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Production and characterization of two monoclonal antibodies to bovine tumour necrosis factor alpha (TNF-α) and their cross-reactivity with ovine TNF-α


Institute for Animal Health, Compton, Near Newbury, RG20 7NN, UK
Moredun Research Institute, International Research Centre, Pentlands Science Park, Bush Loan, Edinburgh EH26 0PZ, UK

The relative paucity of immunological reagents available for veterinary species compared to those available for studies in humans and biomedical species hampers disease pathogenesis studies and consequently the development of safe and effective vaccines for animal species. There remains a need to develop and validate tools and reagents for bovine (bov) and ovine (ov) immunology as prioritised through various research initiatives (Entrican et al., 2009). One such priority is tumour necrosis factor alpha (TNF-α), a pleiotropic cytokine that mediates early inflammatory responses to a variety of physical, environmental, and immunological stimuli. Produced primarily by macrophages and lymphocytes, TNF-α coordinates the inflammatory response through the induction of other cytokines (e.g., interleukin [IL]-1 and IL-6), and through the recruitment of immune and inflammatory cells (Lopez Ramirez et al., 1994; Roach et al., 2002). In addition, TNF-α is important for immune control of intracellular bacteria such as Mycobacterium tuberculosis (Flynn et al., 1995; Havell, 1989; Lin et al., 2007) and Mycobacterium bovis, the causative agent of bovine tuberculosis (Denis et al., 2004; Hope et al., 2004). There are several commercial ELISAs that detect bov TNF-α, with most (but not all) based on polyclonal antisera. There are mAb to bov and ov TNF-α available, but information on their species cross-reactivity, ability to detect intracytoplasmic TNF-α and to neutralize biological activity is limited (Pedersen et al., 2002). We have previously described the development of mAb for the detection and measurement of ruminant cytokines.
such as GMCSF, IL-4, IL-10 and IL-12 (Entrican et al., 1996; Hope et al., 2002, 2005; Kwong et al., 2002). Using a similar approach, we describe here the development of monoclonal antibodies (mAb) to bov TNF-α and their application in ELISA and flow cytometric detection systems in both cattle and sheep. In addition, we describe the neutralization capacity of the mAb against bov TNF-α.

Oligonucleotide primers (forward ATGAGCACCA-AAAGCATGATC, reverse TCACAGGGCGATGATCCCA) were designed to amplify bov TNF-α (accession number NM_173966) from bov cDNA. PCR was performed and the product cloned into pTarget and sequenced by the ABI Prism automated sequencing method (Oxford University). Recombinant (r)bov TNF-α was produced by transfecting COS-7 cells with cDNA encoding bov TNF-α ligated into the pTarget vector (Promega UK Ltd., Southampton, UK). The biological activity of rbov TNF-α was determined by titration in a WEHI-164 cell bioassay (Eskandari et al., 1990). Serial dilutions of rbov TNF-α ranging from 1/4 to 1/1,048,576 were added to wells containing 4 \times 10^4 WEHI-164 cells. Actinomycin D at 2 µg/ml was also added to each well. Following 18 h culture, the cells were fixed, stained with crystal violet and absorbance read at 595 nm. One unit was defined as the reciprocal dilution which induced half maximal cell death. To produce mAb, BALB/c mice were inoculated with the pTarget/bov TNF-α construct on four occasions 1 month apart. Seroconversion was assessed by screening on ELISA plates coated with 50 U/ml rbov TNF-α. Binding of mAb was detected with goat anti-mouse Ig biotin and strepavidin-HRP (SA-HRP) (Amersham Pharmacia Biotech, Uppsala, Sweden). 3,3′,5,5′-Tetramethylbenzidine (TMB, Sigma–Aldrich, Poole, UK) was used as a substrate. A week after the fourth injection, mice that had seroconverted were inoculated intraperitoneally with a final boost of TNF-α plasmid. Spleens were removed three days and fused with SP2/0 cells as previously described (Jones and Howard, 1995). Supernatants from wells containing hybridomas were tested for mAb reactivity to rbov TNF-α by ELISA as described above. Hybridomas were cloned by limiting dilution. Two mAb, termed CC327 (IgG2b) and CC328 (IgG2a), were selected for further characterization. These hybridomas were cultured in a Miniperm™ (Sartorius, Goettingen, Germany) and IgG purified from supernatants by passage through a Sepharose protein G column and transferred to PBS by passage through a PD10 desalting column (Amersham Pharmacia Biotech). Ig was coupled to biotin using NHS-biotin or NHS-long chain-biotin according to the manufacturer’s instructions (Pierce, Rockford, USA). The control mAb used were AV29 (mouse IgG2a) and AV37 (mouse IgG2a), which are directed against chicken CD4⁺ cells and chicken spleen cell subset, respectively. These mAb were kindly provided by Dr. C. Butter (IAH) and their application as isotype control mAb has previously been described for use as isotype control mAb in cattle studies (Hope et al., 2002). To develop a sandwich ELISA, CC327 and CC328 were tested by checkerboard assay for detection of both recombinant and native bov TNF-α. This was performed as described previously for other mAb (Hope et al., 2002, 2005; Kwong et al., 2002). Optimal detection of rbov TNF-α was observed using CC327 as coating mAb at 2 µg/ml and CC328-biotin as detection mAb at 1 µg/ml. Photometric and luminometric readout systems were applied as previously described (Hope et al., 2002; Hope et al., 2005; Kwong et al., 2002). TNF-α concentrations in samples were determined in comparison with a standard curve comprising serial dilutions of rbov TNF-α. Concentrations were expressed as biological units per ml (U/ml). The background level of luminescence (relative light units, RLU) or optical density (OD) in wells with only blocking buffer (PBS-casein) was between 5 and 10 RLU and 0.02–0.05 OD units, respectively. Typical titration curves from the photometric ELISA are shown in Fig. 1a (open symbols), where the limit of detection of rbov TNF-α was approximately 0.05 U/ml. The linear working range of the standard photometric ELISA was 0.05–3.7 U/ml. Comparable detection limits and linear ranges were observed in a number of experiments where luminometric and photometric ELISA detection systems were assessed in
parallel. The intra-assay variation was less than 10%. During these experiments, a commercially available ELISA for bov TNF-α became available (Pierce-Endogen, Rockford, IL, USA). Comparing our material with that ELISA, 1 U of COS cell expressed rbov TNF-α was found to be equivalent to 500 pg, indicating a detection limit of approximately 25 pg/ml and an upper range of 5000 pg/ml for the CC327/CC328 ELISA. Interestingly, this ELISA failed to detect rbov TNF-α expressed in Chinese Hamster Ovary (CHO) cells, although the Pierce-Endogen ELISA did (data not shown). The rbov TNF-α was derived from CHO cells transfected with cDNA encoding ovine TNF-α ligated into the pEE14 expression vector (Lonza Biologics plc, Slough, UK) according to previously published protocols (Entrican et al., 1996).

