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SUMOylation modulates the function of Aurora-B kinase

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Summary
Aurora kinases are central regulators of mitotic-spindle assembly, chromosome segregation and cytokinesis. Aurora B is a member of the chromosomal passenger complex (CPC) with crucial functions in regulation of the attachment of kinetochores to microtubules and in cytokinesis. We report here that Aurora B contains a conserved SUMO modification motif within its kinase domain. Aurora B can bind SUMO peptides in vitro when bound to the IN-box domain of its CPC partner INCENP. Mutation of Lys207 to arginine (Aurora BK207R) impairs the formation of conjugates of Aurora B and SUMO in vivo. Expression of the SUMO-null form of Aurora B results in abnormal chromosome segregation and cytokinesis failure and it is not able to rescue mitotic defects in Aurora-B-knockout cells. These defects are accompanied by increased levels of the CPC on chromosome arms and defective centromeric function, as detected in abnormal chromosome segregation and cytokinesis failure and it is not able to rescue mitotic defects in Aurora-B-knockout cells. These data suggest that SUMOylation of Aurora B modulates its function, possibly by mediating the extraction of CPC complexes from chromosome arms during prometaphase.

Key words: Aurora kinases, Cell cycle, Chromosome segregation, Cancer target, SUMO modifications

Introduction
Aurora B (serine/threonine-protein kinase 12) is the enzymatic core of a multiprotein complex, the chromosomal passenger complex (CPC), which comprises other three non-enzymatic subunits, INCENP, Survivin and Borealin (Carmena and Earnshaw, 2003; Ruchaud et al., 2007). The CPC is an important upstream regulator of centromere-kinetochore function, and is responsible for the recruitment to the kinetochore and centromere of a number of proteins, including inner centromere-kinetochore proteins (Sgo1, Sgo2, MCAK), regulators of the microtubule-kinetochore interaction (such as Hec1 and Plk1, among others) and proteins involved in the spindle-assembly checkpoint (SAC; such as Mad2, BubR1 and Mps1) (Kelly and Funabiki, 2009; Ruchaud et al., 2007). Some kinetochore components, including Hec1, Dam1, MCAK and CENP-A, are Aurora-B substrates, suggesting a crucial role for the CPC in the destabilization of aberrant microtubule-kinetochore attachments and in the SAC-dependent delay of mitotic progression until these defects are corrected. Recent data suggests that substrate phosphorylation depends on the distance of the substrate from Aurora B at the inner centromere, and artificial recruitment of the CPC to the kinetochore prevents the stabilization of improper attachments and activates the SAC to delay the metaphase-to-anaphase transition (Kelly and Funabiki, 2009; Liu et al., 2009). During cytokinesis, Aurora B is localized to the mid-body where its local inactivation is crucial for completion of abscission (Guse et al., 2005; Steigemann et al., 2009).

Activity of Aurora B is regulated through several mechanisms, including controlled interactions with INCENP, Survivin and Borealin. Knockdown of any member of the complex delocalizes the others, disrupts mitotic progression and might destabilize one or more subunits (Jeyaprakash et al., 2007; Ruchaud et al., 2007). In addition, several post-translational modifications are known to be required for proper activity and control of the dynamic behavior of the CPC. Activation of Aurora B is triggered by autophosphorylation of the T-loop residue after association with its substrate, INCENP. Full activation involves a positive-feedback loop after Aurora-B phosphorylation of INCENP in the conserved IN-box region, possibly as a result of structural rearrangements (Bishop and Schumacher, 2002; Honda et al., 2003).

