A two-stage meta-analysis identifies several new loci for Parkinson’s disease

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A Two-Stage Meta-Analysis Identifies Several New Loci for Parkinson’s Disease

International Parkinson’s Disease Genomics Consortium (IPDGC), Wellcome Trust Case Control Consortium 2 (WTCCC2)

Abstract

A previous genome-wide association (GWA) meta-analysis of 12,386 PD cases and 21,026 controls conducted by the International Parkinson’s Disease Genomics Consortium (IPDGC) discovered or confirmed 11 Parkinson’s disease (PD) loci. This first analysis of the two-stage IPDGC study focused on the set of loci that passed genome-wide significance in the first stage GWA scan. However, the second stage genotyping array, the ImmunoChip, included a larger set of 1,920 SNPs selected on the basis of the GWA analysis. Here, we analyzed this set of 1,920 SNPs, and we identified five additional PD risk loci (combined \( p < 10^{-15} \), PARK16/1q32, STX18/16p11, FGF20/8p22, STBD1/4q21, and GPNMB/7p15). Two of these five loci have been suggested by previous association studies (PARK16/1q32, FGF20/8p22), and this study provides further support for these findings. Using a dataset of post-mortem brain samples assayed for gene expression (\( n = 292 \)) and methylation (\( n = 399 \)), we identified methylation and expression changes associated with PD risk variants in PARK16/1q32, GPNMB/7p15, and STX18/16p11 loci, hence suggesting potential molecular mechanisms and candidate genes at these risk loci.

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Introduction

Until the recent developments of high throughput genotyping and genome-wide association (GWA) studies, little was known of the genetics of typical Parkinson’s disease (PD). Studies of the genetic basis of familial forms of PD first identified rare highly penetrant mutations in \( \text{LRRK2} [1,2], \text{PINK1} [3], \text{SNCA} [4], \text{PARK2} [5] \) and \( \text{PARK7} [6] \). Following these findings, GWA scans for idiopathic PD identified \( \text{SNCA} \) and \( \text{MAPT} \) as unequivocal risk loci \([7,8,9,10,11]\) as well as implicated \( \text{BST1} [8], \text{GAK} [12], \) and \( \text{HLA-DR} [13] \). Using sequence based imputation methods [14], the meta-analysis of several GWAS scans \([7,9,10,11]\) conducted by the International Parkinson’s Disease Genomics Consortium (IPDGC) identified and replicated five new loci: \( \text{ACMSD}, \text{STX39}, \text{MCCCI1}/\text{LAMPS}, \text{SYT11}, \text{and CDC62}/\text{HIP1R} [15] \) and confirmed association at \( \text{SNCA}, \text{LRRK2}, \text{MAPT}, \text{BST1}, \text{GAK} \) and \( \text{HLA-DR} [15] \).

We conducted a two-stage association study. Combining stage 1 and stage 2, the data consists of 12,386 PD cases and 21,026 controls genotyped using a variety of platforms (Table 1). Stage 1 used genome-wide genotyping arrays and our initial analysis [15] focused on the subset of SNPs that passed genome-wide significance in stage 1. For stage 2 genotyping, we used a custom content Illumina iSelect array, the Immunochip and additional...
GWAS typing as previously described [15]. The primary content of the ImmunoChip data focuses on autoimmune disorders but, as part of a collaborative agreement with the Wellcome Trust Case Control Consortium 2, we included 1,920 ImmunoChip SNPs on the basis of the stage 1 GWA PD results.

Here, we report the combined analysis for this full set of 1,920 SNPs. This step1+2 analysis identified seven new loci that passed genome-wide significance in the meta-analysis. During the process of analyzing these data and preparing for publication, we became aware that another group was also preparing a large independent GWA scan in PD for publication (Do et al, submitted). Following discussion with this group we agreed to cross validate the top hits from each study by exchanging summary statistics for this small number of loci.

To provide further insights into the molecular function of these associated variants, we tested risk alleles at these loci for correlation with the expression of physically close gene (expression quantitative trait locus, eQTL) and the methylation status (methQTL) of proximal DNA CpG sites in a dataset of 399 control frontal cortex and cerebellar tissue samples extracted post-mortem from individuals without a history of neurological disorders.

