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A Mutation in the Mitochondrial Fission Gene Dnm1l Leads to Cardiomyopathy

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Abstract

Mutations in a number of genes have been linked to inherited dilated cardiomyopathy (DCM). However, such mutations account for only a small proportion of the clinical cases emphasising the need for alternative discovery approaches to uncovering novel pathogenic mutations in hitherto unidentified pathways. Accordingly, as part of a large-scale N-ethyl-N-nitrosourea mutagenesis screen, we identified a mouse mutant, Python, which develops DCM. We demonstrate that the Python phenotype is attributable to a dominant fully penetrant mutation in the dynamin-1-like (Dnm1l) gene, which has been shown to be critical for mitochondrial fission. The C452F mutation is in a highly conserved region of the M domain of Dnm1l that alters protein interactions in a yeast two-hybrid system, suggesting that the mutation might alter intramolecular interactions within the Dnm1l monomer. Heterozygous Python fibroblasts exhibit abnormal mitochondria and peroxisomes. Homozygosity for the mutation results in the death of embryos midway through gestation. Heterozygous Python hearts show reduced levels of mitochondria enzyme complexes and suffer from cardiac ATP depletion. The resulting energy deficiency may contribute to cardiomyopathy. This is the first demonstration that a defect in a gene involved in mitochondrial remodelling can result in cardiomyopathy, showing that the function of this gene is needed for the maintenance of normal cellular function in a relatively tissue-specific manner. This disease model attests to the importance of mitochondrial remodelling in the heart; similar defects might underlie human heart muscle disease.

Introduction

Idiopathic dilated cardiomyopathy (DCM) is characterised by unexplained left ventricular (LV) cavity enlargement with LV systolic impairment [1]. DCM is an important cause of congestive heart failure (CHF) with a prevalence of 36 cases per 100,000 in the United States [2]. Although the molecular pathways responsible for DCM remain largely unknown, it is estimated that between 20–50% of DCM cases are familial in nature, the large majority being inherited as an autosomal dominant trait [3]. Accordingly, the study of single gene disorders that remodel the heart to cause DCM may provide a valuable opportunity to identify critical molecules involved in disease pathways [4].

Over the past decade, DCM-causing mutations have been identified in genes encoding sarcomeric, cytoskeletal, nuclear envelope, intermediary filament, sarcoplasmic reticulum and desmosomal proteins. These findings have implicated pathogenic mechanisms whereby structural integrity, contractile force dynamics, and calcium regulation within the cardiomyocyte are perturbed. Yet such mutations only account for a minority of DCM cases [5] and many of the genes underlying DCM remain to be elucidated. A number of gene knockouts in the mouse produce features of DCM, but these phenotypes are usually recessive and so do not model the human disease. One approach that has been applied successfully to the characterisation of new disease alleles is the use of N-ethyl-N-nitrosourea (ENU) mutagenesis [6]. Treatment of mice with ENU results in a high frequency of predominantly single point mutations in the mouse germ line that recapitulate the spectrum of mutations observed in many human genetic diseases. Screening of offspring reveals phenotypic variants, and the identification of the mutations underlying the abnormal phenotype can reveal new genetic regulators and novel pathways associated with disease pathogenesis. Such an approach is ‘hypothesis neutral’, making no prior assumptions about the
Heart disease is very common. Some cases of heart disease are strongly influenced by lifestyle and diet, whereas others have a strong genetic component. A certain form of heart failure, known as dilated cardiomyopathy (DCM) quite often runs in families suggesting that a defective gene or genes underlie this disease. We describe a new mouse mutant called “Python” which suffers from a heart disease similar to DCM. We were able to pinpoint the defective gene responsible for the disease. This gene is normally involved in the division of mitochondria, the “power plants” of the cell that generate one of the main energy supplies for the cell. This is a unique model that implicates a new gene and mechanism of disease for further investigation.

Results

Python mice develop biventricular DCM

The Python mouse was identified on the basis of rapid size increase, piloerection, and shallow rapid breathing in a visual screen of adult G1 offspring of ENU-mutagenized BALB/cAnN.Crl males crossed with C3H/HeN females (Figure 1A). The phenotype was inherited in an autosomal dominant fashion with complete penetrance in both sexes. The median age of onset of overt symptoms of CHF on a C3H/HeN genetic background was 91 days for females and 83 days for males. A similar phenotype with much later onset occurred on a C57BL/6J background (median age of onset = 164 days for females, 171 days for males; Figure 1B) suggesting that strain-specific genetic modifiers influence disease onset.

The hearts of Python mice were grossly dilated by the time of overt CHF (Figure 1C). The Python hearts exhibited both biatrial and biventricular thinning and dilatation consistent with DCM (Figure 1D). The visible increase in size of Python mice was the terminal phenotype with much later onset occurred on a C57BL/6J background (median age of onset = 164 days for females, 171 days for males; Figure 1B) suggesting that strain-specific genetic modifiers influence disease onset.

Cardiac function was analysed in male Python mice on the C3H/HeN background aged 71–78 days, i.e. approximately 2 weeks before overt clinical signs of CHF become evident. The nature of the genes or pathways involved. Using this approach we describe a novel mouse model of DCM in which a mutation in the Dynamin-1-like gene (Dnm1l) leads to autosomal dominant DCM and congestive heart failure.

