



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

## Ischaemia-reperfusion injury impairs tissue plasminogen activator release in man

**Citation for published version:**

Pedersen, CM, Barnes, G, Schmidt, MR, Bøtker, HE, Kharbanda, RK, Newby, DE & Cruden, NL 2012, 'Ischaemia-reperfusion injury impairs tissue plasminogen activator release in man', *European Heart Journal*, vol. 33, no. 15, pp. 1920-7. <https://doi.org/10.1093/eurheartj/ehr380>

**Digital Object Identifier (DOI):**

[10.1093/eurheartj/ehr380](https://doi.org/10.1093/eurheartj/ehr380)

**Link:**

[Link to publication record in Edinburgh Research Explorer](#)

**Document Version:**

Publisher's PDF, also known as Version of record

**Published In:**

European Heart Journal

**Publisher Rights Statement:**

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

**General rights**

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy**

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact [openaccess@ed.ac.uk](mailto:openaccess@ed.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.



# Ischaemia–reperfusion injury impairs tissue plasminogen activator release in man

Christian M. Pedersen<sup>1,2\*</sup>, Gareth Barnes<sup>1</sup>, Michael R. Schmidt<sup>2</sup>, Hans Erik Bøtker<sup>2</sup>, Rajesh K. Kharbanda<sup>3</sup>, David E. Newby<sup>1</sup>, and Nicholas L. Cruden<sup>1</sup>

<sup>1</sup>Centre for Cardiovascular Science, University of Edinburgh, Edinburgh, UK; <sup>2</sup>Research Unit, Department of Cardiology, Aarhus University Hospital Skejby, Brendstrupgaardsvej 100, 8200 Aarhus N, Denmark; and <sup>3</sup>The John Radcliffe Hospital, OXFORD NIHR Biomedical Research Centre, Oxford, UK

Received 6 May 2011; revised 19 July 2011; accepted 13 September 2011; online publish-ahead-of-print 11 October 2011

## Aims

Ischaemia–reperfusion (IR) injury causes endothelium-dependent vasomotor dysfunction that can be prevented by ischaemic preconditioning. The effects of IR injury and preconditioning on endothelium-dependent tissue plasminogen activator (t-PA) release, an important mediator of endogenous fibrinolysis, remain unknown.

## Methods and results

Ischaemia–reperfusion injury (limb occlusion at 200 mmHg for 20 min) was induced in 22 healthy subjects. In 12 subjects, IR injury was preceded by local or remote ischaemic preconditioning (three 5 min episodes of ipsilateral or contralateral limb occlusion, respectively) or sham in a randomized, cross-over trial. Forearm blood flow (FBF) and endothelial t-PA release were assessed using venous occlusion plethysmography and venous blood sampling during intra-arterial infusion of acetylcholine (5–20 µg/min) or substance P (2–8 pmol/min). Acetylcholine and substance P caused dose-dependent increases in FBF ( $P < 0.05$  for all). Substance P caused a dose-dependent increase in t-PA release ( $P < 0.05$  for all). Acetylcholine and substance P-mediated vasodilatation and substance P-mediated t-PA release were impaired following IR injury ( $P < 0.05$  for all). Neither local nor remote ischaemic preconditioning protected against the impairment of substance P-mediated vasodilatation or t-PA release.

## Conclusion

Ischaemia–reperfusion injury induced substance P-mediated, endothelium-dependent vasomotor and fibrinolytic dysfunction in man that could not be prevented by ischaemic preconditioning.

Clinical Trial Registration Information: Reference number: NCT00789243, URL: <http://clinicaltrials.gov/ct2/show/NCT00789243?term=NCT00789243&rank=1>

## Keywords

Endogenous fibrinolysis • Endothelium • Ischaemia–reperfusion • Preconditioning • Substance P

## Introduction

Acute arterial occlusion can lead to end-organ ischaemia and, ultimately, infarction. Although treatment is usually directed at prompt restoration of flow in the occluded artery, reperfusion itself may trigger additional injury beyond that induced by ischaemia alone, although the mechanism is poorly understood. Impaired endothelium-dependent vasomotor function following ischaemia and reperfusion has been demonstrated in experimental models<sup>1</sup> and *in vivo* in man,<sup>2</sup> and can be prevented by prior exposure to intermittent sublethal ischaemia—*ischaemic preconditioning*—induced either locally in the vascular bed immediately downstream

of the preconditioning stimulus, or remotely in an organ anatomically distant from the preconditioning stimulus.<sup>2,3</sup> The effects of ischaemia–reperfusion (IR) injury and ischaemic preconditioning on other important aspects of endothelial function remain unknown.

In addition to the regulation of vascular tone, the endothelium is intimately involved in the prevention of intravascular thrombosis through the endogenous fibrinolytic pathway.<sup>4</sup> The activation of plasminogen is a critical step in endogenous fibrinolysis, with tissue plasminogen activator (t-PA) being the main plasminogen activator in man. However, only a relatively small proportion of the t-PA present in plasma is functionally active, largely due

\* Corresponding author. Tel: +45 89 49 62 32, Fax: +45 89 49 60 09, Email: [christian.m.pedersen@ki.au.dk](mailto:christian.m.pedersen@ki.au.dk)

Published on behalf of the European Society of Cardiology. All rights reserved. © The Author [2011].

