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The Function of the Conserved Regulatory Element within the Second Intron of the Mammalian Csf1r Locus

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Abstract

The gene encoding the receptor for macrophage colony-stimulating factor (CSF-1) is expressed exclusively in cells of the myeloid lineages as well as trophoblasts. A conserved element in the second intron, Fms-Intronic Regulatory Element (FIRE), is essential for macrophage-specific transcription of the gene. However, the molecular details of how FIRE activity is regulated and how it impacts the Csf1r promoter have not been characterised. Here we show that agents that downmodulate Csf1r mRNA transcription regulated promoter activity altered the occupancy of key FIRE cis-acting elements including RUNX1, AP1, and Sp1 binding sites. We demonstrate that FIRE acts as an anti-sense promoter in macrophages and reversal of FIRE orientation within its native context greatly reduced enhancer activity in macrophages. Mutation of transcription initiation sites within FIRE also reduced transcription. These results demonstrate that FIRE is an orientation-specific transcribed enhancer element.

Introduction

Macrophage colony-stimulating factor (CSF-1) controls the proliferation, differentiation and survival of cells of the mononuclear phagocyte lineage [1,2]. CSF-1 mediates its actions by binding to the CSF-1 receptor (CSF-1R), a type III receptor protein tyrosine kinase. Expression of the Csf1r gene is switched on early in myeloid lineage commitment, and expression levels increase as immature myeloid cells differentiate into mature macrophages [3]. The Csf1r transcript in macrophages is expressed from a purine-rich promoter that lacks a TATA box and other classical elements that specify the transcriptional start site, which are characteristics shared with many other myeloid promoters [4–6].

The proximal promoter of Csf1r is not sufficient to generate maximal expression but requires the enhancer activity of a highly-conserved ~330 bp sequence in the second intron downstream of the macrophage promoter, which we named the Fms-Intronic Regulatory Element (FIRE; [7,8]). This element is functionally conserved between human and mouse [9] and the chicken Csf1r locus also contains a conserved element in an equivalent location within the first intron [10]. Deletion of FIRE from a GFP reporter construct containing the promoter and downstream intron 2 sequence abolishes GFP expression in transgenic mice [8]. FIRE contains multiple binding sites for myeloid transcription factors commonly found in myeloid promoters including PU.1, RUNX1, SPI/3, and AP-1 and lies within open chromatin in macrophages. FIRE was shown to possess anti-sense promoter activity in transient transfections of a macrophage cell line and subsequently, two anti-sense transcription start sites were mapped within FIRE [3]. Antisense promoter activity of FIRE was activated in B cells by PAX5, and was associated with repression of Csf1r expression [11].

Numerous genome-scale studies now indicate that promoter activity associated with enhancers is widespread, producing what have been called eRNA [12–14]. The function of these transcripts appears to be highly variable. In the case of the beta-globin locus, there is some evidence that transcription initiated from an upstream enhancer has an essential function in the generation of a chromatin loop that link enhancer and promoter [15] whereas other eRNAs appear to be bystander products of enhancer activity with as yet unknown functions [16]. In this paper, we present evidence that FIRE is an anti-sense RNA Polymerase II promoter in macrophages and is involved in fine-tuning Csf1r expression in
response to stimuli. We provide evidence that the enhancer activity of FIRE in its native context is orientation-dependent and requires transcription initiation sites.

Results

Anti-sense FIRE Promoter Activity is Increased in the Presence of Stimuli that Down-regulate Csf1r Expression