Results

In addition to eleven loci that passed genome-wide significance in stage 1 [15], we identified over 100 regions of interest defined as 10 kb windows containing at least one SNP associated at \( p<10^{-3} \). We submitted the most associated SNP in each region for probe design and follow-up genotyping using the ImmunoChip platform. For each region of interest, we also added four SNPs in high level of linkage disequilibrium (LD) to provide redundancy where the most associated SNP would not pass the Illumina probe design step or the assay for that SNP would fail. To complete the array design we also added all non-synonymous dbSNPs located in known PD associated regions [1,2,3,4,5,6]. Out of these 2,400 submitted SNPs, 1,920 passed QC and were included in the final array design. For these 1,920 SNPs we combined stage 1 and stage 2 associated data in a meta-analysis of 12,386 cases and 21,026 controls (Table 1) from the IPDGC. We exchanged summary statistics for these most significant hits with an additional large, case-control replication dataset (3,426 PD cases and 29,624 controls) in an attempt to demonstrate independent replication.

On the basis of stage 1+2 results, seven new SNPs passed our defined genome-wide significance threshold \( p<5\times10^{-5} \), Table 2 and Figure 1. These loci are either novel or the previous evidence of association was not entirely convincing in individuals of European descent. We combined these results with the independent replication. Five of these seven loci replicated and showed combined evidence of PD association \( p<10^{-10} \) overall. Taking either the nearest gene (or the strongest candidate when available) to designate these regions, these five loci are 1q32/PARK16[7], 4q21/STBD1, 7p15/GPNMB, 9p22/FGF29[16] and 16p11/STXB.

rs708723/1q32 has been previously reported as PD associated (PARK16[7,8]) but this SNP lacked the unequivocal evidence of association in European samples \( p=9.47\times10^{-10} \) in stage 2 only. To understand the potential biological consequences of risk variation at this locus we tested whether rs708723 was correlated with either gene expression or DNA methylation status of proximal transcripts or CpG sites respectively (Table 3). We found correlations with the expression of NUCKS1 \( p=1.8\times10^{-7} \) and RAB7L1 \( p=7.2\times10^{-5} \). We also found correlations with the methylation state of CpG sites located in the FLJ3269 gene \( p=3.9\times10^{-22} \).

In the case of 16p11/STXB, the proximal gene to the most associated SNP rs4889603 is SETD1L. However, STXB1 is located 18 kb upstream of rs4889603 and is a more plausible PD candidate gene [17] owing to its synaptic receptor function. We therefore used this gene to designate this region. Our methQTL/eQTL dataset identified a correlation between the rs4889603 risk allele and increased methylation of a CpG dinucleotide in STXB1 (Table 3).

The SNP rs591323 in the 8p22 region is located ~150 kb downstream of the FGF29 gene (NCBI build 36.3), for which association with PD has been suggested previously in familial PD samples [16,18] but which remained controversial [19]. Our findings provide further support for a PD association at this locus, but again, whether the functionally affected transcript is FGF29 or not remains unclear.

The regions 4q21/STBD1 and 7p15/GPNMD have not been previously implicated in PD etiology. We found that the risk allele of rs164429, the most associated SNP in the 7p15 region, is associated in our eQTL dataset with decreased expression of the proximal transcript encoded by NUPL2 (Table 3). The same risk allele is also associated with increased methylation of multiple CpG sites proximal to GPNMB itself (Table 3). Neither of these regions contains an obvious candidate gene.

Two additional loci (3q26/NAID3 and 8q11/AMP106) showed strong evidence of association in stage 1 and 2 but were not disease associated in the Do et al dataset. Further replication is required to clarify the role of variation at these loci in risk for PD.

The strongly associated G2019S variant in the LRRK2 gene [20] was included in the Immunochip design and we replicated the published association: control frequency: 0.045% case frequency 0.61%, estimated odds ratio: 13.5 with 95% confidence interval: 5.5–43. However, the case collections have been partially screened for this variant therefore its frequency in cases and the odds ratio is likely to be underestimated.

The ImmunoChip array design provides some power to detect whether multiple distinct association signals exist at individual loci. Indeed, if a SNP showed an independent and sufficiently strong association in stage 1, it would have been included in stage 2 provided that it was not located in the same 10 kb window as the primary SNP in the region. There is precedent for this in PD, with the previous identification of independent risk signals at the SNCA locus [11]. We therefore used the Immunochip data to test whether any of the seven loci in Table 2 showed some evidence of more than one independent signal. None of these seven loci showed any association \( p>0.01 \) after conditioning on the main SNP in the region. In contrast, after conditioning on the most associated SNPs rs356182 in the SNCA region, several SNPs...
remained convincingly associated \( p = 9.7 \times 10^{-8} \) for rs2245801 being the most significant).