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

to binding and inhibition by the serpin, plasminogen activator inhibitor type 1 (PAI-1). In plasma, PAI-1 is present in molar excess over t-PA, hence for active unbound t-PA to reach a thrombus, rapid local release is vital, fibrinolysis being much more effective if t-PA is incorporated during, rather than after, thrombus formation. Thus, the ability of the endothelium to release t-PA rapidly plays a key role in determining local endogenous fibrinolytic activity, with a reduction in local endothelial t-PA release favouring thrombus formation and propagation and, ultimately, vascular obstruction.<sup>5</sup>

Using forearm venous occlusion plethysmography and the endothelium-dependent agonist, substance P, we have demonstrated that substance P-induced t-PA release is impaired in the forearm circulations of smokers<sup>6</sup> and in patients with coronary artery disease—the degree of impairment of t-PA release correlating with the risk of future cardiovascular events.<sup>7</sup> Similarly, endothelial t-PA release is impaired in the coronary circulation of patients with atherosclerotic coronary disease, the extent of impairment being inversely proportional to plaque burden.<sup>8</sup>

Preclinical models report an increase in local t-PA concentrations immediately following IR injury, which return to basal levels within 10 min although these findings may be confounded by intravascular pooling of t-PA.<sup>9–13</sup> In man, the acute effects of IR injury on endogenous fibrinolysis are less clear. Plasma t-PA concentrations are increased in patients with critical limb ischaemia,<sup>14</sup> but reduced in patients with intermittent claudication.<sup>15</sup>

We hypothesized that, in keeping with endothelium-dependent vasodilatation to acetylcholine, IR injury would impair endothelium-

dependent endogenous fibrinolysis and that ischaemic preconditioning would protect against this. To address this, we examined the effect of IR injury and ischaemic preconditioning on substance P-mediated vasodilatation and t-PA release *in vivo* in man.

## Methods

The study was carried out in accordance with the Declaration of Helsinki (2000) of the World Medical Association. The Lothian Research Ethics Committee approved the study and all subjects gave written informed consent.

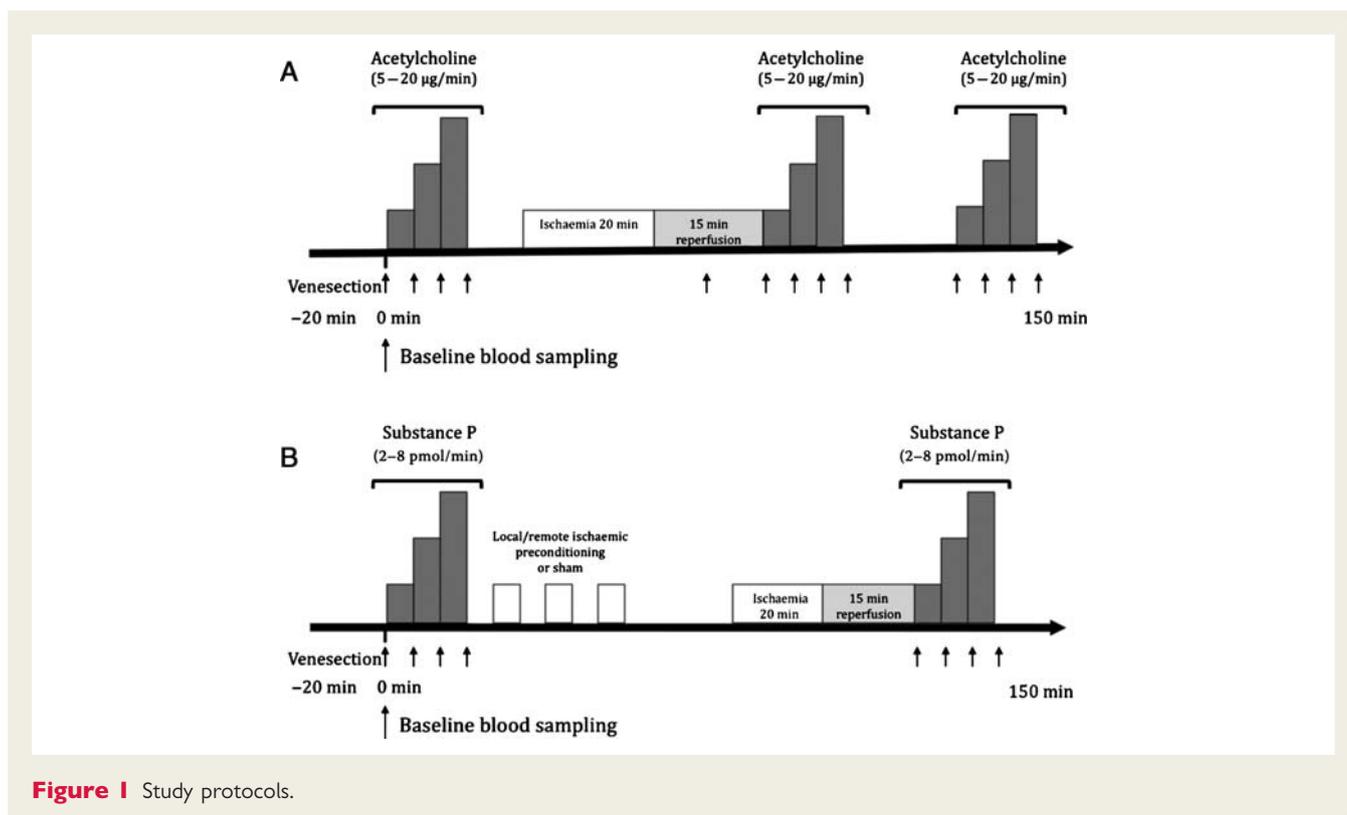
### Subjects

Twenty-two healthy male volunteers were recruited. All subjects were non-smoking, taking no regular medications and clinically well. Subjects abstained from vasoactive drugs for 7 days, caffeine-containing drinks and alcohol for 24 h, and fasted for at least 4 h before each visit. Subjects attended either for one study visit (Protocol 1) or for three study visits (Protocol 2), with at least 2 weeks between each visit.

### Study protocols

#### Protocol 1

Ischaemia–reperfusion injury was induced by cuff inflation around the non-dominant upper arm to 200 mmHg for 20 min in 10 healthy subjects (Protocol 1, *Figure 1A*). Forearm blood flow (FBF) responses to intra-arterial acetylcholine (5–20  $\mu\text{g}/\text{min}$ ) were assessed at baseline and at 15 and 45 min after IR injury.