FIRE was shown previously to exhibit directional promoter activity in transient and stable transfection assays of the macrophage cell line RAW264.7 [7], suggesting a regulatory function of anti-sense transcripts initiating within FIRE. We therefore examined whether treatments that regulate Csf1r transcription acted through FIRE and tested three such stimuli, toll-like receptor agonists, CSF-1 and phorbol esters (PMA) [21] all of which impact Csf1r expression. In transient transfections using a construct carrying only the sense promoter, reporter gene expression was unaffected by TLR agonists (LPS, bacterial DNA) whereas FIRE anti-sense promoter activity was induced 3.2 fold with LPS stimulation and 1.7 fold, (p<0.05) by bacterial DNA treatment (Figure 1A & B). Since RAW264.7 cells express relatively low levels of receptor on the cell surface and do not respond to CSF-1 treatment [22], we generated a stable cell line over-expressing CSF-1R. This cell line was transfected with the Csf1r promoter or the FIRE enhancer cloned in the antisense orientation, and then treated with CSF-1, PMA, or both. CSF-1 actually induced the promoter as it has been reported to do in fibroblasts expressing CSF-1R [23]. Both CSF-1 and PMA induced FIRE promoter activity (Figure 1A & C). Together, these data demonstrate that the anti-sense promoter activity of FIRE is enhanced in the presence of stimuli that normally downregulate Csf1r expression.

Csf1r Primary and Antisense Transcripts Increase in Response to CSF-1 Deprivation While LPS & CSF-1 Cause a Rapid Down-regulation

FIRE is located in open chromatin in mouse and human macrophages [24] and tagged with H3K4Me3 suggesting an active promoter [25]. It has been previously reported that Csf1r mRNA expression is auto-regulated by its ligand. Growth factor withdrawal up-regulates Csf1r expression while inflammatory stimuli, such as LPS, down regulate expression. Both our own data, and others [26,27] indicate that Csf1r down-modulation in macrophages by LPS is associated with diminished production of transcripts as detected in nuclear run-on transcription assays. To confirm that this regulation takes place at the level of transcription initiation, we measured primary transcripts using different primers across the transcribed region. These measurements were combined with chromatin immunoprecipitation (ChIP) experiments testing for the recruitment of the initiating Serine 5 phosphorylated form of RNA-Polymerase II (RNA-Pol II) during a time course of stimulation (Figure 2A & 2B). BMMs were starved of CSF-1 or LPS for 24 hours to allow maximal upregulation of the surface Csf1r, and Csf1r mRNA [28]. They were then stimulated with a combination of LPS & CSF-1 to investigate the events that occur as Csf1r is acutely down-regulated. Both primary RNA levels as well as antisense RNA were measured during a time course of stimulation. These experiments confirmed that the withdrawal of CSF-1 from control cultures led to an up-regulation of Csf1r RNA transcription which was associated with increased RNA Pol II over the promoter. The addition of LPS & CSF-1 caused a rapid down-regulation of RNA levels within two hours (Figure 2A) and an even more rapid loss of initiating RNA Pol II (Figure 2B). Down-regulation of RNA Pol II binding and RNA levels occurred across the promoter as well as the FIRE enhancer, indicating that there was not a primary block in elongation. Low levels of active transcription were maintained for 16 – 24 hours whereas Serine 5 phosphorylated RNA-Pol II was replenished after 24 hours (Figure 2A & 2B). Primary RNA levels after 20 minutes of stimulation were still high while RNA Pol II levels were already strongly reduced. Moreover, increased amounts of RNA were detected over the FIRE enhancer. We therefore considered the possibility that this increase in RNA actually derives from antisense transcripts initiated at FIRE. Detectable anti-sense transcripts found in B cells were previously shown to initiate from two separate anti-sense transcriptional start sites within FIRE [29]. One is in the vicinity of the SP1/3/ETS/Egr-2/RUNX1 conserved element and the other overlaps an AP-1 binding site. To confirm that anti-sense transcripts are produced in primary macrophages, we performed RT-PCR on mRNA from murine macrophages using three distinct primer pairs, each of which produced bands of the predicted size that were below the limit of detection in control samples, but easily detected when cells were stimulated with CSF-1 or LPS (Figure 2C). To confirm that the increased RNA at 20 minutes (Figure 2A) when RNA Pol II levels were strongly suppressed (Figure 2B) is due to increased antisense transcription, we measured the synthesis of FIRE antisense (as) RNA using strand-specific primers (Figure 2D). Our results show a large induction of asRNA after 20 minutes of LPS & CSF-1 treatment indicating that asRNA is responsible for the increase in RNA. Unidirectional anti-sense initiation from FIRE and the location of the major TSS in primary mouse macrophages and in human macrophages is confirmed by genome-scale 5’ RACE (CAGE) data generated by the FANTOM consortium [30–31]. More recent CAGE data, sequenced at greater depth, confirms that FIRE is a unidirectional “broad” promoter, which unlike a typical TATA-less promoter, initiates transcription at multiple sites within a broad window [31]. Although there are two major TSS peaks, numerous minor peaks of initiation are detected within the 150 bp window surrounding the major TSS (Forrest A. et al. Ms submitted). The available data indicate that, in contrast to a major subclass of enhancers [32], FIRE transcription initiation is not bidirectional.