Positional cloning of the Python mutation

The Python mutation was generated on a BALB/cAnN genetic background, and the original mouse exhibiting the CHF phenotype was a BALB/cAnN.Crl x C3H/HeH F1 hybrid male. This individual was backcrossed to C3H/HeH and offspring exhibiting the CHF phenotype were examined with a panel of 53 polymorphic SNP markers spanning the entire genome at regular intervals. A strong linkage signal was observed on chromosome 16 at marker D18Mit215 (LOD score of 3.3 for a recombination fraction = 0.1). Subsequent fine mapping by backcrossing to C3H/HeH, C3H/HeN and, finally, to C57BL/6J mice narrowed the critical region containing the mutation to 787 Kb (Figure 2A). Three genes are located within this region - Phlp2, Fdg4 and Dnm1l. Sequencing of all exons and exon-intron boundaries identified only a single mutation in Python mice; a G/C to T/A transversion in exon 11 of the Dnm1l gene (the official gene name as specified by the International Committee on Standardized Genetic Nomenclature for Mice but often referred to in the literature as Dbp1 or Dlp1) (Figure 2B). Based on the known ENU-induced mutation rate for this strain and ENU dose (1 mutation per 1.8 Mb) [7], the probability of there being an additional intronic or intergenic mutation anywhere in this region is extremely low (P = 0.00001) [8]. There was complete concordance between animals suffering from CHF and the Python mutation. We retrospectively regenotyped 145 DNA samples isolated from C3H and C57BL/6J Python mice that had suffered from CHF and all contained the mutation while it was absent in 189 samples from non-affected littermate control mice. None of the wild type strains examined (BALB/cAnN.Crl, C3H/HeN, C3H/HeH, C57BL/6J, DBA/2J, CBA/J, 101/H, 129/SvJ) contained the mutation. Although PKP2 dominant mutations have been associated with arrhythmogenic right ventricular cardiomyopathy in humans [9] an intronic or intergenic mutation is unlikely to account for the Python mutation.
Figure 1. The Python mutation leads to dilated cardiomyopathy. (A) A 13-week-old Python mouse compared to a littermate control. (B) Kaplan-Meier analysis of onset of overt CHF in Python mice on the C3H/HeN and C57BL/6J genetic backgrounds compared to wild type littermate controls. (C) Photo of the excised hearts from a 13-week-old Python and wild type mouse. Note the grossly enlarged ventricles and atria. (D) H&E section through a wild type and Python heart showing showing clinical signs of CHF. Note the distended ventricles; R = right, L = left. (E) Oedematous
phenotype as all human PKP2 mutations found to date occur in the coding region or splice sites which are not mutated in Python mice, the homozygous phenotype for the null Pkp2 mouse, the homozygous phenotype for the null the coding region or splice sites which are not mutated in Python mouse, the homozygous phenotype for the null the coding region or splice sites which are not mutated in Python mouse. The cysteine residue is with 96% sequence conservation between mouse and zebrafish. The degree of evolutionary conservation of Dnm1l protein is very high. For example, overall homology between human and mouse Dnm1l is 98%, and between zebrafish and mouse Dnm1l is 97% (amino acid numbering according to EBI reference protein Accession No. Q8K1M6) (Figure 2C). This cysteine is located by a phenylalanine at position 452 in the predicted Dnm1l protein (amino acid numbering according to EBI reference protein Accession No. Q8K1M6) (Figure 2C). This cysteine is located within the middle (M) domain of the protein and is fully conserved in all Dnm11 orthologues, and even the yeast dynamin homologue DNM1 (Figure 2D). The degree of evolutionary conservation of the Dnm11 protein is very high. For example, overall homology between human and mouse Dnm11 is 98%, and between zebrafish and mouse is 89%. The M domain conservation is even higher with 96% sequence conservation between mouse and zebrafish over the 291 amino acids of this domain. The cysteine residue is also conserved in the M domain of the mouse homologues of Dnm1, Dnm2 and Dnm3 (Figure 2E) despite overall homology with these domains being less than 40% (Table S1) suggesting that this cysteine plays an important role in M domain function.

The Python mutation impairs intramolecular interaction of Dnm11

There is no available crystal structure of any mammalian dynamin proteins but a crystal structure has been described for a bacterial dynamin-like protein. In this structure the M domain forms an elongated alpha-helical domain where the tip of the M domain helices interact with a similar region of the 'mate' in the dynamin homodimer [12]. Accordingly, a model of mouse Dnm11 was constructed on the basis of comparative sequence homology to the bacterial dynamin-like protein BDLP for which there is a crystal structure [12] and an electron cryomicroscopy reconstruction of BDLP assembled around a lipid tube [13]. A predicted structure could be created for most of the protein, apart from one region where there is no homology in BDLP (indicated by an ‘a’ in Figure 3A). The predicted structure of the dimeric asymmetric repeating unit in the extended confirmation (i.e. after lipid binding) is shown in Figure 3A.

There are six mutations, all dominant or semi-dominant, that have been reported in the M of domain of DNM1L or its yeast homologue DNMI–three in yeast [14], and one each in a human

Table 1. Cardiovascular parameters in 11-week-old male mice.1

<table>
<thead>
<tr>
<th>Organ weights</th>
<th>Littermate controls (n = 6)</th>
<th>Python (n = 6)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (g)</td>
<td>34.0 ± 2.8</td>
<td>32.5 ± 2.1</td>
<td>0.31</td>
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<tr>
<td>LV weight (mg)</td>
<td>94 ± 6</td>
<td>90 ± 6</td>
<td>0.30</td>
</tr>
<tr>
<td>RV weight (mg)</td>
<td>28 ± 4</td>
<td>27 ± 3</td>
<td>0.72</td>
</tr>
<tr>
<td>Lung weight (mg)</td>
<td>138 ± 5</td>
<td>141 ± 16</td>
<td>0.65</td>
</tr>
</tbody>
</table>

| Echocardiography | | | |
| End-diastolic area (cm²) | 0.123 ± 0.015 | 0.132 ± 0.009 | 0.22 |
| End-systolic area (cm²) | 0.069 ± 0.017 | 0.082 ± 0.014 | 0.19 |
| Ejection fraction (%) | 44 ± 8 | 38 ± 9 | 0.24 |
| Wall thickness (mm) | 0.82 ± 0.03 | 0.80 ± 0.03 | 0.19 |
| Heart rate (bpm) | 460 ± 52 | 461 ± 59 | 0.98 |

| Haemodynamics | | | |
| LV systolic pressure (mmHg) | 97 ± 8 | 75 ± 3 | 0.00007 |
| LV end-diastolic pressure (mmHg) | 3.8 ± 1.1 | 17.7 ± 9.2 | 0.004 |
| dP/dtmax (mmHg/s) | 8531 ± 1094 | 5140 ± 1135 | 0.0004 |
| dP/dtmax/instantaneous pressure (s⁻¹) | 149 ± 8 | 107 ± 23 | 0.002 |
| dP/dmax (mmHg/s) | −849 ± 1381 | −4564 ± 1308 | 0.001 |
| Tau (ms) | 8.5 ± 1.5 | 16.6 ± 6.8 | 0.017 |
| Mean arterial pressure (mmHg) | 75 ± 9 | 60 ± 6 | 0.006 |
| Central venous pressure (mmHg) | 1.8 ± 0.2 | 1.6 ± 0.8 | 0.83 |
| Dobutamine dP/dmax (mmHg/s) | 13003 ± 1832 | 6344 ± 1424 | 0.00004 |

1 Mice were on a C3H/HeN genetic background. All data is mean ± SD with comparisons made by Student’s t-test.

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A Mutation in Dnm1l Leads to Heart Failure

A

<table>
<thead>
<tr>
<th></th>
<th>M279</th>
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<th>Mb</th>
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<tbody>
<tr>
<td>D16Mit57</td>
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<td>25.9</td>
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<td>SNP 16.014.985</td>
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<td>15.9</td>
</tr>
<tr>
<td>D16Mit56</td>
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</tr>
</tbody>
</table>

Py-containing region

B

45.3 Kb

AGCATTGTAGCAA

+/+

Py/+ 

Py/Py

C

CAT | TGT | AGC

H   C   S

G→T

D

GTPase

M

PHD

GED

E

Dnm1l

Dnm2

Dnm3

Dnm3

Dnm3

Dnm3

Dnm3

Dnm3

Dnm3
patient [15], a CHO cell line [16], and Python (Figure S1). These were mapped on to the predicted structure (Figure 3A). The Python mutation is located in an alpha-helix that is not predicted to affect interaction between Dnm11 monomers. However, it is located close to several other helical regions of the domain M. Furthermore, a helix-wheel projection of the region around the Python mutation-containing region predicts that one face of the predicted helix contains principally hydrophobic residues (Figure 3B). Taken together, these findings are suggestive of this face being involved in an intramolecular interaction within the Dnm11 monomer.

