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Forkhead Transcription Factor Fd3F Cooperates with Rfx to Regulate a Gene Expression Program for Mechanosensory Cilia Specialization

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SUMMARY
Cilia have evolved hugely diverse structures and functions to participate in a wide variety of developmental and physiological processes. Ciliary specialization requires differences in gene expression, but few transcription factors are known to regulate this, and their molecular function is unclear. Here, we show that the Drosophila Forkhead box (Fox) gene, fd3F, is required for specialization of the mechanosensory cilium of chordotonal (Ch) neurons. fd3F regulates genes for Ch-specific axonemal dyneins and TRPV ion channels, which are required for sensory transduction, and retrograde transport genes, which are required to differentiate their distinct motile and sensory ciliary zones. fd3F is reminiscent of vertebrate Foxj1, a motile cilia regulator, but fd3F regulates motility genes as part of a broader sensory regulation program. Fd3F cooperates with the ciliary transcription factor, Rfx, to regulate its targets directly. This illuminates pathways involved in ciliary specialization and the molecular mechanism of transcription factors that regulate them.

INTRODUCTION
The cilium commonly constitutes a cell’s organelle for environmental sensing in a wide variety of contexts, from developmental signaling pathways (such as Sonic hedgehog) to the specialized receptor processes in sense organs of various sensory modalities (Ishikawa and Marshall, 2011). In other situations, cilia are motile and play many roles connected to fluid movement in the airways, CNS, oviduct, and embryonic node. Despite this structural and functional diversity, cilia share a highly conserved pathway of ciliogenesis, involving basal body docking, axoneme extension, intraflagellar transport (IFT), and ciliary membrane assembly (Silverman and Leroux, 2009). A major question is how this common assembly program is adapted and modified with cell-specific variations to generate cilia diversity (Silverman and Leroux, 2009). It is likely that such specialization requires cell-type-specific gene expression programs, but little is known of the nature of these programs or of the transcription factors that regulate them (Thomas et al., 2010).

In metazoans ciliogenesis broadly depends on regulatory factor X (Rfx) transcription factors (Chu et al., 2010). Their targets are well characterized in functional and bioinformatic studies on C. elegans and Drosophila (Avidor-Reiss et al., 2004; Efimenko et al., 2005; Jacquet et al., 2009; Stubbs et al., 2008; Yu et al., 2008). For instance, Foxj1 knockout mice have left-right asymmetry and airway defects (Brody et al., 2000). Target gene analyses have shown that Foxj1 regulates, directly or indirectly, many genes linked to ciliary motility, including axonemal dyneins, but not core ciliogenesis, such as IFT factors (Jacquet et al., 2009; Stubbs et al., 2008; Thomas et al., 2010; Yu et al., 2008). However, very little is known of its molecular mode of action or its relationship with Rfx function.

In Drosophila the only somatic cells with cilia are bipolar sensory neurons, which have specialized ciliary dendrites for sensory reception and transduction (Figures 1A–1C). Different classes of such neurons have ciliary dendrites that have different morphologies, express different sets of receptor molecules, and respond to different sensory modalities. These neurons present a useful model for investigating how ciliary diversity arises. External sensory (ES) neurons have a short connecting cilium leading to a distal sensory process with an irregular core of microtubules. In contrast, proprioceptive and auditory chordotonal (Ch) neurons have a long sensory cilium with a well-defined 9+0 axoneme (Figures 1B and 1C) (Eberl et al., 2000; Kernan et al., 1994). The Ch cilium has several unique specializations. First, its mechanosensory transduction mechanism uniquely involves TRPV (transient receptor potential) channels encoded by nanchung (nan) and inactive (iav) (Gong et al., 2004; Kim et al., 2003). Second, it can be motile. In the auditory Ch neurons of Johnston’s organ in the antenna (Figure 1B), motility arises from the interplay between transduction channels and adaptation motors (possibly axonemal dyneins). Sensory-induced
motility is thought to underlie a mechanical amplification mechanism to increase auditory sensitivity (Go¨pfert et al., 2005; Go¨pfert and Robert, 2003). Third, the Ch neuron cilium is divided by a ciliary dilation into functionally distinct proximal (motile) and distal (sensory) zones (Figure 1C) (Lee et al., 2008).

We asked, therefore, what regulates gene expression for Ch neuron-specific ciliary specialization. Rfx is expressed in and required for ciliogenesis in both ES and Ch neurons (Dubruille et al., 2002; Laurenc ¸ on et al., 2007), whereas Foxj1 homologs are reportedly absent from Drosophila (Larroux et al., 2008; Mazet et al., 2003). We recently reported, however, that the Fox gene, fd3F, is required for Ch neuron function (Cachero et al., 2011). Here, we show that fd3F does not regulate ciliogenesis per se but directly regulates the genes required for aspects of Ch ciliary specialization. It appears that Fd3F cooperates closely with Rfx to regulate this Ch-specific cohort of genes and, therefore, it acts as a cell-type-specific modulator of Rfx target gene specificity. Comparison with Fox1 and its target genes suggests that fd3F is a highly diverged relative of Fox1.

