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SIWUS1; An X-linked Gene Having No Homologous Y-Linked Copy in Silene latifolia

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ABSTRACT The dioecious plant Silene latifolia has heteromorphic sex chromosomes, and comparison of the positions of sex-linked genes indicates that at least three large inversions have occurred during the evolution of the Y chromosome. In this article, we describe the isolation of a new sex-linked gene from S. latifolia, which provides new information on the evolution of this plant’s young sex chromosomes. By using reverse-transcription polymerase chain reaction degenerate primers based on the Arabidopsis thaliana sequence of WUSCHEL, a flower-development gene, we found two copies in S. latifolia, which we named SIWUS1 and SIWUS2. Southern blot and genetic segregation analysis showed that SIWUS1 is located on the X chromosome and SIWUS2 is autosomal. No Y-linked copy of SIWUS1 was found by either Southern blot analysis under low-stringency conditions or polymerase chain reaction with degenerate primers, so we conclude that SIWUS1 probably has no Y-linked homolog. It is unknown whether the Y chromosome lost the SIWUS1 copy by degeneration of this individual gene or whether deletion of a larger genome region was involved. Several tests lead us to conclude that dosage compensation has not evolved for this sex-linked gene. We mapped the ortholog in the nondioecious relative S. vulgaris (SvWUS1), to compare the location in a species that has no history of having sex chromosomes. SvWUS1 maps to the same linkage group as other fully X-linked genes, indicating that it was not added to the X, but was lost from the Y. Its location differs in the maps from the two species, raising the possibility that the X chromosome, as well as the Y, may have been rearranged.

Sex chromosomes have evolved independently in different groups in both the animal and plant kingdoms; however, they share common evolutionary features (Charlesworth 1996). Sex chromosome evolution probably began with the establishment of male-sterility and female-sterility mutations in a region of a pair of autosomes, followed by recombination becoming suppressed in the region, which led to degeneration, differentiation, and extension of male-specific chromosomal regions in evolving XY systems (and female-specific regions inZW systems). Compared with many animal sex chromosomes, some flowering plant sex chromosomes have evolved recently (Parker 1990), providing opportunities to study the early stages of sex chromosome evolution (for a review, see Jamilena et al. 2008). Genetic degeneration is so far not well characterized in plants, and it is difficult to estimate the extent of gene loss from the male-specific region of plant Y chromosomes or to discover the causes of such losses when they occur.

The sex chromosomes of Spinacia oleracea and Asparagus officinalis evolved very recently, and recombination suppression has either
not occurred, or is confined to regions too small to be detected cyto-
logically (Telgmann-Rauber et al. 2007; Onodera et al. 2011). The sex
chromosomes of Carica papaya are probably further evolved in Y-
chromosome evolution as the YY genotype is inviable, suggesting that
genetic degeneration has started (Liu et al. 2004) even though the male-
specific region of its “Y chromosome” is only 8.4 Mb, or approximately
15% of the chromosome (Ming et al. 2008; Na et al. 2012). In Silene
latifolia, the Y chromosome is 50% larger than the X. The strongly
heteromorphic sex chromosomes of S. latifolia are likely to have evolved
long enough ago for some degeneration to have occurred, and, like
papaya, the YY genotype is inviable (Westergaard 1958; Janousek
et al. 1998), so this species may therefore be suitable for studying de-
generation. Recent studies of expression levels show that Y-linked genes
tend to have lower expression than their X-linked counterparts and also
undergo more (presumably deleterious) nonsynonymous substitutions
(Bergero and Charlesworth 2011; Chibalina and Filatov 2011), but the
ascertainment method used in those studies, based largely on detecting
Y-linked variants in cDNA sequences, could not accurately estimate
gene losses from the Y, and the “X-only” genes inferred have not yet
been shown to have been completely lost from the Y.

The structure of S. latifolia sex chromosomes has been character-
ized using a variety of approaches, including fluorescence in situ hy-
bridization to map repetitive sequences on the sex chromosomes and
reveal different repetitive sequence compositions of the X and Y chro-
mosomes (for reviews, see Kazama and Matsunaga 2008; Keijnsovy
et al. 2009). For example, sequences of the XA3.1 repetitive family are
located at both ends of the X chromosome but only one end of the Y
(Buzek et al. 1997; Matsunaga et al. 1999). Chloroplast sequences have
detectably accumulated on the Y chromosome (Keijnsovy et al. 2006).
Moreover, fluorescence in situ hybridization analyses changed the view
of the position of pseudoautosomal region of the X chromosome. The
pseudoautosomal region of the X chromosome was first characterized
as being on the q-arm (Westergaard 1946) but is now known to be
located on the p-arm (Lengerova et al. 2003; Kazama et al. 2006;
Kazama and Matsunaga 2008).