To test this further, we used the yeast two-hybrid assay based on GAL4 DNA binding and activation domain interactions to examine whether interactions between regions of Dnm11 could be altered by the Python mutation. We used regions of the protein that have been used by others in similar assays [17–22] and examined all possible reciprocal interactions of bait (in pDEST32) and prey (in pDEST22) proteins for regions of Dnm11: full-length, N-terminal region, C-terminal region, M domain and GED (GTPase Effector Domain) (Figure 3C). On the basis of ability to grow on medium lacking histidine and b-galactosidase activity, the only strong interactions we identified were interactions between the full-length proteins, the N terminal and C-terminal regions of the proteins as reported by Zhu et al. [20], and the N-terminal region and the GED (Figure 3D). We found that while the Python mutation had negligible influence on the ability of the full-length proteins to interact (Figure 3D and 3E), it abrogated the ability of the N-terminal region to interact with both the C-terminal region and the GED alone (Figure 3D). Quantitative assays for b-galactosidase activity confirmed the substantial effect the Python mutation had on these interactions (Figure 3E).

The Dnm11PY mutation results in the impairment of mitochondrial and peroxisomal dynamics

Given that the Python mutation occurs in a highly conserved domain of the Dnm11 protein and alters protein interaction in vitro, its effect on in vivo functions associated with Dnm11 were examined. Protein levels of Dnm11 were not altered in either heart or brain (Figure 4A) suggesting that there is no haploinsufficiency (i.e. the Python protein is assumed to be present). Dnm11 was distributed diffusely within the cell in both Python and wild type cultured skin fibroblasts (Figure 4B) suggesting that introduction of the Python protein did not drastically alter trafficking of Dnm11. As Dnm11 function is involved in mitochondrial and peroxisomal dynamics, cultured neonatal skin fibroblasts were examined for morphology of both these organelles. Mitochondrial morphology was altered. Python fibroblasts were highly elongated compared to wild type controls (Figure 4C), as were peroxisomes (Figure 4D). To determine if mitochondrial volume was altered, we utilized a novel assay where cells were loaded with a fluorescent mitochondrial marker and analyzed by flow cytometry. Intensity of fluorescence should reflect mitochondrial volume in the cell. As shown in Figure 4E, there was no difference in overall mitochondrial volume between Python and wild type fibroblasts, despite the significant changes in mitochondrial shape. This indicates that the Python mutation affects the in vivo functional activity of Dnm11, thereby impairing mitochondrial fission.

Homozgyosity for the Python mutation is embryonic lethal; heterozygosity for Python modifies mitochondrial structure

To discern the functional effects of the Python mutation, we intercrossed heterozygotes to obtain homozygous animals. Genotyping of embryos demonstrated that no homozygous Python embryos could be recovered from E12.5 onwards (Figure 5A). Homozygous embryos appear to be normal up to approximately E9.5. At E11.5, Py/Py embryos were severely retarded in growth and exhibited a posterior truncation (Figure 5B). The homozygous embryonic phenotype is very similar to that recently reported for the Dnm11-null mutation [23–24]. Embryos die at a similar stage and their morphology is similar. Mouse Embryonic Fibroblasts (MEFs) cultured from homozygous E9.5 embryos survived poorly in culture. Few cells attached and there was no proliferation (Figure 5C). Mitochondria of Py/+ MEFs were abnormal with numerous long tubular mitochondria (Figure 5C), similar to Py/+ skin fibroblasts. In contrast, homozygous Python MEFs had grossly abnormal mitochondria. Whereas a tubular mitochondrial network evenly distributed throughout the cytoplasm characterized mitochondria in +/+ and Py/+ MEFs, some mitochondria of Py/Py MEFs appeared to be spherical and aggregated (Figure 5C). This could reflect Dnm11 dysfunction alone though it may reflect a general dysfunction of these cells. As mentioned earlier, these cells fail to proliferate. Over several weeks in culture they slowly die. The nuclei staining with Hoechst 33342 demonstrated evidence of chromatin condensation in homozygous Python cells (Figure 5C), suggesting the cells could be dying by necrosis.

The Dnm11PY mutation results in the impairment of energy metabolism in the heart

The observation that Python fibroblasts exhibit abnormal mitochondria coupled with the well-recognized role of DNMI1 in mitochondrial fission [25] and the critical role of mitochondria in both the normal function and the death of cardiomyocytes [26], led us to examine the mitochondrial and energetic phenotype of Python heterozygous mice in greater detail. Aside from the development of CHF, the Python mice did not exhibit any features prominently recognized in mitochondrial cytopathies, such as metabolic, neurological and skeletal muscle defects. They exhibited a normal general behavioral and functional profile as defined by the SHIRPA series of tests (Table S2) [27], which would reveal any major neurological abnormalities. Grip strength (a reflection of muscle strength) was normal, as was muscle histology as assessed from H&E-stained sections (data not shown). Plasma lactate levels, an indicator of general metabolic dysfunction, were not elevated in Python mice aged 5–9 weeks compared to wild type littermate controls (Figure 6A).

Given that the Python phenotype at a gross pathological level appeared to be restricted to the heart, we examined the mitochondrial phenotype of Python cardiomyocytes. There was
A Mutation in *Dnm1l* Leads to Heart Failure

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**A**

- G → R (yeast)
- C → F (Python)
- G → D (yeast)
- G → S/D (yeast/CHO)
- P → L (yeast)
- A → D (human)

**B**

- Hydrophilic
- Hydrophobic

**C**

- GTPase
- Py (C → F)
- M
- 1-742
- 1-495
- 305-523
- 566-742
- 641-742
- 1-742

**D**

- pDEST32
- pDEST22

---

**E**

- % Full-length wild-type interaction
- Expression constructs

---

**Legend**

- SI
- WI
- NI
- Neg
- trp-leu
- trp-leu-his+100mM AT
- YPAD/β-gal
little evidence of morphological change in Python cardiomyocyte mitochondria. For example, we examined the nuclear:mitochondrial DNA ratio in Python hearts as mtDNA is lost when *DNM1L* is down-regulated and mitochondrial fission impaired [28]. However, there was no alteration in the nuclear:mitochondrial DNA ratio in Python hearts at any stage before the development of overt CHF and even at this stage only some Python hearts showed an increased ratio (Figure 6B). This suggests that a major derangement of nuclear:mitochondrial DNA ratios was not a generalized effect of the Python mutation.