RESULTS

fd3F Mutation Results in Uncoordinated Flies with Nonfunctional Ch Neurons

Previously, a transcriptome analysis of embryonic Ch cells revealed that fd3F (CG12632) was highly enriched in developing Ch neurons, and RNA in situ hybridization and immunostaining showed that fd3F is exclusively expressed in Ch cell lineages during embryogenesis (Cachero et al., 2011) (Figures 1D and 1E). In larval imaginal discs, fd3F expression was confined to locations of Ch precursors, including the femoral Ch precursors in the leg disc (Figure 1F) and the Ch precursors of Johnston’s organ in the antennal disc (data not shown). In summary, fd3F expression is unique to developing and differentiating Ch neurons.

Using the fly stock P{EP}fd3FEP1198, we generated a deletion of 1.4 kb (fd3F1) that removes approximately 400 bp of the 3’ end of the fd3F ORF (see Figure S1A available online) (Cachero et al., 2011). fd3F mRNA expression was very strongly reduced in homozygous mutant embryos (Figures S1B and S1C), suggesting that the truncated fd3F transcripts are unstable and that the mutation is close to a genetic null. Consistent with this, little difference was observed between the phenotypes of fd3F1 homozygotes and hemizygotes over a deficiency (fd3F1/Df(1)ED6716) in the analyses below.

fd3F1 homozygote flies are viable and fertile but are uncoordinated (Cachero et al., 2011), and hence, the adult flies perform very poorly in a climbing assay (Figure 2E). This suggests that Ch neurons are defective in fd3F mutants. However, Ch neurons were present in fd3F1 embryos, larvae, and pupae and showed little gross morphological defect (Figures 2A and 2B). When marked by the expression of mCD8-GFP, the Ch sensory cilia
were present and grossly normal (Figures 2C and 2D). To assess the functional integrity of Ch neurons in fd3F mutants, we examined the activity of the auditory Ch neurons of Johnston’s organ. Compound action potentials (CAPs) were recorded in response to auditory stimulus from the antennal nerve of adult flies. Unlike wild-type flies, fd3F mutant flies showed no sound-evoked CAPs in the antennal nerve (Figure 2F). These data suggest that fd3F regulates aspects of Ch neuronal differentiation that are essential for Ch neuron function.

**Figure 2. Mutation of fd3F Results in Functionally Defective Ch Neurons**

(A and B) Cell bodies and dendrites of Ch neuron group (lch5) in third-instar larvae stained with anti-HRP.
(C and D) Adult femoral Ch organ neurons showing sensory cilia (arrows) labeled with mCD8-GFP (elavGal4;UAS-mCD8-GFP). Scale bars, 5 μm.
(E) Climbing assay showing percentage of adult flies climbing above a threshold. Genotypes are wild-type (w1118), flies homozygous for the P element insert at fd3F (p{EP}), fd3F heterozygotes (fd3F/FM6 balancer), and fd3F homozygotes. Error bars are SD. ***p < 0.0001.
(F) Shown on the left are relative CAP amplitudes recorded from the projections of Johnston’s organ neurons in the antennal nerve (inset) as a function of the sound particle velocity. Shown on the right are corresponding absolute values of the maximum CAP amplitudes (n = 5 flies per strain). Error bars are SD.
(G–L) Genes that show reduced expression in fd3F mutant embryos. Wild-type (left) and mutant (right) embryos showing mRNA expression of Ch-expressed genes (named at left) are shown. Expression is either virtually absent (e.g., Dhc93AB) or reduced (e.g., CG11253). Scale bars, 100 μm.

See also Figure S1.

**fd3F Regulates a Subset of Ch-Specific Genes**

As we previously reported (Cachero et al., 2011), fd3F is required for the expression of the genes nan and iav, which encode subunits of a TRPV cation channel uniquely localized to Ch neuron cilia (Gong et al., 2004; Kim et al., 2003). To identify other regulatory targets of fd3F, we began by examining candidate Ch-expressed genes identified previously in our transcriptome analysis (Cachero et al., 2011). Of 16 genes tested initially, 6 showed reduced expression in fd3F mutant embryos.
Thus, fd3F does not regulate all Ch neuron-specific genes. Instead, the nature of the target genes suggests that fd3F regulates specific processes in Ch sensory specialization.

**fd3F Regulates Genes Required for Ch Ciliary Motility**

The fd3F target, Dhc93AB (Figures 2G and 2H), encodes an axonemal outer-arm dynein heavy chain (homolog of human DNAH9/11). Axonemal dyneins form a large family of multisubunit motors that are responsible for ciliary motility. In Drosophila embryos, Ch neurons bear the only cilia with axonemal dynein arms, and correspondingly, Dhc93AB is expressed in a Ch-specific pattern. In Ch neurons of Johnston’s organ, nonlinear mechanical amplification arises from the motility of Ch cilia (Goepfert et al., 2005; Goepfert and Robert, 2003; Nadrowski et al., 2008). Axonemal dyneins are good candidates to contribute to this process. Like vertebrate hair cells, Ch neurons use mechanical amplification to augment the tiny sound-induced vibrations that they transduce. Active amplification can be tested by examining the mechanical response of the antennal sound receiver to tones of different intensities (Goepfert et al., 2006). In wild-type flies this response displays a compressive nonlinearity that enhances sensitivity about 10-fold when sound is faint (Figure 3A). This nonlinearity was completely absent in fd3F mutant flies (Figures 3A and 3B), demonstrating that Ch cilium motility is lost.

