Hyperuricaemia does not impair cardiovascular function in healthy adults

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Objective: To investigate the possibility that uric acid (UA) can impair endothelial function, an important surrogate for atherosclerosis.

Design: UA was administered locally or systemically to healthy adult men and women in a series of randomised placebo controlled studies. This temporarily raised serum UA concentrations, so that the potential effects of hyperuricaemia on mechanisms of cardiovascular disease could be studied.

Main outcome measures: The effects of UA administration on basal blood flow and responses to locally administered acetylcholine, sodium nitroprusside, and L-NAME-monomethylarginine were studied in the forearm vascular bed with venous occlusion plethysmography. The effects of hyperuricaemia on systemic vascular resistance, large artery compliance, and baroreflex sensitivity were examined by validated non-invasive techniques.

Results: UA administration caused a twofold increase in serum concentrations. However, there were no acute effects on haemodynamic variables, basal forearm blood flow, or nitric oxide dependent endothelial function.

Conclusion: Unlike other risk factors associated with endothelial dysfunction, acute exposure to high concentrations of UA does not impair cardiovascular function in healthy men. These findings do not support a causal link between hyperuricaemia and atherosclerosis.
years. Exclusion criteria were increased blood pressure (BP) (>160/100 mm Hg), clinical history of joint, kidney or cardiovascular disease, any medication being taken, regular tobacco use, serum creatinine > 110 µmol/l, or serum UA > 400 µmol/l.

**Intra-arterial drug administration**

The brachial artery of the non-dominant arm was cannulated with a 27 standard wire gauge steel needle (Cooper’s Needle Works Ltd, Birmingham, UK) under local anaesthesia by an aseptic technique. Vasoactive drugs were administered through a 16 gauge epidural catheter (Portex Ltd, Kent, UK) connected to an IVAC P1000 syringe pump (Alaris Medical Ltd, Hampshire, UK). Saline was infused for 30 minutes at the start of the study and for 20 minutes between drugs to establish baseline blood flow. Vasoactive drugs were infused for six minutes at each dose and the rate of infusion was kept constant at 1 ml/min throughout.

**Measurement of forearm blood flow**

Blood flow was measured in both forearms by venous occlusion plethysmography, as previously described. Measurements were taken during the last three minutes of each six minute infusion. The last five recordings were averaged to determine flow in each arm. Responses were expressed as percentage change from baseline, where the ratio of blood flow in the infused to that in the non-infused limbs was considered, to account for any systemic effects.

**Systemic vascular resistance**

BP was recorded in the dominant arm with a validated oscillometric device (HEM-705CP, Omron, Kyoto, Japan). Cardiac index was assessed by transthoracic bioimpedance (NCCOM3-R7, BoMed, Irvine, California, USA). The systemic vascular resistance index was calculated as the mean arterial pressure divided by the cardiac index.

**Pulse wave analysis**

The dominant radial artery pulse waveform was assessed by applanation tonometry (SPC-301 micromanometer, Millar Instruments, Houston, Texas, USA). A corresponding aortic pressure waveform was generated by pulse wave analysis software (SphygmoCor, PWV, Sydney, Australia). The augmentation index is a validated measure of large artery stiffness, calculated as the difference between the first and second central systolic BP peaks, expressed as a percentage of pulse pressure.

**Spontaneous BRS**

Study participants rested supine. Systolic BP was recorded continuously by a Portapres system (TNO, Amsterdam, the Netherlands) and an ECG was measured simultaneously. Signals from both devices were recorded over 15 minutes and analysed off line by Chart HRV software (ADInstruments, New South Wales, Australia). BRS was determined by two independent methods. Parallel increases or decreases in BP and pulse interval (PI) are thought to represent spontaneous baroreflex activity. Sequences of parallel increases or decreases over two or more consecutive beats were analysed and the resulting slope (ΔPI/ΔBP) was used to represent BRS by sequence analysis. Fast Fourier transformation of BP and P1 data gave the total spectral power of the variability of each and the formula (power_hv/power_P1) gave spontaneous BRS by spectral analysis.

**Measurement of biochemical variables**

Blood was collected in gel tubes (Sarstedt Ltd, Leicester, UK), allowed to clot, and centrifuged at 1000 g for 10 minutes. Serum was separated and UA concentration was determined by a colorimetric dry slide method (Vitros, Ortho-Clinical Diagnostics, Amersham, UK).