There was evidence of mitochondrial function changes in hearts of Python mice suffering from overt CHF. Examination by enzyme immunohistochemistry for succinate dehydrogenase (SDH) and cytochrome c oxidase (Complex IV) activities revealed a reduction for both enzymes in Python hearts (Figure 6C). In the case of SDH there is evidence of diminished myocellular enzyme activity as indicated by less-intense staining. However for Complex IV, the enzyme immunohistochemistry suggests that in the failing heart, the change in Complex IV enzyme levels may be due to the substantial fibrosis that occurs late in the disease process rather than a change in enzyme activity. *In vitro* measurement of Complex IV activity and quantity showed that both were proportionately reduced by similar amounts in ailing Python hearts (Figure 6D), indicating that the enzyme remains fully active but is less abundant at 12 weeks of age. This reduction in overall levels at this age most likely reflects the considerable fibrosis and loss of cells that occurs in late-stage Python hearts.

Defects in mitochondrial enzyme activity are recognized as a general phenomenon in CHF [29] and, therefore, the differences observed in failing Python hearts might be secondary to the primary cause of heart failure. To determine if the Python mutation was affecting mitochondrial function prior to major changes in heart structure, we examined heart samples from Python and wild type littermates at 10 weeks of age. This is before there are any overt signs of CHF though the cardiovascular data above (Table 1) indicates that heart function is abnormal at this stage. Electron micrographs of heart samples were examined. Mornorphometric analysis of the proportional area occupied by mitochondria (a reflection of overall volume per cell) revealed no difference between Python hearts and controls (Figure 6E). This in agreement with the flow cytometry findings in Python skin fibroblasts (Figure 4E). Nor were myofilament widths significantly different between Python hearts and controls (data not shown). There was no evidence of the membrane pinching reported in the *Dnm1l*-null mouse fibroblasts [Ishihara2009], nor was there evidence of large aggregates as has been reported for *in vitro* cultured *DNM1L* mutants [30]. However, the average size of a mitochondrion was slightly smaller in Python hearts than wild type hearts (Figure 6E). Smaller cardiomyocyte mitochondria have been previously reported in some cases of heart failure [31–33]. In the case of Python, if mitochondrial volume is not altered but mitochondrial tubules are extended in length, then a smaller cross-sectional transverse area of mitochondria would be consistent with this.

If *Dnm1l* affects mitochondrial dynamics, we predicted that the end point of this would be impairment of respiratory chain function. Examination of mitochondrial enzyme complex activities normalized to citrate synthase activity in heart samples from 10-week-old mice revealed no differences (Figure 6F and 6G). There was a slight reduction in the overall level of mitochondrial citrate synthase activity in Python hearts at this age but it was not statistically significant (Student’s *t* test, *P* = 0.11) (Figure 6G) also suggesting that the total mitochondrial volume in Python hearts was not reduced. As the end-point of respiratory chain function is ATP synthesis, and given that down-regulation of *DNM1L* results in a reduced rate of ATP synthesis [28,34], myocardial ATP and total adenine nucleotide (TAN) levels were measured using HPLC. Python hearts exhibited a dramatic, approximately 50%, reduction in ATP and TAN levels (Figure 6H and 6I) compared to hearts from littermate controls. In liver and brain at the same age, ATP and TAN levels were similar in Python mice and controls (Figure 6H and 6I), indicating that the defect in ATP generation was not a general one. These results were confirmed by using complementary quantitative bioluminescence assays (Figure S2).

A range of metabolites in the heart was examined using high-resolution 1H NMR spectroscopy and Gas Chromatography Mass Spectrometry (GC-MS)-based metabolomic approaches [35] on Python and control mice. The metabolite changes identified are summarized in Table 2. Significant reductions in Python hearts were noted in mitochondrial metabolic intermediates or accessory molecules e.g. succinate, malate, fumarate, pyruvate (all associated with the citric acid cycle), creatine, glucose, AMP and adenosine. Two notable increases were in the amino acids glycine and proline. These account for approximately 50% of the amino acids in collagen, possibly reflecting the fibrosis in Python hearts.

**Discussion**

We report the identification, through ENU mutagenesis, of a novel genetic cause of cardiomyopathy. Our principal finding is that a missense mutation in the middle domain of *Dnm1l*, whose product is critically involved in mitochondrial fission, results in DCM. The resulting defect in mitochondrial remodelling renders the Python hearts progressively energy deficient potentially contributing to the phenotype [36]. The fundamental importance of mitochondrial remodelling in mammalian pathophysiology has been underlined by *in utero* lethality and cerebellar degeneration in mice with homozygous
Figure 4. Mitochondrial and peroxisomal morphology is altered by the Python mutation. (A) Western blot analysis of total heart and liver protein extracts from 5-week-old male mice of the indicated genotypes demonstrating that Dnm1l protein levels are not altered. Western blots were re-examined using an anti-Tim23 antibody, an inner mitochondrial membrane protein, which demonstrate that the level of this mitochondrial protein is also not altered in Python mutants. After stripping, α-tubulin was detected to demonstrate loading levels of protein extracts. (B) Typical example of immunocytochemistry in neonatal mouse skin fibroblasts using an anti-DRP1 antibody. There was no appreciable difference between Python
mutations in Dnm1l itself, as well as Mfn1, Mfn2, or Opa1 [23,37,38]. Given the heart’s manifest dependency on mitochondria [39] as evidenced by its frequent involvement in mitochondrial disorders [40], mitochondrial remodelling defects might be expected to occur in some forms of myocardial disease. Recently it has been reported that mitochondria are smaller in failing hearts and DNM1L protein levels were increased in DCM heart samples [33] but until now, there has been no direct evidence that genes involved in regulating mitochondrial dynamics might be involved in heart failure. Python is the first such example.

DNM1L is a member of the dynamin superfamily. In the higher order spirals formed by dynamin, the basic repeating unit appears to be a dimer [41]. The association between the GED and M domains forms a ‘stalk’ conformation through which strong intramolecular interactions of the dimers occur [41]. Mutations in the M domain have accordingly been shown to adversely affect self-assembly into a higher order oligomeric structures that are critical to Dnm1l function [22] and conformational changes within this region are associated with the constriction of dynamin tubules that facilitates fission [42]. It is not difficult to envisage how the substitution of a bulky hydrophobic phenylalanine residue into a helix-rich region of the M domain (Figure 3A) has the potential to modify interactions necessary for Dnm1l’s effective function. As a corollary, our yeast two hybrid analysis showed Python’s capacity to abrogate Dnm1l’s N-terminal region interacting with the C-terminal region (and GED), while leaving the interaction between Dnm1l monomers unaffected. A similar effect was reported when the S637D mutation was introduced into Dnm1l [43]. This effect of the mutation on Dnm1l higher order structure is consistent with a dominant-negative mode of action. Since there appears to be no change in overall Dnm1l protein levels, we propose that the Python monomer is readily incorporated into dimers with the wild type protein but fails to function effectively within that dimer due to defective intramolecular interactions. If this model is accurate, only 1 in 4 Dnm1l dimers might be expected to be fully functional.