**Figure 1** Study protocols.

## Protocol 2

Local or remote ischaemic preconditioning, or sham, was performed prior to the induction of IR injury in the non-dominant arm in 12 subjects in a randomized, blinded, cross-over fashion (Protocol 2, Figure 1B). Local or remote ischaemic preconditioning was induced by cuff inflation around the non-dominant or dominant upper arm, respectively, to 200 mmHg for 5 min on three occasions 5 min apart. During sham procedures, the upper arm cuff was inflated to 10 mmHg for a similar time period. Forearm blood flow and plasma concentrations of t-PA and PAI-1 antigen and activity during intra-arterial infusion of substance P (2–8 pmol/min) were assessed at baseline and 15 min after IR injury.

## Intra-arterial drug administration

All studies were performed with patients lying supine in a quiet, temperature-controlled (22–25°C) room. Under local anaesthesia, a 27-gauge needle (Cooper Needle Works Ltd) was inserted into the brachial artery of the non-dominant arm. The rate of intra-arterial drug infusion remained constant throughout at 1 mL/min.

## Forearm blood flow and blood pressure

Bilateral arterial FBF was measured using venous occlusion plethysmography (Hokanson EC6 plethysmograph, DE Hokanson, Inc., USA, and Chart v5.0.1 software, ADInstruments Ltd, UK) as previously described.<sup>16</sup> Plethysmographic data were extracted from the Chart data files, and FBF calculated for individual venous occlusion cuff inflations. Usually, the last five flow recordings in each 3 min measurement period were calculated and averaged for each arm. Heart rate and blood pressure were recorded in the non-infused arm at intervals throughout the study using a semi-automated non-invasive oscillometric sphygmomanometer (Takeda UA 751; Takeda Medical, Inc., Japan).

## Venous blood samples and assays

For Protocol 2, 17-gauge venous cannulae were inserted bilaterally into a large antecubital vein. Ten millilitres of blood was withdrawn simultaneously from each arm at baseline and in the last minute of each drug infusion period and collected into acidified, buffered citrate (Biopool Stabilyte, Umeå) for t-PA assays and citrate (Monovette, Sarstedt, Numbrecht) for PAI-1 assays. Estimated net release of t-PA antigen and activity were defined previously as the product of the infused forearm plasma flow (based on the mean haematocrit and the infused FBF) and the concentration difference between the infused and non-infused arms.<sup>16</sup> Samples were kept on ice before being centrifuged at 2000g for 30 min at 4°C. Platelet-free supernatant

was decanted and stored at –80°C before assay. Plasma t-PA (t-PA Combi Actibind Elisa Kit, Technoclone, Vienna, Austria) and PAI-1 antigen and activity (Elitest PAI-1, Hyphen Biomed, Neuville-Sur-Oise, France) concentrations were determined using enzyme-linked immunosorbent assays.

## Data analysis and statistics

The software package GraphPad PRISM 5 was used for analysis (GraphPad Software, Inc., La Jolla, CA 92037, USA). All data are expressed as mean ± SEM unless otherwise stated. Data were analysed using repeated measures two-way ANOVA, one-way ANOVA, and paired Student's *t*-test, where appropriate. Statistical significance was defined as a two-sided *P*-value <0.05.

## Results

There were no differences in the heart rate, blood pressure, haematocrit, FBF, or t-PA release at baseline between or during visits (Table 1; Supplementary material online, Table S1 and Figure S1). Ischaemia–reperfusion injury and ischaemic preconditioning were well tolerated by all subjects, with no reported side effects.

## Vasomotor function

Intra-arterial infusion of acetylcholine caused a dose-dependent increase in the FBF in the infused arm in all studies ( $P < 0.05$  for all) that was attenuated by IR injury ( $P = 0.007$ ; Figure 2).

Substance P caused a dose-dependent increase in FBF in the infused arm in all studies ( $P < 0.05$  for all) that was impaired following IR injury ( $P < 0.01$  for all; Table 2, Figure 3). Compared with sham, neither local nor remote ischaemic preconditioning altered the reduction in substance P-mediated vasodilatation following IR injury (Table 2, Figure 3).

## Fibrinolytic function

Substance P caused a dose-dependent increase in absolute t-PA antigen ( $P < 0.0005$  for all; Table 3) and activity ( $P < 0.0001$  for all; Table 3), and net release of t-PA antigen and activity in the infused arm in all studies ( $P < 0.01$  for all; Figure 4) that was attenuated by IR injury ( $P < 0.05$  for all; Figure 4). Compared with sham, neither local nor remote ischaemic preconditioning altered

**Table 1** Baseline characteristics

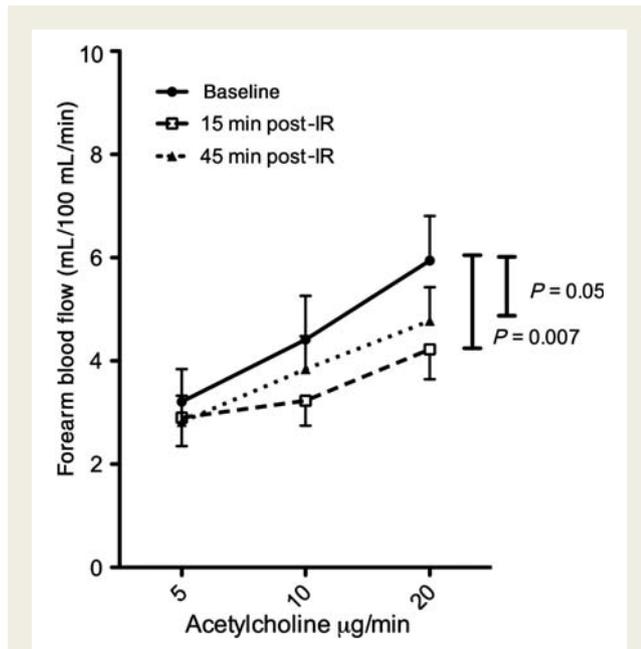
Parameter	Protocol 1	Protocol 2*		
	IR injury	Sham + IR	Local IPC + IR	Remote IPC + IR
Age, years	23.6 ± 1	22.3 ± 1	22.3 ± 1	22.3 ± 1
Body mass index, kg/m <sup>2</sup>	23.8 ± 1	23.5 ± 0.5	23.5 ± 0.5	23.5 ± 0.5
Heart rate, b.p.m.	58.0 ± 3	62.7 ± 2	61.0 ± 2	64.8 ± 2
Mean arterial pressure, mmHg	92.4 ± 2	96.5 ± 2	96.3 ± 2	93.5 ± 3
Baseline haematocrit, %	43.6 ± 1	41.6 ± 1	41.7 ± 1	39.8 ± 2

IR, ischaemia–reperfusion injury; IPC, ischaemic preconditioning.