Another approach to analyzing the sex chromosomes is isolation of
sex-linked genes (Guttman and Charlesworth 1998, Delichere et al.
1999; Atanassov et al. 2001; Moore et al. 2003; Matsunaga et al. 2003;
Filatov 2004; Bergero et al. 2007; Kaiser et al. 2009, 2011). Although this
can potentially detect genes lost from the Y, most of these gene have
both X and Y copies; the one exception for this clade is SlAP3 (Kazama
et al. 2009). X-linked genes have been mapped (Filatov 2005; Kaiser et al.
2009; Nicolas et al. 2005), and their chromosomal arm locations are deter-
mined by a combination of laser microdissection and polymerase chain
reaction [PCR (Hobza et al. 2007)]. Y-linked genes have been physically
mapped using Y chromosome deletion mutants, allowing comparisons of
the gene order on the X and Y (Lebel-Hardenack et al. 2002; Zluvova
et al. 2007; Ishii et al. 2008; Bergero et al. 2008). At least three large
inversions are proposed to have occurred in the S. latifolia Y chro-
mosome during sex chromosome evolution (Keijnsovy and Vyskot 2010).

The isolation of more sex-linked genes will help toward an under-
standing of the mechanism of sex chromosome degeneration. Floral
development differences between S. latifolia male and female
flowers (Grant et al. 1994), and sex-specific expression of some flower
development genes (Zluvova et al. 2006; Kazama et al. 2009), could be
due to involvement of genes in the sex determination mechanism, or
could be downstream consequences of sex determination. Identifying
genes involved in the primary steps in the control of male or female
flower development is difficult because these genes must be located in
the nonrecombining part of the Y chromosome, so that genetic anal-
dates for such genes is to clone homologs of such genes from other
species and test them for sex-linkage. Indeed, the SlAP3 gene, a ho-
molog of APETALA3, has copies on the S. latifolia X and Y chromo-
somes (Cegan et al. 2010; Nishiyama et al. 2010).

Here, we identify the S. latifolia homolog, SIWUS1, of the flower
development gene WUSCHEL (WUS), and demonstrate that SIWUS1
is sex-linked. In flowering plants, floral organ identity is controlled by
the expression and inter-regulation of the ABC genes (Coen and
Meyerowitz 1991; Weigel and Meyerowitz 1994) and several other genes
(Sabolowsky 2007). One non-ABC gene, WUS, is inferred to be
involved in the up-regulation of C-class genes because the size of whorl
four is reduced in the A. thaliana was mutant (Laux et al. 1996). The
WUS gene was the first identified gene in the Wuschel-related homeo-
box (WOX) family (Laux et al. 1996; Haecker et al. 2004). The set of
WOX family genes are classified into three separate clades correspond-
ning with different protein functions: the WUS clade, an intermediate
clad (sometimes called the WOX clade; the terminology we use in
Figure 2), and an ancient clade (van der Graaff et al. 2009). In Arabi-
dopsis thaliana, proteins of the WUS clade contain a WUS box
(TLPLFPMH), and eight genes encoding WOX family proteins have
been identified in the genome of this species, including WUS. Three
WOX family members, WUS, WOX5, and WOX7, also encode proteins
with an EAR-like (for ERF-associated amphiphilic repression) domain,
with the sequence ASLELTN (van der Graaff et al. 2009).

As we detail herein, we have identified the S. latifolia SIWUS1 gene
as an X-linked gene in addition to those already known, and we find
that its Y copy has been lost, making it particularly interesting to test
whether males express the product at half the level of females, or
whether, as in some organisms, dosage compensation has evolved in
response the loss of the Y-linked copy. Recently, different two studies
tested for dosage compensation in S. latifolia using high-throughput
sequencing approaches. Chibalina and Filatov (2011) concluded that
dosage compensation does not occur, whereas Muyle et al. (2012)
concluded that it does. Careful studies of individual loci are therefore
needed to resolve this and to test whether different genes behave
differently (a possibility that must be considered because some mam-
malian X-linked genes escape inactivation; see Carrel and Willard
2005; Park et al. 2010). Moreover, a cytogenetic study found that
DNA methylation levels differ between the two X chromosomes in
S. latifolia females (Vyskot et al. 1993), suggesting the possibility that
one copy of X-linked genes might be inactivated in this plant, as
occurs in the X chromosome inactivation that evolved in mammalian
females, probably as a dosage compensation mechanism.

In this article, we identify the S. latifolia homolog, SIWUS1, of the
flower development gene WUS, and demonstrate that SIWUS1 is Y-
linked but, like Slcyt, does not have a Y-linked copy. Together with
the mapping data for the SIWUS1 ortholog to the Silene vulgaris linkage
group that is homologous with the X chromosome, but probably repre-
sents the ancestral state, before the sex chromosomes evolved, the hemo-
ygosity of SIWUS in males suggests that this gene does not represent an
addition onto the X chromosome. More likely, the S. latifolia Y chro-
mosome has undergone a previously undetected rearrangement during
sex chromosome evolution, resulting in the loss of this gene. The X is
also probably rearranged, although we cannot exclude the possibility
that the difference from the X is due to a rearrangement in S. vulgaris.

