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Myc Inhibits p27-Induced Erythroid Differentiation of Leukemia Cells by Repressing Erythroid Master Genes without Reversing p27-Mediated Cell Cycle Arrest\(^{\dagger,\ddagger}\)

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Inhibition of differentiation has been proposed as an important mechanism for Myc-induced tumorigenesis, but the mechanisms involved are unclear. We have established a genetically defined differentiation model in human leukemia K562 cells by conditional expression of the cyclin-dependent kinase (Cdk) inhibitor p27 (inducible by Zn\(^{2+}\)) and Myc (activatable by 4-hydroxy-tamoxifen). Induction of p27 resulted in erythroid differentiation, accompanied by Cdk inhibition and G1 arrest. Interestingly, activation of Myc inhibited p27-mediated erythroid differentiation without affecting p27-mediated proliferation arrest. Microarray-based gene expression indicated that, in the presence of p27, Myc blocked the upregulation of several erythroid-cell-specific genes, including NFE2, JUNB, and GATA1 (transcription factors with a pivotal role in erythropoiesis). Moreover, Myc also blocked the upregulation of Mad1, a transcriptional antagonist of Myc that is able to induce erythroid differentiation. Cotransfection experiments demonstrated that Myc-mediated inhibition of differentiation is partly dependent on the repression of Mad1 and GATA1. In conclusion, this model demonstrates that Myc-mediated inhibition of differentiation depends on the regulation of a specific gene program.

p27 was originally described as a negative regulator of cell-cycle progression through the inhibition of cyclin-dependent kinases (Cdk) (37, 46). However, p27 has also been involved in biological functions unrelated to cell cycle, including the differentiation of erythroid precursors (14, 48). Regarding he-
matopoiesis, p27 is expressed in CD34+ progenitor cells and in the primitive erythroid precursors (48, 55), but p27-deficient mice do not show gross abnormalities in the hematopoietic lineages (reviewed in reference 34).

A functional antagonism between Myc and p27 in proliferation has been well established: Myc and p27 loss cooperates in animal carcinogenesis models (31), and several reports demonstrate the ability of Myc to abrogate p27 function (7, 43, 51, 54) and expression (54). However, in sharp contrast to the information on the antagonism between Myc and p27 in proliferation, the possible Myc-p27 cross talk in differentiation has not been investigated.

The study of Myc effects on differentiation has been impaired by the complex array of pathways activated by cytokines and chemicals used as differentiation inducers in most model systems. We previously showed that induction of p27 results in erythroid differentiation of K562 (35), and here we have investigated the role of Myc in this genetically defined differentiation model. We found that Myc blocks p27-mediated differentiation but that it cannot rescue the p27-dependent proliferation arrest and Cdk inhibition. We also report that Myc blocks the upregulation of Mad1 and of a set of erythroid-differentiation-determining genes such as the transcription factor GATA1.

MATERIALS AND METHODS

Cell culture, differentiation assays, and transfections. K562 cells were grown in RPMI 1640 medium supplemented with 8% fetal calf serum and antibiotics. Kp27MER and KMERI4 sublines were generated electroporating the pHABE-MycER plasmid (28) into Kp27-5 and KMT sublines, which have been described previously (35). The Kp27pBP and KMTPBP sublines were generated by electroporating Kp27-5 and KMT with pHABE-𝑝𝑝𝑢𝑟𝑜𝑣𝑒𝑟𝑜 puro vector. A total of 107 cells were electroporated with 20 μg of DNA (260 V and 1 mFa in a Bio-Rad gene pulser) and selected with 1 μg of puromycin (Sigma)/ml. Cells (2.5 × 105 cells/ml) were treated with ZnSO4 to induce p27 expression and 4-hydroxy-tamoxifen (4HT; Sigma) to activate MycER. Unless otherwise stated, ZnSO4 and 4HT were used at 75 μM and 100 nM, respectively. Transient transfection of Kp27MER (80,000 cells) was carried out with Lipofectamine 2000 (Invitrogen) with 1.2 μg of pEF-GATA1 (50) and pEF-NFE2 (29) vectors or empty vector. Nucleoelectrons of K562 cells were performed in a Nucleofector (Amaxa) according to the manufacturer’s indications with pCEFL-Myc (32), pCEFL-Mad1 (constructed by inserting human Mad1 cDNA into the pCEFL vector), pCEFL-p27 (35), pCEFL-MycV394D, pME-MycD106-143 (49), CMV-MadMyc (6), pEF-GATA1, pEF-NFE2, and 0.25 μg of human insulin-like growth factor (GFP) vector (pmaxGFP, Amamx) to assess transfection efficiency.

