 and phospho-Erk staining in 4 dpf embryos. (A, B) While Int6 and phospho-Erk signaling overlap in the craniofacial region, they also have distinct patterns, for example in the eye and (C, D) gut. We note that while Int6 and phospho-Erk have overlapping domains of expression in the craniofacial region, Int6 staining in the craniofacial region was stronger than phospho-Erk, and phospho-Erk staining was limited to specific tissues within the craniofacial region. M, Meckel’s; E: Ethmoid plate; CH, ceratohyal; CB, ceratobrancial. Sagittal section, anterior to the left.

Figure S5 Immunohistochemistry of Int6 and phospho-Erk staining in 4 dpf embryos. (A, B) While Int6 and phospho-Erk signaling overlap in the craniofacial region, they also have distinct patterns, for example in the eye and (C, D) gut. We note that while Int6 and phospho-Erk have overlapping domains of expression in the craniofacial region, Int6 staining in the craniofacial region was stronger than phospho-Erk, and phospho-Erk staining was limited to specific tissues within the craniofacial region. M, Meckel’s; E: Ethmoid plate; CH, ceratohyal; CB, ceratobrancial. Sagittal section, anterior to the left.

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Author Contributions
Conceived and designed the experiments: EP CN MG. Performed the experiments: EP JM CA MG DW. Analyzed the data: JA EP CN. Contributed reagents/materials/analysis tools: RK EP CN. Wrote the paper: EP.

REFERENCES


