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1,25-Dihydroxyvitamin D$_3$ Restrains CD4$^+$ T Cell Priming Ability of CD11c$^+$ Dendritic Cells by Upregulating Expression of CD31

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Dendritic cells (DC) are specialized sentinel cells that bridge the innate and adaptive immune response and play a crucial role in shaping the adaptive immune response. Vitamin D, a known epidemiological risk factor for the development of several autoimmune diseases, influences the development of dendritic cells. Consequently, vitamin D metabolites are frequently used in protocols to develop therapeutic dendritic cell therapies for autoimmune diseases. However, the mechanisms by which vitamin D modulates DC function remain poorly understood. We investigated the effects of vitamin D on murine CD11c$^+$ bone marrow derived DC (BMDC) function by analyzing global gene expression in CD11c$^+$ BMDC generated in the presence (VitD-CD11c$^+$ BMDC) or absence (Veh-CD11c$^+$ BMDC) of the active vitamin D metabolite, 1,25-dihydroxyvitamin D$_3$ (1,25(OH)$_2$D$_3$). Seven genes were significantly increased in expression in both immature and LPS-matured VitD-CD11c$^+$ BMDC generated in the presence (VitD-CD11c$^+$ BMDC) or absence (Veh-CD11c$^+$ BMDC) of the active vitamin D metabolite, 1,25-dihydroxyvitamin D$_3$ (1,25(OH)$_2$D$_3$). Seven genes were significantly increased in expression in both immature and LPS-matured VitD-CD11c$^+$ BMDC, one of which was CD31, a member of the immunoglobulin superfamily. Gene knockdown of CD31 enhanced the ability of VitD-CD11c$^+$ BMDC to prime naïve CD4$^+$ T cells in vitro; conversely, increased expression of CD31 on vehicle treated CD11c$^+$ BMDC restrained their T cell priming abilities. Time-lapse imaging of BMDC and CD4$^+$ T cells during in vitro priming revealed that CD31 reduced the BMDC–T cell interaction time. Finally, we confirmed a similar effect of 1,25(OH)$_2$D$_3$ on human CD34$^+$ cell-derived CD11c$^+$ DC, whereby DC generated in the presence of 1,25(OH)$_2$D$_3$ had increased CD31 expression. In summary, we show that both mouse and human DC generated in the presence of 1,25(OH)$_2$D$_3$ upregulate CD31 expression, resulting in a reduced ability to prime CD4$^+$ T cells by impairing a stable cell-cell contact.

Keywords: dendritic cells, vitamin D, 1, 25(OH)$_2$D$_3$, autoimmune disease, CD31, T cell priming
INTRODUCTION

Dendritic cells (DC) are professional antigen presenting cells which play a crucial role in shaping the adaptive immune response (1). They have the ability to either tolerize or activate T cells depending on their activation status and concomitant expression of co-stimulatory or inhibitory molecules. In the absence of activation, antigen presentation by steady-state DC can lead to T cell unresponsiveness and tolerance (1). In the presence of co-stimulatory molecules, antigen presentation by DC results in T cell activation.

A wide range of factors can influence the phenotype and function of DC. One of the most widely studied molecules known to alter the development of DC is the active vitamin D metabolite, 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃) (2–5). The interest in probing the effects of vitamin D on DC function has, in part, been due to the wealth of data which now links suboptimal vitamin D status to the development and progression of many autoimmune diseases (6–8). Several groups have demonstrated that DC generated in the presence of 1,25(OH)₂D₃ from either bone marrow cells or monocytes have a reduced ability to prime T cells in vitro and in many experimental systems can tolerize T cells (9–12). These findings have led to the development of clinical trials of tolerogenic 1,25(OH)₂D₃ conditioned DC in human patients with autoimmune conditions such as rheumatoid arthritis and multiple sclerosis (5, 13–15).

However, the mechanisms by which 1,25(OH)₂D₃ manipulates the phenotype of DC remain incompletely understood. We, and others, have shown that the addition of 1,25(OH)₂D₃ to bone marrow cell cultures leads to the generation of BMDC which have lower MHC class II expression alongside reduced expression of co-stimulatory molecules such as CD80 and CD86 (16, 17). Given the widespread impact that 1,25(OH)₂D₃ can have on immune cells, it would appear likely that additional co-stimulatory or inhibitory pathways may be influenced by exposure to 1,25(OH)₂D₃. To explore this further we performed a global gene expression analysis on CD11c⁺ BMDC generated in the absence (Veh-CD11c⁺BMDC) or presence of 1,25(OH)₂D₃ (VitD-CD11c⁺BMDC). We focused our attention on CD11c⁺ cells for two key reasons; firstly, CD11c⁺ cells are known to have potent antigen presenting capacity and secondly, the addition of 1,25(OH)₂D₃ is known to lower the proportion of CD11c⁺ in murine BMDC cultures (16, 17). Consequently, we wanted to evaluate gene expression in cells which have the capacity to prime antigens and did not want to confound our data by including cells which were CD11c⁻ and did not express MHC class II molecules.

Here, we present microarray results on this defined population which demonstrate that the addition of 1,25(OH)₂D₃ resulted in a large number of differentially expressed genes. Specifically, we discovered that CD31 was one of only seven genes whose expression was upregulated in both immature and LPS-matured VitD-CD11c⁺BMDC. CD31 is a 130-kDa member of the immunoglobulin superfamily, a single-chain transmembrane glycoprotein with six C2-type Ig-like extracellular domains, and a cytoplasmic tail containing two immunoreceptor tyrosine-based inhibitory motifs (ITIMs) (18, 19). CD31 is concentrated at endothelial tight junctions where it supports endothelial cell layer integrity (20), and is also expressed at lower levels on platelets and most leukocytes (21). CD31 mostly facilitates cell-cell adhesion via trans-homophilic interactions (22, 23), but has also been reported to interact in a heterophilic manner via CD177 (24), αvβ3 (25), glycosaminoglycans (26), and CD38 (27). Not surprisingly, CD31 has been implicated in mediating leukocyte migration across the endothelial cell layer (28), but has also drawn attention as a potential immunomodulatory molecule important for communication between immune cells, e.g., as a detachment signal between live neutrophils and macrophages (29), and as a co-inhibitory molecule on T cells (21) and DC (30). Very little is known about the factors which regulate CD31 expression in immune cells.