Proliferation and cell cycle assays. Cell counting was performed in hemacytometer. For DNA synthesis assay, cells were pulsed with 30 μM bromodeoxyuridine (BrdU; Roche Applied Science) for 1 h, fixed in 90% ethanol, treated with RNase and HCl, and incubated with fluorescein isothiocyanate-conjugated anti-BrdU monoclonal antibody (Roche Applied Science) for 30 min as described previously (34). Cells were analyzed by flow cytometry (Excilor; BD Bio sciences). For cell cycle analysis, cells were resuspended in phosphate-buffered saline (PBS)–sodium citrate buffer containing 10 μg of bovine serum albumin/ml, 200 μg of RNase/ml, and 50 μg of propidium iodide (Sigma Chemical Co./ml. The cells were incubated at 37°C in darkness for 30 min and then analyzed by flow cytometry using CellQuest software.

RNA analysis. Total RNA was isolated by using an RNAeasy kit (Qiagen). For reverse transcription–PCR (RT-PCR), first-strand cDNA was synthesized from 1 μg of total RNA by using SuperScript II RNase reverse transcriptase (Invitrogen) with random primers. Quantitative PCR (qPCR) was performed with a QuantiTect Sybr green PCR kit (Qiagen). The sequences of the primers used and amplicon sizes are shown in the Table S1 in the supplemental material. The data were normalized to ribosomal protein S14 (RPS14) mRNA levels. RPS14 has not been described as the Myc target gene (www.mycancergene.org).

Luciferase reporter assays. Three million of Kp27MER cells were electroporated at 260 V and 1 mFa in a Bio-Rad electroporator with 3 μg of pGL2-MLuc reporter (24) and 1 μg of pRL-TK (Promega). After 24 h of incubation, cultures were split into aliquots and further incubated for 24 h with ZnSO4 and/or 4HT. Cells were lysed, and the luciferase activity was measured in duplicate by a dual-luciferase reporter gene assay system (Promega). The data were normalized against the Renilla luciferase activity.

Gene expression profiling. Biotinylated cRNA was obtained from total RNA and hybridized to Affymetrix HG-U133A chip in the Genomic Facility of Centro de Investigación del Cancer (Salamanca, Spain). The data analysis and hierarchical tree clustering were generated by using the dChip software (26; see also http://biosun1.harvard.edu/complab/dchip/). The expression data was filtered so as to include genes with expression changes ≥2.3-fold and with a signal difference ≥50 between compared samples. The analysis was performed with data from two independent experiments and RNA preparations of each treatment (control, ZnSO4, 4HT, and ZnSO4 + 4HT) in Kp27MER and K562. The interaction network for differentially expressed genes was generated by using Ingenuity Pathways Analysis software.