CPC function is also modulated by ubiquitin and ubiquitin-related modifiers. Aurora B is targeted by a Cul3-containing SCF ubiquitin ligase during early mitosis (Sumara et al., 2007). Interestingly, this modification is necessary to remove a fraction of Aurora B from mitotic chromosomes, allowing its accumulation on the central spindle during anaphase. In the absence of this SCF-Cul3 ubiquitin ligase, Aurora B spreads along chromosome arms and fails to dissociate from them during anaphase. Aurora B also requires APC/C-Cdh1 activity to efficiently translocate to the spindle mid-zone in anaphase. In APC/C-Cdh1-depleted cells Aurora B and the CPC prematurely accumulate in the equatorial cortex, resulting in a weak anaphase spindle (Floyd et al., 2008). Finally, after completion of cytokinesis, the remaining pool of Aurora B is targeted for degradation by APC/C-Cdh1 (Floyd et al., 2008; García-Higuera et al., 2008). These post-translational modifications also affect other CPC components. For instance, Survivin is ubiquitylated during mitosis and its deubiquitylation is required for Survivin dynamics at centromeres (Vong et al., 2005).
Post-translational modifications by the covalent attachment of small ubiquitin-related modifiers (SUMOs) are important in several cellular processes, including the cell-division cycle (Geiss-Friedlander and Melchior, 2007; Hay, 2005; Ulrich, 2008). In vertebrates, at least three SUMO forms (SUMO1, SUMO2 and SUMO3) are covalently attached to a lysine residue in the target protein in three enzymatic steps analogous to the ubiquitination cascade. SUMO conjugation requires a SUMO-activating E1 enzyme (SAE1 or SAE2), the SUMO-conjugating E2 enzyme UBC9 and, in some cases, additional E3 SUMO ligases (Hay, 2005). SUMO modification is reversible by the action of SUMO-specific isopeptidases that belong to the family of ubiquitin-like proteases (ULPs), also known as sentrin-specific proteases (SENP) (Hay, 2007). Recent work has revealed the importance of SUMOylation for mitotic progression (Dasso, 2008; Watts, 2007) and has suggested a crucial role of SUMO modifications in kinetochore and centromere function (Dawlaty et al., 2008; Di Bacco et al., 2006; Mukhopadhyay et al., 2010; Zhang et al., 2008), including direct SUMOylation of the CPC component Borealin (Klein et al., 2009).

In this manuscript, we provide evidence for SUMOylation of Aurora B at Lys207. Mutation of this residue results in abnormal mitotic progression and the generation of multinucleated cells. In those mutant cells, Aurora B and INCENP display an abnormal localization at the chromosome arms during prometaphase and metaphase and defective CENP-A phosphorylation, indicating a role for Aurora-B SUMOylation in the proper localization and activity of the CPC during chromosome alignment.

**Results**

A conserved SUMOylation motif in Aurora-B proteins

Aurora B contains a consensus motif for SUMO modification (Ψ-Lys-x-Glu, in which Ψ is a hydrophobic residue and x, any residue) within the kinase domain (Fig. 1A). This motif is readily identified by using different protein- domain-recognition algorithms (see the Materials and Methods). The SUMO motif is conserved among Aurora-B orthologues from yeast to human, and it is also present in the other mammalian members of the Aurora family, but not in other cell cycle kinases, including Polo-like kinases (Fig. 1A).

To test the possible SUMOylation of this domain, we mutated the lysine (K207) predicted to accept the SUMO peptide into a charge-conservative arginine residue. The Aurora-B K207R mutation is predicted to impair covalent binding of SUMO with little effect on the overall kinase structure. Next, we expressed V5-epitope-tagged versions of wild-type (WT) Aurora-B and Aurora-B K207R mutant forms along with the E2 SUMO ligase UBC9 and HA-tagged versions of the three SUMO isoforms (HA-SUMO1 to HA-SUMO3; ~12 kDa) in HEK293 cells (Fig. 1B). The ectopically expressed proteins were immunoprecipitated using anti-V5 antibodies and detected with antibodies against the HA epitope. The predicted SUMO-Aurora-B conjugates (~50 kDa) were detected after expression of all three SUMO isoforms (Fig. 1B), suggesting that Aurora B can be SUMOylated by the three SUMO peptides in this assay. Importantly, the Aurora-B-SUMO conjugates were not observed in cells expressing the Aurora-B K207R isoform, indicating that K207 is required for covalent modification of Aurora B by SUMO residues.

We next tested SUMOylation of endogenous Aurora B by taking advantage of HeLa cells stably expressing histidine-tagged SUMO2 proteins (~16 kDa). These cells were transfected with V5-Aurora-B-expressing vectors and SUMO2 conjugates were exposed to antibodies against V5 or two different antibodies against Aurora B. SUMO2 conjugates were affinity purified under denaturing conditions with Ni-NTA beads. As depicted in Fig. 2A, all these three antibodies detected a band of ~60 kDa, and possibly additional higher-molecular-weight bands in SUMO2 conjugates. This band was detected in taxol-treated cells and it was barely visible in asynchronously growing cells or in parental cells that do not overexpress SUMO2 (data not shown). An additional band of ~55 kDa that might correspond to the endogenous Aurora-B-His(SUMO2) conjugates is detected by a monoclonal antibody against Aurora B. In fact, this band was also observed in similar assays in which the V5-Aurora-B vector was not used (data not shown), thus further suggesting modification of the endogenous protein.
We then used a reconstituted in vitro SUMO modification system to further test the molecular requirements for this SUMOylation. Recombinant Aurora B was incubated with SUMO E1-activating enzymes (SAE1/2), the E2-conjugating enzyme UBC9 and SUMO2 in the presence of ATP. A mutant SUMO2 molecule unable to conjugate to substrates was used as a control. As recently suggested (Klein et al., 2009), we did not detect obvious SUMOylation of Aurora B in this assay, whereas p53 was significantly SUMOylated by SUMO2, but not SUMO2 mutant peptides (Fig. 2B). Since Aurora B seems to be SUMOylated in vivo, we then asked whether additional CPC domains were required for this post-translational modification. Indeed, higher-molecular-weight Aurora B conjugates (Fig. 2B, arrows) were observed after incubation of Aurora B with the IN-box segment of INCENP, a domain that directly binds and activates Aurora B (Adams et al., 2000). INCENP-bound Aurora B was conjugated with SUMO2, but not with SUMO2 mutants, indicating the specificity of this signal. All together, these results suggest that Aurora-B K207 can be conjugated to SUMO residues and this modification requires binding of Aurora B to its activator INCENP.