Besides Dhc93AB, a number of other axonemal dynein components are present in transcriptome of differentiating embryonic Ch neurons (Cachero et al., 2011) (Table S1), and several of these were confirmed to have Ch-specific expression by in situ hybridization (Figures 3B, 3D, and 3F; Table S1). This suggests that axonemal dyneins are important in embryonic (proprioceptive) as well as antennal (auditory) Ch neurons. Of these genes, almost all were strongly reduced or absent in fd3F mutants (L and N), demonstrating that Ch cilium motility is lost.
CG13930 (IC138, WDR78) (Figures 3B–3G; Table S1). Together, these observations suggest that fd3F regulates axonemal dynein genes in general. To check that this also pertains to the Johnston’s organ, Ch neuron ultrastructure in the antenna was examined by transmission electron microscopy (TEM). In transverse sections of mutant ciliary dendrites, normal axoneme arrangement and ultrastructure were observed, but dynein outer and inner arms were missing from the proximal cilium (Figures 3H–3J). Loss of dyneins with consequent loss of motility can account for the loss of mechanical amplification by Johnston’s organ neurons in fd3F mutants, although fd3F might also regulate genes of the sensory transduction apparatus that are required for the process (Nadrowski et al., 2008).

Several genes have been implicated in axonemal dynein assembly. titB is required for transport or assembly of axonemal dyneins (Kavlie et al., 2010), and CG14905 is similar to Chlamydomonas ODA-1, which is required for axonemal dynein assembly (Kamiya, 1988). Ch neuron expression of both genes was reduced in fd3F mutant embryos (Figures 3K and 3L; data not shown). Tektins are required for correct motility of cilia, probably by participating in dynein inner-arm assembly or attachment (Amos, 2008). We found that Tektin-A (TEKT4 ortholog) is expressed exclusively in Ch neurons and that its expression is strongly reduced in fd3F mutants (Figures 3M and 3N). Thus, fd3F regulates genes required for several aspects of dynein arm formation.

### Fd3F Regulates Retrograde Transport Genes to Form Functionally Specialized Ciliary Zones

The fd3F target, CG3769 (Figures 2I and 2J), encodes the ortholog of cytoplasmic dynein 2 light-intermediate chain (DYN2LIC). Cytoplasmic dynein 2 is the motor for retrograde IFT during ciliogenesis, and dynein 2 heavy chain (DYN2HC), encoded by beethoven (btv), is required for Johnston’s organ function (Eberl et al., 2000). Unlike axonemal dyneins, btv expression is not strictly confined to Ch neurons: it is expressed preferentially in Ch neurons but also at a lower level in ES neurons (the so-called Ch-enriched pattern; Cachero et al., 2011) (Figure 4A). This is consistent with retrograde transport being required for ciliogenesis in both sensory neuron subtypes. Like CG3769, btv mRNA is reduced in the Ch neurons of fd3F mutant embryos (Figure 4B). However, some expression remains in Ch neurons, and ES neuron expression appears unaffected.

Retrograde transport also requires proteins of the IFT-A complex, and we previously found that IFT-A genes are also expressed in the Ch-enriched pattern (Cachero et al., 2011). In fd3F mutant embryos, rempA (IFT140) and Oseg6 (IFT144) mRNAs were both reduced in Ch neurons (Figures 4C and 4D; data not shown). Oseg1 was also reduced in whole embryos as shown by RT-PCR (data not shown). In contrast to retrograde transport genes, the antero-IFT-B genes, nompB (IFT88) and CG15161 (IFT48), were not affected in fd3F mutants (Figures 4E and 4F; data not shown).

In Chlamydomonas and other models, disruption of retrograde IFT causes a buildup of proteins at the tip of the cilium, which then becomes swollen (Pazour et al., 1998). This phenotype was present in fd3F mutant Johnston’s organ dendrites: in transverse sections the tips of the cilia fill the dendritic cap (Figures 4G and 4H). However, the cilia did not appear truncated, as is seen in rempA null mutants (Lee et al., 2008), consistent with only a partial loss of retrograde IFT. A second feature of retrograde transport mutants is the accumulation of IFT-B particles in the ciliary tips (Hou et al., 2004). We observed a moderate accumulation of NomBP-GFP fusion protein in the tips of fd3F mutant cilia (Figures 4I and 4J).