Immunofluorescence and immunoblotting. For immunofluorescence, Kp27MER cells were treated with ZnSO4 and/or 4HT for 12 and 24 h and immunostained with the anti-Myc and anti-p27 antibodies, as well as with DAPI (4′,6-diamidino-2-phenylindole) to stain nuclei. A total of 2 × 104 to 5 × 104 Kp27MER cells were centrifuged at 1,500 rpm for 5 min, resuspended in 8 μl of PBS, and dried on microslides. Cells were fixed for 15 min in 3.7% paraformaldehyde in PBS at room temperature and permeabilized with 0.5% Triton X-100 for 30 min. The cells were then incubated with primary (1 h at 4°C), washed, and incubated with secondary antibodies (1 h at room temperature). The primary antibodies used were anti-Myc rabbit polyclonal antibody and anti-p27 mouse monoclonal antibody (both diluted 1:25). Secondary antibodies were conjugated with Texas Red or FITC (Jackson Laboratories). Cells were mounted with antifading medium Vectashield (Vector Laboratories) with DAPI to visualize the nuclei. Images were recorded by using a Bio-Rad MRC 1024 confocal laser microscope. Immunoblots were performed as previously described (35). Blots were revealed with an ECL system (Amersham). The antibodies used are described in Table S2 in the supplemental material.

Immunoprecipitations and kinase activity assays. Protein extracts (500 μg per assay) were immunoprecipitated with 1 μg of anti-cyclin D2, cyclin D3, and Cdk2 and p27 antibodies (see Table S2 in the supplemental material). Immunoprecipitations and kinase assays were performed as described previously (35). Microarray data were deposited in the ArrayExpress database under accession number E-MEXP-1772.

RESULTS

Generation of K562 sublines with conditional expression of p27 and Myc. We previously reported that induction of p27 in K562 human myeloid cells resulted in G1 arrest and erythroid differentiation (35). We took advantage of this differentiation system to ask whether Myc interferes with p27-mediated differentiation and to study the possible mechanisms involved. For this purpose, we first generated cells with conditional expression of both p27 and Myc. Kp27-5 cells (which carry a Zn2+-inducible p27 allele) were stably transfected with an expression vector for the fusion protein MycER, where Myc activity is activated by 4HT (28). One of the transfectants, termed Kp27MER, was selected, and immunoblot analysis confirmed that the cells expressed MycER and retained p27 induction by Zn2+. The activation of MycER by 4HT was assessed, first, by the Myc downregulation with p27-mediated differentiation (35) and to study the possible mechanisms involved. For this purpose, we first generated cells with conditional expression of both p27 and Myc. Kp27-5 cells (which carry a Zn2+-inducible p27 allele) were stably transfected with an expression vector for the fusion protein MycER, where Myc activity is activated by 4HT (28). One of the transfectants, termed Kp27MER, was selected, and immunoblot analysis confirmed that the cells expressed MycER and retained p27 induction by Zn2+. The activation of MycER by 4HT was assessed, first, by the Myc downregulation with p27-mediated differentiation (35) and to study the possible mechanisms involved.
thyroid differentiation mediated by p27. Kp27MER cells were treated with ZnSO₄ to induce p27 and 4HT to activate Myc, and the erythroid differentiation was analyzed first by the fraction of cells containing hemoglobin as determined by the benzidine test. This fraction was reduced by 4HT in Kp27MER cells but not in control Kp27pBP cells (Fig. 2A). Myc effect was further confirmed by the expression of erythroid-cell-specific genes as glycophorin A (GYPA), erythropoietin receptor (EPOR), ζ-globin, and ε-globin. mRNA levels of these genes were induced by p27, and this induction was blocked by 4HT (Fig. 2B). Finally, Myc also reversed the morphological differentiation into erythroid phenotype induced by p27, as shown by the dramatic reduction in the number of basophilic erythroblast-like cells (Fig. 2C). Altogether, the results show that Myc inhibited p27-induced differentiation.