Here, we present data revealing 1,25(OH)₂D₃ as a potent inducer of CD31 expression on BMDC, and identify increased CD31 levels on BMDC as a novel mechanism by which 1,25(OH)₂D₃ restrains the ability of BMDC to prime naïve CD4⁺ T cells.

MATERIALS AND METHODS

Mice, Antigens, and Tissue Culture Medium

B10.PLxC56BL/6 (CD45.2) and Tg4 (CD45.1) mice were bred under specific pathogen-free conditions at the University of Edinburgh. All experiments had local ethical approval from the University of Edinburgh’s Animal Welfare and Ethical Review Body and were performed in accordance with UK legislation. All mice used in the experiments reported were female as this allowed for standardization of experiment groups and permitted the housing of mice from different litters in the same experimental cage. The mice were maintained in individually ventilated cages, and were between 8 and 12 weeks old when used for experiments. The housing facility was compliant with Federation of European Laboratory Animal Science Associations guidelines on screening mice for infectious diseases. Tg4 mice express a transgenic T cell receptor (TCR) recognizing the Ac1-9 peptide of myelin basic protein (MBP) in association with I-A^α (31). The MBP Ac1-9 (4Tyr) analog peptide was obtained from Cambridge Research Biochemicals (Teesside, UK). To obtain cell culture medium, RPMI 1640 medium was supplemented with 2 mM L-glutamine, 100 U/ml penicillin, 100 μg/ml streptomycin, and 5 × 10⁻⁵ M 2-ME (all from Gibco, Paisley, UK) and 10% heat-inactivated FCS (Labtech, East Sussex, UK).

Generation and Stimulation of Murine BMDC

BMDC were generated in the presence of recombinant GM-CSF (Peprotech, London, UK) for 9 days as previously described (32). Briefly, bone marrow was collected from femurs and tibias of B10.PLxC56BL/6 mice, and clusters within the bone marrow suspension were dispersed by vigorous pipetting. Red
blood cells were lysed with RBC lysis buffer (Sigma-Aldrich, Dorset, UK) for 1 min and cells subsequently washed once in cell culture medium. Cells were seeded into the center of 6 well plates at 2 × 10^5/ml in 2 ml of cell culture medium with the addition of 20 ng/ml GM-CSF. At day 3, a further 2 ml of cell culture medium containing 20 ng/ml GM-CSF was added to each well. On day 6, 2 ml of culture supernatant was removed and replaced with 2 ml fresh culture medium containing 20 ng/ml GM-CSF. Vehicle (100% ethanol) or 1,25(OH)_2D_3 was added to the BMDC culture media at the concentration indicated in the text initially and at all subsequent media changes. At day 9, the BMDC cells were harvested and CD11c^+ cells were isolated by FACS (Supplementary Figure 1). The CD11c^+ BMDC were replated at 2 × 10^6 BMDC/ml in cell culture medium, 5 ng/ml of GM-CSF with 0.1 µg/ml Ac1-9(4Tyr) MBP for an additional 18 h. 0.1 µg/ml lipopolysaccharide (LPS) was added to some overnight CD11c^+ BMDC cultures (Sigma-Aldrich, Dorset UK). No 1,25(OH)_2D_3 was added to the culture during the overnight stimulation with LPS and MBP. Cytokines were measured in BMDC supernatants by Ready-SET-Go ELISA as per manufacturer’s instructions (eBioscience, San Diego, USA).

**Generation of Human DC From Mobilized Peripheral Blood CD34^+ Progenitor Cells**

CD34^+ cells were obtained from a commercial source (AllCells, LLC, Alameda, CA, USA). The CD34^+ cells had been isolated by positive selection with an immunomagnetic bead system, from volunteer mobilized peripheral blood CD34^+ stem/progenitor cell donations following informed consent. CD34^+ progenitors were cultured in Iscove Basal Medium (Biochrom) with BIT 9500 Serum Substitute (Stemcell Technologies), 100 ng/ml SCF (Pharmacy), 100 ng/ml FLT3L (Peprotech), and 50 ng/ml TPO (Peprotech). Medium was replenished every 4 days by addition to cultures and splitting into fresh flasks to maintain optimum culture conditions.

After 3 weeks, expanded cells were differentiated to DC by culture in TexMACS medium (Miltenyi Biotech) supplemented with 50 ng/ml GM-CSF (Peprotech) and 15 ng/ml IL-4 (Peprotech), in the presence of 20 nM 1,25(OH)_2D_3 or vehicle (100% ethanol). Seven days later, cells were analyzed by flow cytometry.

**RNA Extraction and Microarray Analysis of BMDC**

RNA was extracted from CD11c^+BMDC using commercially available kits (RNeasy Mini Kit). RNA integrity number (RIN >8) was assessed using the 2100 Bioanalyzer and RNA 6000 Pico kit (Agilent, CA, USA). Sense strand cDNA preparation from RNA samples was generated and labeled using Ambion WT expression kit (Life Technologies, Paisley, UK) and Affymetrix GeneChip-WT terminal labeling kit according to manufacturer’s instructions. Samples were hybridized to Affymetrix Mouse Gene 2.1 ST 16-Array Plate (Affymetrix, CA, USA). Raw data was normalized using robust multichip average method and principal component analysis created using Genomics suite (Partek Incorporated, MO, USA).

**Primary Tg4 CD4^+ T Cell Activation Assays**

To study the primary activation of Tg4 CD4^+ T cells, varying numbers (as stated) of BMDC (CD45.2) were cultured with 2 × 10^4 Tg4 CD4^+ T cells (CD45.1) per well in a round bottomed, 96 well plate. The CD4^+ T cells were purified using microbeads as per manufacturer’s instructions (Miltenyi Biotech, Surrey, UK). Tg4 CD4^+ T cell production of cytokines (IL-2, TNF-α, GM-CSF, and IFN-γ) was assessed in culture supernatants by Ready-SET-Go ELISA (eBioscience, San Diego, USA). IL-2 was measured in supernatants after 48 h of culture and IFN-γ, TNF-α, and GM-CSF were measured after 72 h of culture.