The capability of the CC327/CC328 ELISA to detect native bov TNF-α was assessed using supernatants from bovine monocyte-derived dendritic cells (DC) infected with M. bovis or M. bovis BCG for 24 h (Hope et al., 2004). Briefly, bovine DC were isolated and cultured overnight with M. bovis AF2122 (Garnier et al., 2003) at a multiplicity of infection (MOI) of 1, or with BCG Pasteur (Hope et al., 2000) at MOI of 10. Supernatants removed 24 h later were assessed by ELISA using luminometric readout (Fig. 1b). Significantly higher levels of TNF-α were secreted from DC infected with M. bovis compared to BCG (p < 0.01), or from control, uninfected DC (Fig. 1b).

The capacity of CC327 and CC328 to neutralize the biological activity of rbov TNF-α was assessed by bioassay using WEHI-164 cells as described above (Fig. 2). Doubling dilutions of purified CC327 and CC328 ranging from 10 to 0.04 μg/ml, plus isotype- and concentration-matched control mAb (AV29 and AV37), were added to wells containing 4 × 10^4 WEHI-164 cells in the presence of rbov TNF-α (4 U/ml) for 18 h. In the absence of mAb, rbov TNF-α induced greater than 60% cell death. In the presence of either CC327 or CC328 significantly lower levels of TNF-α-induced cell death were observed (p < 0.05; Fig. 2) indicating that both mAb neutralized the biological activity of rbov TNF-α (Fig. 2). Overall, CC328 was more effective (Fig. 2b), neutralizing TNF-α bioactivity over a wider range of mAb concentrations when compared to CC327 (Fig. 2a).

To detect intracytoplasmic TNF-α, bov peripheral blood mononuclear cells (PBMC) were stimulated for 4.5 h with PMA (50 ng/ml) and ionomycin (1 μg/ml) in the presence of brefeldin-A (10 μg/ml) (all Sigma–Aldrich, Poole, UK). The stable, cloned transfected CHO cell line described above was used for intracytoplasmic detection of ov TNF-α. In all cases, intracytoplasmic expression of TNF-α was assessed by flow cytometric analysis of fixed and permeabilised cells as previously described (Hope et al., 2002, 2005). Isotype-matched control mAb (AV29 and AV37) were included at concentrations that matched CC327 and CC328. Goat anti-mouse Ig isotype specific, FITC labelled, secondary reagents (Southern Biotec, Birmingham, AL, USA) were used. Intracytoplasmic bov TNF-α was detected with both CC327 (Fig. 3a) and CC328 (Fig. 3b). A different pattern of expression was observed between the mAb, with CC327 detecting a larger proportion of cells than CC328. Only mAb CC328 was able to detect intracytoplasmic expression of ov TNF-α (Fig. 3d), there was no evidence for cross-reactivity of CC327 with ov TNF-α (Fig. 3c). This explains the failure of the sandwich ELISA to detect ov TNF-α, despite a predicted 90% amino acid similarity between bov and ov TNF-α (McInnes et al., 1997). Although this ELISA does not detect ovine TNFα, and CC327 does not detect intracellular ovine TNFα by flow cytometry, this mAb has been reported to detect ovine TNFα in fixed tissues, which may be a result of epitope conformational change on fixation (Wheelhouse et al., 2009).

In summary, we have developed a sandwich ELISA for detection of bovine TNF-α and demonstrate the capability of CC328 to detect intracytoplasmic bovine and ovine TNF-α, thereby providing experimental capability to assess the role of this cytokine in ruminant immunity and disease pathogenesis.
Conflicts of interest

The mAb described in this paper are commercially available through AbD Serotec and royalties are paid to Institute for Animal Health.

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References