Myc does not antagonize the proliferation arrest mediated by p27. As proposed for other models, we explored the possibility that Myc could inhibit erythroid differentiation by maintaining the cells in a proliferative state. When p27 was induced with 75 μM Zn²⁺, Kp27MER cells underwent a rapid growth arrest, similar to that induced by p27 in control cells (Kp27pBP) (Fig. 3A). In agreement with the growth determinations, p27 induction was accompanied by S-phase depletion and G₁-phase arrest (Fig. 3B). Surprisingly, the activation of Myc by 4HT in these conditions did not reverse this arrest (Fig. 3A), and cell cycle analysis revealed the G₁-phase arrest provoked by p27 upon treatment with 75 μM Zn²⁺ was not modified by Myc (i.e., by 4HT treatment) (Fig. 3B). Treatment with lower Zn²⁺ concentrations (50 μM) resulted in moderate p27 levels (Fig. 3C) and a less severe proliferation arrest than with higher p27 levels (i.e., with 75 μM Zn²⁺) (Fig. 3A). Consistently, moderate p27 levels (i.e., with 50 μM Zn²⁺) induced a weaker differentiation, which was also blunted by Myc (not shown). In the presence of moderate p27 levels, however, Myc provoked a slight but consistent increase in proliferation (Fig. 3A). This effect depended on Myc since 4HT did not modify the proliferation rate of control Kp27pBP cells (Fig. 3A). Consistently, Myc provoked an increase in the fraction of cells in S phase when the cell cycle was analyzed (Fig. 3B). In agreement with this result, Myc reversed the moderate repression of the mitotic cyclin B1 in cells treated with 50 μM Zn²⁺, as determined by immunoblot analysis (Fig. 3D). In contrast, the ex-
pression of the erythroid gene γ-globin was similarly blunted by Myc in the presence of 75 or 50 μM ZnSO₄. The results were confirmed by measurements of DNA synthesis by BrdU incorporation assays. As expected, DNA synthesis was halted in Kp27MER cells treated with 75 or 50 μM ZnSO₄ (i.e., high p27 levels). Activation of Myc did not relieve this inhibition (Fig. 3E). However, DNA synthesis was not totally inhibited in cells treated with 50 μM ZnSO₄ (moderate p27 levels), and Myc induced a moderate increase in DNA synthesis (Fig. 3E). We conclude that Myc inhibits p27-induced erythroid differentiation without reversing the p27-mediated cell cycle arrest.

**Myc does not reverse p27-mediated inhibition of CdkS.** The best-defined biochemical activity of p27 is the inhibition of Cdk activity. Thus, we sought to determine whether Myc impaired this activity in the K562 model in conditions where Myc is blocking differentiation. We first compared the expression of cyclins D and Cdk4, Cdk6, and Cdk2 in cells arrested and differentiated by p27 (i.e., treated with ZnSO₄) and in cells with the differentiation blocked after Myc activation (i.e., treated with ZnSO₄ and 4HT) (Fig. 4A). p27 induced the accumulation of cyclins D, an effect likely due to protein stabilization since there was no upregulation of cyclins D mRNA (data not shown). Myc also induced a small increase in cyclin A, cyclin D2, Cdk4, and Cdk2. No significant changes were observed in parental K562 cells treated with ZnSO₄ or 4HT (results not shown).

In most models, the G₁-phase arrest by p27 is associated with retinoblastoma (RB) hypophosphorylation. Immunoblot experiments show that this was also the case in Kp27MER treated with ZnSO₄. Interestingly, in the presence of p27, most of the RB remained hypophosphorylated upon 4HT addition (Fig. 4B). We next analyzed the levels of RB phosphorylated in Ser780 and Thr821 using phospho-specific antibodies. Phosphorylation in Ser780 and Thr821 have been reported as specific for Cdk4/6 and Cdk2, respectively (25, 57). Immunoblot results confirmed that p27 profoundly repressed RB phosphorylation and that Myc could reverse none of them.