Lastly, we performed a risk profile analysis to investigate the power to discriminate cases and controls on the basis of the 16 confirmed common associated variants (Table 4). For each locus, we estimated the odds ratio on the basis of stage 1 data and we applied these estimates to compute for each individual in the ImmunoChip cohort a combined risk score. Solely based on these 16 common variants, and therefore not considering rare highly penetrant variants such as G2019S in \( \text{LRK2} \) [20], we found that individuals in the top quintile of the risk score have an estimated three-fold increase in PD risk compared to individuals in the bottom quintile (Table 4). We note however that the effect size of several of these associated variants could be over-estimated (an effect known as winner’s curse, see [21]) but given the consistent estimates of odds ratio across studies (Table 4) we expect this bias to be minimal.

**Discussion**

The combination of GWA scans and imputation methods in large cohorts of PD cases and controls has enabled us to identify five PD associated loci in addition to the 11 previously reported by us. Two of these loci (1q32/\( \text{PARK16} \), 8p22/\( \text{FGF20} \)) implicate regions that had been previously associated with PD risk [8,16]. The 1q32/\( \text{PARK16} \) showed convincing evidence of association in the Japanese population [8] but until now the association P-value had not passed a stringent genome-wide significance threshold in samples of European descent [7]. The 8p22/\( \text{FGF20} \) locus had been previously reported in a study of familial PD [16] and we provide the first evidence of association in a case-control study. The remaining three loci (\( \text{STX1B} \)/16p11, \( \text{STBD1} \)/4q21 and \( \text{GPNMB} \)/7p15) are new.

Adding the eleven previously reported common variants [15] to the five convincingly associated loci identified in this study, common variants at 16 loci have now been associated with PD. Controlling for the risk score based on the 11 SNPs previously identified [15] in the risk profile analysis (Table 4), the addition of these five new loci provides a modest but significant \( p = 2.2 \times 10^{-5} \) improvement of our ability to discriminate PD cases from controls.

Combining eQTL/methylation and case-control data implicates potential mechanisms which could explain the increased PD risk associated some of these variants. In particular, the strong eQTL in the 1q32/\( \text{PARK16} \) region with the \( \text{RAB7L1} \) and \( \text{NUCKS1} \) genes (Table 3) suggests that either one of these genes could be the biological effector of this risk locus. However, existing data show that eQTLs are not always present within the associated region, their known biological function and case-control data are indeed fully consistent.

While we are unable to unequivocally pinpoint the causative genes underlying these associations, their known biological function can suggest likely candidates. At the 1q32/\( \text{PARK16} \) locus our association and eQTL data indicate that \( \text{RAB7L1} \) and \( \text{NUCKS1} \) are the best candidates. The former is a GTP-binding protein that plays an important role in the regulation of exocytotic and endocytotic pathways [23]. Exocytosis is relevant for PD for two main reasons: firstly, since dopaminergic neurotransmission is mediated by the vesicular release of dopamine, i.e. dopamine exocytosis [24], and secondly because it has been shown that alpha-synuclein knock-out mice develop vesicle abnormalities [25], thus providing a potential direct link between genetic variability in the gene and a biological pathway involved in the disease. Less is known regarding \( \text{NUCKS1} \); it has been described to be a nuclear protein, containing casein kinase II and cyclin-dependant kinases phosphor-

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### Table 1. Sample size and genotyping platform for the cohorts included in stage 1 (top set of rows), stage 2 (middle set of rows), and independent replication (bottom row).