\* $P > 0.05$  for all, paired Student's *t*-test; sham + IR vs. local IPC + IR and sham + IR vs. remote IPC + IR.

substance P-mediated release of t-PA antigen or activity following IR injury (Table 3, Figure 4).

There were no differences in plasma PAI-1 concentrations at baseline or following IR injury (Supplementary material online, online data table).



**Figure 2** Effect of ischaemia–reperfusion injury on acetylcholine-mediated vasodilatation. IR indicates ischaemia–reperfusion (Protocol 1). Data analysed using two-way ANOVA with repeated measures.

## Discussion

To our knowledge, this is the first study to examine the effect of IR injury and ischaemic preconditioning on endogenous fibrinolysis in man. Having first confirmed the validity of the forearm model using acetylcholine, we have demonstrated that substance P-induced vasodilatation and endothelial t-PA release are impaired following IR injury. Neither local nor remote ischaemic preconditioning protected against the impairment of substance P-mediated vasodilatation or t-PA release induced by IR injury.

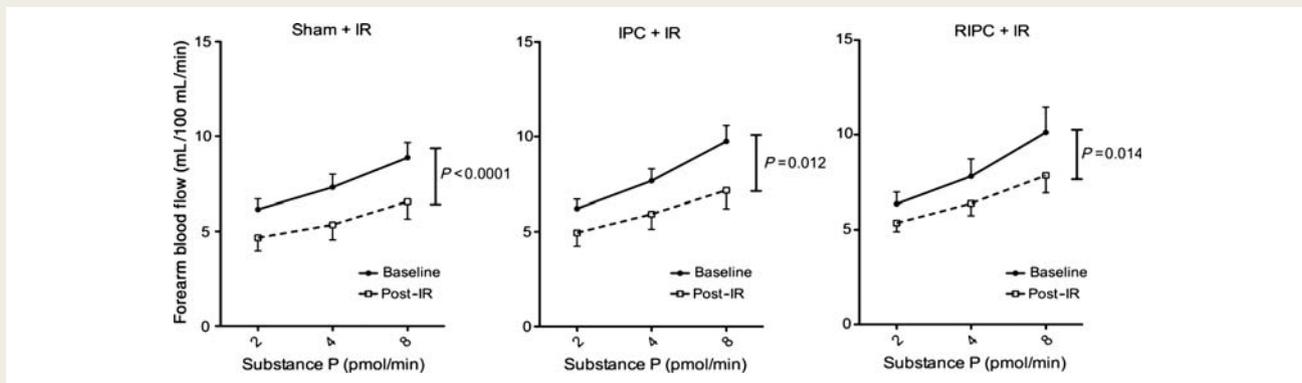
A major finding of this study is that IR injury attenuates the dynamic release of t-PA from the endothelium in the human peripheral vasculature *in vivo* in man. Preclinical data, predominantly from *ex vivo* models, suggest that short periods of IR injury are associated with increased local t-PA concentrations immediately after the onset of reperfusion.<sup>9–13</sup> In these experimental studies, t-PA concentrations peaked 2–5 min after the onset of reperfusion and returned to baseline within 10 min of reperfusion. However, the capacity of the endothelium to release t-PA appeared to decline with repeated episodes of vascular occlusion.<sup>11</sup> The apparent conflict between preclinical data and the current study may be explained, at least in part, by temporal differences in the assessment of t-PA release relative to the onset of reperfusion. We assessed the potential for t-PA release 15 min after the ischaemic insult because we wished to avoid the potential confounding of pooling of t-PA and the effects of the direct ischaemic stimulus itself. These effects underlie the principles of why t-PA concentrations rise during the venous occlusion test.<sup>4</sup>

In contrast to the effects of ischaemic preconditioning on acetylcholine-induced vasodilatation, we found no effect of ischaemic preconditioning on substance P-induced vasodilatation or endothelial t-PA release following IR injury. The reason for these

**Table 2** Effect of ischaemia–reperfusion injury alone or ischaemia–reperfusion injury preceded by local or remote ischaemic preconditioning on substance P-mediated vasodilatation in the infused and non-infused arms

Parameter	Baseline				Post-IR			
	Baseline	Substance P (pmol/min)			Baseline	Substance P (pmol/min)		
		2	4	8		2	4	8
Sham-IPC + IR								
FBF, mL/100 mL/min								
Infused	1.8 ± 0.2	6.2 ± 0.6	7.3 ± 0.7	8.9 ± 0.8*, **	1.9 ± 0.2	4.7 ± 0.7	5.3 ± 0.8	6.6 ± 0.9* **
Non-infused	1.7 ± 0.2	1.7 ± 0.2	1.7 ± 0.2	1.9 ± 0.2	1.7 ± 0.3	1.8 ± 0.4	1.8 ± 0.5	1.8 ± 0.4
Local IPC + IR								
FBF, mL/100 mL/min								
Infused	2.3 ± 0.3	6.2 ± 0.5	7.7 ± 0.6	9.7 ± 0.8*, ***	2.2 ± 0.3	5.0 ± 0.7	5.9 ± 0.8	7.2 ± 1.0* ***
Non-infused	2.0 ± 0.2	2.0 ± 0.2	1.9 ± 0.2	2.2 ± 0.3	2.0 ± 0.3	1.9 ± 0.2	1.9 ± 0.3	2.0 ± 0.3
Remote IPC + IR								
FBF, mL/100 mL/min								
Infused	2.4 ± 0.4	6.4 ± 0.6	7.8 ± 1.0	10.1 ± 1.3*, ****	2.3 ± 0.3	5.4 ± 0.5	6.4 ± 0.6	7.8 ± 0.9* ****
Non-infused	2.0 ± 0.3	1.8 ± 0.3	1.8 ± 0.3	1.9 ± 0.3	1.8 ± 0.3	1.8 ± 0.3	1.7 ± 0.2	1.8 ± 0.3