We next studied the kinase activity in cyclin D2 and cyclin D3 complexes, the most prevalent cyclin D forms in K562 (Fig. 4A and data not shown) using RB protein as substrate. In this setting, the kinase activity assayed is mostly due to Cdk2, Cdk4, and Cdk6. The kinase activity was dramatically inhibited by p27, and Myc only slightly reversed this inhibition (Fig. 4C), which is consistent with the small increase in phospho-RB observed in 4HT-treated cells (Fig. 4B). K562 cells are deficient in p15INK4B and p16INK4A (13), and thus these proteins cannot contribute to Cdk4/6 inhibition in this system. Since RB is also the substrate for Cdk2 kinase activity, we assayed the kinase activity after immunoprecipitation with anti-Cdk2 antibody and in the presence or absence of Myc (i.e., with or without 4HT treatment). The results showed a dramatic inhibition of Cdk2 activity in p27-expressing cells, which was unchanged by Myc (Fig. 4C). Thus, Myc cannot reverse the inhibition of Cdk4/6 and Cdk2 elicited by p27. The possibility still...
Cdk2, Cdk4, and Cdk6 were found in p27 immunoprecipitates, and Myc did not modify their levels (Fig. 4D). However, Myc activation also resulted in greater amounts of cyclins D. Thus, we also tested the possibility that Myc modified the affinity of p27 for cyclin D-Cdk4/6 complexes. However, Myc activation did not change the relative amounts of p27, Cdk4, and Cdk6 present in cyclin D2 and D3 immunoprecipitates (Fig. 4E) and correlated to the amounts present in total extracts (Fig. 4A and data not shown). The results are again consistent with the lack of recovery in Cdk activity (Fig. 4C). Taken together, the data demonstrate that Myc inhibited p27-induced erythroid differentiation without reversing the p27-dependent inhibition of Cdk5, a result that is consistent with the inability of Myc to reverse p27-mediated proliferation arrest.

Myc antagonizes the upregulation of erythroid-cell-specific genes mediated by p27. Since Myc inhibited Kp27MER differentiation without rescuing the cells from the G1 arrest imposed by p27, this model provided an ideal opportunity to identify proliferation-independent targets of Myc that may explain its activity as a differentiation inhibitor. We therefore carried out microarray analysis with the Affymetrix platform testing RNA from Kp27MER cells either untreated or treated for 12 h with ZnSO4, 4HT, or both. The experimental conditions and associated phenotypes, as well as the number of regulated genes, are schematized in Fig. 5A. We focused in the genes regulated by Myc in the presence of p27, comparing the transcriptomes of Kp27MER treated with ZnSO4 with cells treated with ZnSO4 plus 4HT. After subtracting the genes changed in control samples, filtering, and statistical analysis, we found that Myc regulated 200 genes with ≥2.3-fold change, being 121 genes downregulated (see Table S3 in the supplemental material). The clustering analysis of these genes showed that all Myc-expressing cells cluster together (Fig. 5B). This data set was further analyzed with the Ingenuity Pathways software to reveal the network of interactions between differentially regulated genes in the Kp27MER cells after induction of Myc and thus assaying the possible relevance of Myc activation in this model. The results revealed that the top-ranked network had Myc at the most significant node of interactions among proteins whose expression levels (at the mRNA level) changed between cells treated with ZnSO4 and cells treated with ZnSO4 plus 4HT (Fig. 5C). This result argues that Myc is responsible for the phenotypic change of Kp27MER cells upon addition of 4HT.

Myc downregulates erythroid genes. Microarray analysis revealed that a significant fraction of the genes downregulated by Myc in the presence of p27 were erythroid cell related (see Table S4 in the supplemental material), and the same result was observed for other erythroid genes (e.g., ε-globin and ζ-globin). Interestingly, some of these genes downregulated by Myc encode transcription factors able to drive erythroid differentiation in cell culture, as well as in vivo models, such as GATA1, NFE2, STAT5A, STAT3, LMO2, LYL1, and JUNB. Moreover, Myc upregulated genes that block erythroid differentiation, such as the NOTCH ligand JAG2 and the transcription factor MAFA. Myc-mediated regulation of GATA1, NFE2, JUNB, and MAFA mRNA levels was confirmed by RT-qPCR (Fig. 6A). In contrast, Myc did not antagonize the p27-mediated downregulation of genes involved in DNA replication and mitosis, in agreement with the inability of Myc to