If fd3F is responsible for elevated expression of retrograde transport genes in Ch neurons relative to ES neurons, this implies that increased expression has a Ch neuron-specific function. Indeed, although retrograde transport is required for ciliogenesis generally, rempA has an additional Ch-specific role of creating and maintaining the distinct proximal motile and distal nonmotile ciliary zones, separated by the ciliary dilatation (Lee et al., 2008) (Figure 1A). These zones differ in ion channel composition, with Nan and lav (TRPV) localized to the proximal zone, whereas the distal zone contains the candidate force-responsive ion channel, NompC (TRPN1 homolog) (Cheng et al., 2010; Liang et al., 2011) (Figures 4M and 4O). In fd3F mutant embryos, defective ciliary dilatation staining was observed similar to that reported for btv mutants (Eberl et al., 2000) (Figures 4K and 4L). We investigated whether this reflects a disruption of ciliary compartmentalization. As noted above, proteins known to be restricted to the proximal zone (axonemal dyneins, Nan, lav) are themselves fd3F targets, and so their localization cannot readily be analyzed. However, distally localized NompC protein could be examined because its transcription is independent of fd3F (data not shown). In fd3F mutant larvae and adult antennae, NompC protein was not restricted distally but was instead spread along the length of the cilium, suggesting a disruption of ciliary compartmentalization (Figures 4M–4P). Consistent with this, the axonemal ultrastructure of the mutant proximal cilium appears to resemble that of the distal cilium (a dense core is seen in only one microtubule of the doublet) (Figure 3I; data not shown). The RempA-YFP protein itself normally concentrates in the region of the ciliary dilatation (Lee et al., 2008) (Figure 4Q). However, in fd3F mutant neurons, RempA-YFP was dispersed along the proximal cilium (Figure 4R). In conclusion, fd3F not only regulates genes whose products are localized to the proximal motor zone (axonemal dyneins, lav, Nan) but also upregulates genes required to form and maintain the distinct zones (cytoplasmic dynein 2, IFT-A components) (Figure 4S).

### Direct Regulation of nan and iav by Fd3F

We examined the cis regulation of nan and iav. For nan a 557 bp upstream fragment (−561 to −4, relative to ATG) drives GFP reporter expression specifically in embryonic Ch neurons (nan-GFP) (Kim et al., 2003) (Figures 5A and 5B). In fd3F1 mutant embryos, nan-GFP expression was abolished (Figure 5D), showing that this enhancer is responsive to fd3F. Similarly, for iav a 597 bp upstream fragment was able to drive Ch-specific GFP expression in an fd3F-dependent manner (Figures 5A, 5C, and 5E). Fox factors of several subfamilies bind to a common sequence in vitro, RYMAAYA (Benayoun et al., 2011). The nan and iav enhancers contain several such motifs. Mutation of the Fox motifs closest to the iav and nan translational start sites (iav-F1 and nan-F1) abolished iav-GFP and nan-GFP reporter gene expression (Figures 5F and 5G). In contrast, mutation of Fox motifs slightly further upstream (F2 in Figure 5A) had no
Figure 4. *fd3F* Regulates Genes Required for Ciliary Compartmentalization

(A–D) mRNA expression of retrograde IFT-related genes in wild-type and *fd3F* mutant embryos. These genes show a Ch-enriched expression pattern of which the Ch neuron component (ringed and marked with arrows) is reduced in *fd3F* mutants.

(E and F) The expression of an anterograde IFT gene, CG15161 (in a pattern consistent with expression in all ciliated sensory neurons), is unaffected in *fd3F* mutant embryos. Scale bars, 20 μm.

(G and H) TEM cross-section of the tip of a Ch organ unit from adult antennal Johnston’s organ. The Ch cilium tips (ci) are housed within the dendritic cap (c) (schematic shows the location of the cross-section). As shown in (G), the cilium tips are enlarged to fill the cap in the *fd3F* mutant antenna (H).

(I and J) NompB-GFP fusion protein accumulates at the tips of Johnston’s organ Ch neurons in *fd3F* mutant pupal antennae (cell bodies counterstained with 22C10).

(K) In wild-type embryonic Ch neurons, anti-HRP marks the Ch cilia including the ciliary dilation (arrowheads).

(L) The dilation is not apparent in Ch cilia from *fd3F* mutant embryos (arrowheads indicate the expected location of the dilation).

(M–P) NompC protein is localized to the distal Ch cilia in wild-type neurons (M and O) but becomes dispersed along the cilium in *fd3F* mutant neurons (N and P). Lines indicate the approximate extent of the cilium.

(Q) RempA-YFP protein is localized in the vicinity of the ciliary dilation in wild-type Ch cilia.

(R) In *fd3F* mutant cilia, RempA-YFP protein becomes dispersed.

(S) Schematic summary of protein localizations in wild-type and mutant Ch neuron cilia. Scale bars, 4 μm (I, J, and O–R) and 5 μm (K–N). See also Table S1.
effect on reporter gene expression (data not shown). To test whether Fd3F can bind to these F1 motifs, the DNA binding Forkhead domain of Fd3F (Fd3F Fkd) was expressed as a GST fusion protein and isolated from bacterial cells. When used in an in vitro gel mobility shift assay, GST-Fd3F Fkd was able to bind specifically to the F1 sites of both nan and iav (Figure 5H).