**Defective SUMOylation of Aurora B results in abnormal chromosome segregation and reduced cell viability**

We transiently transfected HEK293 cells with GFP-tagged vectors for wild-type Aurora B, kinase-dead mutants (Aurora B\(^{K111M}\) and Aurora B\(^{D205A}\)) and the SUMO-null form (Aurora B\(^{K207R}\)) to understand the relevance of SUMOylation of Aurora B in cell-cycle progression. Overexpression of wild-type Aurora B slightly increased the percentage of 4N and >4N cells, which is consistent with a mitotic delay and defects in completion of cytokinesis. These defects were more pronounced upon expression of the kinase-dead form Aurora B\(^{K111M}\) (19.1% of >4N) and, at an even higher degree with Aurora B\(^{D205A}\) (47.6% of >4N) (Fig. 3A,B). The phenotypic differences observed with the Aurora-B\(^{K111M}\) and Aurora-B\(^{D205A}\) mutants suggest distinct degrees of inhibition of the kinase activity in these mutants (see below). Interestingly, overexpression of the SUMO-null form (Aurora B\(^{K207R}\)) induces polyplody at the same level as the Aurora-B\(^{D205A}\) mutant (44.1% in Aurora-B\(^{K207R}\) mutants vs 47.6% in Aurora-B\(^{D205A}\) mutants). Microscopy analysis indicated that >4N cells overexpressing the Aurora-B\(^{K207R}\) mutant were mostly multinucleated and displayed severe nuclear abnormalities (Fig. 3C), which were similar to those of kinase-dead mutants (not shown). Stable expression of Aurora B\(^{D205A}\) and Aurora B\(^{K207R}\) in U2OS cells severely reduced the number of colonies, suggesting that mutations in these residues impair Aurora-B function in a way that is incompatible with cell proliferation (Fig. 3D,E).

**Altered mitotic progression and cytokinesis in Aurora-B\(^{K207R}\) mutants**

Degregation of the function of Aurora B by chemical inhibition, small interfering RNA (siRNA) or overexpression of the kinase-dead form all lead to similar phenotypes, including chromosome misalignment and failure to complete cytokinesis, resulting in multinucleated cells (Ruchaud et al., 2007). To gain further insights into the cellular consequences of defective Aurora-B SUMOylation, we analyzed stable HeLa cell lines expressing the different mutant constructs. As reported above, proliferation of clones stably expressing GFP-Aurora-B fusion proteins was relatively normal, whereas expression of the kinase-dead and SUMO-null forms resulted in reduced viability. Accumulation of the GFP-Aurora-B fusion protein was mostly observed in G2-M cells (Fig. 4A) in agreement with its role at these cell-cycle stages. The Aurora-B mutants exhibited a slightly lower degree with Aurora B\(^{D205A}\) and Aurora B\(^{K207R}\) respectively, vs 5% in GFP-Aurora-B\(^{WT}\)). Strikingly, the accumulation of these mutant proteins was not restricted to G2-M. This altered behaviour of the mutants might be explained by a higher stability of the mutant proteins or by the existence of aneuploid cells with a DNA content other than 4N.
We then depleted endogenous Aurora B by siRNA in the stable clones expressing GFP-Aurora-B fusion proteins. The mouse Aurkb cDNA, encoding Aurora B, was used to generate these constructs and they were therefore resistant to siRNA treatment directed against the endogenous human transcript (Fig. 4B). The wild-type GFP-Aurora-B construct efficiently rescued the formation of multinucleated cells after depletion of endogenous Aurora B proteins and these rescued cells progressed normally through mitosis (Fig. 4C). However, the expression of Aurora-B^D205A and Aurora-B^K207R mutants failed to rescue the defects caused by knockdown of the endogenous protein. These cultures delayed in prometaphase with unaligned chromosomes, failure to execute cytokinesis and accumulation of multinucleated cells (Fig. 4C,D). Similar results were observed after chemical inhibition of Aurora-B kinase activity with ZM447439 (ZM1; Fig. 4C,D). Quantification of the distribution of mitotic phases in these stable cell lines after depletion of endogenous Aurora B confirmed that expression of GFP-Aurora-B^D205A and GFP-Aurora-B^K207R provokes a similar arrest at prometaphase and a reduction of cells in telophase (Fig. 4E). This was accompanied by a decrease in the percentage of cells in cytokinesis. Furthermore, those cells that were undergoing cytokinesis in cultures expressing Aurora-B^D205A and Aurora-B^K207R were mostly aberrant and frequently displayed chromosomal bridges between the two daughter cells (Fig. 4F).