FIG. 4. Myc does not reverse Cdk inhibition by p27. (A) Expression of G1-phase cyclins and Cdks. Protein extracts of Kp27MER cells treated for 24 h with or without 4HT and ZnSO4 were assayed by immunoblotting for the indicated cyclins and Cdks. (B) Myc does not reverse RB hypophosphorylation. Total RB levels and phospho-specific RB levels were determined by immunoblotting in Kp27MER cells. See Table S2 in the supplemental material for antibody details. The positions of hypophosphorylated RB and hyperphosphorylated RB (pRB) are marked by arrowheads. (C) Cdk activity determination. Lysates were immunoprecipitated with antibodies to cyclin D2, cyclin D3, and Cdk2 and assayed for kinase activity using RB (for cyclins) or histone H1 (for Cdk2) as a substrate. The kinase activity was normalized to the amounts of cyclin D2, D3, and Cdk2 pulled down in the immunoprecipitates. (D) Composition of p27 complexes. Lysates from Kp27MER cells treated with ZnSO4 and/or 4HT were immunoprecipitated with anti-p27 antibodies, and the levels of p27, cyclin D1, D2, D3, Cdk6, Cdk4, and Cdk2 in the immunoprecipitates were assayed by immunoblotting. The corresponding nonimmunoprecipitated samples are shown in panel A. (E) Cyclin D2 and D3 immunoprecipitations. Cells were treated with ZnSO4 and/or 4HT, lysates were immunoprecipitated with cyclin D2 or cyclin D3 antibodies, and the levels of p27, Cdk4, and Cdk6 in the immunoprecipitates were assayed by immunoblotting.

existed that Cdk inhibition was a secondary effect of the p27-induced differentiation rather than the inhibitory interaction of p27 with cyclin-Cdk complexes. Thus, we studied the proteins associated with p27 by immunoprecipitation experiments.

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FIG. 5. Myc-regulated genes in the Kp27MER system. (A) Scheme of the Kp27MER samples subjected to microarray analysis and the resulting phenotypes. Cells were treated for 12 h with ZnSO4 and 4HT. The numbers of genes regulated (≥2.3-fold) are indicated in red. Gene expression changes between samples separated by the red line are subjected to clustering analysis in panel B. (B) Heat map of clustered samples and genes of expression data of duplicated samples of Kp27MER treated with ZnSO4 (i.e., p27) in the presence or absence of 4HT (i.e., Myc). The hierarchical clustering was performed for genes with expression variation ≥2.3-fold between cells expressing p27 and cells expressing p27 and Myc (P < 0.005), after subtraction of the corresponding controls in parental K562 cells. The genes are listed in the Table S3 in the supplemental material. The scale at the bottom shows the relationship between color saturation and the expression ratios. (C) Interaction network for differentially expressed genes in the presence of Myc and p27. A knowledge-based database (Ingenuity Pathways Analysis) was seeded with the differentially expressed genes between cells expressing p27 and cells expressing p27 + Myc shown in panel B. Genes in red were upregulated and those in gray were downregulated in the samples treated with 4HT (i.e., Myc). The figure represents the network with the highest score identified by the program and shows Myc (in blue) at a central node of the network. The meanings of node shape and lines are indicated at the bottom.
FIG. 6. Erythroid genes downregulated by Myc. (A) Kp27MER and K562 cells were incubated for 12 h with ZnSO₄ and/or 4HT, and mRNA levels of the indicated erythroid transcription factor genes were determined by RT-qPCR. Values represent means ± the SEM of three independent experiments. (B) Expression of erythroid genes in K562 cells after transient transfection of vectors for p27, Myc, Myc-V394D (MycVD), and Myc D106-143 (MycD106). The plasmids were nucleasefected, and the mRNA levels of GYPA, EPOR (24 h after transfection), or GATA1 and NFE2 (48 h after transfection) were determined by RT-qPCR. An expression vector for GFP was cotransfected in each case, and the data were normalized to the expression of GFP. Values are means ± the SEM from four independent experiments. (C) Enforced expression of GATA1 in Kp27MER. Cells were transfected by lipofection with GATA1 or empty vector. At 12 h after transfection, cells were treated with ZnSO₄ and/or 4HT. After 24 h of incubation, the expression of GATA1, p27, and Myc was analyzed by immunoblotting. The anti-GATA1 antibody used recognizes the N-terminal domain of mouse protein. (D) GYPA mRNA levels assayed by RT-qPCR in Kp27MER cells transiently transfected with GATA1. At 12 h after transfection the cells were treated for 72 h with ZnSO₄ and 4HT. After 24 h of incubation, the expression of GATA1, p27, and Myc was analyzed by immunoblotting. The anti-GATA1 antibody used recognizes the N-terminal domain of mouse protein. (E) GYPF mRNA levels assayed by RT-qPCR in Kp27MER cells transiently transfected with GATA1. At 12 h after transfection the cells were treated for 72 h with ZnSO₄ and 4HT. The mRNA level was determined by RT-qPCR. Values are means ± the SEM of two independent experiments. (F) Enforced expression of NFE2 in Kp27MER. Cells were transfected with an NFE2 expression vector or empty vector and analyzed by immunoblotting as in panel C. (G) ε-Globin mRNA levels of Kp27MER transfected with NFE2, assayed as in panel D. The data are means ± the SEM of two independent experiments.