Coregulation of Ciliary Specialization Genes by Fd3F and Rfx Transcription Factors

Typically, Rfx factors regulate target genes via a single upstream binding site (X box) (Efimenko et al., 2005; Laurenc¸on et al., 2007). The iav and nan enhancer sequences contain an X box motif similar to that determined biochemically for human Rfx1 (GTNRCC[0-3]RGYAAC; Emery et al., 1996), very close to the functional Fd3F binding sites (Figure 5A). This raises the possibility that although Rfx is a pan-ciliary transcription factor required for both Ch and ES neurons (Dubruille et al., 2002), it contributes to the regulation of Ch-specific Fd3F target genes. Indeed, both nan and iav expression were strongly reduced in Rfx mutant embryos (Figures 5I and 5J; data not shown). In both cases mutation of the X box in the reporter constructs largely abolished enhancer activity (Figures 5K–5N). We therefore conclude that an X box/Fox motif combination is required for nan and iav regulation in Ch neurons and that most likely these sites are bound by Rfx and Fd3F. In support of the conclusion that these transcription factors are coregulators, we

Figure 5. Fd3F Directly Regulates nan and iav with Rfx
(A) Schematics of the nan and iav genes showing locations of Fox motifs (F) and X boxes (X) in their upstream regions. The sequences surrounding the X boxes are shown, with asterisks representing identity in D. pseudoobscura.

(B and C) Stage 16 embryos showing expression of GFP in Ch neurons for nan-GFP (B) and iav-GFP (C) reporter genes.

(D and E) Expression of both reporter genes is lost in fd3F mutant embryos.

(F and G) Expression of both reporter genes is abolished upon mutation of a single Fox motif (F1) in either upstream region.

(H) Gel mobility shift assay with oligonucleotide probes (*) containing the nan-F1 or iav-F1 Fox motifs and purified Fd3F forkhead domain polypeptide (Fd3F Fkh). The DNA-Fd3F Fkh complexes are indicated. Addition of 100-fold excess of cold competitor oligonucleotide (nan-F1, iav-F1) reduces complex formation, but when the Fox motif is mutated (nan-F1m, iav-F1m), the cold competitor has no effect on complex formation.

(i and J) Embryonic expression of iav in Ch neurons (I) is lost in Rfx49 homozygous mutant embryo (J) (embryos stained in parallel).

(K–N) Two abdominal segments from stage 16 embryos stained with anti-GFP (green) and 22C10 (magenta). Ch neuron expression of nan-GFP (K) and iav-GFP (M) reporter genes is abolished when the upstream X box is mutated (L and N).

Scale bars, 100 µm (B–J) and 20 µm (K–M). See also Table S1.
found no evidence of a regulatory hierarchy between Rfx and fd3F: in embryos we found that fd3F transcription does not depend on Rfx function and vice versa (data not shown).

When we examined a selection of other fd3F target genes (including Dhc93AB, rempA), we found that their expression was also strongly reduced in Rfx mutant embryos (Table S1). Moreover, we observed that almost all fd3F target genes contain a closely spaced combination of conserved Fox and X box motifs, usually within 50 bp of the transcriptional or translational start site (Figure 6A; Table S2). Some of these genes were suspected Rfx direct targets from other bioinformatic analyses (Avidor-Reiss et al., 2004; Laurençon et al., 2007) (Table S2).

However, most of these genes were not previously predicted to be Rfx targets, partly due to the fact that the X box motif often does not conform completely to the palindromic consensus sequence previously considered for Drosophila targets: GTTGGCCATGGCAAC (Avidor-Reiss et al., 2004); GYTRYYN(1-3)RRHRAC (Laurençon et al., 2007) (Figure 6B). The juxtaposition of Fox motifs and X boxes suggests that Fd3F and Rfx are coregulators of a subset of Ch ciliary genes. Subsequent to the bioinformatic analysis, we confirmed that the regulation of Dhc93AB indeed mirrors that of nan and iav. An 820 bp promoter fragment supports Ch neuron-specific reporter gene expression, and mutation of the proximal-most X box and Fox motifs reduced this expression. However, in this case expression was not completely abolished, possibly due to redundancy with other binding sites that are present in this longer enhancer (Figures 6C–6F).

### Target Genes Are Ectopically Activated upon Fd3F Misexpression

Using flies transgenic for an inducible UAS-fd3F construct, we ectopically expressed Fd3F in embryonic sensory precursor cells using a scaGal4 neuroectodermal driver line (Figures 7A and 7B). This induced the misexpression of several fd3F target genes in ES neurons, including CG8800 (DNAL1), Tektin-A, CG11253, and CG31320 (Figures 7C–7H; data not shown). In addition the nan, iav, and Dhc93AB GFP reporter gene constructs were all ectopically expressed upon fd3F misexpression (Figures 7I–7L; data not shown). Thus, fd3F is sufficient to activate aspects of Ch-specific gene expression in other sensory neurons. Interestingly, although scaGal4 drives expression widely in neuroectodermal cells, misexpression of target genes appears to be confined to ES neurons (Figures 7K and 7L). This is consistent with the idea that Fd3F function is limited to cells expressing its coregulator, Rfx.