**Lentiviral Transduction of BMDC for Modulation of CD31 Expression**

To investigate the effects of CD31 expression on the ability of CD11c^+BMDC to prime CD4^+ T cells, lentiviral constructs expressing GFP and murine CD31 or murine CD31 siRNA were developed. For the CD31 overexpression construct (pLent6-cppt-CMV-mCD31-RES-empGF-opre), murine platelet/endothelial cell adhesion molecule-1 (Pecam-1; mCD31) isoform 3 (NM_001305157.1) was synthesized by Integrated DNA Technologies (IDT, Leuven, Belgium). The synthetic gene was flanked by attB1 and attB5r GATEWAY cloning sites (Thermo Fisher Scientific: Invitrogen UK) and a Kozak consensus overlapped the start codon. Using GATEWAY technology this was used to create the lentiviral shuttle vector pLent6-cppt-CMV-mCD31-RES-empGF-opre (33). Negative control lacked mCD31. By partially concentrating lentivirus in OptiMEM (Invitrogen, UK) titers of 10^7 transduction units/ml (TU/ml) were obtained.

For siRNA-expressing vectors, knock down target sequences within mCD31 were identified by a combination of literature searches and the output of the Invitrogen Block-iT RNAi designer web program. A scrambled sequence was used as a negative control. The synthesized oligonucleotides were annealed, then inserted into pcDNA6.2-GW_emGFP-miR 285, 816, and 1827, with numbers identifying the target distance from the start codon of mCD31 (Table 1). These were shuttled into pLent6.2-cppt-CMV-DEST-opre using GATEWAY (Invitrogen, UK), and packaged. Titers of 10^8 TU/ml were routinely obtained.

Six days following BMDC induction, cells were seeded at 5 × 10^5 cells/well in a 24 well cell culture dish in 1 ml cell culture medium and lentiviral constructs added at an MOI of 20. For CD31 overexpression lentiviral constructs, 6 µg/ml DEAE were added to enhance transduction efficiency. Two days later, BMDC were stimulated for 18 h as described above and CD11c^+GFP^+ BMDC isolated by FACS prior to further experimentation.

**Preparation of Spleen Mononuclear Cells and FACS Analysis**

Single cell suspensions were made from the spleen and draining lymph nodes, red blood cells were lysed using an ammonium chloride buffer (Sigma Aldrich, Dorset, UK), and cells were then re-suspended in FACS buffer (PBS, 2% fetal calf serum, 0.01% sodium azide (Sigma Aldrich, Dorset, UK). Fc receptors were blocked with supernatant from
TABLE 1 | Synthetic oligonucleotides used for miR.

<table>
<thead>
<tr>
<th>miRNAi</th>
<th>Name</th>
<th>DNA Sequence (5′-3′)</th>
</tr>
</thead>
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<tr>
<td>Neg</td>
<td>Scr top</td>
<td>TGCTGAAATGACTGCGCTGGAGAAGCTTGGGC</td>
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<td></td>
<td>Scr bottom</td>
<td>AAATGTCGCTGCGTAAGCTCAGTGCC</td>
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<tr>
<td>CD31</td>
<td>285 top</td>
<td>TGCTGATCGTCGTTTGGGC</td>
</tr>
<tr>
<td></td>
<td>285 bottom</td>
<td>CGGCGTTTGGGC</td>
</tr>
<tr>
<td></td>
<td>816 top</td>
<td>TGCTGTAAGAAGCTTGTTGGGC</td>
</tr>
<tr>
<td></td>
<td>816 bottom</td>
<td>TGCTGAGAAGCTTGTTGGGC</td>
</tr>
<tr>
<td></td>
<td>1827 top</td>
<td>TGCTGTTCCAGGAAAGCTGGGGG</td>
</tr>
<tr>
<td></td>
<td>1827 bottom</td>
<td>GGGCCAAAACCAAATGCTACTTCCTGAGAAC</td>
</tr>
</tbody>
</table>

the hybridoma 2.4G2. All antibodies were from eBioscience, Hatfield, UK, except where stated; live/dead fixable cell stain conjugated to ef455 (Life Technologies), anti-CD4-APC, anti-CD45.1-FITC, anti-CD44-APC-Cy7, anti-CD80-PE, anti-CD11c-ef450, anti-Ki67-PE-Cy7, anti-CD11b-AF700, CD4-AF700 (BD Pharmingen, Oxford, UK), anti-CD11c-PE-Cy7, anti-CD31-PE, anti-CD86-bv785, and anti-Foxp3-eF450. FACS data were collected using a 6 laser LSR Fortessa (BD Biosciences, New Jersey, USA) and analyzed using FlowJo software (Tree Star, Olwen, Switzerland).

PrimeFlow RNA Assay
Murine BMDC were stained with fixable viability dye for 30 min at 4°C, washed once, and stained for surface markers of interest (anti-CD11c-PE-Cy7, anti-CD11c-eF450, anti-Kit67-PE-Cy7, anti-CD11b-APC, anti-CD45.1-FTTC, anti-CD44-APC-Cy7, anti-CD80-PE, anti-CD86-APC, anti-CD62L-PerCP-Cy5.5, and anti-Foxp3-eF450). BMDC are shown in Figure 1A. The genes with the greatest upregulation and downregulation in VitD-CD11c+ BMDC compared to Veh-CD11c+ BMDC by a Fisher’s exact test. Significance was set at p < 0.05.

RESULTS
Addition of 1,25(OH)2D3 to BMDC Cultures Leads to Altered Gene Expression Profiles in CD11c+ BMDC
1,25(OH)2D3 and vehicle conditioned BMDC were generated as described in materials and methods. After 9 days of culture, CD11c+ cells were isolated by FACS and cultured overnight in 1,25(OH)2D3 free media in the presence or absence of LPS. Global gene expression analysis revealed that 101 genes were downregulated and 24 were upregulated in non-LPS stimulated VitD-CD11c+ BMDC compared to Veh-CD11c+ BMDC (Figure 1A). The number of genes differentially expressed in VitD-CD11c+ BMDC increased following LPS stimulation, with 254 genes downregulated and 178 upregulated (Figure 1A). The genes with the greatest upregulation and downregulation in VitD-CD11c+ BMDC compared to Veh-CD11c+ BMDC are shown in Tables 2–5. The gene expression datasets for this study can be found in the Gene Expression Omnibus, accession number GSE114768.