**Altered chromosomal dynamics of the CPC in Aurora-B^K207R mutants**

Endogenous Aurora B concentrates in the centromeric region of chromosomes during prometaphase, similarly to other CPC components such as INCENP. In the absence of endogenous Aurora B, both INCENP and exogenous wild-type GFP-Aurora-B localized properly to these centromeric structures, consistent with the functional rescue by this wild-type protein (Fig. 5A,B). Interestingly, the Aurora B^K207R mutant (as well as the kinase-dead isofrom) was dispersed along the entire chromosome arms in prometaphase (PM) and metaphase (M) cells. INCENP also failed to concentrate in inner centromeres, suggesting that there are CPC localization defects upon lack of Aurora B SUMOylation. Further analysis of the distribution of Aurora B using Metamorph algorithms confirmed that Aurora-B^D205A and Aurora-B^K207R were dispersed along the chromosome arms (Fig. 5C-E). Quantification of the integrated optical density (OD), a measure of the opacity of an object when exposed to light, showed that the chromosomal staining of the SUMO-null Aurora B mutant is significantly (P<0.001) less opaque than Aurora-B^WT staining. This means that Aurora B^K207R does not concentrate at any chromosomal region (Fig. 5E). These defects were reduced in the kinase-dead mutant and did not reach statistical significance in this assay. We also tested whether Aurora-B kinase activity was important for proper localization of Aurora B in the centromere, by treating GFP-Aurora-B^WT cells with the ZM1 inhibitor. In agreement with previous reports (Ditchfield et al., 2003; Xu et al., 2009), chemical inhibition of Aurora B kinase activity did not perturb localization of Aurora B or INCENP to the centromere, indicating that an active kinase is not required for their centromeric localization (Fig. 5F).

Since the presence of endogenous Aurora B might interfere with the localization of exogenous Aurora-B^K207R mutants, we decided to further study these mutants in Aurora-B-knockout cells. These cells derive from Aurora-B conditional-knockout mice in which exons 2 to 6 are flanked by loxP sites [Aurkb(lox) allele; G.F.-M., I.P.C. and M.M., unpublished results]. Upon expression of the Cre recombinase (Fig. 6A), exons 2 to 6 are excised, resulting in a null allele [Aurkb(ΔΔ)]. Aurkb(ΔΔ) MEFs progress normally through prophase and prometaphase. However, whereas 65% of mitotic cells (n=40 of each genotype) were in metaphase, anaphase or telophase in Aurkb(lox/lox) cultures, these figures were reduced to 8% in Aurkb(ΔΔ) cells (most of these anaphases and telophases showed immunoreactivity against Aurora B, probably as a consequence of inefficient gene deletion; data not shown). Aurkb(ΔΔ) prometaphases displayed several misaligned
chromosomes and abnormal spindles (Fig. 6B). These defects could be rescued by infection with retroviruses expressing wild-type Aurora B, and *Aurkb*(ΔΔ); Aurora B*WT* cells displayed normal mitotic progression (about 60% mitotic cells were in metaphase, anaphase or telophase) and resulted in normal cytokinesis. In these *Aurkb*(ΔΔ) MEFs, the exogenous Aurora B was properly located at centromeres and migrated to the midzone during anaphase (Fig. 6B). By contrast, *Aurkb*(ΔΔ); Aurora B*K207R* cells displayed abnormal spindles and misaligned chromosomes and did not progress to anaphase (about 10% of mitotic figures were in metaphase, anaphase or telophase). In these cells, the exogenous Aurora B*K207R* displayed an abnormal centromeric concentration and remained diffuse at chromosome arms (Fig. 6B).