<table>
<thead>
<tr>
<th>Cohort</th>
<th>Controls</th>
<th>Cases</th>
<th>Genotyping platform</th>
</tr>
</thead>
<tbody>
<tr>
<td>United Kingdom</td>
<td>5,200</td>
<td>1,705</td>
<td>Illumina 660W-Quad</td>
</tr>
<tr>
<td>USA-NIA</td>
<td>3,034</td>
<td>971</td>
<td>Illumina HumanHap 550</td>
</tr>
<tr>
<td>USA-dbGAP</td>
<td>857</td>
<td>876</td>
<td>Illumina 370 K</td>
</tr>
<tr>
<td>French</td>
<td>1,984</td>
<td>1039</td>
<td>Illumina 610-Quad</td>
</tr>
<tr>
<td><strong>Total Stage 1</strong></td>
<td><strong>12,019</strong></td>
<td><strong>5,333</strong></td>
<td></td>
</tr>
<tr>
<td>Icelandic</td>
<td>1,427</td>
<td>479</td>
<td>Illumina HumanHap 300</td>
</tr>
<tr>
<td>Dutch</td>
<td>2,024</td>
<td>772</td>
<td>Illumina 610-Quad</td>
</tr>
<tr>
<td>USA</td>
<td>2,215</td>
<td>2,807</td>
<td>ImmunoChip</td>
</tr>
<tr>
<td>United Kingdom</td>
<td>1,864</td>
<td>1,271</td>
<td>ImmunoChip</td>
</tr>
<tr>
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<td>402</td>
<td>304</td>
<td>ImmunoChip</td>
</tr>
<tr>
<td>French</td>
<td>363</td>
<td>267</td>
<td>ImmunoChip</td>
</tr>
<tr>
<td>German</td>
<td>712</td>
<td>1,153</td>
<td>ImmunoChip</td>
</tr>
<tr>
<td><strong>Total Stage 2</strong></td>
<td><strong>9,007</strong></td>
<td><strong>7,053</strong></td>
<td></td>
</tr>
<tr>
<td><strong>Stage 1+Stage 2</strong></td>
<td><strong>21,026</strong></td>
<td><strong>12,386</strong></td>
<td></td>
</tr>
<tr>
<td>Do et al- USA</td>
<td>29,624</td>
<td>3,426</td>
<td></td>
</tr>
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</table>

DOI:10.1371/journal.pgen.1002142.t001
Two-Stage Meta-Analysis of Parkinson’s Disease

rs708723

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

rs34016896

dbGAP–GWAS
FR–GWAS
GER–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

rs6812193

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

rs156429

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

rs591323

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

chr8:89442157

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio

rs4889603

dbGAP–GWAS
FR–GWAS
GER–GWAS
ICE–GWAS
NL–GWAS
UK–GWAS
US–GWAS
US–Immuno

Summary

Odds Ratio
PD that have a complex genetic component. We therefore expect that further and larger association analyses, perhaps using dedicated high-throughput genotyping arrays like the ImmunoChip, will continue to yield new insights into PD etiology.

**Material and Methods**

Genotyping and case control cohorts

Participating studies were either genotyped using the ImmunoChip as part of a collaborative agreement with the ImmunoChip Consortium, or as part of previous GWA studies provided by members of the IPDGC or freely available from dbGaP [7,9,10,11]. Genotyping of the UK cases using the Immunochip was undertaken by the WTCCC2 at the Wellcome Trust Sanger Institute which also genotyped the UK control samples. The constituent studies comprising the IPDGC have been described in detail elsewhere [15], although a summary of individual study quality control is available as part of Table S1. In brief all studies followed relatively uniform quality control procedures such as: minimum call rate per sample of 95%, mandatory concordance between self-reported and X-chromosome-heterogeneity estimated sex, exclusion of SNPs with SNP Chrom Gene(s) Alleles MAF OR (95%CI) OR (95%CI) OR (95%CI) OR (95%CI) P 1.06–1.18 0.839–0.937 0.891–0.925 1.08–1.18

**Association test statistics**

Single SNP test statistics were combined across datasets using a score test methodology, essentially assuming equal odds ratio across cohorts. In addition, fixed and random effects meta-analyses were implemented in R (version 2.11) to confirm that the score test approximation does not affect the interpretation of the results. We also tested the relevant SNPs heterogeneity across cohorts and no significant heterogeneity was detected (Table S2).

**Data exchange**

We communicated to our colleagues in charge of the independent study (Do et al) the seven SNPs listed in Table 2. For this subset of SNPs they selected the marker with the highest $r^2$ value on their genotyping platform and provided us with the following summary statistics: odds ratio, direction of effect, standard error for the estimated odds ratio and one degree-of-freedom trend test P-value.

**eQTL analysis and methylation analysis**

Quantitative trait analyses were conducted to infer effects of risk SNPs on proximal CpG methylation and gene expression. For the five replicated SNP associations (Table 2), all available CpG probes and expression probes within +/-1 MB of the target SNP were