**Drugs and reagents**

UA and lithium carbonate (Ultrapure preparations, Sigma Chemical Company, Poole, UK) were reconstituted in a sterile dextrose solution (Baxter Healthcare, Norfolk, UK) and filtered (0.22 µm Millipex, Millipore, Molsheim, France). The drugs used were acetylcholine (ACH; CIBAVision-Opthalmics, Southampton, UK), sodium nitroprusside (SNP; David Bull Laboratories, Warwick, UK), and L-NMMA (L-N-monomethylarginine (L-NMMA; Calbiochem-Novabiochem, Nottingham, UK).

**Basal forearm blood flow**

Six healthy men aged 29 (4) years (SEM) were recruited to a two way randomised placebo controlled study. Participants underwent intra-arterial administration of saline for 20 minutes to establish baseline blood flow, followed by infusion of 0, 0.5, 1.0, 2.0, and 4.0 mg/min UA in 4% dextrose/0.1% lithium carbonate vehicle for six minutes at each dose and for 12 minutes at the maximum dose. Forearm blood flow was assessed at baseline and during each infusion.

**Local hyperuricaemia and endothelial function**

Ten healthy men were recruited to a two way randomised placebo controlled study. They underwent intra-arterial administration of saline for 30 minutes to establish baseline blood flow, followed by ACh 7.5, 15, and 30 mmol/min, SNP 2, 4, and 8 mmol/min, and L-NMMA 2 and 4 µmol/min, where the order of infusion of ACh and SNP was randomised between subjects. Drug infusions were separated by saline for 20 minutes to allow restoration of basal blood flow. Drugs were infused for six minutes at each dose. UA 2.0 mg/min in 4% dextrose/0.1% lithium carbonate vehicle or vehicle alone was co-infused locally. Effluent venous blood (5 ml) was collected from each forearm during infusion for UA measurement.

**Systemic hyperuricaemia and endothelial function**

Ten healthy men were recruited to a two way randomised placebo controlled study. An 18 standard gauge venous cannula was inserted into a suitable vein in each antecubital fossa under local anaesthetic. Participants underwent systemic administration of 1000 mg UA in 4% dextrose/0.1% lithium carbonate vehicle or vehicle alone over one hour through the cannula in the non-dominant forearm. Immediately after infusion, endothelial function was studied as described above. Venous blood (5 ml) was drawn from the non-infused forearm cannula at baseline, immediately after infusion, and one hour after infusion for UA measurement.

**Systemic haemodynamic variables**

Eight healthy men were recruited to a three way randomised placebo controlled study. An 18 standard gauge venous cannula was inserted into a suitable vein in each antecubital fossa under local anaesthetic. Portapres finger cuff and BoMed electrodes were applied. Participants rested supine for 30 minutes, then underwent systemic administration of 100 mg UA in 500 ml 4% dextrose/0.1% lithium carbonate vehicle, vehicle alone, or 0.9% saline over one hour through the non-dominant forearm cannula. Electrocardiogram and Portapres signals were recorded for BRS determination at baseline and at the end of the infusion. BP, cardiac index, and pulse wave analysis (PWA) were measured at baseline and at 15 minute intervals. Venous blood (5 ml) was drawn through the non-infused forearm cannula at baseline and immediately after infusion for UA measurement.
Data analysis and statistics

Numbers of study participants were determined to give at least 80% power to detect a 10% difference in the primary outcome variables (forearm blood flow response to ACh, augmentation index, and BRS). Responses were compared by two way analysis of variance and paired Student’s t tests, where appropriate. Significance was accepted at the 5% level in all cases. All values are reported as mean (SEM).

RESULTS

Table 1 shows baseline characteristics of the study participants.

Basal blood flow

Local administration of neither vehicle nor UA had any effect on basal forearm blood flow (fig 1). Local UA and vehicle administrations caused systemic UA concentrations to rise by 62 (13) and −4 (3) μmol/l, respectively (p < 0.001); 69 mg (about 410 mol/l) UA was administered to each subject and the mean volume of distribution was calculated to be 22.6 (2.0) l.

Local hyperuricaemia and endothelial function

Venous effluent UA concentrations in the infused and non-infused forearms were 384 (7) and 280 (1) μmol/l, respectively, during vehicle administration. This was an increase of 33 (3)% and of −1 (0)% during UA and vehicle administration, respectively (p < 0.001). Despite this, responses to ACh, SNP, and L-NMMA were unaltered in the forearm vascular bed (fig 2).

Systemic hyperuricaemia and endothelial function

Serum UA concentrations before, immediately after, and one hour after infusion were 227 (8), 534 (18), and 452 (11) μmol/l, respectively, for UA administration and 224 (27), 220 (27), and 217 (27) μmol/l, respectively, for vehicle administration. These were increases of 145 (19)% and of −4 (1)% for UA and vehicle, respectively (p < 0.001). Forearm blood flow responses to ACh, SNP, and L-NMMA were not altered by systemic hyperuricaemia (fig 3).