**MATERIALS AND METHODS**

**Plant materials**

An inbred Silene latifolia line, the K line (Kazama et al. 2003), was used in
all of our molecular experiments. Plants were grown from seeds in
pots in a regulated chamber at 23°C. The Berlin (B) plant was obtained from the seed collection of the Royal Botanic Gardens (Kew, United Kingdom). F1 progeny for use in segregation analysis were produced by crossing a K line male and a B female plant. For genetic mapping, 90 F2 progeny (21 male and 69 female) were generated by crossing single F1 males and females to generate a full-sib family. For sequencing analysis of S. vulgaris WUS genes, genomic DNA isolated from the ggK line (Kaiser et al. 2011) was used. To genetically map the SIWUS1 ortholog in S. vulgaris, 64 offspring of the SV2 family (Kaiser et al. 2009) were used. Leaves and flower buds were frozen in liquid nitrogen and stored at −80°C before their use in DNA or RNA preparation.

**Isolation of S. latifolia WUSCHEL (SIWUS) gene and orthologs in other Silene species**

Total RNA was extracted from male and female flower buds using an RNasey Plant Mini Kit (QIAGEN, Hilden, Germany). cDNA was synthesized from 3 mg of total RNA using Superscript II reverse transcriptase (Invitrogen by Life Technologies, Carlsbad, CA). A fraction of the cDNA preparation was then subjected to 30 cycles of PCR amplification (94°C for 1 min, 50°C for 1 min, and 70°C for 1 min) with the degenerate primers 5’T-TCY BCR TGC ATN GGR AAT AGT-3 and 5’T-TGG TTY CAR AAY CAT AAA GC-3. The PCR products were then cloned into the pGEM-T Easy vector (Promega Corporation, Madison, WI) and sequenced using a Big Dye Terminator v. 3.1 Cycle Sequencing Kit (Applied Biosystems by Life Technologies, Foster City, CA) and a 3730xl DNA Analyzer (Applied Biosystems by Life Technologies) with M13 Forward and Reverse primers.

To determine the sequence of the full-length SIWUS cDNA, both 5’ and 3’ RACE (Rapid Amplification of cDNA Ends) were performed using GeneRacer (Invitrogen by Life Technologies). Primers for the 5’ and 3’ regions of the isolated fragment were designated as SIWUS1_5RACE1 (5’T-CTG CTA ATG CAA GGG AAT ATA GTG TTC TC-3’) and SIWUS1_3RACEF1 (5’T-ACA CTT TAC AAT GGA CTT CTA TGG GGA GTA-3’) for SIWUS1, and SIWUS2_5RACE1 (5’T-GTT CTG AAA GTG AGG ACT TGA GTT GTC TGC AGT-3’) and SIWUS2_3RACEF1 (5’T-AGT AGT CAG TGG AAT CCC GTA GGT GGT A-3’) for SIWUS2. PCR conditions were 94°C for 2 min; five cycles at 94°C for 30 sec and 72°C for 2 min; five cycles at 94°C for 30 sec and 70°C for 1 min; 25 cycles at 94°C for 30 sec, 65°C for 30 sec and 72°C for 1 min; and then 72°C for 10 min. Nested PCR with SIWUS1_5RACE2 (5’-AAC ATG TAG TAG TAG TAG TGT AGG GAG TAA GC-3’), SIWUS1_3RACEF2 (5’-ACA AGA TCA CTA TGT TGC AGT ATC CGA GA-3’) and SIWUS2_5RACE2 (5’T-TGC AGT GTT ATT ATG AGG GGG AGT AGG-3’), and SIWUS2_3RACEF2 (5’T-CGA CAT CAT CAC TAT GTA GCA CAG CCA GA-3’) primers was carried out using the same PCR conditions. The PCR products were subcloned into the pGEM-T Easy vector and sequenced. Subclones of the 3’ RACE products of SIWUS1 and SIWUS2 were used as probes for Southern blot analyses.

To isolate orthologs of SIWUS genes, genomic DNA samples were obtained from S. dioica, S. denticulata, and S. vulgaris. For isolation of genomic fragments of SIWUS1 orthologs, a primer set of SIWUS1F9 and SIWUS1R7 was used in S. dioica and S. denticulata. SIWUS1F8 and SIWUS1R6 were used for isolation of the SIWUS1 ortholog in S. vulgaris. For isolation of genomic fragments of SIWUS2 orthologs, SIWUS2F9 and SIWUS2R5, SIWUS2F10 and SIWUS2R5, and SIWUS2R10 and SIWUS2R4 were used in S. dioica, S. denticulata, and S. vulgaris, respectively. The clones were cultured as described above, then sequenced using sequencing primers. All primers used for the sequencing analysis are listed in supporting information, Table S1.

**Phylogenetic tree analysis**

The amino acid sequences of the WUSCHEL homeobox proteins were aligned using CLUSTAL X version 1.81 (Thompson et al. 1997). The alignment was used in a neighbor-joining (NJ) analysis (Saitou and Nei 1987) in the software MEGA4 (Tamura et al. 2007). The resulting tree was tested by 1000 bootstrap replications using the MEGA program. For the phylogenetic analysis of the SIWUS1 and SIWUS2 orthologs in Silene species (Figure 6), the deduced amino acid sequences were aligned using the corresponding amino acid sequences of SIWUS1 and SIWUS2 as guides. The alignment was also used in NJ analyses using MEGA4 software.

