Liver repopulation with bone marrow derived cells improves the metabolic disorder in the Gunn rat

Citation for published version:

Digital Object Identifier (DOI):
10.1136/gut.2007.127969

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Gut

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and/or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Liver repopulation with bone marrow derived cells improves the metabolic disorder in the Gunn rat

M Muraca, C Ferraresso, M T Vilei, A Granato, M Quarta, E Cozzi, M Rugge, K A Pauwelyn, M Caruso, I Avital, D Inderbitzin, A A Demetriou, S J Forbes, G Realdi

Background: Reversible ischaemia/reperfusion (I/R) liver injury has been used to induce engraftment and hepatic parenchymal differentiation of exogenous β2-microglobulin+/Thy1− bone marrow derived cells. Aim: To test the ability of this method of hepatic parenchymal repopulation, theoretically applicable to clinical practice, to correct the metabolic disorder in a rat model of congenital hyperbilirubinaemia.

Methods and results: Analysis by confocal laser microscopy of fluorescence labelled cells and by immunohistochemistry for β2-microglobulin, 72 hours after intraportal delivery, showed engraftment of infused cells in liver parenchyma of rats with I/R, but not in control animals with non-injured liver. Transplantation of bone marrow derived cells obtained from GFP-transgenic rats into Lewis rats resulted in the presence of up to 20% of GFP positive hepatocytes in I/R liver lobes after one month. The repopulation rate was proportional to the number of transplanted cells. Infusion of GFP negative bone marrow derived cells into GFP positive transgenic rats resulted in the appearance of GFP negative hepatocytes, suggesting that the main mechanism underlying parenchymal repopulation was differentiation rather than cell fusion. Transplantation of wild type bone marrow derived cells into hyperbilirubinuric Gunn rats with deficient bilirubin conjugation after I/R damage resulted in 30% decrease in serum bilirubin, the appearance of bilirubin conjugates in bile, and the expression of normal UDP-glucuronyltransferase enzyme evaluated by polymerase chain reaction. Conclusions: I/R injury induced hepatic parenchymal engraftment and differentiation into hepatocyte-like cells of bone marrow derived cells. Transplantation of bone marrow derived cells from non-affected animals resulted in the partial correction of hyperbilirubinemia in the Gunn rat.
the liver (corresponding to about 30% of total liver mass) was occluded with an atraumatic vascular clamp for 60 minutes. The body temperature was maintained at 37 ± 0.3°C by a heating lamp. Reperfusion was initiated by removal of the clamp. Under these experimental conditions, a significant increase in necrotic and apoptotic hepatocytes is observed one hour after reperfusion. The animals received 1 ml of sterile saline intraperitoneally, the wound was closed in layers with 4-0 silk and wound staples, and the rats were allowed to recover. Twenty four hours after reperfusion, the animals were reanaesthetised, the abdomen reopened, and BMDCs suspended in 0.9% saline solution were seeded in the test animals through the portal vein into the right lobe of the liver. Control rats received saline solution alone.

Evaluation of liver parenchymal repopulation

The degree of hepatic engraftment achieved was monitored by two different criteria: fluorescent staining of BMDCs with a membrane bound dye, and genetic labelling (GFP-transgenic animals). A description of the experimental groups is given in table 1.

Fluorescent staining of BMDCs

BMDCs were stained with PKH26 red fluorescent cell linker (6 μmol/l). Three experimental groups of Lewis inbred male rats were studied: group A (n = 9), animals with intraportal infusion of 5 × 10⁶ BMDCs in the absence of I/R injury; group B (n = 16), animals with intraportal infusion of BMDCs following I/R injury; group C (n = 6), animals with I/R injury without BMDCs infusion.