FBF, forearm blood flow; IPC, ischaemic preconditioning; IR, ischaemia–reperfusion injury. ANOVA dose response \* $P < 0.0001$  for all; ANOVA baseline vs. post-IR \*\* $P < 0.0001$ , \*\*\* $P = 0.012$ , \*\*\*\* $P = 0.014$ .



**Figure 3** Effect of ischaemia–reperfusion injury alone and ischaemia–reperfusion injury preceded by local or remote ischaemic preconditioning on substance P-induced vasodilatation (Protocol 2). IR indicates ischaemia–reperfusion; IPC, local ischaemic preconditioning; and RIPC, remote ischaemic preconditioning. Data analysed using two-way ANOVA with repeated measures.

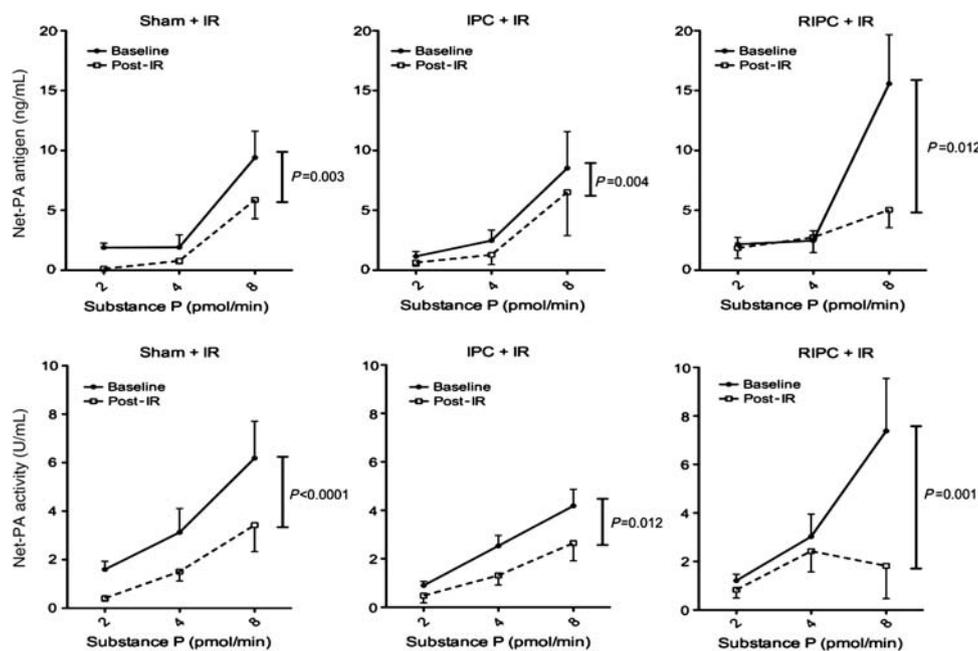
**Table 3** Effect of ischaemia–reperfusion injury alone or ischaemia–reperfusion injury preceded by local or remote ischaemic preconditioning on substance P-mediated tissue plasminogen activator antigen and activity in the infused and non-infused arms

Parameter	Baseline				Post-IR			
	Baseline	Substance P (pmol/min)			Baseline	Substance (P pmol/min)		
		2	4	8		2	4	8
<b>Sham-IPC + IR</b>								
t-PA antigen, ng/mL								
Infused	3.2 ± 0.7	3.8 ± 0.7	3.8 ± 0.6	5.2 ± 0.7*	2.9 ± 0.7	3.0 ± 0.6	3.4 ± 0.7	4.7 ± 0.8*
Non-infused	3.0 ± 0.7	3.3 ± 0.7	3.6 ± 0.7	3.4 ± 0.6	2.8 ± 0.6	3.2 ± 0.6	3.3 ± 0.7	3.1 ± 0.6
t-PA activity, U/mL								
Infused	0.6 ± 0.1	0.9 ± 0.1	1.1 ± 0.2	1.7 ± 0.3*	0.6 ± 0.05	0.8 ± 0.1	1.1 ± 0.1	1.5 ± 0.3*
Non-infused	0.4 ± 0.1	0.4 ± 0.1	0.4 ± 0.1	0.6 ± 0.1	0.6 ± 0.1	0.7 ± 0.05	0.6 ± 0.05	0.6 ± 0.05
<b>Local IPC + IR</b>								
t-PA antigen, ng/mL								
Infused	3.1 ± 0.6	3.4 ± 0.7	3.8 ± 0.8	4.8 ± 1.1*	3.0 ± 0.7	3.2 ± 0.7	3.6 ± 0.9	4.7 ± 1.4**
Non-infused	3.1 ± 0.6	3.1 ± 0.6	3.2 ± 0.7	3.2 ± 0.6	2.9 ± 0.6	3.0 ± 0.6	3.2 ± 0.6	3.0 ± 0.6
t-PA activity, U/mL								
Infused	0.5 ± 0.1	0.7 ± 0.1	1.0 ± 0.1	1.4 ± 0.2*	0.6 ± 0.1	0.8 ± 0.1	1.1 ± 0.2	1.4 ± 0.2*
Non-infused	0.5 ± 0.1	0.5 ± 0.1	0.4 ± 0.1	0.6 ± 0.1	0.6 ± 0.05	0.6 ± 0.05	0.6 ± 0.05	0.7 ± 0.05
<b>Remote IPC + IR</b>								
t-PA antigen, ng/mL								
Infused	3.5 ± 0.6	3.9 ± 0.6	4.5 ± 0.9	6.2 ± 1.2*	3.0 ± 0.6	3.3 ± 0.7	3.6 ± 0.6	4.3 ± 0.9**
Non-infused	3.2 ± 0.6	3.3 ± 0.6	3.9 ± 0.7	3.4 ± 0.6	3.3 ± 0.7	3.0 ± 0.6	3.1 ± 0.7	3.2 ± 0.6
t-PA activity, U/mL								
Infused	0.6 ± 0.1	0.8 ± 0.1	1.2 ± 0.2	1.8 ± 0.3*	0.6 ± 0.1	0.7 ± 0.1	1.0 ± 0.2	1.1 ± 0.3*
Non-infused	0.4 ± 0.1	0.5 ± 0.1	0.5 ± 0.1	0.3 ± 0.1	0.5 ± 0.1	0.5 ± 0.1	0.4 ± 0.1	0.6 ± 0.1