<table>
<thead>
<tr>
<th>SNP</th>
<th>Chrom</th>
<th>Gene(s)</th>
<th>Alleles</th>
<th>MAF</th>
<th>OR (95%CI)</th>
<th>OR (95%CI)</th>
<th>OR (95%CI)</th>
<th>OR (95%CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs708723</td>
<td>1q32</td>
<td>RAB7L1/PARK16</td>
<td>T&gt;C</td>
<td>0.439</td>
<td>0.905 (0.862–0.95)</td>
<td>6.68×10⁻⁵</td>
<td>0.863 (0.824–0.905)</td>
<td>9.47×10⁻¹⁰</td>
</tr>
<tr>
<td>rs34016896</td>
<td>3q26</td>
<td>NMD3</td>
<td>C&gt;T</td>
<td>0.305</td>
<td>1.14 (1.09–1.2)</td>
<td>3.00×10⁻⁷</td>
<td>1.08 (1.02–1.14)</td>
<td>0.00399</td>
</tr>
<tr>
<td>rs6812193</td>
<td>4q21</td>
<td>STBD1</td>
<td>C&gt;T</td>
<td>0.36</td>
<td>0.886 (0.843–0.932)</td>
<td>2.52×10⁻⁶</td>
<td>0.906 (0.864–0.95)</td>
<td>5.29×10⁻⁵</td>
</tr>
<tr>
<td>rs156429</td>
<td>7p15</td>
<td>GPNNB</td>
<td>A&gt;G</td>
<td>0.403</td>
<td>0.894 (0.849–0.942)</td>
<td>2.15×10⁻⁵</td>
<td>0.893 (0.852–0.917)</td>
<td>3.86×10⁻⁶</td>
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<td>rs513123</td>
<td>8p22</td>
<td>FG20</td>
<td>G&gt;A</td>
<td>0.271</td>
<td>0.884 (0.836–0.935)</td>
<td>1.59×10⁻⁵</td>
<td>0.875 (0.83–0.923)</td>
<td>8.49×10⁻⁷</td>
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<td>chr8:89442157</td>
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<td>1.29 (1.12–1.49)</td>
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<tr>
<td>rs4889003</td>
<td>16p11</td>
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<td>1.12 (1.06–1.18)</td>
<td>4.13×10⁻⁵</td>
<td>1.15 (1.11–1.21)</td>
<td>8.21×10⁻⁶</td>
</tr>
</tbody>
</table>

1q32/PARK16 has been reported previously but is included because these data provide for the first time unequivocal evidence of association. P-values are computed using a one-degree-of-freedom trend test regression models were utilized to quantify associations with PD incorporating allele dosages as the primary predictor of disease. Imputed data was analyzed using MACH2DAT, and genotyped SNPs were analyzed using PLINK. All models were adjusted for covariates of components 1 and 2 from either principal components or multidimensional scaling analyses to account for population substructure and stochastic genotypic variation (except in the UK-GWAS data which were not adjusted for population substructure).
investigated as candidate QTL associations in frontal cortex and cerebellar tissue samples. 399 samples were assayed for genome-wide gene expression on Illumina HumanHT-12 v3 Expression Beadchips and 292 samples were assayed using Infinium HumanMethylation27 Beadchips, both per manufacturer’s protocols in each brain region. A more in depth description of the sample series comprising the QTL analyses, relevant laboratory procedures and quality requirements may be found in [15]. The QTL analysis utilized multivariate linear regression models to estimate effects of allele dosages per SNP on expression and methylation levels adjusted for covariates of age at death, gender, the first 2 component vectors from multi-dimensional scaling, post mortem interval (PMI), brain bank from where the death, gender, the first 2 component vectors from multi-dimensional scaling, post mortem interval (PMI), brain bank from where the

doi:10.1371/journal.pgen.1002142.t003

<table>
<thead>
<tr>
<th>Study</th>
<th>Assay Region</th>
<th>SNP</th>
<th>Probe Region</th>
<th>Gene Tagged by Probe</th>
<th>Illumina Probe</th>
<th>Alleles</th>
<th>Effect Estimate</th>
<th>Standard Error</th>
<th>Unadjusted P</th>
<th>Adjusted P</th>
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<td>Expression</td>
<td>rs156429</td>
<td>7p15/GPNMB</td>
<td>NUP1L2</td>
<td>ILMN_1789616</td>
<td>A&gt;G</td>
<td>0.083</td>
<td>0.018</td>
<td>3.6E-06</td>
<td>1.0E-04</td>
</tr>
<tr>
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<td>Expression</td>
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<td>A&gt;G</td>
<td>0.078</td>
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<td>1.0E-04</td>
</tr>
<tr>
<td>Germany</td>
<td>Expression</td>
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<td>NUCKS1</td>
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<td>1.5E-05</td>
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<tr>
<td>France</td>
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<td>rs156429</td>
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<td>NUP1L2</td>
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<td>1.2E-02</td>
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<td>3.4E-03</td>
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Table 4. Estimated PD risk profile for the five cohorts genotyped using the Immunochip.