GFP labelled cells

Two different experimental protocols were used:

First series (GFP positive cells into GFP negative animals)

BMDCs were isolated from GFP positive transgenic rats and transplanted into Lewis rats. This series included four experimental groups: group D (n = 5), rats with I/R injury and transplantation of 7 × 10⁶ BMDCs, and control group E (n = 3), rats with I/R injury without cell transplantation (both groups were studied 72 hours after cell infusion). Group F (n = 7), rats with I/R injury and transplantation of different amounts of BMDCs (1 × 10⁶, n = 2; 7 × 10⁶, n = 3; and 10 × 10⁶, n = 2, respectively) and control group G (n = 6), rats with I/R injury without cell transplantation. In group F, two female rats received 10 × 10⁶ BMDCs isolated from male animals, in order to use the Y chromosome as a second independent marker of transplanted cells. Both group F and group G were studied one month after cell infusion. All animals received daily immunosuppressive treatment with ciclosporine A, 15 mg/kg.

Second series (GFP negative cells into GFP positive animals)

To evaluate the occurrence of fusion, 7 × 10⁶ GFP negative BMDCs were infused into each GFP positive animal following I/R (group H, n = 6), while controls received vehicle only (group I; n = 4). In these conditions, differentiation of BMDCs should produce GFP negative hepatocytes, while fusion between haematopoetic cell lineages and liver parenchymal cells should result in GFP positive hepatocytes. The animals were studied one month after cell infusion. All animals received daily immunosuppressive treatment with ciclosporine A, 15 mg/kg.

Collection of samples

The animals were killed by exsanguination under anaesthesia and liver samples were collected from the postischaemic and the non-ischaemic lobes. Samples were either fixed in phosphate buffered formalin for immunohistochemical analysis or frozen in dry ice and stored at ~80°C until fluorescent microscopy analysis.

Histology, immunohistochemistry, and FISH

Liver tissue was fixed in 10% formalin and embedded in paraffin. Sections 5 μm thick were cut and stained with haematoxylin-eosin for morphological analysis and for counting the fluorescent stained cells. Immunohistochemistry was undertaken with antibodies against β₂m (Santa Cruz, Santa Cruz, California, USA), GFP (Abcam, Cambridge, UK), or CK18 (Progen, Heidelberg, Germany). Fluorescent stained cells were analysed using confocal laser microscopy. At least 500 hepatocytes were counted per animal, and the number of fluorescent cells was expressed as per cent of counted hepatocytes. Tissue sections were deparaffinised, rehydrated, and either treated for 15–20 minutes at room temperature with 0.06% Triton X-100 and 0.1% H₂O₂ for β₂m and GFP antibodies, or microwaved at 750 W for 10 minutes in 10 mM citrate buffer, pH 6.0, for CK18 antibody, and cooled down for at least 45 minutes. After washing in phosphate buffered saline (PBS), sections were incubated with normal serum in PBS for one hour at room temperature and for 30 minutes in an avidin/biotin blocking kit (Vector, Vinci-Biochem, Firenze, Italy). Sections were then incubated with the primary antibodies for two hours at 37°C (β₂m 1:20; CK18 1:20) or for one hour at 20°C (GFP 1:50).

<table>
<thead>
<tr>
<th>Group</th>
<th>Procedure</th>
<th>Harvest</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Transplantation of fluorescent labelled BMDCs (PKH26)</td>
<td>72 h</td>
<td>9</td>
</tr>
<tr>
<td>B</td>
<td>5 × 10⁶ BMDCs ip in the absence of I/R injury</td>
<td>72 h</td>
<td>16</td>
</tr>
<tr>
<td>C</td>
<td>Vehicle only after I/R injury</td>
<td>72 h</td>
<td>6</td>
</tr>
<tr>
<td>D</td>
<td>Transplantation of GFP positive BMDCs</td>
<td>72 h</td>
<td>5</td>
</tr>
<tr>
<td>E</td>
<td>7 × 10⁶ BMDCs after I/R injury</td>
<td>72 h</td>
<td>3</td>
</tr>
<tr>
<td>F</td>
<td>Vehicle only after I/R injury</td>
<td>1 m</td>
<td>7</td>
</tr>
<tr>
<td>G</td>
<td>1 × 10⁶, 7 × 10⁶, and 10 × 10⁶ BMDCs after I/R injury</td>
<td>1 m</td>
<td>6</td>
</tr>
<tr>
<td>H</td>
<td>Transplantation of BMDCs from wild type into GFP positive rats</td>
<td>1 m</td>
<td>6</td>
</tr>
<tr>
<td>I</td>
<td>Vehicle only after I/R injury</td>
<td>1 m</td>
<td>4</td>
</tr>
<tr>
<td>K</td>
<td>Transplantation of BMDCs from wild type into Gunn rats</td>
<td>1 m</td>
<td>6</td>
</tr>
<tr>
<td>L</td>
<td>Vehicle only after I/R injury</td>
<td>1 m</td>
<td>6</td>
</tr>
</tbody>
</table>

