Is Bacille Calmette-Guérin (BCG) Vaccine a Known Risk Factor for Latent Tuberculosis Infection?: A Cross-sectional Study on 180 New Immigrants from BCG-vaccinated Countries to Kuwait

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Abstract

The Bacille Calmette-Guérin (BCG) vaccine has existed for 90 years and is the most widely used of all current national childhood immunization programmes. The impact of BCG vaccination on transmission of Mycobacterium tuberculosis is not clear or whether BCG confers lifelong immunity through sufficient protection against formation and diagnosis of latent tuberculosis infection (LTBI) is recently questionable especially in tuberculosis endemic regions and vaccinated high risk individuals, which was discussed along the paper work following a new evidence-based criteria for LTBI diagnosis.

Keywords: Bacille Calmette Guerin (BCG); Tuberculosis (TB); Latent tuberculosis infection (LTBI); Tuberculin skin test (TST)

Introduction

Since the 20th century the Bacille Calmette-Guérin (BCG) vaccine there is only one defensive tool against Mycobacterium tuberculosis (MTB) introduced as a shot of a 90-year-old vaccine (attenuated virulence of Mycobacterium bovis). BCG limits the growth and protect against MTB and aids in clearance of bacillary loads during chemotherapeutic treatment in the active post-infective stage [1]. BCG vaccination has obvious public health advantages. It can lessen the burden of TB-related disorders such as LTBI prevalence and active TB incidence in children [2]. TB incidence rose again after abandoning routine BCG in the mid-1980s even with the introduction of other preventive measures [3]. But the efficacy impacts of BCG vaccination whether confers lifelong immunity or adversely causing tuberculosis (TB) through indirectly forming latent tuberculosis infection (LTBI) is still unclear. Worldwide the tuberculin skin test (TST) has been the standard diagnostic test for detection of LTBI for more than 100 years. It was first described by Robert Koch in 1890, and involves injecting intradermally a purified protein derivative (PPD) of 0.1 ml tuberculin (which is a glycerol extract of the bacillus) followed by measurement of the localized delayed-type hypersensitivity (DTH-IV) reaction [4]. The TST has several limitations and cannot distinguish between previous BCG vaccination and current TB infection due to poor specificity in BCG negative people and poor sensitivity in BCG-vaccinated individuals. Meta-analysis has shown that previous BCG administration increases the likelihood of TST false-positive results up to 15 years after-vaccination [5].

BCG vaccination might alter the interpretation of a positive PPD, and therefore TST interpretation is extremely limited as a diagnostic tool to prove whether a positive result is caused by MTB infection or cross-reactivity resulting from BCG vaccination (containing live attenuated M. bovis antigens), and has to be interpreted taking into consideration both the pre-test risk of TB infection and BCG vaccination status.

Objective

To assess the interference effect of BCG vaccination on the results of the tuberculin skin test.

Sub-objective

Assess with evidence-based the cross-reactive effects of BCG vaccine on Mantoux test (used for LTBI diagnosis) according to a new defined categories of latent TB infection using four TB diagnostic testing implemented on healthy new immigrants.

Materials and Methods

The study design was a cross-sectional study involving a sample of 180 new immigrants to Kuwait during four months (between February and May) 2010 during their compulsory registration at Al Farwaniya Immigration Centre (the most populous region in Kuwait covering 907,321 out of the 3,442,945 total residential population). Included participants were randomly chosen as one in every thirty four expatriates without selection bias.

The study screening tools were a structured questionnaire to assess the socio-economic characteristics and past history of BCG vaccination with careful examination for the presence of characteristic BCG scar on both arms. Four tests were performed on each immigrant simultaneously. Two blood samples for both IGRAs (three 1 ml tubes for Quantiferon Gold In-Tube and one 8ml tube for T-SPOT.TB test) were taken. Thirdly, this was followed by performing TST through intradermal administration of five tuberculin units (5TU) of 0.1 ml of purified protein derivative (PPD) (Tubersol: Sanofi Pasteur Ltd, Toronto, Ontario, Canada), using 1 ml disposable tuberculin syringes.

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Table 1: New classification representing categorization criteria for diagnosis of latent tuberculosis infection cases using a combination of four-tuberculosis diagnostic tests and score for latent tuberculosis infection case diagnosis.