### DISCUSSION

Fd3F is a cell-type-specific transcriptional regulator of ciliary sensory specialization. It is exclusively expressed in mechanosensory Ch neurons where it regulates aspects of Ch neuron ciliogenesis and ciliary function. Fd3F is not a Ch neuron identity factor or “master regulator” of Ch neuron differentiation: neural differentiation and ciliogenesis occur largely normally in fd3F mutant Ch neurons as attested by general morphology, and many Ch-specific genes are fd3F independent. Instead, fd3F regulates a program of gene expression for mechanosensory specialization that is unique to Ch neuron cilia and is absolutely required for their response to sensory stimulation. Specifically, fd3F targets are concerned with the structurally and functionally distinct ciliary zones of Ch neuron dendrites: fd3F regulates genes for both the machinery for construction and delineation of the zones (retrograde transport) and the proteins that populate the specialized proximal motor zone (axonemal dyneins, tektin, TRPV proteins). fd3F mutation has two consequences: ciliary motility of JO neurons is lost (reflected by loss of mechanical amplification), and sensory transduction is lost (reflected by loss of electrical response to stimulus). Effects on sensory transduction are both direct (loss of TRPV expression) and indirect (disrupted localization of candidate force-gated channel, TRPN1). Loss of axonemal dyneins might underlie the loss of motility directly, or their role may be indirect through their potential function as adaptation motors in sensory transduction (Nadrowski et al., 2008).

Mutation of fd3F does not lead to transformation of Ch cilium morphology to that expected for ES neurons—only motility and compartmentalization-related specializations are lost. Similarly, fd3F misexpression does not convert ES cilia to a Ch cilium morphology (unpublished data). We suggest that ES and Ch neuron cilia are both specialized derivatives of a default “nonspecialized” cilium structure, such that loss of fd3F results only in loss of specific Ch ciliary specializations. There may be other aspects of specialization regulated by other factors in both Ch neurons and other sensory neurons.

Consideration of fd3F target genes illuminates the outstanding question of how ciliogenesis pathways are modulated to produce specialized cilia. One might surmise that ciliary specialization requires subtype-restricted gene products. This is true to some extent, as exemplified by the TRPV proteins and axonemal dyneins. However, our findings suggest that quantitative differences in gene expression between sensory neuron subtypes are also important for ciliary specialization. Compartmentalization of Ch cilia into specialized motile and sensory zones requires retrograde transport proteins (Eberl et al., 2000; Lee et al., 2008). Our results suggest that a “basal” level of retrograde transport is sufficient for ciliogenesis in all sensory neurons (as is present in ES neurons and fd3F mutant Ch neurons), but compartmentalization of the Ch cilium requires a higher level of activity, as is provided by fd3F regulation of the genes involved. Thus, a quantitative difference in gene expression programs might underlie a qualitative feature of Ch ciliary specialization. This can be seen as a variation of the idea that differences in IFT activity might contribute to ciliary specialization (Silverman and Leroux, 2009).

One model for transcriptional regulation of ciliary specialization is that Rfx is required for pan-ciliary gene expression, whereas other more restricted transcription factors regulate subtype-specific gene expression (Silverman and Leroux, 2009). However, some targets of Rfx are expressed only in subsets of ciliated cells (Efimenko et al., 2005), raising the question of how a pan-ciliary factor can be responsible for subtype-specific gene expression (Silverman and Leroux, 2009; Wang et al., 2010). Rfx is required for all ciliated neurons in Drosophila, but its target gene specificity is modulated in Ch neurons by fd3F (Figure 7M). This suggests a general mechanism in which Rfx cooperates with cell-type-specific transcription factors to regulate the genes required for cilia diversification. Most fd3F/Rfx
targets have a conserved X box/Fox motif combination, demonstrating that specificity factors (in this case Fd3F) may act in very close molecular cooperation with Rfx, perhaps entailing cooperative binding. Conversely, most if not all fd3F target genes are also Rfx dependent, such that Rfx appears to be an obligate cofactor of Fd3F. Interestingly, the X boxes associated with...
targets often do not conform to the classic palindromic Rfx binding site (e.g., GTTGCCATGGCAAC; Avidor-Reiss et al., 2004) but, instead, show a strong match in only one half-site (RGYAAC). Half-sites and modified (nonpalindromic) X box sites have been noted for a variety of other cilia genes and might be especially associated with cell-type-restricted targets (Efimenko et al., 2005; Piasecki et al., 2010).

Interestingly, the same X box/Fox site combination is shared by both Ch-specific and Ch-enriched targets. Therefore, Fd3F acts as an obligatory cofactor of Rfx for Ch-specific genes, but for Ch-enriched genes it only enhances a basal level of Rfx-dependent regulation that is already existent in Ch and ES neurons (Figure 7M). To extend our model above, Rfx regulates low-level retrograde transport activity sufficient for
its “basal” ciliogenesis role, whereas fd3F/Rfx regulate the higher activity in Ch neurons that is required for ciliary compartmentalization.