Combined CSF-1 and LPS Treatment Induces Transient Occupation of Sp1 and AP1 Binding sites within FIRE

Transcription factor occupancy on the Csf1r promoter and FIRE was assessed by dimethyl sulfate (DMS) in vivo footprinting over a time course of treatments with CSF-1 or LPS, as indicated in Figure 3A-C. These assays indicate protein contacts at the N7 position of guanines that protect these residues from being methylated and also highlight hyper-reactivity to modification at guanines juxtaposed to such a contact [33]. Untreated macrophages displayed the characteristic and complex transcription factor binding pattern at the promoter and FIRE described previously. We observed protein-DNA contacts over binding sites for PU.1 (ETS), C/EBP, Sp and EGR family members as well as RUNX1 consensus sequences [3] and this pattern remained unchanged when the cells, after overnight growth factor withdrawal, were treated with CSF-1 alone (Figure 3A). LPS treatment of CSF-1 starved cells did not change transcription factor occupancy at the promoter, but resulted in weak alterations of DMS-reactivity at FIRE (Figure 3B). Here the LPS-induced changes were restricted to an AP1 consensus sequence and to the RUNX1/Sp1/ETS element as indicated by a reduction in DMS reactivity (white squares). These two elements are juxtaposed to the two start sites for anti-sense transcription [29]. Combined
treatment with CSF-1 and LPS produced more pronounced changes (Figure 3C). DMS reactivity at the RUNX1/Sp1/ETS element and at the AP-1 site was reduced within 30 minutes of stimulation and returned to a pattern indistinguishable from unstimulated cells after sixteen hours. As previously described, no alterations of transcription factor occupancy were seen at the Csf1r promoter [3]. These data suggest that the RUNX1/Sp1/ETS and the AP-1 sites are maximally bound by protein in the presence of CSF-1 and LPS.

Binding of AP-1 family proteins to the urokinase enhancer has previously been shown to be induced by CSF-1 and PMA in bone marrow-derived macrophages (BMM) [34]. Since we found that the FIRE AP-1 site was occupied in our DMS footprint analysis in cells stimulated with CSF-1 and LPS, we sought to determine if a macrophage nuclear protein complex actually bound to the site. There are two AP-1 consensus binding sites within FIRE (Figure 4A). The consensus for the 5' AP-1 site (TGAATCA) and the 3' AP-1 site (TGAGTTC) conform imperfectly to the optimal AP1 consensus (TGA[G/C]TCA). Extract from both stimulated and unstimulated RAW264.7 cells as well as BMMs were compared to determine whether inducible macrophage nuclear proteins bind to either of the putative AP-1 elements in FIRE (Figure 4B). EMSA of the two candidate AP-1-like elements within FIRE showed binding of an inducible protein complex that could be competed for with an oligonucleotide competitor containing the AP-1 consensus sequence from the stromelysin promoter. Both sequences showed identical protein complexes of the same relative mobility and abundance (Figure 4B, data not shown). Binding of the AP-1-like protein complex was inducible by LPS in RAW264.7 cells and by LPS, CSF-1 or PMA in BMMs (Figure 4B). These data show that binding activity that recognizes the AP-1 element at FIRE is induced by agonists that repress full-length Csf1r. Together, the data demonstrate that maximal down-regulation of Csf1r transcription requires CSF-1 and LPS signalling and that these signals terminate at FIRE.