t-PA, tissue-plasminogen activator; IPC, ischaemic preconditioning; IR, ischaemia–reperfusion injury. ANOVA dose response \**P* < 0.0001 for all, \*\**P* = 0.0004.

apparent differences is unclear but is likely to reflect differences in intracellular signalling. Previous work has implicated the mitochondrial ATP-sensitive potassium channel in the protection of

acetylcholine-mediated vasodilatation afforded by ischaemic preconditioning.<sup>17,18</sup> In contrast, thrombin-stimulated release of t-PA from endothelial cells appears to occur independently of both



**Figure 4** Effect of ischaemia–reperfusion injury alone and ischaemia–reperfusion injury preceded by local or remote ischaemic preconditioning on substance P-induced net release of tissue plasminogen activator antigen and activity (Protocol 2). IR indicates ischaemia–reperfusion; IPC, local ischaemic preconditioning; RIPC, remote ischaemic preconditioning; and t-PA, tissue-plasminogen activator. Data analysed using two-way ANOVA with repeated measures.

nitric oxide and  $K^+$  channels *in vitro*,<sup>19</sup> and if anything, in contrast to acetylcholine-mediated vasodilatation, nitric oxide inhibition augments thrombin-induced t-PA release *in vivo* in man.<sup>20</sup>

Limited preclinical data have suggested a potential role for substance P in the mechanism of ischaemic preconditioning.<sup>21</sup> We have previously demonstrated that forearm vascular responses to substance P are highly reproducible at 30 min and 7 days.<sup>22</sup> Taken together, our results do not support the hypothesis that substance P-induced signalling protects against endothelial dysfunction induced by IR injury, although we acknowledge that the current study was not designed to specifically address this issue.

## Clinical relevance

Our findings have potentially significant clinical implications. Despite optimal antithrombotic therapy, early arterial reocclusion and microvascular obstruction remain recognized complications following reperfusion therapy for acute myocardial infarction and are associated with adverse clinical outcomes.<sup>23–26</sup> A number of mechanisms have been implicated in the pathophysiology of microvascular obstruction, or ‘no reflow’, including vasomotor dysfunction and vasospasm, localized oedema and inflammation,<sup>27</sup> coronary micro-embolization,<sup>28</sup> and *in situ* thrombus formation.<sup>23,29</sup> Our findings suggest a novel mechanism, namely an inducible defect in local endogenous fibrinolytic activity, favouring local thrombus formation in the pathophysiology of microvascular obstruction and re-infarction. In support of this hypothesis, streptokinase reduces myocardial congestion and improves microvascular perfusion in a canine model of IR injury<sup>30</sup> and acutely improves

indices of coronary microvascular perfusion when administered immediately following primary percutaneous coronary intervention in patients with ST-segment elevation myocardial infarction.<sup>31</sup>

Direct assessment of coronary endothelial function *in vivo* in man necessitates complex invasive studies with obvious limitations. The accessibility of the forearm vascular bed makes it an attractive model by which to assess vascular function *in vivo*. Although less susceptible to atherosclerosis and thrombosis, consistent findings between the peripheral<sup>2,6,32</sup> and coronary circulations<sup>8,33,34</sup> provide support for the forearm model as a surrogate for the coronary circulation.<sup>35</sup>

## Limitations

We employed a robust, randomized, cross-over study design. Although study investigators were blinded to the intervention performed at each visit (IR injury, ischaemic preconditioning, or sham), for obvious reasons it was not possible to ensure blinding of the study subjects. Forearm venous sampling during local infusion of agonists is a well established technique in the assessment of endogenous t-PA release.<sup>4,6,7,36</sup> To minimize the potential for regional variation in venous effluent, venous sampling is always performed from a large vein located in the antecubital fossa.

Intravascular thrombus formation is dependent on a number of factors, including the endothelial function, the coagulation cascade, and platelets.<sup>5</sup> We have previously demonstrated that IR injury increases markers of platelet activation and that remote ischaemic preconditioning protects against this platelet activation.<sup>37</sup> The current study was designed to examine the effects of IR injury

and ischaemic preconditioning on a specific aspect of endothelial function, t-PA release, intimately involved in thrombus formation. In addition to t-PA antigen, we assessed enzymatic activity using a standard immunological technique. We acknowledge that this model may be relatively simplistic in the context of the complex *in vivo* response to thrombus formation. There are, however, obvious methodological difficulties in directly assessing the effect of endogenous t-PA release on arterial thrombosis *in vivo* in man. Further work is required to examine the effects of ischaemic preconditioning and IR injury on the coagulation cascade and markers of inflammation *in vivo* in man.