This study cannot exclude a systemic (i.e. non-cardiac) impact of the C452F mutation, though from a clinical perspective, most tissues and organ functions appeared to be grossly spared. The liver was the one organ that did show evidence of extra-cardiac involvement but this might reflect congestive cardiac heart failure caused by heart disease. The predominant cardiac phenotype in the Python heterozygotes contrasts with the marked skeletal muscle and metabolic abnormalities in the infant with a dominant mutation in DNM1L [15]. It is unclear why the Python mutation manifests an overt phenotype only in heart. The reported human mutation in domain M, a dominantly acting alanine to aspartate acid mutation, appears to be much more severe than the Python mutation appeared to have a widespread metabolic defect. This may be related to the location of the amino acid within the structure and/or the chemical nature of the amino acid substitutions involved. An inter-species difference in Dnm1l functionality is possible but unlikely given the extremely high degree of inter-species conservation. All 7 mutations identified in the M domain that affect mitochondrial dynamics are conserved not only in the M domain of DNM1L orthologues across multiple species (Figure S1), but are also conserved across the M domain of DNM1, DNM2, DNM3 and DNM1L. (Figure 2E). The human alanine to aspartate mutation occurs in a region of the protein that might interact with the membrane (Figure 3A). When overall compatibility of the amino acid replacements are compared the Python amino acid replacement scores significantly higher in compatibility with its original amino acid in terms of hydrophobicity, size and charge compatibility (15.8, 9.4, 13.17 for the Python mutation compared to 12.8, 8.6 and 12.82 for the A400D human DNM1L mutation) [44]. Given that there is no evidence that this cysteine is involved in disulphide bonding in DNM1L, and its effect in the yeast two-hybrid system on intramolecular but not intermolecular Dnm1l interactions, its effect on Dnm1l function may be relative mild, still enabling the formation of dimers but with reduced functionality.

Although the gross impact of the heterozygous C452F Dnm1l mutation on cardiac morphology was relatively subtle at ~70 days, the corresponding functional and, even more so, the biochemical phenotypes were apparent. This was reflected in uniform lethality shortly thereafter. The degree of ATP deficiency in the Python hearts at this age is, to the best of our knowledge, unprecedented. The apparent fall of ATP levels to 44% of wild type levels in hearts from 10-week-old mice (15.1 vs. 6.6 nmol ATP/mg/protein in +/- and P0/+ hearts at 10 weeks of age confirmed by two independent means) may have been exaggerated by increased fibrosis, yet still seems beyond the range by which it is thought that cardiac [ATP] is allowed to diminish (~25% to 30%) even in CHF [39]. We used an exacting method involving HPLC to measure these levels to ensure they were as accurate as possible. However a major caveat to these observations, is that the degree of difference in ATP levels observed herein will inevitably be exaggerated by the technical limitations associated with measuring ATP levels in tissue samples after excision from the body. The resulting rapid depletion of ATP pools may have precluded accurate measurements. However, a difference in ATP levels between Python and control hearts (that is not found in liver and brains) may indicate that the Python mutation does espose an aberrant and progressive impact on myocardial energetics.

One possible explanation for the Dnm1l-mediated cardiomyopathy is cardiac energy deficiency. It is recognized that CHF is associated with, and in many cases exacerbated by, cardiac energy deficiency [36,39,45,46]. Indeed, the commonality of CHF to numerous primary mitochondrial diseases represents an excellent source of evidence that primary energy deficiency can and does cause CHF [36,47]. As reduction of Dnm1l function is known to impair cellular energetics [28,34] and as this pattern of cellular energetic impairment is profoundly and progressively manifested in Python hearts even in advance of gross cardiac dysfunction, cardiac energy deficiency may contribute to the phenotype of Python hearts and may be the proximate cause.

Detailed questions remain regarding the impact of the C452F substitution. Further biochemical and cellular studies are needed to investigate the hypothesis that such M domain mutations alter the intra/inter-molecular interactions and alter the homooligomerisation properties of Dnm1l [48]. Such studies will have to explain why in the context of the subtle reticular mitochondrial changes in Python, there is a discrepancy between the lack of an
Figure 5. Homozygosity for the Py allele results in embryonic lethality. (A) Distribution of genotypes of embryos recovered from Py/+ x Py/+ intercrosses. Embryo numbers genotyped at each time point were: E8.5–26, E9.5–48, E10.5–20, E11.5–19, E12.5–20, E13.5–21, E15.5–10 and E19.5–14.
A Mutation in Dnm1l Leads to Heart Failure

overall decrease in mitochondrial volume or function and the degree of ATP depletion. There are a number of other metabolic and cellular processes that could be altered by the Python mutation. Calcium cycling, for example, is altered in cells lacking the mitochondrial fission protein Mitofusin 2, reflecting its role in endoplasmic reticulum (ER)-mitochondria tethering [49]. Inhibition of Dnm1l alters ER structure [50], though we could find no alteration in the morphological appearance of the ER in Python fibroblasts (data not shown). Nevertheless, further investigation of Ca\textsuperscript{2+} uptake into mitochondria and release from ER is warranted. An energy defect not reflected in altered respiratory complex enzyme activity might also result from other disturbances such as uncoupling of electron transport and ATP production, perturbation of supercomplexes [51], cell cycling [52] organelle quality control through autophagy, or generation of reactive oxygen species [28]. It is possible that Python has effects on Dnm1l function, unrelated to mitochondrial dynamics.

The tissue specificity of the Python defect warrants further investigation. This may reflect unique properties of cardiomyocytes, their mitochondria, or a unique role for Dnm1l in cardiomyocytes. For example, a role for DN1M1L in the heart has been inferred by Ong et al. who observed that mitochondrial fission protects the heart against ischemia/reperfusion [53]. One strategy to address this question would be to effect conditional inactivation of Dnm1l in cardiomyocytes, in a similar manner to mitochondrial fusion factor elimination in skeletal muscle [54]. In conclusion, we report the first model of mitochondrial remodeling to be associated with cardiomyopathy. It is likely that the C452F substitution in the M domain of Dnm1l alters the balance of mitochondrial fission and fusion. The impairments in mitochondrial remodeling and function result in a relatively tissue specific disease as manifested by rapidly progressive cardiomyopathy. It is plausible that mutations in Dnm1l that are similarly subtle and hence do not represent a barrier to viability, will be identified and prove to be of importance in human disease.