Statistical Analysis
For the bioinformatic analyses, a total of 20 arrays (n = 3 Veh-CD11c+ BMDC no LPS, n = 3 VitD-CD11c+ BMDC no LPS, n = 6 Veh-CD11c+ BMDC LPS, n = 8 VitD-CD11c+ BMDC LPS) were assessed for quality control using the arrayQualityMetrics package in Bioconductor (www.bioconductor.org). log2 expression values for the high quality arrays were quantile normalized. Pairwise group comparisons were undertaken using linear modeling with the limma package in Bioconductor. Subsequently, empirical Bayesian analysis was applied, including vertical (within a given comparison) p-value adjustment for multiple testing, which controls for false-discovery rate. An adjusted p-value of <0.05 was applied as the significance threshold. Functional enrichment analyses of KEGG pathways associated with the significant loci were performed using hypergeometric tests (34). FACS data were collected using a 6 laser LSR Fortessa (BD Biosciences, New Jersey, USA) and analyzed using FlowJo software (Tree Star, Olwen, Switzerland).

Time-Lapse Imaging
BMDC transduced with control or CD31 overexpressing lentiviral vector were stimulated for 4 h with 0.1 µg/ml LPS and 0.1 µg/ml 4Y peptide. CD11b+CD11c+GFP+ cells were FACs sorted and re-plated onto poly-L-lysine coated glass-bottom plates at a density of 3 × 104 cells/ml in cell culture medium. After overnight adhesion, BMDC were carefully washed twice in PBS before naïve CD4+ T cells from the spleen of a Tg4 (CD45.1) mouse were slowly added at a 4:1 ratio in phenol red-free HBSS. Brightfield images (5 randomly selected fields in each well) were acquired every 30 s using an Axiovert 200 microscope in a CO2-supplemented chamber at 37°C. Movies of cells were tracked in Fiji ImageJ to determine the duration of interaction between BMDC and CD4+ T cells.
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FIGURE 1 | CD31 is upregulated by 1,25(OH)\textsubscript{2}D\textsubscript{3} in BMDC. Bone marrow derived dendritic cells (BMDC) were generated over 9 days in the presence of 1,25(OH)\textsubscript{2}D\textsubscript{3} (VitD) or absence (Veh) of 20 mM 1,25(OH)\textsubscript{2}D\textsubscript{3}. (A) Veh and VitD BMDC were matured for 18 h in the presence or absence of 0.1 µg/ml LPS. mRNA was extracted and microarray performed. Differential gene expression caused by 1,25(OH)\textsubscript{2}D\textsubscript{3} in the presence (left) or absence (right) of LPS stimulation is depicted as volcano plots. (B) Number of genes differentially regulated by 1,25(OH)\textsubscript{2}D\textsubscript{3} in CD11c\textsuperscript{+}BMDC in the presence or absence of LPS stimulation. (Continued)

CD31 Expression Is Increased on VitD-CD11c\textsuperscript{+}BMDC

There were 47 genes which were differentially expressed in VitD-CD11c\textsuperscript{+}BMDC (in both immature and LPS-matured cells), of which 7 were upregulated and 40 were downregulated (Figure 1B). The 7 upregulated genes were Pecam1, Cd300ld, Adgre1, Ebp41l3, Srgap3, Klk1b1, and Mmp8. The increased expression of CD31 in VitD-CD11c\textsuperscript{+}BMDC was striking given recent studies which have indicated that CD31 plays an important role in regulating a wide range of leukocytes (30, 35). The increased mRNA expression of CD31 in non-LPS stimulated VitD-CD11c\textsuperscript{+}BMDC was confirmed by flow cytometric evaluation of CD31 mRNA (Figure 1C). Protein expression of CD31 on VitD-CD11c\textsuperscript{+}BMDC was subsequently examined by flow cytometry which demonstrated an increase in CD31 expression on BMDC which had been conditioned with 20 nM 1,25(OH)\textsubscript{2}D\textsubscript{3} (Figure 1D). This increase was dependent on the dose of 1,25(OH)\textsubscript{2}D\textsubscript{3}, with sequential increases in CD31 expression occurring in CD11c\textsuperscript{+}BMDC which had been generated in the presence of 5, 10, and 15 nM of 1,25(OH)\textsubscript{2}D\textsubscript{3} (Supplementary Figure 2A). The increase in expression of CD31 on VitD-CD11c\textsuperscript{+}BMDC was an early event with high levels of CD31 detectable on VitD-CD11c\textsuperscript{+}BMDC as early as day 3 of the BMDC cultures (Supplementary Figure 2B). When 1,25(OH)\textsubscript{2}D\textsubscript{3} was added to the BMDC culture on day 3 or day 6, rather than on day 0, CD31 levels were still upregulated on day 9, although at lower levels compared to BMDC which were cultured in the presence of 1,25(OH)\textsubscript{2}D\textsubscript{3} since the beginning of the culture (Supplementary Figure 2C). However, when 1,25(OH)\textsubscript{2}D\textsubscript{3} was added to or removed from Veh-CD11c\textsuperscript{+}BMDC or VitD-CD11c\textsuperscript{+}BMDC, respectively, during overnight stimulation with LPS, no change in CD31 levels was observed (Supplementary Figure 2D), indicating that CD31 levels are determined during the generation, not the maturation, of BMDC.

CD31 Expression on VitD-CD11c\textsuperscript{+}BMDC Restrains Their Ability to Prime CD4\textsuperscript{+} T Cells

CD31 is expressed on both naïve and activated CD4\textsuperscript{+} T cells (Supplementary Figure 3). Consequently, it would be challenging to dissect the role of CD31 expression in the ability of BMDC to prime CD4\textsuperscript{+} T cells in vitro by adding a blocking CD31 antibody to the BMDC-T cell co-cultures, as this would also affect CD31 availability on T cells. To precisely explore the role of CD31 expression in CD11c\textsuperscript{+}BMDC function, we used lentiviral constructs to either upregulate or downregulate
### TABLE 2 | List of top 10 genes upregulated in VitD-CD11c<sup>+</sup> BMDC compared to Veh-CD11c<sup>+</sup> BMDC without LPS stimulation.