<table>
<thead>
<tr>
<th>Mantoux Score grade</th>
<th>LTBI diagnosis</th>
<th>Final result n (%)</th>
</tr>
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<tbody>
<tr>
<td>0</td>
<td>Negative (no reaction)</td>
<td>Normal 170 (96.04%)</td>
</tr>
<tr>
<td>1 - &lt; 5</td>
<td>Normal (reactive)</td>
<td>Normal 6 (3.38%)</td>
</tr>
<tr>
<td>5 - &lt; 10</td>
<td>Borderline</td>
<td>Normal 1 (0.05%)</td>
</tr>
<tr>
<td>10 - &lt; 15</td>
<td>Positive</td>
<td>Abnormal 0 (0.00%)</td>
</tr>
<tr>
<td>15 - &lt; 20</td>
<td>Highly positive</td>
<td>Abnormal 0 (0.00%)</td>
</tr>
<tr>
<td>≥ 20</td>
<td>Definite</td>
<td>Abnormal 0 (0.00%)</td>
</tr>
<tr>
<td>Excluded</td>
<td>Cannot be judged</td>
<td>-</td>
</tr>
</tbody>
</table>

Modified 'cut-off' point value of TST induration was more than 10 mm, score (>)no score for excluded case (due to missed follow-up or result reading after 5days), mm=millimetre induration

Table 2: Skin test response and score frequency of latent tuberculosis infection of the 180 new immigrants to Kuwait, February-May, 2010.

- Mantoux (induration) reaction was measured in 98.33% (177/180) who fulfilled the inclusion criteria for TST results. The TST results of three excluded immigrants were as follows: two participants came on day 7 (a male Indian electrician and a female Nepali housemaid) and one participant did not return for a TST reading (a female Ethiopian housemaid). The frequency of LTBI reactions in the new immigrants, including the three missed follow-up participants, is shown in Table 2.

- According to the induration reaction, the prevalence of LTBI in the sample of new immigrants was zero (0/177) using the TST method. Applying the new classification criteria of LTBI, all participants belonged to the normal results within score I (zero=non reaction) and score II (≤ 5 mm induration) in 97.78% (176/180). Immigrants had score III (> 5 - ≤ 10 mm induration) were 0.56% (1/180) with no observed case for positive abnormal reaction above the modified 'cut-off' value (>10 mm) (LLR χ(2)=33.865, p<0.001) as shown in the next Table 3.

- Average LTBI category is zero (means no detected cases), Missed=excluded case due to missed follow-up or came after 5 days for reading of TST reaction, cannot be judged means excluded cases (3 missed follow-up for TST results and one pregnant female not preformed chest X-ray due to contraindication)

- Analysis of the influence of BCG vaccination status on test results
is shown in Table 3. BCG vaccination status was ascertained from the participants through availability of reliable past history of vaccination and/or presence of a characteristic BCG scar. Analysis of the effects of BCG vaccination status (either vaccinated or non-vaccinated) represented in relation to the defined cases of latent tuberculosis infection categories is shown in Table 4.

The overall median age of BCG vaccination for the 180 participants was 5 years (IQR=2 years). The 'high' LTBI group had been vaccinated at older ages (median=7.5 years, IQR=6 years). The minimum (youngest) age of vaccination was one year versus the oldest maximum age of 13 years was answered (Table 4). However these findings did not show a statistically significant difference between normal vaccination ages for LTBI development (KW \( \chi^2(4)=3.330, p=0.504 \)) (Figure 1).

Presence of BCG vaccination scar was not a statistically significant risk factor for LTBI development or to cause positive a TST reaction, even in those expatriates classified as 'high' and 'extremely high' LTBI cases who were having forearm scar.

A positive scar was detected in 86.11% (155/180) of participants (LLR \( \chi^2(4)=6.104, p=0.192 \)). The overall median age of BCG vaccination for the participants was 5 years (IQR=2 years) (Table 4).

Immigrants having a history of exposure to poultry animals e.g.
here, duck and/or dairy animals e.g. goat, buffalo and/or dog in their mother country were 52.22% (94/180), and had no significant statistical association with LTBI suspicion except at the 10% level of significance according to the LTBI categories. Table 4 shows the distribution of the majority for those considered as ‘negligible’ LTBI cases in 56.67% (51/90) and ‘extremely high’ LTBI individuals in 52.17% (24/46) (LLR $\chi^2$(4)=7.928, $p=0.094$). On the other hand, a correlation between owning and/or exposure to cattle (cow) in mother countries and LTBI development was detected only in 15.56% (28/180) of total immigrant cases, but was not statistically significant (LLR $\chi^2$(4)=3.114, $p=0.539$) (Table 4).