**Sp1 Binding Sites and Transcription Start Sites are Important for Promoter and Anti-sense Promoter Activity within FIRE**

Two major TSS within FIRE, located +2706 bp and +2760 bp downstream of the translational ATG start site (Figure 4A), were shown previously to be functionally active in B cells [11,20]. To test the function of the TSS and putative control elements in macrophages we generated a number of deletions and mutations of FIRE in a luciferase reporter and tested them in RAW264.7 cells (Figure 4C). The minimal promoter activity of FIRE resides on a small fragment (+2664 to +2723) containing two Sp1/3 sites.
which have been shown to be functional [3]. Progressive removal of other transcription factor binding sites from the 3′ end of FIRE, such as the AP-1 site overlapping the distal TSS, led to a progressive reduction in activity. Removal of the Sp1 sites by deletion of sequences upstream of +2725 severely reduced promoter activity.

We also examined six-base pair deletions spanning each antisense TSS (Figure 4D, left). The 5′ antisense transcriptional start site (AS TSS1; +2706) was dispensable for anti-sense promoter activity whereas the 3′ antisense start site (AS TSS2; +2760), was absolutely required. The six base pair deletion adjacent to ΔAS TSS2, Δ2753–2758 3′ in reverse orientation, were also required for anti-sense promoter activity but the six base pairs on the other side (Δ2765–2770) were not (Figure 4D, right). To confirm that the effect of deletion Δ2753–2758 on anti-sense promoter activity was not due to the distance and position relative to AS TSS1, we tested 3 other 6 bp deletions in the sequence between AS TSS1 and AS TSS2. Deletion of Δ2747–2752 or Δ2741–2746 had no effect on anti-sense promoter activity while deletion of Δ2735–2740 was required for anti-sense promoter activity. This indicates that the effect of deletion Δ2753–2758 is driven by the base pair motif and not the distance to AS TSS1 or 2 and that there are other necessary sites for anti-sense promoter activity.

The Enhancer Activity of FIRE is Orientation Dependent and Requires the Transcription Start Sites

FIRE enhancer activity is essential for basal Csf1r transcription so dissection of the role of antisense transcription by mutational analysis in mice is not straightforward. The traditional view of enhancers is that their function is orientation-independent. To test this, we produced a luciferase reporter construct in which the...
Figure 3. In vivo DMS footprinting of the Csf1r promoter and FIRE in stimulated BMM. Macrophages were differentiated from mouse bone marrow under the influence of CSF-1 (+) and subjected to DMS footprinting after either starving cells of CSF-1 (−) or restimulation with CSF-1 (A), LPS (B), or CSF-1 & LPS (C) for the indicated time points. G: Maxam-Gilbert reaction of naked genomic DNA are shown as vertical bars on the right hand side of the gel images. Nucleotide positions relative to the ATG start are designated by numbers on the left. Macrophage specific footprints are indicated as circles (black: enhancement, white: inhibition) while LPS responsive footprints are indicated as squares (black: enhancement, white: inhibition). L-shaped arrows are the position of antisense RNA.

The observation in B cells that PAX5 acts to repress Csf1r transcription by blocking the main promoter but also inducing the antisense promoter [11]. The enhancer activity of FIRE is associated with the deposition of active histone marks likely to require PU.1, C/EBP and RUNX1 (AML1), all of which bind to FIRE in both mouse and human. The latter protein binds to two sites within mouse FIRE, and expression of Ranl7 mRNA and nuclear binding activity is repressed by CSF-1 [28] which would be consistent with the up-regulation of Csf1r mRNA in the absence of CSF-1 (Figure 4A).

We previously provided evidence that the Csf1r proximal promoter was active in non-macrophage tumour cells, and was growth factor-responsive [23]. We suggested that FIRE acted in part to prevent read through of the intron, producing a block to transcription elongation, and that this might also explain the repression by LPS, PMA and CSF-1 [21]. However, this mechanism of regulation is incompatible with the data in Figure 2. Instead, the data demonstrates a complex interplay between the main promoter and FIRE in response to inflammatory stimuli which leads to direct and rapid repression of transcription initiation. That conclusion is consistent with earlier studies of LPS action using nuclear run-on transcription [37]. CSF-1 starvation of BMM produced an increase in primary Csf1r transcripts from the promoter and also from FIRE, paralleled by increased association of active RNA Pol II. Addition of LPS and CSF-1 caused an immediate increase of the antisense transcripts followed by a rapid decrease of active RNA Pol II occupancy and a slow decrease in primary transcripts across the coding region. This precise temporal correlation suggests that the interplay between sense and antisense transcription is of relevance for the regulation of Csf1r mRNA levels. However, while the effects of transcriptional interference have been described in yeast [38] there have been few studies of the consequences of bidirectional transcription initiation in mammals and more elaborate experimentation is necessary to dissect the precise molecular details at Csf1r.