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>Description</th>
<th>EntrezID</th>
<th>Log FC</th>
<th>Adjusted p-value</th>
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<tbody>
<tr>
<td>Epb41l3</td>
<td>Erythrocyte protein band 4.1-like 3</td>
<td>13823</td>
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<tr>
<td>Srgap3</td>
<td>SLIT-ROBO Rho GTPase activating protein 3</td>
<td>259302</td>
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<td>5.40E-06</td>
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<tr>
<td>Adgre1</td>
<td>EGF-like module containing, mucin-like, hormone receptor-like sequence 1</td>
<td>13733</td>
<td>1.5</td>
<td>0.0013</td>
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<tr>
<td>Mmp8</td>
<td>Matrix metallopeptidase 8</td>
<td>17394</td>
<td>2.3</td>
<td>0.0041</td>
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<tr>
<td>Serpinb2</td>
<td>Serine (or cysteine) peptidase inhibitor, clade B, member 2</td>
<td>18788</td>
<td>3.2</td>
<td>0.0081</td>
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<tr>
<td>Sfca3</td>
<td>Stefin A3</td>
<td>20863</td>
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<tr>
<td>Tanc2</td>
<td>Tetratricopeptide repeat, ankyrin repeat and coiled-coil containing 2</td>
<td>77097</td>
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<td>0.012</td>
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<tr>
<td>Pecam1</td>
<td>CD31 antigen</td>
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<tr>
<td>S100a9</td>
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<tr>
<td>Syne1</td>
<td>Spectrin repeat containing, nuclear envelope 1</td>
<td>64009</td>
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<td>0.019</td>
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</table>

### TABLE 3 | List of top 10 genes downregulated in VitD-CD11c<sup>+</sup> BMDC compared to Veh-CD11c<sup>+</sup> BMDC without LPS stimulation.

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>Description</th>
<th>EntrezID</th>
<th>Log FC</th>
<th>Adjusted p value</th>
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<td>Ccl5</td>
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<td>Fabp4</td>
<td>Fatty acid binding protein 4, adipocyte</td>
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<td>Ifi27l2a</td>
<td>Interferon, alpha-inducible protein 27 like 2A</td>
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<td>Gbp5</td>
<td>Guanylate binding protein 5</td>
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<td>Gbp2</td>
<td>Guanylate binding protein 2</td>
<td>14469</td>
<td>−1.6</td>
<td>9.00E-05</td>
</tr>
<tr>
<td>Ace</td>
<td>Angiotensin I converting enzyme (peptidyl-dipeptidase A) 1</td>
<td>11421</td>
<td>−2.3</td>
<td>9.00E-05</td>
</tr>
<tr>
<td>Lira5</td>
<td>Leukocyte immunoglobulin-like receptor, subfamily A (with TM domain), member 5</td>
<td>232801</td>
<td>−1.7</td>
<td>0.000011</td>
</tr>
<tr>
<td>Adam23</td>
<td>A disintegrin and metallopeptidase domain 23</td>
<td>23792</td>
<td>−2.8</td>
<td>0.00021</td>
</tr>
<tr>
<td>Lgals3bp</td>
<td>Lectin, galactoside-binding, soluble, 3 binding protein</td>
<td>19039</td>
<td>−1.4</td>
<td>0.00039</td>
</tr>
<tr>
<td>Slc35g1</td>
<td>Solute carrier family 35, member G1</td>
<td>240660</td>
<td>−1.4</td>
<td>0.00059</td>
</tr>
</tbody>
</table>

### TABLE 4 | List of top 20 genes upregulated in VitD-CD11c<sup>+</sup> BMDC compared to Veh-CD11c<sup>+</sup> BMDC with LPS stimulation.

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>Description</th>
<th>EntrezID</th>
<th>Log FC</th>
<th>Adjusted p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Epb41l3</td>
<td>Erythrocyte protein band 4.1-like 3</td>
<td>13823</td>
<td>3</td>
<td>9.20E-09</td>
</tr>
<tr>
<td>Srgap3</td>
<td>SLIT-ROBO Rho GTPase activating protein 3</td>
<td>259302</td>
<td>1.8</td>
<td>2.50E-08</td>
</tr>
<tr>
<td>Litr</td>
<td>Leukemia inhibitory factor receptor</td>
<td>16880</td>
<td>2.7</td>
<td>5.70E-08</td>
</tr>
<tr>
<td>Angpt2</td>
<td>Angiopoietin-like 2</td>
<td>26360</td>
<td>2.1</td>
<td>0.00002</td>
</tr>
<tr>
<td>Adgre1</td>
<td>EGF-like module containing, mucin-like, hormone receptor-like sequence 1</td>
<td>13733</td>
<td>1.4</td>
<td>0.000028</td>
</tr>
<tr>
<td>Mmp8</td>
<td>Matrix metallopeptidase 8</td>
<td>17394</td>
<td>2.4</td>
<td>0.00043</td>
</tr>
<tr>
<td>Gstp1</td>
<td>Glutathione S-transferase, pi 1</td>
<td>14870</td>
<td>1.5</td>
<td>0.00046</td>
</tr>
<tr>
<td>Acpp</td>
<td>Acid phosphatase, prostate</td>
<td>56318</td>
<td>1.3</td>
<td>0.000066</td>
</tr>
<tr>
<td>Fam110c</td>
<td>Family with sequence similarity 110, member C</td>
<td>104943</td>
<td>1.3</td>
<td>0.00012</td>
</tr>
<tr>
<td>Cts2b</td>
<td>Cytotoxic T lymphocyte-associated protein 2 beta</td>
<td>13025</td>
<td>2.4</td>
<td>0.00012</td>
</tr>
<tr>
<td>Nppc</td>
<td>Natriuretic peptide type C</td>
<td>18159</td>
<td>2.6</td>
<td>0.00014</td>
</tr>
<tr>
<td>Prlbp2</td>
<td>PTPRF interacting protein, binding protein 2 (lprin beta 2)</td>
<td>19024</td>
<td>1.2</td>
<td>0.00017</td>
</tr>
<tr>
<td>Qja1</td>
<td>Gap junction protein, alpha 1</td>
<td>14609</td>
<td>1.9</td>
<td>0.0002</td>
</tr>
<tr>
<td>Ripk3</td>
<td>Receptor-interacting serine-threonine kinase 3</td>
<td>56532</td>
<td>1</td>
<td>0.00022</td>
</tr>
<tr>
<td>Hgf</td>
<td>Hepatocyte growth factor</td>
<td>15234</td>
<td>2</td>
<td>0.00023</td>
</tr>
<tr>
<td>B3glct</td>
<td>Beta-3-glucosyltransferase</td>
<td>381694</td>
<td>1.3</td>
<td>0.00028</td>
</tr>
<tr>
<td>Nucb2</td>
<td>Nucleobindin 2</td>
<td>53322</td>
<td>0.8</td>
<td>0.0003</td>
</tr>
<tr>
<td>Dapk2</td>
<td>Death-associated protein kinase 2</td>
<td>13143</td>
<td>1.7</td>
<td>0.00042</td>
</tr>
<tr>
<td>Mmp13</td>
<td>Matrix metallopeptidase 13</td>
<td>17386</td>
<td>1.3</td>
<td>0.00079</td>
</tr>
<tr>
<td>It13</td>
<td>Interleukin 13</td>
<td>16163</td>
<td>2.9</td>
<td>0.0011</td>
</tr>
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</table>
CD31 expression. Initially, we produced three siRNA lentivirus constructs. Both control vectors and siRNA constructs were effective in transducing BMDC (Figure 2A). The transduction efficiency increased as multiplicity of infection (MOI) range was increased from 1 to 50 (Supplementary Figure 4A). Construct 1827 was able to reduce the expression of CD31 most effectively and was used in all subsequent experiments (Figure 2B). There was no increase in the extent of CD31 downregulation with increasing MOI (Supplementary Figure 4B).