Systemic haemodynamic variables

Baseline versus post-infusion serum UA concentrations were 336 (12) v 350 (1) μmol/l, 370 (17) v 361 (17) μmol/l, and 370 (16) v 627 (23) μmol/l (p < 0.001) for saline, vehicle, and UA administration, representing increases from baseline of −4 (1)%, −2 (1)%, and 79 (7)%, respectively (p < 0.001). Augmentation index, central systolic BP, BRS by sequence analysis, and BRS by spectral analysis were not altered by systemic hyperuricaemia (table 2). The systemic vascular resistance index increased during all infusions, with a non-significant trend towards lower increases during UA infusion (table 2).

DISCUSSION

The importance of high serum UA concentration as a marker of increased cardiovascular risk has been recognised for more than 50 years.27 However, no biologically plausible causal link to atherosclerosis has been shown in vivo. In the current study, UA administration had no effect on basal forearm blood flow or response to L-NMMA, indicating that short lived hyperuricaemia does not have a direct impact on resting vascular tone or basal nitric oxide release. Lack of effect on endothelium dependent and endothelium independent vasodilator responses suggests that high UA concentrations do not affect vascular smooth muscle nitrate responsiveness or stimulate nitric oxide release in health.

There is a strong association between disease states characterised by loss of vascular nitric oxide activity and high serum UA concentrations. The present findings indicate that acute hyperuricaemia does not directly influence constitutive, or stimulated, nitric oxide liberation from the vascular endothelium. A previous study has shown that enhancing vascular nitric oxide bioavailability by L-arginine supplementation causes a reduction in circulating UA concentrations.13 Therefore, UA may be responsive to vascular nitric oxide activity, consistent with a non-causal association between endothelial dysfunction and increased serum UA concentrations. Furthermore, UA is an important intracellular free radical scavenger during metabolic stress,24 for example, in vascular smooth muscle and endothelial cells,25 and circulating concentrations are thought to be responsive to the local redox state.26 Therefore, it is possible that increased UA concentrations are a compensatory response, in view of the antioxidant properties of UA.

Hyperuricaemia coexists with impaired large artery compliance in several disease states characterised by reduced vascular nitric oxide bioavailability. UA had no impact on the large conduit vessels that determine pulse waveform conduction, suggesting that high concentrations are not causally linked to increased vascular stiffness. BRS, determined by two discrete methods,25 was unaffected by UA, consistent with the lack of effect on constitutive nitric oxide activity and large artery compliance, which are important vessel wall properties that can influence BRS. The lack of effect on BRS also indicates that baroreceptor function, and indeed cardiac sensitivity to autonomic outflow, was not affected by local

Table 1 Study subjects baseline characteristics

<table>
<thead>
<tr>
<th></th>
<th>Local UA and endothelial function</th>
<th>Systemic UA and endothelial function</th>
<th>Systemic UA and haemodynamic variables</th>
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<tbody>
<tr>
<td>Number/men</td>
<td>10/10</td>
<td>10/6</td>
<td>8/8</td>
</tr>
<tr>
<td>Age (years)</td>
<td>23 (1)</td>
<td>24 (1)</td>
<td>30 (4)</td>
</tr>
<tr>
<td>Systolic BP (mm Hg)</td>
<td>110 (4)</td>
<td>107 (6)</td>
<td>108 (3)</td>
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<tr>
<td>Diastolic BP (mm Hg)</td>
<td>68 (5)</td>
<td>71 (4)</td>
<td>72 (3)</td>
</tr>
<tr>
<td>Heart rate (beats/min)</td>
<td>59 (2)</td>
<td>58 (3)</td>
<td>60 (2)</td>
</tr>
<tr>
<td>Body mass (-kg/m²)</td>
<td>23 (1)</td>
<td>22 (1)</td>
<td>23 (0)</td>
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<tr>
<td>Creatinine (μmol/l)</td>
<td>84 (4)</td>
<td>84 (6)</td>
<td>86 (3)</td>
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<tr>
<td>Glucose (mmol/l)</td>
<td>4.6 (0.2)</td>
<td>4.8 (0.1)</td>
<td>4.7 (0.3)</td>
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<tr>
<td>Cholesterol (mmol/l)</td>
<td>4.1 (0.2)</td>
<td>4.1 (0.2)</td>
<td>4.8 (0.2)</td>
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<tr>
<td>Uric acid (μmol/l)</td>
<td>258 (13)</td>
<td>252 (14)</td>
<td>366 (12)</td>
</tr>
</tbody>
</table>

Data are mean (SEM). BP, blood pressure; UA, uric acid.
increases in UA concentration, irrespective of vessel wall conditions.