Fluorescence recovery after photobleaching (FRAP) was then used to compare the relative mobility of Aurora-B isoforms at metaphase chromosomes and the mid-body. Wild-type Aurora B showed a more dynamic behavior in the chromosomes located at the metaphase plate than the other mutant isoforms (Fig. 7). Although the recovery half-life (*t*1/2) was 35.6±10.2 seconds for wild-type Aurora B (*n*=7), the kinase-dead mutant Aurora B*BD205A* displayed a higher value (40.8±17.4 seconds; *n*=4), as previously reported in similar assays (Murata-Hori and Wang, 2002). A significantly higher *t*1/2 value (119.9±36.8 seconds; *n*=4) was scored in the Aurora-B*K207R* mutant, suggesting defective dynamics in the SUMO mutant protein. A similar recovery pattern for all the Aurora B isoforms was observed at the mid-body (*t*1/2=44.00±30.5 seconds in wild-type Aurora B (n=4); *t*1/2=38.9±7.4 seconds in Aurora B*KD205A* (n=3); *t*1/2=22.5±12.9 seconds in Aurora B*K207R* (n=4)] (Fig. 7B,C) indicating a specific chromosomal defect in the dynamics of Aurora B following mutation of the SUMOylation motif.

All together, these data indicate that Aurora B*K207R* is functionally defective and suggest that SUMOylation at K207 is crucial to regulate Aurora-B and INCENP dynamics at chromosomes.

**The Aurora B*K207R* mutant displays no compromised kinase activity**

The differences in localization described above suggest that the molecular defects caused by the kinase-dead and SUMO-null mutations of Aurora B are not completely identical. We therefore tested the effects of these two mutations on kinase activity. Staining with phospho-specific antibodies against CENP-A, a known Aurora-B substrate, reveals that phosphorylation of CENP-A was significantly decreased in cells expressing either Aurora B*KD205A* or Aurora B*K207R* (Fig. 8A). By contrast, GFP-Aurora-B*WT*, which was concentrated at centromeres, efficiently phosphorylated CENP-A, resulting in positive staining for phospho-CENP-A at kinetochores flanking the GFP-Aurora-B*WT* signal (Fig. 8A). We next directly tested the kinase activity of V5-tagged Aurora-B proteins immunoprecipitated from transiently transfected HEK293 cells (Fig. 8B). Aurora B*K111M* and Aurora B*KD205A* failed to efficiently phosphorylate the substrate MBP in vitro, although Aurora B*K111M* still displayed some kinase activity,
in agreement with the previous phenotypic analysis of these kinase-dead mutants.

Strikingly, the kinase activity of the Aurora B-K207R mutant was not reduced at all and indeed it was slightly increased when compared with the activity of the wild-type protein. Similar results were obtained when a similar SUMO-defective mutant (K141R) of the highly related kinase Aurora C was used (Fig. 8C). To further corroborate whether lack of SUMOylation modulates the intrinsic kinase activity of these proteins, we performed similar kinase assays in cells transfected with Aurora B, SUMO2 and wild-type or dominant-negative forms of UBC9. Interestingly, promoting Aurora-B SUMOylation by overexpressing SUMO2 and UBC9 resulted in a slight overall decrease in Aurora-B activity, whereas preventing SUMOylation by expressing a dominant-negative form of UBC9 led to increased Aurora-B kinase activity (Fig. 8D). Finally, we also tested kinase activity in human recombinant proteins produced in baculovirus. As shown in Fig. 8E, the recombinant Aurora-B-K207R mutant (corresponding to mouse K207) was also active and displayed a slight increase in kinase activity. Whether this increase represents differences in the intrinsic kinase properties of the K202,K207 mutants, or is a consequence of impaired SUMOylation of the wild-type molecule in mammalian or insect cells is not clear at present. In any case, these results indicate that the functional defects in the Aurora-B-K207R mutant are not a consequence of decreased kinase activity, but rather suggest that SUMOylation modulates Aurora-B function by impairing its proper removal from chromosome arms.
Discussion

Global mapping of the SUMO network by protein-protein interactions and genetic-network analysis has linked SUMOylation to many different biological processes, including chromosome segregation and the cell cycle (Makhnevych et al., 2009). In recent years, the SUMO pathway has been implicated in several aspects of mitosis, including chromosome structure, cell-cycle progression, kinetochore function and cytokinesis (Dasso, 2008; Watts, 2007).

In mammals, cells depleted of the SUMO E2-conjugating enzyme UBC9 show major deficiencies in chromosome condensation and segregation, along with severely altered nuclear morphology (Hayashi et al., 2002; Nacerddine et al., 2005), revealing the importance of this post-translational modification for mitotic progression. The CPC has been widely proposed to be regulated by SUMOylation. The yeast CPC component Bir1 (Survivin) is known to be modified by SUMO (Montpetit et al., 2006) and Sli15 (INCENP) is also a candidate for SUMO modifications in this organism (Wohlschlegel et al., 2004). Similarly, Borealin is regulated by binding to SUMO2 and SUMO3 in human cells (Klein et al., 2009).