In being required for Ch cilium motility, fd3F has a strikingly similar role to vertebrate Foxj1 genes, albeit that Foxj1 mutation has wider phenotypic consequences due to the many roles performed by motile cilia in vertebrates. Is fd3F related to Foxj1? The FoxJ subfamily is ancient, but bioinformatic analyses previously detected no Drosophila or C. elegans orthologs, suggesting that FoxJ genes have been lost from ecdysozoans (Mazet et al., 2003). Conversely, such analyses also failed to assign fd3F to any Fox subfamily (Lee and Frasch, 2004). However, a recent comprehensive analysis of mosquito Fox genes tentatively placed fd3F and its mosquito equivalent in the FoxJ subfamily (Hansen et al., 2007). Although the sequence evidence is equivocal, we suggest that fd3F is a highly diverged representative of the FoxJ subfamily.

Support for this relationship comes from consideration of target genes. Target gene analyses have indicated many candidate direct or indirect targets for FoxJ1 in mouse and Xenopus (Jacquet et al., 2009; Stubbs et al., 2008). Several fd3F target genes are homologs of FoxJ1-dependent genes in multiple vertebrate species (Thomas et al., 2010) (Table S1), including Dhc93AB (DNAH9 in human), CG9313 (WDR68), tektin-A (TEKT4), CG6971 (DNAH1), CG13930 (WDR78), Dhc62B (DNAH3), Dhc16F (DNAH6), CG10064 (WDR16), and CG34192 (DNA1R82). The shared targets are all directly concerned with motility, whereas retrograde transport genes are not FoxJ1 targets in vertebrates. This suggests that regulation of axonemal motor genes is an ancestral function of FoxJ1/fd3F, whereas regulation of retrograde transport was acquired later in the Drosophila lineage, coinciding with the emergence of distinct ciliary zones in the evolution of Ch neurons. Other fd3F target genes have no known function: we suggest they provide a potential source of new ciliary motility genes.

Cilia are classically classified as either being sensory (primary) with a 9+0 microtubule axoneme or propulsive (motile) with a 9+2 axoneme. Ch cilia are 9+0 sensory cilia with a limited set of motility-related features that are intimately linked to the Ch sensory transduction mechanism. These cilia therefore differ greatly from most propulsive cilia. Several FoxJ1-dependent cell types also bear 9+0 motile cilia somewhat reminiscent of those on Ch neurons. These include the mouse embryonic node, which is required for left-right asymmetry (Brody et al., 2000), related cells in Xenopus and zebrafish (e.g., Kupffer’s vesicle), and long 9+0 cilia of the neural tube floor plate in zebrafish (Stubbs et al., 2008; Yu et al., 2008).

In vertebrates, Rfx3 is required in many ciliated cells that also require Foxj1, including the mouse embryonic node, CNS ependymal cells (Baas et al., 2006), and chick neural tube floor plate (Cruz et al., 2010). Moreover, in the embryonic node Rfx3 regulates genes involved not only with ciliogenesis but also with cilium mobility (Bonnafe et al., 2004), and some Foxj1-dependent genes are also affected in Rfx3 mutants, including Dnahc9, the ortholog of Dhc93AB (El Zein et al., 2009). We suggest that Rfx3/Foxj1 may work in combination to regulate directly a subset of ciliary targets in the way we have identified for Rfx/fd3F.

EXPERIMENTAL PROCEDURES

Fly Stocks

Flies were maintained on standard media at 25°C. We used w1118 as wild-type control. The stocks elavGal4, UAS-mCD8-GFP, pEP[Fd3F]GFP, and Df(1)ED6716, ec1 were obtained from the Bloomington Stock Center (Indiana University, Bloomington, IN, USA). Other stocks used were Rfx69 (Dubrulle et al., 2002), rempA-YFP (Lee et al., 2008), and nompB-GFP (Han et al., 2003).

Immunohistochemistry

Primary antibodies used were mAb-22C10 (1:200), RbAb-HP (1:500), RbAb-GFP (1:500) (Molecular Probes), and mAb-NompC (Jiang et al., 2011). For generating the Fd3F antibody, synthetic peptides matching the C-terminal region of Fd3F were injected into two rabbits to produce anti-Fd3F antiserum (CovalAb/Eurogentec). The serum was purified using Melon Gel IgG Spin Purification Kit (Thermo Scientific) and was used at 1:100. Secondary antibodies were from Molecular Probes. mRNA in situ hybridizations to whole embryos by standard diogenin method. Gene fragments were amplified from genomic DNA by PCR (oligonucleotides shown in Supplemental Experimental Procedures) and digoxigenin-labeled RNA probes prepared using T7 RNA polymerase. For bright-field microscopy, embryos were observed using an Olympus AX70 microscope and captured with a DP50 camera. For immunofluorescence microscopy, embryos were observed using a Zeiss Pascal confocal microscope. Images were processed for contrast and size in Adobe Photoshop.