Materials And Methods

Mice

Mice were maintained in high health status facilities with access to food and water ad libidum. All work was approved by the Animal Ethical Review Committees of MRC Harwell, University of Sheffield and University of Leeds, and the UK Home Office, and conducted with the highest quality of animal care and in accordance with the 3Rs.

ENU mutagenesis

Adult BALB/cAnNcrI mice aged 10 weeks were mutagenized by intraperitoneal injection of two weekly doses of 100 mg/kg ENU, mated with C3H/HeH and F1 offspring screened for abnormalities. Python mice were further backcrossed to C3H/HeN and C57BL/6J. All tissue samples used for metabolic and cellular analysis were derived from a line generated by inbreeding of an N11 backcross to C3H/HeN.

Genetic mapping

Genome-wide low-resolution mapping was performed using DNA samples from 15 N2 C3H/HeH backcross animals that were identified as carriers based on the development of congestive heart failure. Genomic DNA samples isolated from tail biopsies of these animals were screened by PCR amplification and gel electrophoresis with 53 microsatellite markers spaced at regular intervals across the genome. Samples were genotyped as either homozygous C3H or heterozygous BALB/c-C3H for each marker. For finer mapping, crosses of Python mice with both C3H and C57BL/6J were used, and further microsatellite markers polymorphic between either BALB/c and C3H or BALB/c and C57BL/6J were used to identify Python mice that were recombinant in the critical region. Single nucleotide polymorphisms (SNPs) were genotyped by sequencing of PCR products amplified with primers flankng the SNP site. Sequencing of candidate genes involved designing primer pairs to amplify individual exons as well as flanking splice donor/acceptor sequences. All exons and splice sites in the critical region were sequenced using homozygous as well as heterozygous Python DNA, ensuring that no base changes were missed.

Measurement of blood lactate

A small drop of blood was obtained from the lateral tail vein and processed using a Lactate Pro analyzer (HabDirect, Southam, UK).

Histology and TEM

Tissues for light microscopy were emersion fixed in 10% neutral buffered formalin and wax-embedded 3 \( \mu \)m sections were stained with haematoxylin and eosin or Mauritius Scarlet Blue (MSB). Mice were perfusion fixed for TEM. In situ staining for succinate dehydrogenase and Complex IV activities were a previously described [47,55,56].

In vivo cardiac phenotyping

Conscious ECG measurements were obtained in unrestrained male Python mice and littermate controls (n = 6 of each) at 10 weeks of age using the non-invasive AnonyMOUSE ECG screening tool (Mouse Specifics Inc). One week after, at 75 ± 2 days of age, the same mice were anaesthetised with isoflurane and placed on a homeothermic blanket. Parasternal short- and long-axis views were obtained under 1.25% isoflurane anaesthesia using an Agilent Sonos 5500 with 15 MHz transducer. The LV was cannulated via the right carotid artery with a 1.4F Mikro-tip conductance cannula (SPR-839, Millar Instruments). The superior vena cava was cannulated with a second 1.4F Millar cannula (SPR-671) to measure central venous pressure. Mice were allowed at least 15 minutes equilibration before baseline aortic and ventricular pressure measurements were obtained. Dobutamine was given by intraperitoneal injection (1.5 \( \mu \)g/g body weight) and pressure measurements obtained under maximal b-adrenergic stimulation. Mice were then killed by cervical dislocation and organs washed in heparinised saline, blotted and weighed. Two experiments were performed daily, alternating between genotypes for morning and afternoon experiments.

Enzymatic assays

All activities were determined at 30°C. Prior to analysis cells were subjected to three cycles of freezing and thawing to lyse.
A Mutation in Dnm1l Leads to Heart Failure

A

B

C

D

E

F

G

H

I

SDH
Complex IV
H&E
MSB

Mt Area

Mt Size

Activity
Quantity

Complexes

Level normalized to CS activity

mmol ATPing protein

mmol TANing protein

Genotype

Genotype

Py+/+

Genotype

Genotype

Genotype
membranes. Enzyme activities were assessed were assessed using a UVikon 940 spectrophotometer (Kontron Instruments Ltd, Watford, UK). Complex I activity was measured according to the method of Ragan et al. [57]. Complex II-III activity was measured according to the method of King [58]. Complex IV activity was measured according to the method of Wharton and Tzagoloff [59]. Citrate synthase (CS; EC 1.1.1.27) activity was determined by the method of Shepherd and Garland [60]. Enzyme activities were expressed as a ratio to citrate synthase to compensate for mitochondrial enrichment in cell samples [61].

**NMR and mass spectroscopy**

Tissues were extracted using a methanol: chloroform: water extraction procedure to separate aqueous soluble metabolites from lipids as previously described [62]. Briefly, ~100 mg tissue were pulsed with dry ice. 600 µl methanol:chloroform (2:1) was added and the samples were sonicated for 15 min. Water and chloroform were added (200 µl of each). The resulting aqueous and organic layers were separated from the protein pellet. The organic layer was dried overnight in a fume hood whilst the aqueous extracts were evaporated to dryness using an evacuated centrifuge (Eppendorf, Hamburg, Germany).

**NMR spectroscopy.** The aqueous extracts were rehydrated in 600 µl D2O, buffered in 0.24 M sodium phosphate (pH 7.0) containing 1 mM (sodium-3-(tri-methylsilyl)-2, 2, 3, 3-tetradeteriopropionate (TSP) (Cambridge Isotope Laboratories Inc., Andover, MA, USA). The samples were analysed using an AVANCE II+ NMR spectrometer operating at 500.3 MHz for the 1H frequency (Bruker GmbH, Germany) using a 5 mm ATMA TXI probe. Spectra were collected using a solvent suppression pulse sequence to saturate the residual 1H water proton signal (noesypr1d pulse sequence; relaxation delay = 2 s, τ1 = 3 µs, mixing time = 150 ms, solvent presaturation applied during the relaxation time and the mixing time). 128 transients were collected into 16 K data points over a spectral width of 12 ppm at 27°C.