In this work, we have identified a SUMO target site in mammalian Aurora B and analyzed the consequences of lack of SUMOylation in Aurora-B regulation and function. The SUMOylation site is highly conserved among Aurora B orthologs and in the other members of the Aurora family, Aurora A and Aurora C. Our data indicate that Aurora B can be SUMOylated in vivo by the three SUMO isoforms at Lys207 (K207). According to observations in Xenopus egg extracts and human cells, SUMO2 and SUMO3 are found at centromeres and chromatin during prometaphase and metaphase, whereas SUMO1 localizes to the mitotic spindle and the spindle mid-zone during these stages.
negative controls of MBP phosphorylation. Quantification of [32P]MBP signal increased. Aurora-B kinase-dead mutants K111M and D205A were included as dominant-negative UBC9 form (C93S mutant). Whereas the overexpression of Aurora-B in green and DAPI (DNA) in blue. The inset represents proper localization of SUMO-null Aurora B, despite exhibiting normal kinase activity in vitro, fails to phosphorylate CENP-A in vivo. Thus, the lack of SUMOylation of Aurora B seems to affect a small fraction of the Aurora B pool, although it is not clear at present whether this is a consequence of the transient nature of SUMOylation or technical problems owing to strong demodifying activity in mammalian protein extracts (Geiss-Friedlander and Melchior, 2007). Rescue experiments performed with the Aurora-B SUMO-deficient form (Aurora B<sup>B<sub>207R</sub></sup>) demonstrates that both Aurora B and INCENP fail to concentrate in inner centromeres during prometaphase or metaphase, thus suggesting that lack of Aurora-B SUMOylation affects the localization of other CPC components. Other examples of mitotic proteins whose localization at kinetochores or centromeres is regulated by SUMO include the kinesin CENP-E (Zhang et al., 2008), the CENP-H/I/K kinetochore complex (Mukhopadhyay et al., 2010) and the GTPase-activating protein RanGAP1 (Joseph et al., 2002). It has been recently confirmed that phosphorylation of Aurora-B substrates is influenced by the relative locations of the substrate and kinase at the inner centromere (Liu et al., 2009). This mechanism offers an efficient mechanism to selectively destabilize improper microtubule-kinetochore attachments (Kelly and Funabiki, 2009). As described in the present manuscript, the SUMO-deficient form of Aurora B, despite exhibiting normal kinase activity in vitro, fails to phosphorylate CENP-A in vivo. Thus, the lack of dynamic localization of SUMO-null Aurora B might impair its interaction with its substrates, resulting in failure to phosphorylate them in vivo but not in vitro.

SUMOylation of Aurora B might also directly modulate the intrinsic kinase activity. Aurora-B D205 is crucial for kinase function and it is only two residues upstream from the SUMO-binding site (K207). Indeed, both residues are in the same structural loop (Fig. 9B). Rescue experiments with a kinase-dead mutant (Aurora B<sup>D205A</sup>) result in phenotypes that are similar to those obtained with the SUMO-null (Aurora B<sup>B<sub>207R</sub></sup>) mutant. This led us...
SUMOylation of Aurora B

Materials and Methods

Sequence analysis and gene constructs
For protein-domain analysis, we first used the ELM (Eukaryotic Linear Motif, http://elm.eu.org) algorithm. To further confirm the presence of SUMO motifs we used the specific SUMO prediction software SUMOplot™ Analysis Program (http://www.abgent.com/tools/SUMOplot). Sequence alignments were obtained with the Clustal software (http://www.ebi.ac.uk/Tools/clustalw2/index.html). Mouse Aurkb cDNA was amplified by PCR from the plasmid pAA0155 (Mammalian Gene Collection), cloned into pENTR-D-TOPO vector using TOPO technology (Invitrogen) and transferred to a destiny vector coexpressing GFP or V5 tag by a LR recombination reaction of the Gateway system (Invitrogen). Aurora-B mutants (Aurora BK207R, Aurora B D205A and Aurora B K207R) were prepared using the Quik-Change site-directed mutagenesis kit (Stratagene) and were verified by DNA sequencing. Plasmids expressing histidine- or HA-tagged SUMO1, SUMO2 and SUMO3 as well as wild-

unknown pathway that recognizes the SUMO residues (Fig. 9C). In the Aurora-B\textsuperscript{K207R} mutant, the Aurora-B kinase might be hyperactive, as a result of the lack of SUMO residues, but the CPC is diffuse over the chromatin, resulting in deficient localized phosphorylation of its substrates.