Climbing Assay

A total of 20 mated female flies were placed in a measuring cylinder. After a 1 min recovery period, the cylinder was banged firmly once on the bench, and the percentage of flies passing a 10 cm threshold within 1 min of banging was recorded. This was repeated 5 times each for four groups of flies from each line.

Electron Microscopy

Whole-adult heads were removed and rinsed in 0.5% Triton X-100. The proboscis was removed to facilitate infiltration of the fix, and the heads were then fixed in 2.5% glutaraldehyde and 2% paraformaldehyde in 0.1 M phosphate buffer (pH 7.4) overnight at 4°C. Heads were then washed in 0.1 M phosphate buffer (pH 7.4), postfixed with OsO4, dehydrated in an ethanol series, and embedded in Polybed812. Ultrathin (75 nm) sections of the antennae were then stained with aqueous uranyl-acetate and lead citrate and examined with a Hitachi 7000 electron microscope (Electron Microscopy Research Services, Newcastle University Medical School).

Promoter Fusions

Frames were amplified from genomic DNA and cloned into pHStinger GFP reporter vector. Site-directed mutagenesis of these constructs was carried out using the QuikChange II Kit (Stratagene). Fox motifs were changed from ACGG to CGGG in the reporter vector. Site-directed mutagenesis of these constructs was carried out using T7 RNA polymerase. For bright-field microscopy, embryos were observed using a Zeiss Pascal confocal microscope. Images were processed for contrast and size in Adobe Photoshop.

Expression and Purification of Fd3F Forkhead Domain

Total RNA was extracted from 5–21 hr embryos using RNAeasy kit (QiAGEN). This RNA was reverse transcribed to produce total embryonic cDNA using ImProm-II Reverse Transcription System (Promega). The forkhead box of the gene (Fd3F4m) was amplified from the cDNA and cloned in pGEX-2T (Supplemental Experimental Procedures). The pGEX-Fd3F4m plasmid was used to transform BL21-pLysS E. coli. The cells were grown to OD600 of 0.35, and expression of Fd3F4m was then induced with 0.5 M IPTG for 2 hr at 20°C. GST-Fd3F4m was purified from the soluble cell lysate using the GST binding resin. The GST-Fd3F4m fusion protein was cleaved using the thrombin protease. GST-Fd3F4m was purified from the soluble cell lysate using the GST binding resin.
glutathione Sepharose beads and eluted with 50 mM reduced glutathione and 0.4% deoxycholate in 250 mM Tris- HCl (pH 8).

**Gene Mobility Shift Assay**

Oligonucleotides (Supplemental Experimental Procedures) were labeled with \( \gamma^{32P}ATP \) (Amersham) using T4 polynucleotide kinase (New England Biolabs). Unincorporated \( \gamma^{32P}ATP \) was removed using ProbeQuant G-50 microcolumns (GE Healthcare). \( \gamma^{32P} \)-labeled duplexes were used at 0.1 nM in 20 μl reaction mixtures in binding buffer (10 mM Tris- HCl, 1 mM dithiotreitol, 1 mM EDTA, and 100 mM NaCl). The duplexes were incubated with purified Fd3F \( ^{\text{th}} \) (12.5 μM in binding buffer) for 20 min on ice. The reaction mixture was then electrophoresed on a native 6% polyacrylamide gel in 0.5 x TBE at 40 mA for 1 h before exposure to a phosphorimager screen. For competition assays 10 nM unlabeled DNA duplex was mixed with 0.1 nM labeled DNA prior to adding Fd3F \( ^{\text{th}} \).

**Misexpression of Fd3F**

The fd3F ORF was amplified by RT-PCR on embryonic mRNA using a primer from the 5’ noncoding exon and the 3’ end of the annotated fd3F ORF (Supplemental Experimental Procedures). This yielded a fragment of 2.5 kb, which is 1 kb larger than expected from current genome annotation (http://www.flybase.org). Sequencing revealed a large ORF consisting of the coding regions of both fd3F and CG32779, which is currently annotated as a separate transcription unit within the first intron of fd3F. Inspection of fd3F orthologs from other Drosophila species and other insects suggests that CG32779 is indeed part of fd3F (unpublished data). This ORF was therefore inserted into pUAS-attB, and transgenic flies were obtained by microinjection of syncytial blastoderm embryos.

**Electrophysiological and Biophysical Analyses**

For evaluating Johnston’s organ function, flies were exposed to pure tones of different intensities at the individual best frequency of their antennal sound receiver (Goepfert et al., 2005), and the resulting mechanical displacements and antennal nerve potentials were simultaneously recorded. Receiver displacements were measured with an OFV-400 Polytec laser Doppler vibrometer at the tip of the antennal arista. Nerve responses were recorded by inserting an electrolytically tapered tungsten wire between the first antennal segment and the head capsule, and an indifferent electrode placed in the thorax (Effertz et al., 2011). Sound particle velocities were measured with an Emkay NR 3158 pressure gradient microphone at the position of the fly.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes one figure, two tables, and Supplemental Experimental Procedures and can be found with this article online at doi:10.1016/j.devcel.2012.05.010.

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**REFERENCES**