BMDC, bone marrow derived cell; GFP, green fluorescent protein; ip, intraportal; I/R, ischaemia/reperfusion.
room temperature (GFP 1:1200). The biotin conjugated secondary antibody was applied followed by incubation with peroxidase-streptavidin reagent, both for 30 minutes at room temperature (peroxidase vectastain elite ABC kit universal for GFP and CK18; peroxidase vectastain ABC kit, goat IgG, for β2m (Vector, Vinci-Biochem)). The immunoreaction was visualised by developing sections with diaminobenzidine (Dako Cytomation, Milan, Italy). Controls consisted of incubation with normal serum instead of the primary antibody and of tissue sections expressing or not the specific antigens.

**Dual staining protocol GFP/β2m**
The anti-GFP antibody was detected by biotin labelled swine anti-rabbit (1:400, Dako, Glostrup, Denmark) after avidin/biotin and serum blocking, followed by streptavidin-alkaline phosphatase (1:50, Dako) and visualisation with Alexa 488 (Invitrogen, Paisley, UK). The goat polyclonal anti-β2m antibody (1:20) was used after avidin/biotin and protein blocking followed by donkey anti-goat labelling with vector red (Vector Labs, Burlingame, California, USA). DAPI (4’6-diamidino-2-phenylindole) counterstaining was carried out before mounting.

**Dual staining protocol, GFP/albumin**
The polyclonal rabbit anti-albumin antibody (1:2000, Dako) was used, after the blocking steps, followed by biotin labelled swine anti-rabbit, streptavidin-alkaline phosphatase, and vector red development, as above. Following multiple washing steps and blocking with avidin/biotin and total protein block (Dako) the GFP antibody was applied (with PBS/normal goat serum) followed by goat anti-rabbit Alexa 488 (Invitrogen). DAPI counterstaining was carried out as above.

At least 500 hepatocytes were counted at high power field (200×), and the presence of BMDC derived hepatocyte-like cells (GFP positive or negative) was confirmed by examination at 400×. The number of BMDC derived hepatocyte-like cells was expressed as per cent of total counted hepatocytes.

The bone marrow origin of hepatocyte-like cells was confirmed by fluorescent in situ hybridisation (FISH) for the Y chromosome using a rat Y/12 chromosomal paint (Y-Cy3; 12-FITC, CA-1631, Cambio, Cambridge, UK) combined with immunohistochemistry for GFP. Sections were dewaxed and then microwaved for 10 minutes in sodium citrate (2.94 g/l). The primary antibody was an anti-GFP (1:500) (catalogue No...
A11122, Molecular Probes, Eugene, Oregon, USA) and the secondary antibody was an Alexa Fluor 647-anti-rabbit (Molecular Probes). Sections were incubated in 1 M sodium thiocyanate for 10 minutes at 80°C, washed in PBS, and then digested in pepsin (0.4% wt/vol) in 0.1 M HCl, at 37°C for four minutes. After quenching in glycine (0.2% vol/vol), the sections were rinsed in PBS, post-fixed in parafomaldehyde (4% wt/ vol), dehydrated through graded alcohols, and air dried. The chromosome paints were added to the sections, sealed under glass with rubber cement, heated to 60°C for 10 minutes, and incubated overnight at 37°C. The slides were rinsed in 0.5×SSC (NaCl/sodium citrate) and then in PBS and mounted in Vectashield (Vector Laboratories) for confocal microscopy.