Following transduction with lentiviral constructs there was little change in the numbers of viable cells in the cultures. There was no difference in the number of viable cells in VitD-CD11cBMDC cultures transduced with negative control vector containing a scrambled siRNA compared to vectors encoding CD31 siRNA (Supplementary Figure 4C). Following the transduction with CD31 siRNA lentivirus, CD31 expression was successfully decreased with no significant changes to MHC class II, CD40, CD80 and CD86 expression compared to control vector treated CD11cBMDC (Supplementary Figure 5A).

To probe the functional effects of CD31 expression on VitD-CD11cBMDC, activated and peptide-pulsed VitD-CD11cBMDC transfected with negative control vector containing a scrambled siRNA or CD31 siRNA construct were co-cultured with naïve cognate CD4+ T cells. IL-2 production was significantly increased in co-cultures of CD4+ T cells and VitD-CD11cBMDC which had CD31 expression reduced by the siRNA encoding vector (Figure 2C). In addition, production of IFN-γ, GM-CSF, and TNF-α was also increased following priming by VitD-CD11cBMDC with reduced CD31 expression (Figure 2C).

**Overexpression of CD31 Expression on Veh-CD11c+BMDC Reduces Their Ability to Prime CD4+ T Cells**

To examine the effects of increased CD31 expression on CD11c+BMDC, a GFP-CD31 lentiviral construct was developed. The construct was readily incorporated into a proportion of host cells and GFP+ cells had a marked increase in CD31 expression (Figure 3A). There was a reduction in IL-2 production by CD4+ T cells that were co-cultured with Veh-CD11cBMDC over expressing CD31 (Figure 3B). In addition, pro-inflammatory cytokine production was also reduced from CD4+ T cells co-cultured with Veh-CD11cBMDC over expressing CD31 (Figure 3C). Lentiviral induction of CD31 overexpression had no influence on the levels of MHC class II, CD40, CD80 and CD86 expression in Veh-CD11cBMDC (Supplementary Figure 5B) or increase the proportion of T cells which expressed Foxp3 (Supplementary Figure 5C). Taken together, the genetic manipulation experiments, which allowed us to both overexpress and downregulate CD31, demonstrated that CD31 on BMDC is a potent regulator of CD4+ T cell priming.

**Increased Levels of CD31 on BMDC Reduces BMDC—CD4+ T Cell Interaction Time During in vitro Priming**

CD31 has been implicated in mediating cell detachment between live neutrophils and phagocytes, thereby inhibiting binding and engulfment of non-apoptotic neutrophils (29). To understand...
1,25(OH)\textsubscript{2}D\textsubscript{3} Enhances CD31 Levels in Human CD11c\textsuperscript{+} Cells

In order to assess whether 1,25(OH)\textsubscript{2}D\textsubscript{3} could have a comparable effect on CD31 expression in human DC, CD34\textsuperscript{+} progenitor cells were isolated from mobilized blood and differentiated toward DC in the presence or absence of 1,25(OH)\textsubscript{2}D\textsubscript{3}. CD11b\textsuperscript{+}CD11c\textsuperscript{+} cells from 7-day cultures showed a marked increase in CD31 expression when generated in the presence of 1,25(OH)\textsubscript{2}D\textsubscript{3}, suggesting that 1,25(OH)\textsubscript{2}D\textsubscript{3} could modulate DC function through CD31 not only in mice, but also in humans (Figures 5A,B). When CD11b\textsuperscript{+}CD11c\textsuperscript{+} cells were sorted and re-plated overnight with or without LPS, cells generated in the presence of 1,25(OH)\textsubscript{2}D\textsubscript{3} were able to maintain high CD31 levels whereas control cells showed a considerable reduction in CD31 expression upon stimulation (Figure 5C).