Several lines of evidence link SUMO modifications with ubiquitin-mediated processes (Geoffroy and Hay, 2009). Recent studies have shown that ubiquitylation of Aurora B by the Cul3-Klhl9/Klhl13 E3-ligase complex during early mitosis might serve as a signal for its removal from chromosomes (Sumara et al., 2007). The AAA-ATPase p97 complex specifically interacts with ubiquitylated Aurora B and extracts it from chromosomes, leading to inactivation of the kinase on chromatin (Ramadan et al., 2007). In the absence of Cul3-Klhl9/Klhl13 activity, Aurora B is not ubiquitylated and accumulates on chromosomes (Sumara et al., 2007). The fission yeast Aurora kinase (Ark1p) interacts with two RING-finger proteins that possess SUMO-interacting motifs (SIMs), called Rfp1p and Rfp2p (Sun et al., 2007). These E3 ubiquitin ligases recognize SUMOylated proteins and heterodimerize with Slx8, another RING-finger protein, to form a functional ubiquitin ligase that is functionally similar to vertebrate RNF4 (Geoffroy and Hay, 2009; Sun et al., 2007; Uzunova et al., 2007). It is therefore possible that, in mammals, SUMOylation of Aurora B facilitates the formation of a functional Cul3-Klhl9/Klhl13 ligase complex during mitosis. Indeed, we have found three predicted SIMs in each coadaptor, Klhl9 and Klhl13, and one additional SIM in the Cul3 ligase that are conserved in human and mouse in all cases (data not shown). It is also possible that SUMOylation of Aurora B modulates its ubiquitylation by the APC/C-Cdh1. However, Aurora B does not accumulate on chromosomes upon RNA interference to knock down Cdh1 (Floyd et al., 2008), suggesting that these two pathways are not linked. Future experiments will be necessary to test and validate these hypotheses.

In summary, our data suggest that Aurora-B function is modulated by SUMOylation at K207. This modification is likely to interfere with Aurora-B kinase activity and it might participate in the extraction of the CPC from chromatin during prometaphase and the metaphase-to-anaphase transition. The effects of SUMOylation on the dynamic localization of Aurora B might be linked to ubiquitylation by the Cul3-Klhl9/Klhl13 ligase, because prevention of either SUMOylation or ubiquitylation by this complex results in a similar phenotype where the CPC is delocalized on chromosome arms. The regulation of Aurora B by SUMOylation might also have relevant implications, not only in preventing genomic instability, but also in tumorigenesis and cancer therapy (Girdler et al., 2006; Keen and Taylor, 2004; Mo and Moschos, 2005; Perez de Castro et al., 2007; Perez de Castro et al., 2008).
type UBC9 or the UBC9 C93S mutant were kindly provided by Ron Hay (Wellcome Trust Centre for Gene Regulation and Expression, University of Dundee, UK).

**Cell culture, transfection and cell-cycle analysis**

HEK293, U2OS and HeLa human cells were maintained in DMEM medium supplemented with 10% fetal bovine serum and antibiotics and were grown at 37°C in a humidified 5% CO₂ atmosphere. The generation of conditional alleles at the mouse Auklloc locus will be described elsewhere. Mouse embryonic fibroblasts were obtained from E14.5 Auklloc(lox/lox) embryos and cultured using routine protocols (Garcia-Higuera et al., 2008). Human Aurora B was silenced using validated siRNA oligos from Qiagen with the following sequence: 5'-AACCACGGCCTTACCA-ATGTTA-3'. HEK293, U2OS and HeLa cells were transfected with 1 μg plasmid DNA using Effectene transfection reagent (Qiagen) according to the manufacturer’s instructions. For silencing experiments, siRNA oligos were nucleofected in HeLa cells using Amaxa kit and the conditions provided by Amaxa (113 program. Cell-cycle distribution was determined by flow cytometry after DNA staining with Propidium Iodide (Sigma) incubated with purified E1- activating enzyme (Sigma AL2), E2- conjugating enzyme (UBC9) and either SUMO2 or mutant SUMO2 protein (harbouring a proprietary mutation that impedes SUMO conjugation) in the presence of ATP and following the manufacturer’s instructions (Active Motif).

**Expression and analysis of recombinant proteins**

Recombinant baculovirus-expressing human His-Aurora B WT, His-Aurora B- K106R and His-Aurora B- K207R were generated using the pFastBacHTA vector and the Bac-to-Bac Expression System (Invitrogen), as described previously (Gassmann et al., 2004). For in vitro SUMOylation assays, His-Aurora B WT or a complex of His-Aurora B WT and the IN-box fragment (human INCENP amino acids 821 to 1117) were incubated with purified E1-activating enzyme (Sigma AL2), E2-conjugating enzyme (UBC9) and either SUMO2 or mutant SUMO2 protein (harbouring a proprietary mutation that impedes SUMO conjugation) in the presence of ATP and following the manufacturer’s instructions (Active Motif).