Correction of the metabolic disorder in the Gunn rat
Gunn j/j rats with congenital deficiency of bilirubin glucuron-onyltransferase were subjected to I/R injury. Forty eight hours after the procedure, the animals received an intraportal infusion of either 10×10^6 BMDCs isolated from wild type syngeneic animals (group K, n = 6) or saline solution (group L, n = 6). Blood was collected from the tail vein before surgery and 30 days after cells infusion. After 30 days, the animals were anaesthetised and bile was collected on ice and in the dark. Alkaline methanolysis—HPLC analysis of bile pigments in bile and plasma—was carried out as described previously. Liver tissue was obtained for RNA extraction as described below.

RNA extraction and PCR analysis
Total RNA was extracted from snap frozen liver tissue using Tri Reagent solution (Sigma-Aldrich, Milan, Italy), according to the manufacturer’s instructions. One microgram of total RNA was reverse transcribed with Moloney murine leukaemia virus reverse transcriptase (2.5 U/µl) in a final volume of 20 µl containing: buffer (500 mmol/l KCL, 100 mmol/l Tris-HCl, pH 8.3), MgCl_2 (5 mmol/l), dNTPs (1 mmol/l), random examiners (2.5 µmol/l), and RNase inhibitor (1 U/µl). After an incubation period of 10 minutes at room temperature, the cDNA synthesis was carried out in a thermalcycler (Perkin Elmer, Milan, Italy) for 30 minutes at 42°C.

Polymerase chain reaction (PCR) was carried out using the following specific primers that amplified a 600 base pair (bp) fragment of the sequence of rat UGT1A1, the only uridine diphosphate glucuronosyltransferase isoform that preferentially binds bilirubin: UGT1A1 forward 5'-GAA AGA TTA CCC CAG GCC CAT CAT GCC-3'; UGT1A1 reverse 5'-GCC AGG TCC AGA GGC TCG ATA GGA CG-3'. The specificity and efficiency of the PCR reaction was increased by using the Eppendorf HotMasterMix (Eppendorf, Milan, Italy), according to the manufacturer's instructions. The cycle employed was: two minutes at 94°C, 38 cycles of one minute at 95°C, 40 seconds at 58°C, and 40 seconds at 72°C, followed by seven minutes at 72°C, before ending at 4°C. PCR products were digested with MspI (Roche Diagnostics, Monza, Milan, Italy) and analysed on 7% acrylamide gel, visualised by ethidium bromide and then silver stained.

Statistical analysis
Analyses were done using the Mann–Whitney test (two tailed) and the correlation analysis in the GraphPad Prism software program (San Diego, California, USA).

RESULTS
PKH26 labelled BMDCs
More fluorescent cells were observed in livers of group B (rats that underwent I/R) than of sham operated group A rats (mean (SD): 0.28 (0.11) vs 0.04 (0.01) PKH26 labelled cells/100 hepatocytes; p<0.0001; fig 1). Immunohistochemistry for βM showed the presence of βM negative cells integrated in the liver parenchyma of I/R rats and having morphological features of hepatocyte differentiation (group B, fig 1). A few βM negative cells were occasionally observed in sinusoidal spaces. In group A, as well as in non-ischaemic liver lobes of group B, small βM cells were sporadically observed only inside portal branches or sinusoids (fig 1). No βM negative cells were observed in the non-infused controls (group C).