In summary, we have demonstrated that substance P-induced endothelium-dependent vasodilatation and t-PA release are impaired following IR injury in the human forearm. In contrast to acetylcholine, neither local nor remote ischaemic preconditioning protected against the impairment of substance P-mediated vasomotor and fibrinolytic function induced by IR injury. Our findings support an inducible defect in endogenous fibrinolysis as a novel mechanism in the pathophysiology of IR injury in man.

## Supplementary material

Supplementary material is available at *European Heart Journal* online.

## Acknowledgements

We would like to thank the staff of the Wellcome Trust Clinical Research Facility at the Royal Infirmary of Edinburgh for their help with this study.

## Funding

This work was supported by Foundation Leducq (CV06) and the British Heart Foundation (PG/08/093/26020). C.M.P. has been personally funded by the Danish Agency for Science, Technology and Innovation, Region Midtjyllands Sundhedsvidenskabelige Forskningsfond, Det Classenske Fideicomis Jubilæumsfond, Snedkermester Sophus Jacobsen og hustru Astrid Jacobsen's Fond, Civilingeniør Stenild Hjorth's enke Else Hjorth's Fond, The A.P. Møller Foundation for the Advancement of Medical Science, Kirsten Antonius' Mindelegat and Institute of Clinical Medicine, University of Aarhus. R.K.K. is supported by the OXFORD NIHR Biomedical Research Centre. D.E.N. is supported by the British Heart Foundation, and the Wellcome Trust Clinical Research Facility is supported by NHS Research Scotland (NRS) through NHS Lothian. Funding to pay the Open Access publication charges for this article was provided by British Heart Foundation.

**Conflict of interest:** none declared.

## References

1. Tiefenbacher CP, Chilian WM, Mitchell M, DeFily DV. Restoration of endothelium-dependent vasodilation after reperfusion injury by tetrahydrobiopterin. *Circulation* 1996;**94**:1423–1429.
2. Kharbanda RK, Peters M, Walton B, Kattenhorn M, Mullen M, Klein N, Vallance P, Deanfield J, MacAllister R. Ischemic preconditioning prevents endothelial injury and systemic neutrophil activation during ischemia-reperfusion in humans *in vivo*. *Circulation* 2001;**103**:1624–1630.
3. Kharbanda RK, Mortensen UM, White PA, Kristiansen SB, Schmidt MR, Hoschitzky JA, Vogel M, Sorensen K, Redington AN, MacAllister R. Transient limb ischemia induces remote ischemic preconditioning *in vivo*. *Circulation* 2002;**106**:2881–2883.
4. Oliver JJ, Webb DJ, Newby DE. Stimulated tissue plasminogen activator release as a marker of endothelial function in humans. *Arterioscler Thromb Vasc Biol* 2005;**25**:2470–2479.
5. Rosenberg RD, Aird WC. Vascular-bed-specific hemostasis and hypercoagulable states. *N Engl J Med* 1999;**340**:1555–1564.
6. Newby DE, Wright RA, Labinjoh C, Ludlam CA, Fox KA, Boon NA, Webb DJ. Endothelial dysfunction, impaired endogenous fibrinolysis, and cigarette smoking: a mechanism for arterial thrombosis and myocardial infarction. *Circulation* 1999;**99**:1411–1415.
7. Robinson SD, Ludlam CA, Boon NA, Newby DE. Endothelial fibrinolytic capacity predicts future adverse cardiovascular events in patients with coronary heart disease. *Arterioscler Thromb Vasc Biol* 2007;**27**:1651–1656.
8. Newby DE, McLeod AL, Uren NG, Flint L, Ludlam CA, Webb DJ, Fox KA, Boon NA. Impaired coronary tissue plasminogen activator release is associated with coronary atherosclerosis and cigarette smoking: direct link between endothelial dysfunction and atherothrombosis. *Circulation* 2001;**103**:1936–1941.
9. Aspelin T, Eriksen M, Lindgaard AK, Lyberg T, Ilebakk A. Cardiac fibrinolytic capacity is markedly increased after brief periods of local myocardial ischemia, but declines following successive periods in anesthetized pigs. *J Thromb Haemost* 2005;**3**:1947–1954.
10. Valen G, Eriksson E, Risberg B, Vaage J. Reactive oxygen intermediates and ischemia-reperfusion injury release tissue plasminogen activator from isolated rat hearts. *Thromb Res* 1993;**71**:113–121.
11. Winnerkvist A, Wiman B, Valen G, Vaage J. Release of tissue plasminogen activator during reperfusion after different times of ischaemia in isolated, perfused rat hearts. *Thromb Res* 1996;**82**:533–542.
12. Schoots IG, Levi M, van Vliet AK, Declercq PJ, Maas AM, van Gulik TM. Enhancement of endogenous fibrinolysis does not reduce local fibrin deposition, but modulates inflammation upon intestinal ischemia and reperfusion. *Thromb Haemost* 2004;**91**:497–505.
13. Roelofs JJ, Rouschop KM, Claessen N, de Boer AM, Frederiks WM, Lijnen HR, Weening JJ, Florquin S. Tissue-type plasminogen activator modulates inflammatory responses and renal function in ischemia reperfusion injury. *J Am Soc Nephrol* 2006;**17**:131–140.
14. Treska V, Valenta J, Pecen L, Topolcan O. Endogenous fibrinolysis in patients with lower extremity ischemia. *Ann Vasc Surg* 2000;**14**:356–359.
15. Killewich LA, Gardner AW, Macko RF, Hanna DJ, Goldberg AP, Cox DK, Flinn WR. Progressive intermittent claudication is associated with impaired fibrinolysis. *J Vasc Surg* 1998;**27**:645–650.
16. Newby DE, Wright RA, Ludlam CA, Fox KA, Boon NA, Webb DJ. An *in vivo* model for the assessment of acute fibrinolytic capacity of the endothelium. *Thromb Haemost* 1997;**78**:1242–1248.
17. Garlid KD, Paucek P, Yarov-Yaroyov V, Murray HN, Darbenzio RB, D'Alonzo AJ, Lodge NJ, Smith MA, Grover GJ. Cardioprotective effect of diazoxide and its interaction with mitochondrial ATP-sensitive K<sup>+</sup> channels. Possible mechanism of cardioprotection. *Circ Res* 1997;**81**:1072–1082.
18. Broadhead MW, Kharbanda RK, Peters MJ, MacAllister RJ. KATP channel activation induces ischemic preconditioning of the endothelium in humans *in vivo*. *Circulation* 2004;**110**:2077–2082.
19. Muldowney JA III, Painter CA, Sanders-Bush E, Brown NJ, Vaughan DE. Acute tissue-type plasminogen activator release in human microvascular endothelial cells: the roles of Galphaq, PLC-beta, IP3 and 5,6-epoxyeicosatrienoic acid. *Thromb Haemost* 2007;**97**:263–271.
20. Gudmundsdottir IJ, Lang NN, Boon NA, Ludlam CA, Webb DJ, Fox KA, Newby DE. Role of the endothelium in the vascular effects of the thrombin receptor (protease-activated receptor type 1) in humans. *J Am Coll Cardiol* 2008;**51**:1749–1756.
21. Zhong B, Wang DH. TRPV1 gene knockout impairs preconditioning protection against myocardial injury in isolated perfused hearts in mice. *Am J Physiol Heart Circ Physiol* 2007;**293**:H1791–H1798.
22. Newby DE, Sciberras DG, Mendel CM, Gertz BJ, Boon NA, Webb DJ. Intra-arterial substance P mediated vasodilatation in the human forearm: pharmacology, reproducibility and tolerability. *Br J Clin Pharmacol* 1997;**43**:493–499.
23. Jaffe R, Dick A, Strauss BH. Prevention and treatment of microvascular obstruction-related myocardial injury and coronary no-reflow following percutaneous coronary intervention: a systematic approach. *JACC Cardiovasc Interv* 2010;**3**:695–704.
24. Gibson CM, Karha J, Murphy SA, James D, Morrow DA, Cannon CP, Gugliano RP, Antman EM, Braunwald E. Early and long-term clinical outcomes associated with reinfarction following fibrinolytic administration in the Thrombolysis in Myocardial Infarction trials. *J Am Coll Cardiol* 2003;**42**:7–16.
25. Grines C, Patel A, Zijlstra F, Weaver WD, Granger C, Simes RJ. Primary coronary angioplasty compared with intravenous thrombolytic therapy for acute myocardial infarction: six-month follow up and analysis of individual patient data from randomized trials. *Am Heart J* 2003;**145**:47–57.