DISCUSSION

1,25(OH)\textsubscript{2}D\textsubscript{3} Modulates Gene Expression in Immature and Mature BMDC

This study demonstrates that 1,25(OH)\textsubscript{2}D\textsubscript{3} markedly influences gene expression in CD11c\textsuperscript{+} BMDC, and has unveiled a novel mechanism by which 1,25(OH)\textsubscript{2}D\textsubscript{3} alters BMDC function—via elevated expression levels of CD31. We focused our approach on studying the effects of 1,25(OH)\textsubscript{2}D\textsubscript{3} on CD11c\textsuperscript{+} BMDC since we have previously established that 1,25(OH)\textsubscript{2}D\textsubscript{3} leads to the development of a higher proportion of CD11c\textsuperscript{−} cells in GM-CSF-supplemented bone marrow cell cultures (16). Many of the CD11c\textsuperscript{−} cells in 1,25(OH)\textsubscript{2}D\textsubscript{3} conditioned BMDC do not express cell surface markers such as MHC class II molecules which are considered to be a critical feature of DC (16). Consequently, the gene changes we observed are within the BMDC subset.
FIGURE 3 | Upregulation of CD31 in VitD BMDC leads to dampened in vitro priming of CD4⁺ T cells. Bone marrow derived dendritic cells (BMDC) were generated over 9 days with 20 ng/ml GM-CSF in the absence (Veh) of 1,25(OH)₂D₃. On day 6, BMDC were transduced with indicated lentiviral constructs, all expressing GFP. (A) CD31 protein expression in GFP⁺ Veh CD11c⁺ BMDC transduced with a lentiviral vector expressing CD31 (dotted line) compared to Veh CD11c⁺ BMDC transduced with a negative control vector Veh. (Continued)

(B) Veh CD11c⁺ BMDC transduced with negative control vector containing a scrambled siRNA (black) or CD31 expressing (white) lentiviral vector were matured in the presence of 0.1 µg/ml LPS for 4 h and pulsed with 0.1 µg/ml 4Y peptide, and were subsequently co-cultured at indicated cell numbers with 2 × 10⁴ Tg4 CD4⁺ T cells for 48 h (IL-2) or 72 h (all other cytokines). Cytokine release was assessed by ELISA. (C) CD4⁺ T cells were analyzed by flow cytometry for CD44 and Ki-67 expression following 72 h of co-culture with indicated Veh CD11c⁺ BMDC. n = 3, representative of 2 experiments (*p < 0.05, **p < 0.01, and ***p < 0.001).

FIGURE 4 | High levels of CD31 on BMDC result in reduced interaction times with CD4⁺ T cells in vitro. Bone marrow derived dendritic cells (BMDC) were generated over 9 days with 20 ng/ml GM-CSF in the absence (Veh) of 1,25(OH)₂D₃. On day 6, BMDC were transduced with negative control vector containing a scrambled siRNA or CD31 expressing lentiviral constructs, all expressing GFP. (A) Experimental layout; on day 9, transduced BMDC were sorted for CD11c⁺ GFP⁺ cells and matured for 4 h in the presence of 0.1 µg/ml LPS and pulsed with 0.1 µg/ml 4Y peptide. Matured BMDC were rested for 4 h and Tg4 CD4⁺ T cells added at a 1:4 ratio prior to time-lapse imaging. (B) The length of BMDC—T cell interaction was calculated based on images taken in 30 s intervals for the duration of 3 h and were classified as short (<5 min), intermediate (5-15 min) and long (>15 min). Data pooled from three mice. Representative of 2 experiments.

which have the capacity to prime CD4⁺ T cells. Within the hundreds of genes that were differentially expressed in VitD-CD11c⁺ BMDC, a range of biochemical pathways were altered when CD11c⁺ BMDC were generated in the presence of 1,25(OH)₂D₃, as demonstrated in KEGG pathway analysis (data not shown).

An extensive body of work has revealed the dampening effects of 1,25(OH)₂D₃ on DC maturation via downregulation...
of co-stimulatory molecules, MHC-II, and pro-inflammatory cytokines (36). We were therefore particularly interested in genes whose expression was upregulated by 1,25(OH)2D3, and their functional relevance in DC biology. For further investigation, we focused on genes which were upregulated robustly in both immature and LPS-matured VitD-CD11c+ BMDC; one of the seven genes fitting this criterion was CD31. CD31 is a member of the Ig superfamily and highly expressed on endothelial cells but also on platelets and the majority of leukocytes (37) and plays an important role in transendothelial migration (38, 39). Beyond its role in cell-cell contact and migration, CD31 has also been found to act as a co-inhibitory molecule on both T cells (21) and DC (30).

**1,25(OH)2D3 as a Novel Transcriptional Regulator of CD31 Expression in BMDC**

Very little is known about the regulation of CD31 expression in immune cells. In naïve T cells, downregulation of CD31 expression has been observed in association with homeostatic proliferation (40). Human memory T cells, on the other hand, have been reported to acquire CD31 expression following trans-endothelial migration in vitro (41). Conversely, in mouse leukocytes, CD31 expression has consistently been found to be reduced upon transmigration through the endothelial cell layer, both in vitro (42) and in vivo (43, 44), although the details of the mechanisms involved and the functional implications of these expression changes are at present unclear. The data presented here reveal a strong influence of 1,25(OH)2D3 on the expression levels of CD31 in mouse and human DC and thereby provide a first insight into transcriptional control of this multifunctional glycoprotein. In human DC, CD31 is known to be downregulated upon maturation (45, 46). The reduction in the expression of this co-inhibitory molecule upon DC maturation is thought to allow for an adequate immune response to be mounted upon encounter of infective agents and is likely to play an important role in balancing immune tolerance vs. immunity. In line with this, we observed a downregulation of CD31 expression on human mBDC when matured in the presence of LPS. Strikingly, the presence of 1,25(OH)2D3 enhanced CD31 levels in immature DC and stabilized CD31 expression during DC maturation with LPS. This suggests that the stably enhanced CD31 expression on human DC through 1,25(OH)2D3 could contribute to the tolerogenic effects of 1,25(OH)2D3, as observed in mouse BMDC. Interestingly, we did not observe a reduction in CD31 expression on mouse BMDC matured in the presence of LPS, which does not align with results from Clement et al. (30). In their study, they observe that following LPS stimulation, a subgroup of BMDC that exhibit a more pronounced activation status as defined by higher MHC-II and CD80 expression than the majority of BMDC have slightly lower CD31 expression. However, an overall comparison of CD31 expression between immature and LPS-matured BMDC was not made, and this difference in analysis may account for the distinct observations made. In 1,25(OH)2D3-conditioned BMDC, however, CD31 expression was greatly enhanced, and high CD31 expression levels were retained upon stimulation with LPS. The details of this 1,25(OH)2D3-dependent transcriptional control of CD31 expression in mouse and human DC deserve further investigation, and more extensive studies are required to address the transcriptional regulation of CD31 in other cell types which express the Vitamin D receptor and could respond to changes in 1,25(OH)2D3 concentrations.
CD31 Expressed by DC Controls DC—T Cell Interaction Time and T Cell Priming