GFP labelled BMDCs
At immunohistochemistry, GFP was homogeneously expressed in liver parenchymal cells of transgenic animals. Seventy two hours after transplantation, GFP positive, CK18 positive hepatocyte-like cells were found in ischaemic lobes of Lewis rats with I/R at a frequency of 0.39 (0.31) (n = 5), as per cent of total hepatocytes in the liver (group D; fig 2, panels A, B, C, D), while no GFP positive cells were present in controls without cell infusion (group E). The frequency of repopulation at 72 hours with GFP labelled hepatocytes was comparable to that observed with PKH26 labelled cells (group A, 0.28 (0.11); group D, 0.39 (0.31); p>0.05). Immunofluorescence analysis showed that after 72 hours some GFP positive cells stained negative for βM (fig 3D). The extent of repopulation with GFP positive hepatocyte-like cells was further evaluated one month after transplantation (group F), and was found to be proportional to the number of infused cells (linear regression: r² = 0.9117; exponential growth: r² = 0.9575; fig 4), up to approximately 20% of total liver parenchymal cells after infusion of 10×10^6 BMDCs (fig 5, panels A to E). Immunofluorescence analysis showed that GFP positive hepatocyte-like (bone marrow derived) cells stained more weakly for albumin than the surrounding native hepatocytes (fig 6). GFP positive endothelial-like cells were also observed in the ischaemic lobes of Lewis rats one month after transplantation, while they were not found in non-ischaemic parenchyma (fig 5F). The use of a double marker (that is, FISH for Y chromosome and co-staining with anti-GFP antibody) further confirmed the bone marrow origin of the GFP positive liver parenchymal cells. Liver parenchyma of female rats receiving male BMDCs showed the presence of clusters of GFP positive hepatocytes incorporating the Y chromosome (fig 7). Again, no GFP positive cells were present in controls without cell infusion (group G). Infusion of BMDCs isolated from Lewis rats into GFP-transgenic rats was associated with the appearance of GFP negative, CK18 positive hepatocytes (group H, fig 8), while no GFP negative hepatocytes were observed in controls (group I). The proportion of repopulation of GFP negative hepatocytes in GFP positive livers infused with 7×10^6 BMDCs was 5.38 (1.49)% and, it was not significantly different from the percentage of GFP positive hepatocytes observed in GFP negative livers infused with the same quantity of cells (8.2 (1.9)%).

Gunn rats
Analysis of blood and bile bilirubin concentrations
Thirty days after cell transplantation, the level of plasma unconjugated bilirubin was 35% lower than in basal conditions in rats receiving cell infusions (group K), while it was unchanged in control animals (group L) (fig 9). The decrease in plasma bilirubin was associated with the appearance of monoconjugated bilirubin in bile (0.9 (0.19) µmol/l; n = 6), while no conjugates were detected in controls (fig 10). The alkaline methanolysis procedure adds specificity to the identification of bilirubin conjugates, as the trans-esterification reaction yielding bilirubin methylesters from bilirubin glucuronoxides takes place only in an alkaline environment. When plain methanol was substituted for alkaline methanol, the peak of bilirubin monomethylesters was selectively absent from the chromatogram. Such verification confirmed the presence of
bilirubin conjugates in the Gunn rats transplanted with BMDCs.

PCR analysis
PCR analysis of uridine diphosphate glucuronosyltransferase (UDP-GTA) gene expression followed by digestion with the restriction enzyme MvaI is illustrated in fig 11. The amplification product of the normal donor cDNA yields four bands (12, 67, 123, and 384 bp) after digestion. As the 12 bp band is lost during chromatography, only three bands are visible. The PCR product of Gunn rat cDNA lacks a restriction site on position 1271 for this enzyme, and therefore the digestion products yield

Figure 2  Bone marrow derived cells (BMDCs) isolated from green fluorescent protein (GFP) positive rats acquire morphological features and phenotypic markers of differentiated hepatocytes after transplantation into liver injured Lewis recipients (group D). Serial sections of livers removed 72 hours after cell transplantation were stained for GFP (A) (C) and for CK-18 (B) (D). GFP positive cells are also positive for the epithelial marker CK-18. Several GFP positive cells with hepatocyte-like morphology are visible. The arrow indicates a binucleated cell, a feature typical of hepatocytes. Some GFP positive cells are smaller and have smaller nuclei (arrowheads) than the surrounding host hepatocytes. (Original magnification ×40.)

The two figures in the lower panel are: (E) a liver section from a Lewis rat (group E, GFP negative control) and (F) a liver section from a GFP transgenic animal (GFP positive control), respectively. (Original magnification ×10.)