**Southern blot analysis**

Genomic DNA was extracted from S. latifolia leaves using the Nucléo PhytoPure Genomic DNA Extraction Kit (GE Healthcare UK Ltd, Buckinghamshire, England). Genomic DNA (15 μg) was digested with EcoRV or HindIII for 12 h. The concentrations of the digests were measured and loaded equally onto a 1.0% (w/v) agarose gel and then transferred to an Immobilon-Ny+ membrane (Millipore). For the analysis shown in Figure 3A, inserts of subclones of 3’ RACE products of SIWUS1 and SIWUS2 were used as probes. For the analysis shown in Figure S1, the homoeo domain regions were amplified using SIWUS1-specific primer set (5’-AGT AGC ACA AGG TGG TAC ACA CCT A-3’ and 5’-CTT GTA GTG TGG TTA GAG GGA GTA-3’) and SIWUS2-specific primer set (5’T-AGT AGC ACA AGG TGG TAC ACA CGG A-3’ and 5’T-AGT GAC TAC TTC AGT TTG CAG TTG CAG-3’) and used as probes. Hybridization and detection were carried out as described previously (Kazama et al. 2009). Hybridization and signal detection were performed using the Gene Images AlkPhos Direct Labeling and Detection System (GE Healthcare UK Ltd). Posthybridization washes were carried out under relatively high stringency conditions at 60°C for 2 × 10 min (Figure 3A) and low stringency conditions at 50°C for 2 × 10 min (Figure S1). The hybridized membranes were visualized using Hyperfilm ECL (GE Healthcare UK Ltd) at room temperature with appropriate exposure times.

**Expression analysis**

RNA purification and cDNA synthesis were performed as described previously. Quantitative RT-PCR (qRT-PCR) was performed using a LightCycler instrument (Roche Diagnostics GmbH, Mannheim, Germany) with SYBR-Green detection chemistry (Applied Biosystems by Life Technologies). The SIWUS1-specific primers (5’T-CTT TTCT CGT CTT CGC CGC TA-3’ and 5’T-AGG AGG TGT CTC TAT CTC GG-3’) and the SIWUS2-specific primers (5’T-CAA CTA CCA CGG ATG TTC CGT-3’ and 5’T-CCA TCA TGG GTT CTT CTC GC-CC-3’) were used to amplify a fragment. 18S rRNA was used as a reference control by performing QRT-PCR with 100,000-fold cDNA dilutions as templates in each sample. The 18S rRNA-specific primers were as follows: 5’T-GGC AAC GGA TAT CTC GGC TG-3’ and 5’T-GTA GCC GGG AGA CTG GC-3’. Primer specificity was confirmed by sequencing each fragment after qRT-PCR. The qRT-PCR data shown are average relative quantities ± SE from at least three biological replicates. RT-PCR and qRT-PCR for allele-specific expression analysis were performed by the same method using K-line specific primers (5’T-AAG AAA AGG CTT ACT CCT TCC TAC A-3’ and 5’T-AGT ACT CCC CAT AGA AGT CTA TTC T-3’) and B-line-specific primers (5’T-AAG AAA AGG CTT ACT CCC TCA AAC T-3’ and 5’T-AGT ACT CCC CAT AGA AGT CCA CGT G-3’). In Figure 4B, primers specific for the S. latifolia actin gene (5’T-TTA CGG TAA
AGG TTC CTG AT-3' and 5'-AGC TTC GTG TCT CTG AAG A-3') were used as a control.

Testing for sex linkage and genetic mapping
Genomic DNA fragments of SlWUS1 and five other previously known X-linked genes (SlX1, SlX3, SlX4, SlX7, and DD4X) were amplified by PCR using the primers listed in Table S2. PCR products were cloned using the TOPO TA Cloning Kit (Invitrogen by Life Technologies) for sequencing. Using polymorphisms differing between the K-line parent and the B parent was 940 bp. To genotype DD4X, a cleaved amplified polymorphic sequence marker was designed to map the DD4X ortholog in S. vulgaris (SlWUS1). A fragment was amplified by PCR using gene-specific primers for SlWUS1 (5'-TTC CTG GAT GCA ACC AGT TCT C-3') and S. latifolia (SlWUS1) sequence (e-values: 0.01 and 0.013). The full-length cDNA sequences of the amplified fragments (using RACE) were 1212 bp coding a protein of 282 amino acid residues and a 1283 bp encoding a 317-amino acid protein. The former cDNA (which we named SlWUS1) was used for computing genetic distances in centimorgans (Kosambi 1944). The PCR parameters were 94°C for 5 min, followed by 30 cycles at 94°C for 30 sec, 60°C for 30 sec, and 72°C for 30 sec. The amplified fragments were digested by HincII and separated by agarose gel electrophoresis (1%). Another cleaved amplified polymorphic sequence marker was also designed to map the SlWUS1 ortholog in S. vulgari S (SlWUS1). A fragment was amplified Phusion enzyme (Finnzymes, Espoo, Finland) in a PIKO 24 thermal cycler (Finnzymes) using primers specific for SlWUS1 (5'-TTT GTC TCC GTG CCT GCC AGT ATG-3' and 5'-GTG TTC TTC ACC ACA AAC AGA GGT CAG-3'). PCR with the primers specific for the K line parent was then performed on F1 male and female progeny, allowing us to score the segregation of these genes. The PCR parameters used were 94°C for 3 min, followed by 33 cycles at 94°C for 30 sec, 53°C for 30 s, and 72°C for 30 sec.