Discussion

The influx of BCG-vaccinated immigrants in non-endemic countries from TB high-incidence regions represents a potential pool for new TB infection and highlights the need for LTBI early and accurate detection (reducing TB reservoirs) by using other higher specific diagnostics such as interferon gamma release assays (IGRAs) [10]. A common confounder, which affects TST reactivity, is prior vaccination with BCG vaccine, and/or the strain and dose of BCG used, and/or method of vaccine administration and age at vaccination. Successful vaccination can be assessed by the presence or absence of a BCG scar [11]. Similar finding was emphasized on skin testing in BCG-vaccinated populations to predict LTBI suspicious carriers [12]. Therefore TB skin testing at immigrant entry centres in Kuwait that serve large foreign-born populations can be effective and might add suspicion for LTBI carriers. Li et al. (2010) [13] detected LTBI prevalence of positive TST in 24.4% (higher among foreigners) in BCG-vaccinated which help to target TST testing before starting immigrant’s chemoprophylaxis.

Exposure to previous BCG vaccination did not significantly reduce the risk of being diagnosed as active or latent TB was also revealed by Caley et al. (2010) [14], in common with our findings.

This study proved that the presence of a BCG scar and related TST negative results of all immigrants were not significantly associated with a higher prevalence of LTBI as also noted by Demkow et al. (2008) [15]. Other similar significant findings was detected the absence of risk difference in TB patient contacts having positive BCG scar compared to other normal control contacts without scars [16]. Also Kik et al. (2009) [17] showed no association between the presence of BCG scar with recent exposure to TB in immigrants having positive IGRAs and TST results.

Since BCG-TST positive reactivity wanes with time (if more than five years have elapsed since administration of BCG vaccine) - a positive TST reaction is most likely a result of exposure to MTB infection [18]. Gomes and colleagues (2011) [19] have recently concluded that waning of the BCG-induced protection was associated with raised risks of TB morbidity in children aged between three and five years. Soysal et al. concluded that absence of an immunization scar can determine the likelihood for TB infection and is significantly associated with TB disease severity, which can be related to the level of exposure to MTB [2].

TST reactions can be interpreted regardless of BCG vaccination history [6]. Absence of positive TST reactions in our study strengthens the inference that BCG is not interfering with TST interpretation even though 86.11% (155/180) of participants had already been vaccinated against TB with BCG around pre-school ages. Similar related finding we recorded by Minodier et al. (2010) [20], namely that a positive TST is more likely to be related to an increased duration of TB exposure in the TB-endemic country of birth rather than to previous BCG vaccination. TST cannot be positive in certain biological factors due to suppression of DTH-IV reaction and T-lymphocytes such as malignancies and viral infections (e.g. HIV) [21].

Bradshaw et al. (2011) [22] suggested that advancing age increases the likelihood of exposure to unpasteurized dairy products and cross-reactions of environmental mycobacterial antigens using IGRA tests. A similar significance in our research results of those participants having past history of exposures to various animals and positive BCG scar without LTBI diagnosis. On the contrary Grafein et al. (2011) [23] concluded that there was an absence of a significant association between the presence of LTBI and a BCG scar, but presence of a significant association with consumption of unpasteurized (cows) milk in the past 6 months, which can be related to Mycobacterium bovis.

Similar to the majority of BCG-related publications, the size of BCG scar was not measured in our research because did not correlate with protection against TB, and also is not an indication for diagnosis (or presence) of LTBI. A similar result was also achieved by Crampin et al. (2009) [24].

TST reactivity caused by BCG vaccine generally wanes with the passage of time, but periodic skin testing may prolong reactivity in vaccinated persons, the phenomenon of ‘boosting reactivity’ [6] (CDC, 2010). On the contrary, TST reaction size and TST results were not affected by the time since the last dose of BCG vaccination, the number of BCG scars or BCG vaccination schedule in children, and significantly affected the decision for LTBI prophylaxis [25].

Study Limitation and Strength

The absence of a gold standard test for latent TB infection and TB can be considered as a limiting factor. Another limitation is the small sample size which might constrain generalization of the results to a larger population and wider community. On the other hand presenting and comparatively testing the new evidence-based diagnostic criteria for LTBI in addition to absence of any previous data on LTBI prevalence in immigrants or Kuwait residents add strength for future comparison.
Conclusions

Tuberculin skin test is extremely limited as a diagnostic tool for latent tuberculosis infection to prove whether a positive result is caused by cross-reactivity resulting from BCG vaccination. Still a history of BCG and/or presence of vaccination scar (those answered ‘No’ or ‘Unknown’ vaccination status) is not a contraindication for tuberculin skin testing or LTBI chemoprophylaxis in suspected individuals.

Recommendations

The relationship between immunogenicity against tuberculin skin test in the response to BCG vaccination needs to be elucidated. Considering the confounding effect of Mycobacterium tuberculosis risk factors, BCG efficacy and protection need further evaluation using interferon gamma release assays to compare between LTBI detected in both vaccinated and non-vaccinated populations. Emphasis on skin testing in BCG-vaccinated populations should be considered in the appropriate clinical setting to predict LTBI suspicious carriers.

References

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