Figure 3  Sections of livers removed 72 hours after cell transplantation (group D) were stained for GFP (A, fluorescent green) and β2m (B, fluorescent red) combined with DAPI nuclear staining (C, fluorescent blue). Some smaller (presumably less differentiated) GFP positive cells stain negative for β2m (arrow) while some larger (presumably more differentiated) GFP positive cells stain positive (arrowhead) (D).
DISCUSSION

There has been much interest in cell therapy for liver metabolic disorders in the last decade, and several investigators have reported a significant clinical improvement following hepatocyte transplantation as an alternative to liver transplantation. Major limitations are the limited availability of human hepatocytes and the limited long term efficacy. In theory, the use of BMDCs as an alternative to hepatocytes could solve both problems. The present work indicates first, that a significant parenchymal repopulation with hepatocyte-like cells derived from BMDCs can be induced even in metabolic disorders not associated with increased parenchymal cell turnover; second, that probably such repopulation is mostly due to differentiation rather than to cell fusion; and third, that such repopulation results in partial correction of a congenital hepatic metabolic disorder.

The extent of parenchymal repopulation with exogenous bone marrow derived, GFP positive hepatocyte-like cells averaged 0.4% in the liver lobes submitted to I/R 72 hours after infusion, but it increased by more than 20-fold after one month, suggesting proliferation of the exogenous cells. The histological finding showing large clusters of GFP positive hepatocytes supports this hypothesis. Continuous propagation of stem/progenitor cells in the absence of ongoing liver damage can be driven by cell–cell competition, with increased proliferation and reduced apoptosis of the transplanted cells when

![Figure 4](image)

**Figure 4** Percentage of green fluorescent protein (GFP) positive hepatocytes (Hcs) one month after GFP positive bone marrow derived cell (BMDC) transplantation, as a function of the number of infused cells (group F; n=8).

three bands (311, 67, and 12 bp, the last one not visible). In Gunn j/j rats, which received infusion of BMDCs from non-affected, wild type animals, four bands are visible, including those derived from digestion of the normal enzyme.

![Figure 5](image)

**Figure 5** Bone marrow derived cells (BMDCs) isolated from green fluorescent protein (GFP) positive transgenic rats acquire morphological features and phenotypic markers of differentiated hepatocytes after transplantation into liver injured Lewis recipients (group F). Serial sections of livers removed four weeks after cell transplantation were stained for GFP (A) and for CK-18 (B). (C–E) Extensive repopulation of liver parenchyma with GFP positive hepatocyte-like cells four weeks after transplantation of $10^6$ GFP positive BMDCs (group F). Large clusters of GFP positive hepatocyte-like cells are mainly located in proximity of blood vessels. (F) GFP positive endothelial-like cells (arrows) are also visible in the ischaemic lobes. (Original magnification: (A), (B) ×20; (C), (D), (E) ×10; (F) ×40.)
compared with host hepatocytes. Previous work reported low or insignificant repopulation in several animal models of liver injury, with the notable exception of a study by Jang et al., which demonstrated 7.6% repopulation 48 hours after cell infusion. Possible explanations for such differences include the selected stem cell population, the model of liver damage (also involving the endothelium), direct intraportal infusion vs systemic administration, and the survival advantage of transplanted cells in a milieu of endogenous cells injured by the previous I/R injury.

Figure 6  Sections of livers removed one month after cell transplantation (group F) were stained for green fluorescent protein (GFP) (A, fluorescent green) and albumin (B, fluorescent red) combined with DAPI nuclear staining (C, fluorescent blue). GFP positive (bone marrow derived) hepatocyte-like cells stain more weakly or negatively for albumin when compared with the surrounding autologous (GFP negative) hepatocytes (D).

Figure 7  Confocal microscopy analysis of liver tissue of a female rat one month after infusion of bone marrow derived cells (BMDCs) isolated from male GFP-transgenic rats (A) (B). Tissue has been processed with fluorescent in situ hybridisation (FISH) for the Y chromosome (Cy3, red nuclear signals) and immunostained for GFP (Cy5 labelling, blue cytoplasmic stain). Nuclei are dark as there is no nuclear DAPI labelling. Clusters of GFP positive hepatocytes incorporating the Y chromosome signal are visible throughout the parenchyma (60×); interestingly there are frequently multiple Y signals in a nucleus, denoting polyploidalisation, a typical hepatocyte feature. (B) An enlarged section of the area indicated by an asterisk (*) in the panel (A). (C, D) Fluorescent microscopy of control liver tissue: here DAPI (blue) is used to label the nuclei as no cytoplasmic fluorochrome is used; again Y chromosome detection is with the Cy3 labelled chromosome paint (red). (C) Male control liver shows nuclear Y chromosome labelling (red signals). (D) Female control livers have no nuclear Y chromosome labelling.
The infused BMDC population was selected on the basis of negativity for B2m and of positivity for Thy-1, a haematopoietic stem cell marker shared by hepatic oval cells. B2m participates as an integral part in molecules of the major histocompatibility complex (MHC I). Indeed, BMDCs share MHC I negativity.