**GC-MS.** Aqueous samples were derivatized using the procedure reported [63]. 150 µl of the D2O sample used for 1H NMR spectroscopy was evaporated to dryness in an evacuated centrifuge and 30 µl methoxyamine hydrochloride (20 mg ml−1 in pyridine) was added. The samples were vortex mixed for 1 minute, and derivatised at room temperature for 17 hours. Samples were then silylated with 30 µl of N-(tert-butyldimethylsilyl)-trifluoroacetamide (MSTFA) for 1 hour at room temperature. The derivatised samples were diluted (1:10) with hexane prior to GC-MS analysis. Organic phase metabolites were derivatised by acid catalyzed esterification [64]. Lipids were dissolved in 0.25 ml of chloroform/methanol (1:1 v/v), 0.10 ml BF3/methanol (Sigma-Aldrich) was added and the vials were incubated at 80°C for 90 minutes. Once cool 0.3 ml H2O (mQ) and 0.6 ml hexane was added and each vial vortex mixed for one minute. The aqueous layer was discarded and the remaining organic layer was evaporated to dryness before reconstitution in 200 µl hexane for analysis. The derivatised aqueous samples were injected into a Thermo Electron Trace GC Ultra equipped with a 30 m x 0.25 mm ID 5% phenyl polysilphenylene-siloxane column with a chemically bonded 25 µm TR-5MS stationary phase (Thermo Electron Corporation; Injector temperature = 220°C, helium carrier gas flow rate = 1.2 ml min−1). The initial column temperature was 70°C; this was held for 2 min then increased by 5°C min−1 to 310°C and was held for 20 min. The derivatised organic metabolites were injected onto a ZB-WAX column (30 m x 0.25 mm ID 0.25 µm df; 100% polyethylene glycol). The initial column temperature was 60°C; this was held for 2 min then increased by 10°C min−1 to 150°C and then by 4°C min−1 up to a temperature of 230°C where it was held for 7 min. The column eluent was introduced into a DSO quadrupole mass spectrometer (Thermo Electron Corporation) (transfer line temperature = 310°C for aqueous metabolites and 240°C for lipid metabolites, ion source temperature = 250°C, electron beam = 70 eV). The detector was turned on after a solvent delay of 240 s and data was collected in full scan mode using 3 scans s−1 across a mass range of 50–650 m/z.

**Data Analysis.** NMR spectra were processed using ACD SpecManager 1D NMR processor (version 8, ACD, Toronto, Canada). Spectra were Fourier transformed following multiplication by a line broadening of 1 Hz, and referenced to TSP at 0.0 ppm. Spectra were phased and baseline corrected manually. Each spectrum was integrated using 0.04 ppm. integral regions between 0.5–4.5, and 5.1–10.0 ppm. To account for any difference in concentration between samples each spectral region was normalised to a total integral value of 10000. GC-MS chromatograms were analysed using Xcalibur, (v. 2.0, Thermo Fisher Corp), integrating each peak individually. Peaks were normalised so that the total sum of peaks was set to 10000.
Table 2. Changes in metabolite levels measured in hearts from Python mice and littermate controls.1

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>wt ratio</th>
<th>Python</th>
<th>P value (t test)</th>
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</thead>
<tbody>
<tr>
<td><strong>Increased in Python</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aspartate</td>
<td>2.11</td>
<td>4.2x10^-9</td>
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</tr>
<tr>
<td>Glutamate</td>
<td>1.15</td>
<td>5.1x10^-4</td>
<td></td>
</tr>
<tr>
<td>Glycine</td>
<td>1.67</td>
<td>4.8x10^-15</td>
<td></td>
</tr>
<tr>
<td>Proline</td>
<td>1.24</td>
<td>1.4x10^-8</td>
<td></td>
</tr>
<tr>
<td>Valine/leucine/soleucine</td>
<td>1.69</td>
<td>6.3x10^-18</td>
<td></td>
</tr>
<tr>
<td><strong>Decreased in Python</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Choline</td>
<td>0.80</td>
<td>1.6x10^-4</td>
<td></td>
</tr>
<tr>
<td>Creatine</td>
<td>0.69</td>
<td>2.3x10^-1</td>
<td></td>
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<tr>
<td>Lactate</td>
<td>0.80</td>
<td>2.6x10^-5</td>
<td></td>
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<tr>
<td>Succinate</td>
<td>0.58</td>
<td>3.2x10^-8</td>
<td></td>
</tr>
<tr>
<td>Taurine</td>
<td>0.92</td>
<td>1.1x10^-3</td>
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<tr>
<td><strong>Gas chromatography-mass spectrometry</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Increased in Python</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Asparagine</td>
<td>1.62</td>
<td>2.3x10^-4</td>
<td></td>
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<tr>
<td>Glycine</td>
<td>1.57</td>
<td>1.3x10^-8</td>
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<tr>
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<tr>
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<tr>
<td>Uric acid</td>
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<tr>
<td>Decreased in Python</td>
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<td></td>
<td></td>
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<tr>
<td>α-glycerophosphoric acid</td>
<td>0.69</td>
<td>3.5x10^-7</td>
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<tr>
<td>Adenosine</td>
<td>0.37</td>
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<tr>
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<tr>
<td>Glucose</td>
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<td><strong>1H NMR spectra</strong></td>
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<tr>
<td>1H-Indole-2,3-dione</td>
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<tr>
<td>1H-Purine-6-amine</td>
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<td>Malate</td>
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<td>Myo-inositol</td>
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<td>Pyruvate</td>
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<tr>
<td>Ribose</td>
<td>0.40</td>
<td>1.4x10^-3</td>
<td></td>
</tr>
<tr>
<td>Succinate</td>
<td>0.54</td>
<td>5.9x10^-3</td>
<td></td>
</tr>
</tbody>
</table>

1 Mice were on a C3H/HeN genetic background. Average of individual measurements from each heart sample of mice aged 10–11 weeks (n = 10 Py/+; n = 10 C).

doi:10.1371/journal.pgen.1001000.t002

Isolation of embryonic fibroblasts

For skin fibroblasts, 2–4-day-old pups were humanely culled, and a portion of the skin removed, washed in PBS and finely minced using a razor blade. A small piece of tissue was retained for genotyping purposes. Numerous small segments of tissue approximately 1 mm² were placed well spaced on 10 cm Petri dishes and allowed to air dry for 10 minutes, DMEM containing Glutamax and 10% FCS (Invitrogen Ltd., Paisley, UK) was then carefully added to the dish so as to avoid dislodging the tissue pieces. Plates were cultured for 7 days at 37°C, 5% CO₂, then outgrowing cells were harvested with trypsin and passaged in the same medium. For embryonic fibroblasts, embryos were dissected from the uterus and the yolk sacs were removed for use in genotyping the embryos. The embryos were placed in 30 ml 0.05% trypsin and macerated using a 20 μl disposable pipette tip. After incubation for 15 minutes at 37°C, the cells were counted and plated in one well of a 6-well plate in DMEM containing Glutamax and 10% FCS and cultured at 37°C, 5% CO₂.
Labeling of mitochondria in fibroblasts

For microscopy, fibroblasts were cultured on glass coverslips in 50–200 nM MitoTracker Orange CM-H2TMRos in DMEM containing Glutamax and 10% FCS for 45 minutes at 37°C. Cells were washed in PBS, then fixed in 4% paraformaldehyde for 20 minutes at room temperature. Cover slips were mounted on glass slides in a drop of ProLong Gold mounting medium (Invitrogen Ltd., Paisley, UK) containing 10 μg/mL Hoechst 33258. For FACS analysis, cells were harvested by trypsinization, washed with PBS, then fixed for 20 minutes at 4°C in 4% paraformaldehyde. Cells were subsequently centrifuged and resuspended in PBS.