CD31 contains six extracellular Ig C2-type domains. These enable transhomophilic binding of CD31 on adjacent cells (22, 23), but have also been reported to allow heterophilic binding of αVβ3 (25), glycosaminoglycans (26) and CD38 on lymphocytes (27). Mechanistically, we were able to show that high CD31 expression on BMDC led to reduced interaction times with CD4+ T cells during in vitro priming. This observation is consistent with results published by Brown et al. who elegantly showed that neutrophil CD31 engagement with macrophages induced a detachment signal in neutrophils, thereby avoiding engulfment by the APC (29). Apoptotic neutrophils, however, lacked the intracellular signal and were not able to detach from macrophages, resulting in apoptotic neutrophil engulfment.

In the case of BMDC, as presented in the present study, 1,25(OH)2D3-induced elevated CD31 expression led to reduced interaction time and consequently reduced priming of CD4+ T cells. Whether this mechanism involves transhomophilic or heterophilic binding of CD31 has not been established in the present study, but we show that CD31 is expressed on both naive and activated CD4+ T cells. This does not exclude the possibility that CD31 interaction between BMDC and T cells does not also lead to inhibitory downstream signaling in BMDC themselves or T cells in this setting. The cytosolic ITIM domains of CD31, when phosphorylated, recruit Src homology 2 (SH2) domain-containing proteins, e.g., SH2 domain containing protein tyrosine phosphatases SHP-1 and SHP-2 (47). In CD4+ T cells, CD31 downstream signaling has been shown by several groups to have inhibitory effects on TCR downstream signaling (35). The reduced activation of T cells during in vitro priming with CD31-enriched BMDC may therefore well be the result of a combination of reduced interaction time and CD31 signaling in T cells.

In DC, CD31 has more recently been proposed to act as a co-inhibitory molecule via SHP-1 (30). Clement et al. used a stimulatory CD31 peptide to show that sustained CD31 signaling in DC led to reduced expression of co-stimulatory molecules such as CD40 and CD86, and reduced pro-inflammatory cytokine production (30). In the present study, CD31 expression was modulated, rather than changing its signaling, and altered CD31 expression patterns did not result in changes in surface expression of co-stimulatory molecules. DC tolerized by culture with 1,25(OH)2D3 have been shown to induce regulatory T cells, and would therefore be a potential mechanism by which CD31 could confer its tolerogenic effect. However, no increase in Treg induction during in vitro priming of T cells with CD31-enriched BMDC was observed in this study. In contrast, Clement et al. reported an increase in the Treg pool in vivo when mice were injected with DC matured in the presence of stimulating CD31 peptide, attributed to the enhanced secretion of TGF-β and IL-10 (30). It appears that CD31 signaling induced by a soluble CD31 peptide alters the phenotype and cytokine expression pattern of DC toward a more tolerogenic phenotype, which consequently induce regulatory T cells and have an attenuated ability to prime conventional T cells. The present study shows that the overexpression of CD31 in BMDC per se, using lentiviral constructs, does not induce a change in BMDC phenotype. The observed reduction in T cell priming by CD31-enriched BMDC is therefore likely a direct result of CD31 binding and not confounded by an altered BMDC phenotype indirectly leading to changes in T cell priming.

1,25(OH)2D3-Induced CD31 Expression on DC: Implications for Treatment of Autoimmune and Chronic Inflammatory Disease?

The use of tolerogenic DC as cell therapy in autoimmune and chronic inflammatory diseases is considered a potential breakthrough in personalized medicine and antigen-specific therapy (5, 15). Although numerous protocols have been described to generate tolerogenic DC, 1,25(OH)2D3 is one of the most widely used molecules to generate DC with a tolerogenic phenotype (15). A key attribute of tolerogenic DC is to suppress autopathogenic T cells which may be through the induction of anergy or apoptosis or via the induction of regulatory T cells (5). Prevention of further activation of effector T cells is clearly an important attribute of tolerogenic DC; our study indicates that 1,25(OH)2D3 mediated increase in CD31 expression may be one way in which 1,25(OH)2D3 could restrain priming ability in tolerogenic DC used in clinical therapies. Our work suggests that evaluation of CD31 expression on tolerogenic DC may be informative on their priming ability prior to administration to patients. In addition, genetic manipulation of CD31 expression may further improve the safety profile of tolerogenic DC by lessening their capacity to activate effector T cells.

CONCLUSION

The factors that control CD31 expression on BMDC have been poorly explored to date. This is the first study to demonstrate that the active vitamin D metabolite 1,25(OH)2D3 can enhance CD31 expression in both mouse and human DC. CD31 acts as a co-inhibitory molecule and prevents effective T cell priming at least in part by reducing the interaction time between BMDC and naïve T cells. This finding is an important demonstration that the failure of 1,25(OH)2D3 conditioned BMDC to effectively prime T cells is not simply due to the reduced expression of MHC class II and co-stimulatory molecules such as CD40, CD80, and CD86. Exposure of BMDC during development to 1,25(OH)2D3 is critical in the upregulation of inhibitory pathways which further restrain the ability of BMDC to prime T cells.

DATA AVAILABILITY

The datasets generated for this study can be found in Gene Expression Omnibus, GSE114768.

AUTHOR CONTRIBUTIONS

LS, IM, and RM designed and analyzed experiments. LS and IM performed experiments. AI, DS, and NK...
REFERENCES

31. 1,25-Dihydroxyvitamin D3 Upregulates CD31 on DCs

ACKNOWLEDGMENTS

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2019.00600/full#supplementary-material


Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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