The Activity of FIRE in situ is Orientation Dependent

Enhancers are supposed to act in a position and orientation-independent manner. However, reversing an enhancer in its native context has rarely formally tested this definition. Our experiments clearly demonstrate that in its native context the enhancer activity of FIRE is orientation dependent. Experiments with stably transfected cells and transgenic mice have shown that the Csf1r proximal promoter is active in non-macrophage tumour cells, and was growth factor-responsive [23]. We suggested that FIRE acted in part to prevent read through of the intron, producing a block to transcription elongation, and that this might also explain the repression by LPS, PMA and CSF-1 [21]. However, this mechanism of regulation is incompatible with the data in Figure 2. Instead, the data demonstrates a complex interplay between the main promoter and FIRE in response to inflammatory stimuli which leads to direct and rapid repression of transcription initiation. That conclusion is consistent with earlier studies of LPS action using nuclear run-on transcription [37]. CSF-1 starvation of BMM produced an increase in primary Csf1r transcripts from the promoter and also from FIRE, paralleled by increased association of active RNA Pol II. Addition of LPS and CSF-1 caused an immediate increase of the antisense transcripts followed by a rapid decrease of active RNA Pol II occupancy and a slow decrease in primary transcripts across the coding region. This precise temporal correlation suggests that the interplay between sense and antisense transcription is of relevance for the regulation of Csf1r mRNA levels. However, while the effects of transcriptional interference have been described in yeast [38] there have been few studies of the consequences of bidirectional transcription initiation in mammals and more elaborate experimentation is necessary to dissect the precise molecular details at Csf1r.
regulation of the lysozyme locus [39]. Here transcription from an inducible antisense promoter upstream of the normal transcription start site suppresses the activity of a silenter element by reorganising chromatin architecture and altering transcription factor binding. Cook et al. [40] first suggested that transcription arising from enhancers may act to focus enhancers and target promoters into active transcription factories, facilitating ciselement interaction and concentrating transcriptional regulators around the start site. The data herein could support such a mechanism of action for FIRE. A cis-acting mechanism is also favoured by a lack of trans-acting effects of the intron. Co-transaction of a 10-fold excess of the 7.2 kb Csf1r promoter fragment containing FIRE, had no effect on expression of the 7.2 kb Csf1r-luciferase reporter gene in RAW264.7 cells (data not shown). The fact that many enhancers generate transcripts makes it likely that the activity of such elements to stimulate transcription in their native chromatin environment could also be orientation dependent.

Materials and Methods

Ethics Statement

Animals were allowed free access to food and water and were maintained under temperature, humidity and light-controlled conditions. In accordance with the United Kingdom Animal (Scientific Procedures) Act of 1986, this study did not require a Home Office project license because no regulated procedures were carried out. Mice were humanely killed at a designated establishment by exposure to carbon dioxide gas and/or dislocation of the neck, which are appropriate methods under Schedule 1 of the Act.

Cell Culture

Bone marrow cells were isolated from the femurs and/or tibias of adult C57Bl/6 mice exactly as in Schroder, et al. [17] and cultured in RPMI or IMDM media containing penicillin-streptomycin, GlutaMAX-I supplement, and 10% fetal bovine serum. Cells were differentiated to macrophages by a 6–7 day treatment with 1000 units of human CSF-1 (Chiron Corp., Emeryville, CA) or with 10% L929 conditioned medium as a source of mouse CSF-1. RAW264.7 cells (obtained from the American Type Culture Collection) were cultured in RPMI medium with the above supplements.