Western blot analysis

Brain and heart samples from 5-week-old mice were snap frozen in liquid nitrogen then homogenized in RIPA (50 mM Tris-HCl pH 8, 150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate, 0.1% SDS) containing 1× protease inhibitor cocktail (P8340; Sigma-Aldrich, Dorset, UK). Equal amounts of protein were separated by Laemmli stacking/separating SDS PAGE and blotted to Hybond-P (GE Healthcare, Little Chaelfont, UK), then individual proteins were detected using antibodies to Drp1 (Clone 8; BD Biosciences, Oxford, UK), Tim23 (Clone 32; BD Biosciences, Oxford, UK), and α-tubulin (T6074; Sigma-Aldrich, Dorset, UK). An anti-mouse HRP-conjugated antibody (7076; Cell Signaling, NEB, Hitchin, UK), was used for detection in combination with ECL Western Blotting Detection Reagents (GE Healthcare, Little Chaelfont, UK).

Immunocytochemistry

For peroxisome labeling, a primary rabbit anti-catalase IgG (ab16731, Abcam) was used with a goat anti-rabbit FITC-conjugated IgG (ab6717, Abcam). For examining Dnm1l distribution in fibroblasts, an anti-Drp1 primary antibody (Clone 8; BD Biosciences, Oxford, UK) with a Daylight549-conjugated anti-mouse Ig was used.

Fluorescent imaging

All images were taken on a Zeiss AxioCam MRc5 microscope using Axiovision Release 4.7.2 software. For imaging mitochondria with MitoTracker Orange and Dnm1l using a Daylight549-labelled secondary antibody, a filter with an excitation of 530–585 nm, emission 600–660 nm was used. For Imaging peroxisomes with a FITC-labeled secondary antibody, a filter with an excitation of 450–490 nm, emission 510–560 nm was used. For FACS analysis, cells were incubated in 200 nM MitoTracker Orange CM-H2TMRos in DMEM containing Glutamax and 10% FCS for 45 minutes at 37°C, harvested using trypsin, washed with PBS and fixed by incubation in 4% paraformaldehyde for 20 minutes at 4°C. Cells were washed in PBS and resuspended in PBS containing 1% FCS and analyzed for fluorescence on a CyAnADP O2 flow cytometer (Dako). The gate for MitoTracker Orange-positive cells was set using control cells that were not labelled.

ATP and total adenine nucleotide pool measurements

Hearts were excised, washed in heparinised normal saline, blotted and weighed, before being snap frozen in liquid nitrogen and stored at −80°C. Total adenine nucleotide (TAN) content and myocardial adenosine triphosphate (ATP) were measured by High Performance Liquid Chromatography (HPLC) as previously described [65]. For the purpose of calculating activity per mg of protein, the Lowry method was used.

Homology modelling

The structure of the human DNML protein (Uniprot [66] accession number O00429) was predicted by homology modelling as follows. The structure of the bacterial dynamin-like protein BDLP [13] (PDB [67] identification code 2w6d) was used as template. Given the low sequence identity (around 12%), the alignment between target sequence and template was performed using a profile-to-profile alignment method implemented in the FFAS03 server [68] (http://ffas.nlm.nih.gov). Profile-to-profile alignments are superior in terms of sensitivity and alignment quality compared to traditional pair wise alignments (and in particular at low sequence identity). The FFAS03 score for the alignment was −33.5 implying significant similarity (FFAS03 scores below −9.5 reported less than 3% false positive under benchmarking conditions [60]). The alignment was manually inspected and the structural model was derived using MODELLER [69] as previously described [70]. Structure representations were prepared using the molecular visualization program PyMOL (http://www.pymol.org).

Software and databases

DNA sequence alignments for detecting mutations were performed using DNASTAR (DNASTAR Inc. Madison, WI). Protein alignments were performed using CLUSTALW2 and ALIGN software available online from the European Bioinformatics Institute (www.ebi.ac.uk). Prism software (GraphPad Software, Inc., La Jolla, CA) was used throughout for statistical analysis. Morphometric analysis used ImageJ [71]. For measurement of mitochondrial area and size, contrast of micrographs was enhanced by 0.5%. After setting the scale, the threshold was altered such that only mitochondria were marked. The thresholded image was then analyzed for area, and area fraction for particles greater than 50 pixels. For measurement of area stained in MSB sections, the image type was set to RGB stack and the threshold set to detect areas positive for collagen. The thresholded image was then analyzed for area above threshold.

Supporting Information

Figure S1 Alignment of domain M in multiple species and mutations found in Dnm1l protein homologues that have an effect on mitochondrial morphology. Identi cal or functionally very similar amino acids are shown grouped together by colour on the basis of size and hydrophobicity. Reported mutations that result in amino acid substitutions are 1, G269R yeast [14]; 2, G385D yeast [14]; 3, G398S yeast [14]; 4, G369D hamster CHO cell line [16]; 5, A400D human [15]; 6, P444L yeast [14]; 7, C452F mouse (Python). Found at: doi:10.1371/journal.pgen.1001000.s001 (9.96 MB TIF).

Figure S2 ATP measurements (mean plus/minus SD) in heart and brain samples expressed as level relative to wild type mean, measured using a quantitative bioluminescent method. Tissue samples were minced and then digested for 45 minutes in 10 mM Tris pH 8.0 and 100 μg/ml proteinase K at 55°C. Protein concentrations were estimated using a Bio-Rad DC Protein Assay kit according to the manufacturers instructions. Samples were then normalized based on total protein. ATP was measured using an ATP Luminescence Assay Kit (Invitrogen Corp.) according to the manufacturers instructions. Found at: doi:10.1371/journal.pgen.1001000.s002 (0.71 MB TIF).

Table S1 Degree of similarity of domain M of mouse dynamin proteins.

Found at: doi:10.1371/journal.pgen.1001000.s003 (0.03 MB DOC).
Table S2  SHIRPA Assessment of Python mice.  

Found at: doi:10.1371/journal.pgen.1001000.s004 (0.04 MB DOC)

Acknowledgments

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Author Contributions

Conceived and designed the experiments: HA CAL JL PD JLG IH NFF MC TND. Performed the experiments: HA LD VL CT MN VS CAL JLG IH MC TND. Wrote the paper: HA HW TND. Conceived and designed the experiments: HA CAL JL PD JLG IH NFF MC TND. Performed the experiments: HA LD VL CT MN VS CAL JLG IH MC TND. Wrote the paper: HA HW TND.

References


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