To construct the X map, primers for high-resolution melting curve (HRM) analysis were designed for SlWUS1, SlX1, SlX3, SlX4, SlX7, and DD4X. The primers were then tested by HRM using genomic DNAs of the K-line parent, the B parent, and F1 plants. HRM was performed using a LightCycler 480s (Roche Diagnostics GmbH) as described previously (Kazama et al. 2011). Among the primer sets designed, the primer sets for SlWUS1, SlX1, SlX3, SlX4, and DD4X could distinguish polymorphisms differing between the K-line parent and the B parent. These primers were used for genotyping analysis (Table S2). To genotype SlX3, PCR primers for a simple sequence-length polymorphism marker were designed (5'-CAA TGA AGT TCT GTG CAC TGT TGA-3' and 5'-ATC CGG CGG TCT TCC GA-3'). The PCR parameters were 94°C for 5 min, followed by 30 cycles at 94°C for 30 sec, 60°C for 30 sec, and 72°C for 60 sec. Amplified segments were separated by agarose gel electrophoresis (1%). The K-line derived fragment was 749 bp, whereas fragment inherited from the B parent was 940 bp. To genotype SlX7, a cleaved amplified polymorphic sequence marker was designed. Genomic DNA fragments were amplified by PCR using gene-specific primers for SlX7 (5'-TTT GTC GAT GCA ACC AGT TCT C-3' and 5'-CCA GCA AAT ACA ACT CCC TCT T-3'). The PCR parameters were 94°C for 5 min, followed by 30 cycles at 94°C for 30 sec, 60°C for 30 sec, and 72°C for 30 sec. The amplified fragments were digested by HincII and separated by agarose gel electrophoresis (1%). Another cleaved amplified polymorphic sequence marker was also designed to map the SlWUS1 ortholog in S. vulgaris (SlWUS1). A fragment was amplified Phusion enzyme (Finnzymes, Espoo, Finland) in a PIKO 24 thermal cycler (Finnzymes) using primers specific for SlWUS1 (5'-TTT GTC TCC GTG CCT GCC AGT ATG-3' and 5'-GTG TTC TTC ACC ACA AAC AGA GGT CAG-3'). PCR with the primers specific for the K line parent was then performed on F1 male and female progeny, allowing us to score the segregation of these genes. The PCR parameters used were 94°C for 3 min, followed by 33 cycles at 94°C for 30 sec, 53°C for 30 s, and 72°C for 30 sec.

RESULTS
Identification of WUS homologs in S. latifolia
Using degenerate primers based on the A. thaliana WUS sequence, we performed PCR on S. latifolia male and female cDNAs, which amplified two fragments: 475 bp and 539 bp. A BLAST search identified homologous sequences with the A. thaliana WUS sequence (e-values: 0.01 and 0.013). The full-length cDNA sequences of the amplified fragments (using RACE) were 1212 bp encoding a protein of 282 amino acid residues and a 1283 bp encoding a 317-amino acid protein. The former cDNA (which we named SlWUS1) has 67.5% amino acid identity with the A. thaliana WUS sequence and the latter (SlWUS2) 55.8%. Homologs of the gene classified into the WUS-clade have been identified in many flowering plants, and the sequences are similar, not only in the homeobox domain but also in other regions. Alignment of the deduced amino acid sequences with WUS homologs in other plants showed high conservation of the homeobox domain,
were detected, and the to the highly conserved homeodomain regions. Both low-stringency Southern blot analyses using probes complementary linked and using all site types, not just synonymous ones, because the sequences SlWUS sequences fell into the clade with other plants ZmWOX3B (NP_001106240), ZmWOX3A (NP_001105160.1), OsWOX3 (Q33DK1.1), WOX4 (NP_175145.2), and WOX7 (NP_196196.1). Both (CAT02936.1), PhWOX2 (ACA64094.1), WOX5 (NP_187735.2), NjWOX5 (CAT03216.1), WOX3 (NP_180429), PhWOX3 (ACU68503.1), ZmWUS2 (NP_001105961.1), MAEWEST (ACA64093.1), WOX1 (NP_188428), WOX2 (NP_200742.2), GgWOX3 (CAT02931.1), PsWOX3 (CAT02936.1), PhWOX2 (ACA64094.1), WOX5 (NP_187735.2), NjWOX5 (CAT03216.1), WOX3 (NP_180429), PhWOX3 (ACU68503.1), ZmWOX3B (NP_001106240), ZmWOX3A (NP_001105160.1), OsWOX3 (Q33DK1.1), WOX4 (NP_175145.2), and WOX7 (NP_196196.1). Both SIWUS sequences fell into the clade with other plants’ WUS homologs, not the WOX clade.

and also the C-terminal domains (Figure 1), which include an acidic domain (Mayer et al. 1998), the WUS box (TLPLFPMH; Haecker et al. 2004), and the EAR-like domain (ASLELTLN; Ohta et al. 2001).