There are several important limitations of these findings. This series of acute, mechanistic studies did not address the effects of chronic exposure to increased serum UA concentrations, which may be a more important determinant of future cardiovascular risk. However, it is less feasible to examine chronic hyperuricaemia in a controlled manner because of the potential risks of joint and kidney disease. A further potential limitation is that the current studies addressed the effects of an acute increase of UA in a young, healthy population, free from major cardiovascular risk factors. Nonetheless, if UA were an independent causal risk factor for atherosclerosis, then its presence should be expected to impair endothelial function. Other atherosclerotic risk factors cause acute impairment of endothelial function in young, healthy people—for example, increased homocysteine concentrations after methionine administration\(^3^2\) or ingestion.

### Table 2

<table>
<thead>
<tr>
<th></th>
<th>Saline</th>
<th>Vehicle</th>
<th>UA</th>
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<tbody>
<tr>
<td></td>
<td>Baseline</td>
<td>Postinfusion</td>
<td>Baseline</td>
</tr>
<tr>
<td>SBP (mm Hg)</td>
<td>96 (3)</td>
<td>97 (3)</td>
<td>93 (2)</td>
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<tr>
<td>DBP (mm Hg)</td>
<td>61 (2)</td>
<td>65 (2)</td>
<td>63 (3)</td>
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<tr>
<td>HR (beats/min)</td>
<td>55 (3)</td>
<td>53 (3)</td>
<td>57 (4)</td>
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<tr>
<td>Alx (%)</td>
<td>−6.7 (6.0)</td>
<td>−3.1 (6.1)</td>
<td>−2.6 (6.0)</td>
</tr>
<tr>
<td>CI (/min/m(^2))</td>
<td>3.3 (0.2)</td>
<td>3.0 (0.2)</td>
<td>3.3 (0.4)</td>
</tr>
<tr>
<td>SVRI (au)</td>
<td>12.0 (1.0)</td>
<td>13.7 (0.9)</td>
<td>10.2 (1.5)</td>
</tr>
<tr>
<td>BRS(_{seq}) (ms/mm Hg)</td>
<td>22.3 (4.7)</td>
<td>21.2 (4.0)</td>
<td>19.7 (4.9)</td>
</tr>
<tr>
<td>BRS(_{spec}) (ms/mm Hg)</td>
<td>24.3 (5.8)</td>
<td>19.4 (3.4)</td>
<td>22.2 (6.6)</td>
</tr>
</tbody>
</table>

Data are mean (SEM).

Alx, augmentation index; au, arbitrary units; BRS\(_{seq}\), baroreflex sensitivity by sequence analysis; BRS\(_{spec}\), baroreflex sensitivity by spectral analysis; CI, cardiac index; DBP, diastolic blood pressure; HR, heart rate; SBP, systolic blood pressure; SVRI, systemic vascular resistance index.
of a fatty meal. Additionally, amelioration of established risk factors allows rapid restoration of endothelial function, including correction of hypertension, hypercholesterolaemia, or hyperhomocysteinaemia.

A further limitation is that the effects of raising UA concentrations on endothelial function were studied only in healthy men. Previous studies have shown that the relation between hyperuricaemia and increased cardiovascular risk is more apparent in women, in both unselected populations and those with established coronary artery disease. Additional research is required to investigate the effects of raised serum UA concentrations on endothelial function in women. Furthermore, the effects of raised serum UA concentrations merit further investigation in men and women with established cardiovascular risk factors.

The effects of raising UA concentrations may be modest because of the comparatively high background concentrations to which humans, as a species, are ordinarily exposed. Further work is required to establish the cardiovascular effects of lowering serum UA concentrations. Xanthine oxidase inhibitors—for example, allopurinol—cause a modest reduction in circulating UA concentrations. Xanthine oxidase activity produces hydrogen peroxide, an important source of free radicals in vivo, which confounds the relation between xanthine oxidase inhibitors and UA lowering. Methods that lower UA directly should allow more useful interpretation of any cardiovascular effects—for example, using urate oxidase, which causes rapid and substantial reductions in circulating UA concentrations.

In summary, high serum UA concentrations, at least in the acute setting, do not impair cardiovascular function in healthy men. These findings do not support a causal role for UA in the development of atherosclerosis.

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REFERENCES


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