Plasmids

The pGL0.5 fms, pGLFIRE-, pGL-7.2 fms, pGL-3.5 fms, pGL-7.2 fms ΔFIRE plasmids have been described previously [7]. All reporter constructs in Figure 4C carrying deletions or point mutations within FIRE have been described previously [11]. The pGL7.2 fms FIRE- plasmid was produced by deletion of the FIRE region using splice overlap PCR on an XhoI fragment at the end of intron 2 excised from the pGL7.2 fms plasmid. Primers for splice overlap introduced an AscI site into the sequence and FIRE was cloned in the inverted orientation before reinserion of the XhoI fragment into the pGL7.2 fms plasmid. In Figure 4D, the generation of pGLFIRE- six base pair (bp) deletion mutants were generated across the 337 bp FIRE sequence using splice overlap PCR using a modified protocol based on published reports [18,19]. Universal external PCR primers were designed with flanking AscI sites to facilitate cloning. TAGC AscFire CB F: 5'TACGCCGCCGCCCCAGGACGCGAGGAGGAGG' TAGC FIRE CB ASC R: 5'TACGCCGCCGCGGTACCAGTCTGCCCGTCCG3' Internal primers containing six bp deletions were systematically designed across FIRE with eighteen bp flanking each side of the deletion. PCR reactions contained a final concentration: 1 mM MgSO4, 0.3 mM each dNTP, 0.4 mM each primer and 0.5 U Platinum PfX Polymerase (Invitrogen, Carlsbad, IN). Cycling parameters were 94°C for 2 min; 94°C for 15 sec, 60°C for 30 sec, 68°C for 30 sec for 30 cycles followed by 68°C for 1 minute once. PCR products were gel purified using Qiagen Extraction Gel Kit (Qiagen, Valencia, CA) as per manufacturer’s instructions. Two microliters of each PCR reaction for each mutation was used for second round PCR. Second round PCRs were performed and cycled as above but no primers were added and fifteen cycles were performed. After fifteen cycles, the external primers (TAGC AscFire F2 and Asc Univ FIRE Rev) and 0.2 U Taq were added and cycled for 30 rounds of PCR. PCR products were purified using Qiagen Gel Extraction Kit, as above and then digested with AscI (NEB), and heat inactivated for 20 min at 65°C. These products were directly ligated into PGL2B that had an AscI site inserted into the Smal site to facilitate cloning purposes. Correct clones were confirmed by AscI digest and sequence analysis. In Figure 5D, the generation of pGL7.2 fms six base pair (bp) deletion mutants were generated using splice overlap PCR using a modified protocol based on published reports [18,19]. Universal external PCR primers were designed within FIRE as well as internal primers containing the same four 6 bp deletions from Figure 4D with eighteen bp flanking each side of the deletion. PCR conditions were the same as above. These products were directly ligated into PGL2B using native XhoI sites. Correct clones were confirmed by restriction enzyme digest and sequence analysis.

Transfection

Stable transfections were performed by electroporation of 5 × 10⁶ cells in 250 μl RPMI media containing 20 mM HEPES.
Figure 5. The enhancer activity of FIRE is orientation dependent and requires the transcription start sites. (A) Schematic of FIRE constructs: the entire Csf1r regulatory region plasmid (pGL-7.2 fms), pGL-7.2 fms with FIRE subcloned into the reverse orientation (pGL-7.2 fms FIRE-), pGL-7.2 fms with FIRE deleted (pGL-7.2 fms ΔFIRE), and pGL-7.2 fms with intron 2 deleted leaving 3.5 Kb of the Csf1r promoter (pGL2-3.5 fms). (B) RAW264.7 cells were transfected with pGL-7.2 fms, pGL-7.2 fms FIRE-, or pGL-7.2 fms ΔFIRE constructs and luciferase activity was assessed. Data is shown as a percentage of pGL2-7.2 fms (100%) and error bars represent the SEM. Statistically significant differences versus pGL-7.2 fms are indicated.
and 10 μg reporter with 1 μg pPNT neomycin resistance plasmid at 280 volts and 1000 μFarad capacitance on a Bio Rad Gene Pulser (Bio Rad), followed by selection with 250 μg/ml Geneticin. For stable transfection, antibiotic resistant cells with or without 48 hours of LPS treatment were analysed on a Facstar flow cytometer (Becton Dickinson). The extended treatment with LPS was required to see repression of the stable EGFP protein. Transient transfections were performed as above with 10 μg of reporter plasmid without the additional antibiotic resistance plasmid. In Figure 4C, RAW264.7 cells were transfected using a 1:3 ratio of lipofectamine and plasmid (0.6 μg of FIRE constructs reporter vectors, 0.25 ng of Renilla and 0.2 μg of pBluescript) in 250 μL Opti-mem (Gibco; Invitrogen). Cells were transfected for 24 hours.