Our Nj tree showed that both SIWUS sequences belong to the clade that included the WUS homologs of other plants, not to the WOX gene clade (see Introduction and Figure 2). This result suggests that SIWUS1 and SIWUS2 are orthologs of the A. thaliana WUSCHEL gene. The synonymous divergence between the two S. latifolia sequences is 0.628 (using the method of Pamilo and Bianchi (1993); non-synonymous divergence is 0.149). SIWUS1 and SIWUS2 therefore diverged long before the sex chromosomes started diverging: silent-site divergence estimates between the sequences of many S. latifolia X- and Y-linked gene pairs range from 1.7% (for SfiX1/Sfy1, at one end of our X genetic map) to 16% (for SfiX4/Sfy4, which is distal in the X map; see Filatov and Charlesworth (2002) and Nicolas et al. (2005)), and the largest synonymous divergence values for gene pairs with X- and Y-linked copies is 25%. The divergence time of the duplicates also greatly predates that between S. latifolia and S. vulgaris, a species used below as an outgroup; based on 12 gene sequences, silent divergence values between orthologs of these two species never exceeds 20%, and the mean is 11.2% (standard error 1.3%).

Sex-linkage of the SIWUS1 gene
To determine whether SIWUS1 or SIWUS2 is located on the sex chromosomes, we used genomic Southern hybridization analyses of these genes. Specific regions from each gene, downstream of the conserved homeodomain sequences, were used as probes. The SIWUS1-specific probe gave weaker signals with males than females (Figure 3A), suggesting that SIWUS1 may be located on the X chromosome. In contrast, the SIWUS2-specific probe showed equal signal intensities in both sexes. To further test X-linkage of SIWUS1, segregation analysis was performed on F1 progeny from the cross between a K line male and a B female parent (see Materials and Methods). PCR on F1 male and female plants was performed using primers specific for the K line sequences. The F1 males X chromosome is inherited from the B female parent and their Y chromosome from the K line male, whereas the F1 females have one X chromosome from the B parent and the other from the K line. SIWUS1 amplified only from females, whereas SIWUS2 amplified from progeny of both sexes (Figure 3B). The SIAP3Y gene, a known Y-linked gene (Matsunaga et al. 2003), also amplified only from the F1 males. We conclude that SIWUS1 is X-linked and SIWUS2 is autosomal.

To test whether there is also a Y-linked copy of SIWUS1, we did low-stringency Southern blot analyses using probes complementary to the highly conserved homeodomain regions. Both SIWUS1 and SIWUS2 were detected, and the SIWUS1 probe and the SIWUS2 probe cross-hybridize weakly with one another; the nucleotide divergence between these two homeodomain region probes (which we estimated using all site types, not just synonymous ones, because the sequences are too short for accurate synonymous site estimates) is 16%. The divergence between the SIWUS1 sequence and its Y-linked copy, if one exists, is probably similar to this value, based on the genetic map location of SIWUS1 close to the S3X3 gene (see the section Genetic mapping of the X-linked SIWUS1 gene on the X chromosome and the ancestral chromosome), in the older evolutionary stratum of Bergero et al. (2007), with silent- or synonymous-site divergence up to 25% (see the section Identification of WUS homologs in S. latifolia). Because total divergence will be less than silent-site divergence, this experiment should thus be able to detect a Y copy, if it exists. However, no male-specific signal was detected (Figure S1). We conclude that there is no homologous SIWUS1 copy on the Y chromosome. Below, we discuss the divergence.

To further test whether a homologous Y-linked SIWUS1 copy exists, we used genomic DNA and cDNA from male individuals of S. latifolia and two other closely related dioecious species, S. dioica and S. dicianis, in PCR amplification tests using primers designed to match regions of the homeodomain that are conserved between SIWUS1 and SIWUS2 in S. latifolia and the A. thaliana sequence. All amplicons were sequenced and only the two SIWUS1 and SIWUS2 copies were detected (Figure S2). We conclude that SIWUS1 has no homologous Y-linked copy.