26. Keeley EC, Boura JA, Grines CL. Primary angioplasty vs. intravenous thrombolytic therapy for acute myocardial infarction: a quantitative review of 23 randomised trials. *Lancet* 2003;**361**:13–20.
27. Nanobashvili J, Neumayer C, Fuegl A, Blumer R, Prager M, Sporn E, Polteraer P, Malinski T, Huk I. Development of 'no-reflow' phenomenon in ischemia/reperfusion injury: failure of active vasomotility and not simply passive vasoconstriction. *Eur Surg Res* 2003;**35**:417–424.
28. Heusch G, Kleinbongard P, Bose D, Levkau B, Haude M, Schulz R, Erbel R. Coronary microembolization: from bedside to bench and back to bedside. *Circulation* 2009;**120**:1822–1836.
29. Reffelmann T, Kloner RA. The no-reflow phenomenon: a basic mechanism of myocardial ischemia and reperfusion. *Basic Res Cardiol* 2006;**101**: 359–372.
30. Woo KS, Armiger LC, White HD, Norris RM. Can streptokinase produce beneficial effects additional to coronary recanalization? Quantitative microvascular analysis of critically injured reperfused myocardium. *Microvasc Res* 2000;**60**: 8–20.
31. Sezer M, Oflaz H, Goren T, Okcular I, Umman B, Nisanci Y, Bilge AK, Sanli Y, Meric M, Umman S. Intracoronary streptokinase after primary percutaneous coronary intervention. *N Engl J Med* 2007;**356**:1823–1834.
32. Okorie MI, Bhavsar DD, Ridout D, Charakida M, Deanfield JE, Loukogeorgakis SP, Macallister RJ. Postconditioning protects against human endothelial ischaemia-reperfusion injury via subtype-specific KATP channel activation and is mimicked by inhibition of the mitochondrial permeability transition pore. *Eur Heart J* 2011;**32**:1266–1274.
33. Richard V, Kaeffer N, Tron C, Thuillez C. Ischemic preconditioning protects against coronary endothelial dysfunction induced by ischemia and reperfusion. *Circulation* 1994;**89**:1254–1261.
34. Laskey WK, Yoon S, Calzada N, Ricciardi MJ. Concordant improvements in coronary flow reserve and ST-segment resolution during percutaneous coronary intervention for acute myocardial infarction: a benefit of postconditioning. *Catheter Cardiovasc Interv* 2008;**72**:212–220.
35. Heusch G, Schulz R. Preservation of peripheral vasodilation as a surrogate of cardioprotection? The mechanistic role of ATP-dependent potassium channels and the mitochondrial permeability transition pore. *Eur Heart J* 2011;**32**: 1184–1186.
36. Brown NJ, Gainer JV, Murphey LJ, Vaughan DE. Bradykinin stimulates tissue plasminogen activator release from human forearm vasculature through B(2) receptor-dependent, NO synthase-independent, and cyclooxygenase-independent pathway. *Circulation* 2000;**102**:2190–2196.
37. Pedersen CM, Cruden NL, Schmidt MR, Lau C, Botker HE, Kharbanda RK, Newby DE. Remote ischemic preconditioning prevents systemic platelet activation associated with ischemia-reperfusion injury in humans. *J Thromb Haemost* 2011;**9**:404–407.