**Immunofluorescence and FRAP analysis**

For immunofluorescence, cells were fixed with 4% PFA and permeabilized with 0.15% Triton X-100. These cells were then blocked with 3% BSA and incubated with the primary antibodies against the following proteins: α-tubulin (Sigma), Aurora B (Abcam, BD Transduction), INCENP (Human GLPL serum, a gift from Manuel Valdivia), phospho-Centromeric Protein H3 (Cytocell, Spain), UPF1 (Upstate Biotechnology). The matching secondary antibodies, with different Alexa Fluor dyes (488, 594, 647), are from Molecular Probes (Invitrogen). Images were obtained using a confocal ultramicroscope (Leica TCS-SP5) or an Olympus IX70 microscope controlled by Delta Vision SoftWorx (Applied Precision, Issaquah, WA). Image stacks were deconvolved, quick-projected and saved as tiff images to be processed using Adobe Photoshop.

Fluorescence recovery after photobleaching (FRAP) was performed on HeLa stable cell lines expressing GFP-Aurora-B wild-type and mutants (D205A and K207R). All experiments were conducted using a FRAP-enabled DeltaVision. Images of the centromeres were captured at 20-30 second intervals. Intensity values were normalized using the following equation: \(I(t) = I(0)\times\left(1 - \frac{I_t}{I_0}\right)\), where \(I(t)\) is the intensity normalized at time \(t\), \(I_0\) is the actual intensity at time \(t\), \(I_t\) is the intensity immediately following the photobleach (when \(t\) is equal to zero), and \(z\) is the intensity at the final time point. This sets the initial post-bleach intensity (at \(t=0\)) to zero and the final intensity to 1 arbitrary units. Regression analysis was performed using the GraphPad Prism software by calculating the least squares fit using the standard first order logarithmic equation. The \(t_1/2\) value was the time at which the normalized intensity reaches 0.5 arbitrary units.

**Protein extraction and analysis**

For immunodetection in protein lysates, cells were washed twice with ice-cold PBS and lysed in RIPA lysis buffer (37 mM NaCl, 0.5% NP-40, 0.1% SDS, 1% Triton X-100, 20 mM Tris-HCl, pH 7.4, 2 mM EDTA, 10% glycerol 1 mM PMSF supplemented with protease and phosphatase inhibitor cocktails (Sigma). Additionally, for detecting SUMOylated proteins, an inhibitor of SUMO proteases, N-ethylmaleimide (NEM), was added at 10 mM final concentration. After 30 minutes on ice, samples were cleared by centrifugation. Proteins were separated on SDS-PAGE, transferred to nitrocellulose membranes (Bio-Rad), probed using specific antibodies and detected using goat anti-rabbit or anti-mouse secondary antibodies (D scrap for ECL (Perkin Elmer) detection. After transfer of the protein lysates, we probed nitrocellulose membranes with primary antibodies against Aurora B (BD Transduction and Abcam), INCENP (Abcam), SUMO2/3 (MBL), Cyclin B1 (Santa Cruz Biotechnology), p53 (Active Motif), HA tag (Abcam), V5 tag (Invitrogen), His tag (Roche) and α-tubulin (Sigma).

**Immunoprecipitation and in vitro kinase assays**

Total protein lysates extracted from HEK293 cells transiently transfected with V5-tagged Aurora-B vectors were preclarified with protein-G agarose bead suspension for 1 hour (Amersham). Supernatants were first incubated for 2 hours at 4°C on a rotating wheel with mouse anti-V5 (Invitrogen) and later with 50 μl of the blocked beads. Immunoprecipitation for an additional hour. Immunoprecipitates were then washed three times in RIPA buffer and, in case of the kinase assay, one extra wash was performed with RIPA plus 0.5 M NaCl to remove possible bound unspecific kinases. One-fourth of the immunoprecipitates bound to the beads were used for V5 detection by western blot. For the kinase assay, four-fifths of the beads from the previous immunoprecipitations or recombinant proteins (Millipore) were washed in kinase assay buffer [10 mM HEPES-NaOH (pH 7.4), 150 mM NaCl, 10 mM MgCl₂, and 1 mM EGTA] supplemented with 0.5 mM DTT and phosphatase inhibitor cocktail (Calbiochem) and later incubated in 25 μl of 2× kinase buffer supplemented with 0.2 mM ATP, 2.5 μCi [γ-32P]ATP and 3 μg MBP (Sigma) or histone H3 (New England Biolabs) as an Aurora substrate. The reactions were incubated for 30 minutes at 30°C, stopped by addition of 5 μl of 1× Laemmli sample buffer, separated by 12.5 % SDS-PAGE and analyzed by autoradiography.

**References**