Expression of SIWUS1 in flower buds and leaves and tests for sex differences in expression and dosage compensation
To examine the tissues expressing SIWUS1 and SIWUS2, qRT-PCR analyses were performed with SIWUS1- and SIWUS2-specific primers (Figure 4A) using RNA extracted from leaves and flower buds of male and female plants corresponding to stages 1—8 (small flower buds) and 9—11 (large flower buds). The SIWUS transcript levels, relative to the 18S rRNA control, were calculated from at least three independent experiments. We used this small tissue panel because we focused on WUS function(s) that might be involved in floral development in S. latifolia. Both SIWUS1 and SIWUS2 were expressed in developing flower buds of both sexes but not in leaves. Both genes showed greater expression levels in large flower buds than the small ones. In the small flower bud sample, females showed greater SIWUS1 expression than males (P < 0.05, Student’s t-test). In contrast, large female buds tended to show lower expression levels of SIWUS1 and SIWUS2 than large male ones, although the difference for SIWUS2 transcription level was not statistically significant. The somewhat-lower expression levels of both SIWUS genes observed in large female flower buds, compared with male ones, is consistent with the previous report that Arabidopsis WUSCHEL is required for anther development (Deyhol et al. 2007).

Due to the complete X-linkage of SIWUS1, with no Y-linked copy, the S. latifolia male genome has one SIWUS1 copy, whereas females have two. We therefore tested whether dosage compensation occurs to
increase expression in males. First, we found roughly 2-fold greater expression level of SIWUS1 in small female flower buds than male ones and therefore concluded that the X-linked SIWUS1 gene is not expressed from only one of the females' alleles, whereas the other is inactivated. We further examined expression of each X-linked SIWUS1 allele in F1 females from an interstrain cross between the K-line male and a B female parent by RT-PCR, using K-line and B parent-specific primers. Both parental SIWUS1 X-linked alleles were detected in the female progeny (Figure 4B). Finally, we also checked the expression levels of both alleles by qRT-PCR. The expression levels did not differ significantly (Table S3).

These results indicate that there is no plant-wide inactivation of one SIWUS1 allele in S. latifolia females, as has been suggested from the observed methylation of one of the two X chromosomes (see Introduction). These results do not, of course, exclude random X-inactivation in individual cells, or sets of cells within the buds studied (like the situation in mammalian females). With such inactivation, both alleles would still be expressed. However, it seems incompatible with the much greater expression of SIWUS1 in small female flower buds than male ones; there is therefore no evidence that this gene is dosage compensated, although it cannot be ruled out that dosage compensation might have evolved, increasing expression in males, but that expression in female buds is also greatly increased, perhaps because the gene has a function specific to female buds.

**Genetic mapping of the X-linked SIWUS1 gene on the X chromosome and the ancestral chromosome**

We genetically mapped SIWUS1 using segregation analysis of 90 F2 progeny from the interstrain cross between a K-line male and a B female parent. We identified haplotypes of SIWUS1 and several other known X-linked genes, SIX1 (Delichere et al. 1999), SIX3 (Nicolas et al. 2005), SIX4 (Atanassov et al. 2001), DD44X (Moore et al. 2003), and SIX7 (Bergero et al. 2007) in the K-line and the B parent, and used them in segregation analysis. Recombination rates and a genetic map of the X chromosome are shown in Figure 5. SIWUS1 maps...
between these alternatives, we genetically mapped the from an autosome after the sex chromosomes evolved. To test be-
mosomes, using 64 F2 progeny of the mapping family SV2 (Kaiser in the related species SlWUS1 deleted during the inversions.
lost Y copy of Bergero et al. lost in plants with Y chromosome deletions (Zluvova SlX3 1276 | Y. Kazama
a phylogenetic tree using the function (Kosambi 1944). There is no X-inactivation of We sequenced the ortholog sequences (in order of distance from SlWUS1/SlY3, and SlX3; see Figure 5) with only one recombi-
from the divergence data (see the section Identification of WUS homologs in S. latifolia) that SIWUS1 and SIWUS2 duplicated long before the split between the ancestors of S. latifolia and S. vulgaris. We therefore did not map SIWUS2, since its location is not relevant for the questions we study here.

DISCUSSION

Despite their sequence divergence, degenerate primers can sometimes be used to identify genes in S. latifolia using A. thaliana sequences. We used this approach to sequence and study SIWUS1, a new X-linked gene in S. latifolia. Although the A. thaliana homolog is important in flower development, it may not be related to sex determination in S. latifolia. The SIWUS1 is X-linked, but has no homologous Y-linked copy, and sex is genetically determined in S. latifolia by a strongly male-determining Y chromosome (Westergaard 1958). This is only the third sex-linked gene with no Y homolog known from this plant. There is currently a strong bias toward discovering plant sex-linked genes through transcripts from Y-linked alleles, leading to underestimation of the extent of gene loss. However, loss of genes from plant Y chromosomes may really be much less than in animals, because some Y-linked genes will be under selection pressure to preserve their function because they are expressed and have important functions in haploid male gametophytes. Of the three genes so far known without Y copies, the function of the SlCyt gene is not currently known. Although the Y copy of the MROS3 gene is nonfunctional, this gene is expressed only in developing male flowers, so there is no reason to expect selection on MROS3-Y in pollen (Guttman and Charlesworth 1998).