<table>
<thead>
<tr>
<th>Study</th>
<th>1st quintile</th>
<th>2nd quintile</th>
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<tr>
<td>USA</td>
<td>0.614</td>
<td>1.29−1.84</td>
<td>1.92</td>
<td>1.61−2.29</td>
<td>2.21</td>
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<tr>
<td>UK</td>
<td>0.636</td>
<td>1.05−1.71</td>
<td>1.79</td>
<td>1.41−2.28</td>
<td>2.35</td>
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<tr>
<td>Germany</td>
<td>0.692</td>
<td>0.98−1.79</td>
<td>1.88</td>
<td>1.38−2.58</td>
<td>1.88</td>
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<tr>
<td>France</td>
<td>0.675</td>
<td>0.99−2.92</td>
<td>1.13</td>
<td>0.65−1.98</td>
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<tr>
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<td>0.65−1.74</td>
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<td>0.83−2.20</td>
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<td>1.26−1.61</td>
<td>1.79</td>
<td>1.58−2.02</td>
<td>2.22</td>
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</tbody>
</table>

% Cases per Quintile: 37.90, 46.06, 51.15, 56.56, 63.75

Table 3. Significant eQTL associations (p<0.01) between the five SNPs with positive replication data (Table 2) and proximal (cis) changes in gene expression/methylation in frontal cortex and cerebellar tissue.

<table>
<thead>
<tr>
<th>Assay Region</th>
<th>SNP</th>
<th>Region</th>
<th>Gene Tagged by Probe</th>
<th>Alleles</th>
<th>Effect Estimate</th>
<th>Standard Error</th>
<th>Unadjusted P</th>
<th>Adjusted P</th>
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<td>Expression</td>
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<td>A&gt;G</td>
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<td>0.018</td>
<td>3.6E-06</td>
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<tr>
<td>Expression</td>
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<td>NUP1L2</td>
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<td>A&gt;G</td>
<td>0.078</td>
<td>0.017</td>
<td>3.1E-06</td>
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<tr>
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<td>NUCKS1</td>
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<td>T&gt;C</td>
<td>0.155</td>
<td>0.03</td>
<td>1.8E-07</td>
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<tr>
<td>Methyltion</td>
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<td>NUCKS1</td>
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<td>T&gt;C</td>
<td>-0.062</td>
<td>0.018</td>
<td>7.2E-04</td>
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<td>NUCKS1</td>
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<td>A&gt;G</td>
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<td>3.4E-03</td>
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<td>0.066</td>
<td>0.022</td>
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</tbody>
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Two-Stage Meta-Analysis of Parkinson’s Disease

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samples were provided and in which preparation/hybridization batch the samples were processed. A total of 670 candidate QTL associations were tested: 57 expression QTLs in the cerebellum samples, 83 expression QTLs in the frontal cortex samples, 249 methylation QTLs in the cerebellum samples and 249 methylation QTLs in the frontal cortex samples. Multiple test correction was undertaken using false discovery rate adjusted p-values<0.05 to dictate significance, with the p-value adjustment undertaken in each series separately, stratified by brain region and assay. A complete list of all QTL associations tested is included in Table S3.

Supporting Information

Table S1 Summary of results for fixed and random effects meta-analysis, as estimates of effect heterogeneity across cohorts and SNP used at the Do et al replication stage. (XLSX)

Table S2 Summary of the quality control parameters applied to the GWA datasets included in this study. (XLSX)

Table S3 Complete list of tested QTL associations (expression and methylation). (XLSX)

Text S1 Membership of the Wellcome Trust Case Control Consortium 2. (DOC)

Acknowledgments

This study utilized the high-performance computational capabilities of the Biowulf Linux cluster at the National Institutes of Health, Bethesda, Maryland (http://biowulf.nih.gov). DNA panels and samples from the NINDS Human Genetics Resource Center DNA and Cell Line Repository (http://ccr.coriell.org/sections/Collections/NINDS/?SsId=10). DNA panels and samples from the Wellcome Trust Case-Control Consortium 2 (WTCCC2) on UK PD cases and on UK controls from the 1958 Birth Cohort (58C) and National Blood Service (NBS). UK population control data was made available through WTCCC1. We thank Jeffrey Barrett for assistance with the design of the Immunochip.

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**References**