Fetal hepatoblasts as well as neuronal stem cells are negative for MHC class I antigens, suggesting that such a feature could be a general marker of stemness/immaturity. A subpopulation of B2m negative BMDCs was shown to differentiate into cardiomyocytes and to reconstitute the myocardium in a model of myocardial infarction. Interestingly, B2m negative cells appeared to be selected (or B2m downregulated) following transplantation of human cord blood stem cells in the liver of SCID mice. More recently, a population of B2m negative, c-Met positive cells was shown to differentiate into hepatocyte-like cells in culture. That B2m negative cells are the true population responsible for the liver parenchymal repopulation in the present work is suggested by the presence of B2m negative cells integrating in liver parenchyma and showing morphological signs of hepatocytic differentiation 72 hours after infusion.

I/R is a powerful stimulus for stem cell homing and repair of the injured tissue, probably mediated through the rapid induction of the stem cell factor in various tissues, including the myocardium, the retina, and the liver. Moreover, macrophage activation secondary to the ischaemic damage is associated with the release of cytokines, activating endothelial cells with release of adhesion molecules, facilitating the anchorage and translocation of circulating stem cells. Ischaemic damage is also associated with the induction of SDF-1, a key molecule in stem cell recruitment to injured organs and tissues. In addition, the endothelial damage associated with I/R injury might facilitate translocation and

---

**Figure 8** Immunohistochemistry (IHC) on serial sections for green fluorescent protein (GFP) (A) (C) (E) and for CK-18 (B) (D) (F) one month after infusion of GFP negative bone marrow derived cells (BMSC) into GFP positive transgenic rats (group H). Animals were subjected to hepatic ischaemia/reperfusion (I/R) injury before cell infusion. Several GFP negative (arrows), CK-18 positive hepatocytes are visible within the liver parenchyma, suggesting differentiation from GFP negative BMDCs into hepatocyte-like cells rather than cell fusion between BMDCs and host hepatocytes. (Original magnification: (A), (B) ×20; (C), (D), (E), (F) ×40.)

**Figure 9** Serum bilirubin immediately before and 30 days after bone marrow derived cell (BMDC) transplantation in Gunn rats. Black bars represent animals transplanted with 10 × 10^6 BMDCs from wild type non-jaundiced rats. Controls (white bars) received vehicle only. Data are expressed as means, error bars = standard deviation, n = 6. * Different from basal conditions, p<0.05.
parenchymal engraftment of infused cells. Indeed, BMDCs were invariably observed only within the blood vessels 72 hours after infusion into normal livers, while engraftment of β2m negative cells was evident at the same time in I/R injured livers (figs 1 and 3).

In pilot experiments, we did not observe significant homing of fluorescent labelled BMDCs when the cells were administered immediately after releasing the vascular clamps. Thus the activation of biochemical signals was necessary in order to induce homing and engraftment of BMDCs.

Previous studies demonstrating insignificant liver parenchymal repopulation with BMDCs involved either bone marrow transplantation with subsequent random migration to the liver of an unknown cell population or the intravenous systemic delivery of a selected cell phenotype. As the behaviour of stem cells is strongly determined by their niche, direct intraportal infusion of BMDCs might have contributed to the relatively high hepatic repopulation with bone marrow derived hepatocytes observed in the present study. The importance of the local concentration of transplanted cells is also supported by the finding of a positive relation between the number of administered cells and the extent of parenchymal repopulation (fig 4).