it has been reported that Mad1 induces erythroid differentiation in Friend murine erythroleukemia cells (10). Thus, we sought to determine whether the functional antagonism between Myc and Mad1 could also contribute to explain the Myc-mediated inhibition of differentiation in the K562 system. We assayed the expression of Mad proteins in the Kp27MER system and found that Mad1 protein (Fig. 7A) and mRNA (Fig. 7B) was upregulated upon p27-induced differentiation. Interestingly, Mad1 upregulation was reduced when Myc was activated by 4HT (Fig. 7A and B). In contrast, other members of the Mad family such as Mnt and Mxi1 were not upregulated by p27 (Fig. 7A). Moreover, transient transfection of p27 in K562 resulted in an increase in Mad1 mRNA (not shown). We next sought to determine whether Mad1 could contribute to p27-induced differentiation in this model. Cells were transiently transfected with expression vectors for Mad1 and Myc, and the erythroid differentiation was assayed by the benzidine test and the expression of ε-globin and EPOR. The results showed that Mad1 increased the expression of differentiation markers (Fig. 7D and E). We also transfected the MadMyc construct, which carries the transrepression domain of Mad1 linked to the HLH-LZ domain of Myc and acts as a strong repressor of Myc-responsive transcription (6). MadMyc was transfected into K562, and its expression demonstrated by immunoblotting (Fig. 7C). We found that MadMyc efficiently induced differentiation as assessed by the upregulation of EPOR and ε-globin genes (Fig. 7E). Finally, coexpression of Myc blunted the differentiation induced by Mad1 and by Mad-Myc in transient-transfection experiments (Fig. 7C and D). We conclude that Mad1 upregulation is part of the differentiation response and that Myc also functions as a Mad1 antagonist in this system.

DISCUSSION

One of the first biological activities described for Myc was differentiation inhibition, namely, the chemically induced erythroid differentiation of FMEL cells (8, 16, 44). This effect is consistent with the erythroleukemia induced in transgenic mice with enforced Myc expression in erythroid precursors (47). However, the mechanisms by which Myc blocks differentiation are poorly understood. Proliferation stimulation can certainly contribute to Myc-mediated inhibition of differentiation, but the hypothesis that Myc exerts proliferation-independent mechanisms to block differentiation has received little attention. Several studies have shown that Myc antagonizes the cell cycle arrest effect of p27, and the prevalent mechanism proposed is the sequestration of p27 in type D cyclins complexes (7, 43, 51). In contrast, there is little information on whether and how Myc impairs p27-induced differentiation. In this context, it has been shown that the monocytic differentiation induced by retinoic acid in U937 myeloid cells is mediated by p27 and inhibited by Myc (15) and that inactivation of p27 gene rescues the cerebellar development otherwise impaired in N-Myc-deficient mice (59).