Luciferase Assay

Luciferase activity was assayed according to the manufacturer’s protocol (Roche Biochemical), pGL2 basic and pGL2 luciferase reporter vectors were used as controls. The concentration of protein was determined by BCA protein assay (Pierce) and the level of luciferase activity was given as relative light units (RLU), calculated as light units/μg of protein assayed. Samples in Figure 4C were assayed for luciferase activity using the Promega Dual Luciferase assay system.

Antisense RNA Assay

Antisense RNA-expression was assayed exactly as described in [20]. Briefly, cDNA was synthesised from 2 μg of DNaseI-treated total RNA by using 400 U of M-MLV reverse transcriptase and biotinylated primers specific for antisense-Cysf4 transcript ([Bu]GGTCAGCAAAACAGGAAGAGCCGACAGAC-GAG) (2 pmol) and rDNA ([Bu]GGACCGGCAAGACGGACAGGACAGA-GAG) (0.2 pmol) in one reaction. Synthesised cDNA was immobilised on Dynabeads (DynaL, M-280). RNA and trace amounts of genomic DNA were removed by alkaline denaturation and serial washing. cDNA was eluted by heating the bead suspension in 0.1TE for 15 min at 95°C and was followed by qPCR using antisense-Cysf4 (FIRE4 for: TGTTGCCAGTCTGCTTTCA and FIRE4 rev: CTCCTGCGCATTGCTCTTCTC) and rDNA (rDNA for: CTTGCCTTTTTCTTAAACT and FIRE4 rev: GAA- TAGGCTGGACAGCAAAACA) primers. Primers amplification efficiency was calculated using genomic DNA as a standard. Antisense-Cysf4 expression was normalised against rDNA signals.