Studies with hepatocyte transplantation have shown that a metabolic liver deficiency can be corrected by transplanting as little as 2% of the total hepatocyte mass, 25–50% of which will probably engraft. According to our results in group F, transplantation of 10×10⁶ BMDCs results in 20% repopulation of the infused lobes, corresponding to about 30% of the total liver mass. Therefore the magnitude of the repopulation relative to the whole liver is about 6–7%. Such parenchymal repopulation with hepatocytes derived from healthy BMDCs was able to partially correct a metabolic liver disease, as demonstrated by the decrease in serum bilirubin associated with the appearance of conjugated bilirubin in the bile of Gunn rats. The metabolic improvement was relatively small when compared with previous reports on hepatocyte transplantation, where a similar repopulation rate with exogenous wild type hepatocytes in Gunn rats resulted in up to a 50% decrease in blood bilirubin. Collectively, these results confirm that even the appearance of a low metabolic activity can produce dramatic changes in serum bilirubin levels.

Our results do not exclude a fusion process as a possible mechanism partially contributing to the observed repopulation of liver parenchyma with “normal” hepatocytes. However, the extent of liver parenchymal repopulation with exogenous cells was similar in the GFP positive to GFP negative and in the GFP negative to GFP positive groups. Moreover, fusion was
described as a late phenomenon,\(^2\) while in the present study, evidence of hepatocytic differentiation of BMDCs was observed already 72 hours after infusion (figs 1 and 2), as in the observations reported by Jang et al in liver injured mice.\(^3\) More recent work further supports the hypothesis that, through subjection to local growth factors and cell–cell contact, the BMDCs may undergo changes in cell fate, mimicking the phenotype of the surrounding cells in the absence of cell fusion.\(^4\) Interestingly, the BMDCs had high levels of CK-18 expression, similar to oval cells.\(^5\) However, the expression of albumin was lower than in the surrounding indigenous non-bone-marrow derived hepatocytes (fig 6), suggesting that these hepatocyte-like cells were less metabolically competent, in keeping with a lower than expected metabolic effect in the Gunn rat.

A limitation of the present study is the short observation period following the metabolic improvement in treated animals. Even if exogenously derived cells showed normal appearance at histology one month after transplantation of BMDCs, we cannot exclude the possibility that longer observation times would result in improved expression of metabolic activity. Another important issue is the life span of transplanted BMDCs, which differentiated into hepatocyte-like cells and showed specific (although lower than expected) metabolic activity. These results provide preliminary proof-of-principle that such a procedure could be used as a therapeutic tool in patients with inherited disorders of liver metabolism.

**ACKNOWLEDGEMENTS**

We thank Professor Johan Fevery for helpful discussion. The financial support of Telethon–Italy (grant No GG02051), of the Italian Association for the Study of the Liver (AISF), and of the Regione Veneto (Centro Regionale per la Terapia Cellulare delle Malattie Metaboliche and grant No 03/03/01) is gratefully acknowledged.

**Authors’ affiliations**

M Muraca, M Caruso, Ospedale Bambino Gesù, Laboratory Medicine, Rome, Italy

C Ferrarossa, M T Vilei, A Granato, M Quarta, G Realdi, University of Padova, Internal Medicine, Padova, Italy

E Cozzi, University of Padova, Surgery, Padova, Italy

M Rugge, ICRCS-IOV University of Padova, Pathology, Padova, Italy

K A Pauwelyn, Catholic University of Leuven, Internal Medicine, Leuven, Belgium

I Avital, Memorial-Sloan Kettering Cancer Center, Surgery, New York, USA

D Inderbitzin, University of Bern, Transplantation and Abdominal Surgery, Bern, Switzerland

A A Demetrio, Cedars-Sinai Medical Center, Surgery, Los Angeles, USA

S J Forbes, The Tissue Fibrosis and Repair Laboratory MRC/University of Edinburgh Centre for Inflammation Research, The Queen’s Medical Research Institute, Edinburgh, UK

*CF, MTV, and AG contributed equally to this work.

Competing interests: None declared.

**REFERENCES**


www.gutnl.com