We have established a genetically defined model system where differentiation can be induced in K562 human myeloid leukemia cells by p27 induction and Myc can be conditionally activated. The molecular mechanisms by which p27 induces differentiation in this model are unknown. However, they do
not entirely depend on cell cycle arrest since p21 also arrests proliferation of K562 while inducing a different type of differentiation (34). In the present study we show that Myc impairs the erythropoiesis of K562 cells and Myc coexpression impairs Mad1 and MadMyc-induced differentiation. This suggests that the impairment of Mad1 function by Myc is contributing to differentiation inhibition induced by Myc, indicating that Myc effect is mediated, in part, by antagonizing GATA1 induction. However, GATA1 is only partly rescuing the anti-differentiation effect of Myc, since Myc is likely impairing differentiation by repressing several erythroid-determining genes.

The molecular mechanism by which Myc represses GATA1 is unclear. Actually, about half of Myc target genes are repressed, but the mechanism of repression is unknown for the vast majority of the genes. In a small subset of genes, Myc-mediated repression depends on the interaction with Miz1 (52). In the case of GATA1 in the K562 model, this effect seems to be Miz1-independent, since a Myc mutant unable to bind Miz1 antagonized the upregulation of GATA1 and erythroid differentiation as efficiently as did wild-type Myc. In contrast, a Myc mutant lacking Myc box II (a conserved region required for transformation, transactivation, and repression activities of Myc) cannot efficiently antagonize p27-induced differentiation in our model. This result has also been observed for other differentiation models (4, 20) and suggests that Myc is using a common molecular mechanism to transform and to abrogate differentiation. On the other hand, it has been reported that GATA1 represses Myc and induces p27 in murine embryonic cells (45), suggesting the possibility of a regulatory p27-GATA1 loop. However, GATA1 does not repress Myc in our K562 model (data not shown).

Myc and Mad1 have antagonist activities in transcription upon binding to the same E-boxes and in cell proliferation (5, 21). We found that p27-induced differentiation of K562 is accompanied by Mad1 upregulation and that Mad1 is able to promote K562 erythroid differentiation, although less efficiently than p27. Moreover, MadMyc, a chimeric protein that represses gene expression upon binding to Myc-binding sites (6), is a potent differentiation inducer. Thus, Myc would antagonize this Mad1 effect at the transcriptional level. Furthermore, Myc activation in our model provokes Mad1 downregulation in K562 cells and Myc coexpression impairs Mad1 and MadMyc-induced differentiation. This suggests that the impairment of Mad1 function by Myc is contributing to differentiation inhibition in the K562p27MER model. These results are consistent with Mad1 inducing erythroid differentiation of murine erythroleukemia cells (10). Moreover, it has been shown that Mad1 and p27 cooperate for a correct development of the myeloid compartment in vivo (33). Similar scenery seems to operate in K562 differentiation, and our data suggest that Myc might impair the Mad1-p27 functional interaction.

In summary, the results in the K562 model demonstrate that the differentiation-inhibitory effect of Myc depends on its gene regulatory activity and is uncoupled from its effect as proliferation stimulator. Importantly, tumor cell redifferentiation is the mechanism for the tumor regression after Myc deactivation in some transgenic mice models (reviewed in reference 3).
Moreover, Myc is one of the four transcription factor set capable of conferring pluripotent stem cell properties to differentiated adult cells, and Myc is reactivated in the tumors arising in the chimeric mice derived from such cells (38). Thus, it is conceivable that Myc displays common pathways as a "stem cell-ness" keeper and as an oncogene, preventing differentiation in both cases. Our results in the K562 model support the hypothesis that inhibition of cell differentiation is a Myc tumorigenic mechanism independent of Myc effects in cell proliferation.

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REFERENCES