Chromatin Immunoprecipitation Experiments

Sub-confluent bone marrow derived macrophages were harvested and washed in cold PBS. Chromatin was cross-linked with 1% formaldehyde (Thermo Fisher Scientific, IL USA) for 1 hour at 4°C. The cells were lysed in 10 mM HEPES (pH 8), 10 mM EDTA, 0.5 mM EGTA, 0.25% Triton X100 and protease inhibitor cocktail (PIC). The nuclei were collected by centrifugation and lysed in 10 mM HEPES (pH 8), 200 mM NaCl, 1 mM EDTA, 0.5 mM EGTA, 0.01% Triton X100 and PIC. Chromatin was transferred to IP buffer containing 25 mM Tris-HCl (pH 8), 2 mM EDTA, 150 mM NaCl, 1% Triton X100, 0.25% SDS, PIC and sheared using a Bioruptor sonication water bath (Diagenode) to an average length of 200–500 bp. The chromatin solution was diluted in 25 mM Tris-HCl (pH 8), 2 mM EDTA, 150 mM NaCl, 1% Triton X100, 7.5% glycerol and antibody precipitations were performed. Chromatin from 10⁶ cells and 10 μl of Dynabeads protein G (Invitrogen) coupled with 1 μg of RNP II S5 antibody (ab131; Abcam, Cambridge, United Kingdom) were incubated for 2 hours at 4°C with rotation. The immune complexes were collected using a magnet separator and washed with low salt wash buffer 1 (20 mM Tris-HCl (pH 8), 2 mM EDTA, 1% Triton X100, 0.1% SDS, 150 mM NaCl), high salt wash buffer 2 (20 mM Tris-HCl (pH 8), 2 mM EDTA, 1% Triton X100, 0.1% SDS, 500 mM NaCl), LiCl buffer (10 mM Tris-HCl (pH 8), 1 mM EDTA, 0.5 mM EGTA, 250 mM LiCl), LiCl wash buffer (25 mM Tris-HCl (pH 8), 0.5 mM EDTA, 250 mM LiCl) and LiCl wash buffer 2 (25 mM Tris-HCl (pH 8), 0.5 mM EDTA, 250 mM LiCl, 1% SDS, 150 mM NaCl). After precipitation, DNA was quantified using a NanoDrop spectrophotometer (Thermo Fisher Scientific, IL USA). Chromatin was immunoprecipitated with anti-C57 (ab5131; Abcam, Cambridge, United Kingdom) before the ChiP assay.
EDTA, 0.25 M LiCl, 0.5% NP40, 0.5% Na deoxycholate) and TE/NaCl buffer (10 mM Tris-HCl (pH 8), 1 M EDTA, 50 mM NaCl). Precipitated DNA was eluted in 1% SDS and 100 mM NaHCO3 and followed by crosslink reversal by heating at 65°C overnight. Chromatin DNA was purified using Ampure PCR purification kit (Agencourt, AMP00130) and quantified using real-time qPCR with SYBR Green. Primers used in this assay were Prom -1.5 kb (For: CAGCCTGGCTGTAGTGTCTGT and Rev: TCCACGTTAGATGGTGTCAGCAT), c-fms prom (For: CTGCTGTTGCGACAGTTT and Rev: CAGC- GATGCCCCCTTGTGCTG), Fire 5’ (For: GGAACCCCT- GAACTGCTCTTAAAG and Rev: TGACCCCGCAAGT- CAACC), Fire 3’ (For: CAGACTGGGTACCTCTCTCCT- CTAC and Rev: GCGGAAAAACGTTGTTGCTCAT) and intron 2 (For: TGACAAAGCACCATTAGGCAA and Rev: GATCAAAGGCTGTACGCT).

**In vivo DMS Footprinting**

Bone marrow macrophages (BMM) were deprived of CSF-1 for 24 hours, and then restimulated with CSF-1 (10% L929 cell conditioned medium) or 20 ng/ml LPS or the combination as indicated. At the end of each incubation time, cells were washed with PBS and treated with 0.2% DMS/PBS for 5 minutes at room temperature. After three washes with ice-cold PBS, genomic DNA was extracted from cells and subjected to pipeline cleavage and Ligation-mediated PCR (LM-PCR) amplification. LM-PCR was performed as described previously [3].

**EMSA**

Isolation of nuclei and extraction of protein from BMMs and RAW264.7 cells was performed according to established protocols and the concentration of protein in nuclear extracts was determined by BCA protein assay. The double stranded oligonucleotide probes used contained the following sequences; CSF-1R Sp1: CAGGGTCGCGGAAACCCA; CSF-1R Ap-1: AGGCCTGTGATCAGCCTACA. Binding reactions contained 20 mM HEPES buffer pH 7.4, 40 mM NaCl, 15% glycerol, 1 mg of protein from nuclear extract, 25 ng pol (dT-ol) and approximated 80 fmol of labelled probe for 20 minutes at room temperature. Antibody supershifts were performed by pre-incubating binding reactions with α-Sp1 antibody (Santa Cruz Biotechnology) for 15 minutes at 4°C before addition of probe and protein binding. Competitions were performed by addition of unlabeled oligonucleotide corresponding to the strongbinding AP-1 consensus element at a ratio of 20 to 1 immediately before addition of labeled probe. Protein binding was resolved on 5% mini-polyacrylamide gels (Bio Rad) in 0.5 X TBE (45 mM Tris base, 45 mM boric acid, and 1 mM EDTA) for 1 hour at 100 volts.

**Author Contributions**

Conceived and designed the experiments: KS AB JO DS HT RI CP CB DH. Performed the experiments: KS AB JO DS HT RI CB DH. Analyzed the data: KS AB JO DS HT RI CB DH. Contributed reagents/materials/analysis tools: CB DH. Wrote the paper: KS AB JO DS HT RI CB DH.

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