Each example, of a plant X-linked gene without a Y copy is of great interest because these genes are the most relevant for testing whether dosage compensation has evolved in these young sex chromosomes. There is no X-inactivation of SIWUS1 in females, despite the observed apparent methylation of one of the X chromosomes, and no evidence for dosage compensation (small male buds have considerably lower apparent methylation of one of the X chromosomes, and no evidence dosage compensation (small male buds have considerably lower expression than female ones, see Results). To the extent that a single gene is informative, this finding is consistent with the conclusion that dosage compensation has not yet evolved in the S. latifolia sex chro-

SIWUS1 appears to have been lost from the Y chromosome during the evolution of dioecy. However, it is not known whether this was a degenerative loss, i.e., a deleterious change that would select for dosage compensation. Assuming that SIWUS1 has a function similar to the Arabidopsis WUSCHEL (See results and Figure 1 and Figure 2), loss of the gene from the Y chromosome creates a dosage difference between the sexes, which might lead to differences in floral phenotypes between male and female flowers. In A. thaliana, WUSCHEL is expressed in developing anthers and is required for development of

**Figure 5** Genetic map of six X-linked genes in S. latifolia. Recombination rates between the genes in this study are shown (A). (B) Genetic map; genetic distances were estimated using the Kosambi mapping function (Kosambi 1944). SIWUS1 is closely linked to SX3.

<table>
<thead>
<tr>
<th></th>
<th>SIX4</th>
<th>SIX7</th>
<th>SIX3</th>
<th>SIWUS1</th>
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**Figure 6** NJ tree (Saitou and Nei 1987) of WUS homologs in Silene species. The tree was estimated using an amino acid alignment of cDNA sequence from S. latifo-
and a genomic sequence from S. vulgaris, or using both (in S. dioica and S. clinica). The A. thaliana WUS amino acids were used as an out-group. Bootstrap values, based on 1000 replications, are shown.
the stamium (Deyhle et al. 2007). Although expression in anthers might suggest that loss of the SIWUSIY copy is likely to be deleterious (predicting that dosage compensation should evolve), it also complicates investigation of sex differences in the dosage of SIWUSI in large flower buds because only male flower buds have developing anthers at these stages.

It remains possible, however, that the loss of the SIWUSIY copy was adaptive. In S. latifolia, a sex difference in the size of floral whorl four is seen early in development, with a smaller size in males than females (Grant et al. 1994), consistent with the reduced size of whorl four in the A. thaliana wus mutant (Laux et al. 1996). Our qRT-PCR (Figure 4A) showed that female flower buds express SIWUSI at twice the level in male buds in stages 1–8, and the dosage difference may lead to the reduced gynoecium size of male flowers. This may be beneficial in S. latifolia males, releasing resources that allow them to produce more flowers, thus becoming more attractive to pollinators and increasing male reproduction success. Under such an adaptive hypothesis for the loss of the SIWUSIY copy, dosage compensation would not be predicted to evolve for the SIWUSI gene.

Alternatively, the SIWUSIY copy may have evolved a function in female flower development and have evolved elevated expression in female buds that more than compensates for any increase in males (assuming that dosage compensation has evolved, increasing male expression). A S. latifolia homolog of the CLAVATA1 gene is differentially expressed in male and female flower development (Koizumi et al. 2010). CLAVATA1 is involved in stem cell maintenance in shoot and flower meristems together with WUS in A. thaliana (Brand et al. 2000; Ogawa et al. 2008). It is possible that SIWUSI contributes to female flower development through the WUSCHEL-CLAVATA pathway.

The mechanism by which the Y copy of SIWUSI has disappeared could be deletion of an entire Y region, deleting several or many genes, or by a specific deletion of just this gene, or by loss-of-function mutations followed by sequence divergence, so that no Y-linked copy remains detectable even in our low-stringency Southern blotting experiments, including in S. dioica and S. diclinis (see Results). We therefore conclude that the Y-linked copy of the SIWUSI gene was lost before these species diverged, in a deletion event in their common ancestor, before these dioecious species split, additional to the inversions currently detected (Kejnovsky and Vyskot 2010).

Because our results exclude the possibility that SIWUSI was translocated to the X chromosome from an autosome after the sex chromosomes evolved, they suggest that gene losses are occurring, an important form of genetic degeneration that has not previously been documented in the S. latifolia Y. An addition to the X was reported for SICyt, based on evidence that the S. vulgaris ortholog is unlinked to the other sex chromosome homologs in that species (Kaiser et al. 2009). However, genetic mapping has subsequently shown that this was due to a gap in the genetic map, and that all genes that are fully sex-linked in S. latifolia map to a single S. vulgaris linkage group (R. Bergero, S. Qiu, A. Forrest, H. Borthwick, and D. Charlesworth, unpublished results).

Farther mapping of sex-linked genes will be needed to test whether the loss of the Y copy involved deletion of other genes and to learn the details of the rearrangement during sex chromosome evolution in S. latifolia and its close relatives. Deleted genes would have to have been dispensable, either because they are expressed in the sporophyte stage (as just discussed), or else because the chromosome must have carried genes that were already inactive, so that their function was adequately carried out by a single X-linked copy. In A. thaliana, the WUSCHEL gene is required for both meristem size regulation of whorl 4, and for anther development (Deyhle et al. 2007), making it unlikely that this male function is dispensable in S. latifolia.

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