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A rare de novo nonsense mutation in OTX2 causes early onset retinal dystrophy and pituitary dysfunction

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Purpose: To describe the clinical findings of a patient with an early onset retinal dystrophy and a novel mutation in OTX2, and to compare these findings with previously reported cases.

Methods: Using direct sequencing, we screened 142 patients, who had either Leber congenital amaurosis (LCA) or early onset retinal dystrophy (EORD), for mutations in OTX2. All patients received a detailed ophthalmic examination including electroretinography and retinal imaging.

Results: Only one mutation in OTX2 was identified. A novel heterozygous p.S138X stop mutation was identified in a seven-year-old male who had an infantile onset retinal dystrophy. The mutation was not present in either parent or in 181 blood donor samples. There was a history of failure to thrive in infancy, poor feeding, and growth hormone deficiency. Poor vision and nyctalopia was present from the first year. Funduscopy revealed a hyperpigmented peripapillary ring with a fine granular pigmentation of the RPE throughout the fundus. The scotopic bright flash ERG a-wave was subnormal and the waveform electronegative, in keeping with dysfunction both at the level of the photoreceptor and postphototransduction. Visual function has been stable to date.

Conclusions: Mutations in OTX2 have been reported in association with major developmental malformations of the eye, with retinal dystrophies such as LCA, and with pituitary dysfunction and seizure activity in some cases. This case adds further support for a role of OTX2 both in retinal development and pituitary function, and highlights a novel retinal dystrophy phenotype seen in association with mutations in OTX2.

The orthodenticle protein homolog 2 (OTX2) is a homeobox gene that plays a critical role in retinal photoreceptor development. The gene, on chromosome 14q23.1, is organized into five exons, only three of which are translated. It contains a highly conserved homeodomain. OTX2 is expressed during development in the neuroepithelium of most of the forebrain and midbrain, including the eye domain [1]. Complete elimination of OTX2 function in mice by gene targeting results in the absence of photoreceptor function genes, including opsins. Ot2−/− mice can be normal, have developmental eye anomalies including anterior segment malformations, severe eye abnormalities such as microphthalmia or anophthalmia, or head abnormalities [2,4]. In early development Ot2, specifies the anterior neuroectoderm during gastrulation [2] and is expressed throughout the forebrain and midbrain in the developing embryo [1]. Once eye development is specified, expression of Ot2 narrows. It is required for the development of the presumptive retinal pigment epithelium (RPE) [5], where it has a continuing maintenance role throughout adulthood [6]. Analysis of Ot2−/− mice has shown that it is essential for specification of the RPE; loss of OTX2 function results in the outer layer of the optic cup developing into ectopic neural retina [7]. However, OTX2 is also required for the development and maintenance of the neural retina. Ot2 protein is found in bipolar and in ganglion cells [6,8,9], while in the outer nuclear layer it regulates the expression of the closely related Crx gene that controls the expression of a suite of photoreceptor function genes, including opsins. Ot2 and Crx can also bind to common DNA-binding sequences [10]. Ablation of OTX2 in retinal progenitor cells has been shown to affect RPE development and leads to a significant decrease in rod photoreceptors and bipolar cells, suggesting that OTX2 is also involved in retinal bipolar cell development [8]. It has recently been demonstrated that OTX2 protein is transported from the retina to the visual cortex in the developing rat, and appears to modify postnatal synaptic plasticity [9].

OTX2 mutations in humans have been associated with a range of ophthalmological phenotypes including anophthalmia, microphthalmia, developmental anomalies of the optic nerve and chiasm, and Leber congenital amaurosis (LCA). As OTX2 plays a key role in retinal development, and
a mutation in OTX2 has been reported in association with an infantile onset retinal dystrophy [11], we screened a large panel of patients with LCA and other forms of childhood onset retinal dystrophy for mutations in this gene.

METHODS

A panel of DNA from 142 patients with LCA and severe childhood onset retinal dystrophy were recruited at Moorfields Eye Hospital and The Hospital for Children, Great Ormond Street, London, as part of an ongoing molecular genetic study of childhood retinal dystrophies. The research has local research ethics committee approval and conformed to the tenets of the Declaration of Helsinki. A full clinical history was taken and examination performed in all subjects. In older children and adults this included, where possible, best corrected logMAR visual acuity and color vision (Hardy Rand Rittler, Richmond Products, Albuquerque, NM); slit-lamp biomicroscopy or indirect ophthalmoscopy; Goldmann visual fields; autorefraction (Luneau L62–3D autorefractometer - LUNEAU Chartres, France), axial length measurement, and auto-keratometry (IOL Master, Carl Zeiss Meditec, Dublin, CA); optical coherence tomography (Stratus OCT™ - software version 3; Carl Zeiss Meditec, Inc.); fundus photography (TRC-50IX with IMAGEnet 2000 system software–TOPCON Corporation, Japan), and fundus autofluorescence (Heidelberg Retinal Angiograph-II, Heidelberg Engineering, Heidelberg, Germany). All patients had electroretinography performed either at our institutions or at the referring hospital. Most patients had tests performed at diagnosis which, in the older patients, predated the ISCEV standards. Patients seen more recently, or in whom we wished to confirm the diagnosis, had electroretinography conducted in accordance with current International Society for Clinical Electrophysiology of Vision standards using gold foil or DTL electrodes [12,13]. ERG testing in infants and young children was performed using skin electrodes in accordance with previously published protocols [14,15].

DNA was extracted from whole blood obtained from each affected individual and immediate family members using the Nucleon BACC-2 kit (GE Healthcare) performed as per manufacturer’s instructions. DNA samples from all patients were sent to Asper Ophthalmics (Tartu, Estonia) for analysis using the LCA chip [16]. This is a microarray which contained (at the time of the study) 344 disease associated SNPs and several common variant polymorphisms in six LCA associated genes and 2 early onset genes (GUCY2D, CRX, RPE65, CRB1, RPGRIP1, AIPL1, LRAT, and MERTK). Genorama genotyping software (Asper Ophthalmics Ltd) can subsequently produce a bidirectional allele call at each SNP being interrogated. The panel subsequently underwent direct sequencing of PCR products [11] performed using an ABI 3130 36 cm capillary sequencer (Applied Biosystems, Inc., Foster City, CA). 181 control DNA samples isolated from anonymous Aberdeen-based blood donors were screened for DNA mismatches using the Idaho Technology Inc. Light Scanner (Salt Lake City, UT). Fragments revealing a nucleotide mismatch were fully sequenced to identify the mutations.

RESULTS

A de novo heterozygous c.413C>G p.S138X stop mutation was identified in a seven-year-old male (Figure 1; Pt 13424) who was negative for all variants assayed by the LCA chip. The mutation was not present in either parent. This mutation was not detected in 181 controls. Known polymorphic single nucleotide polymorphisms (SNPs), were observed at similar frequencies in the disease cohort and the blood donor controls, indicating that the ethnic background of the patient cohort and the controls was comparable.
The patient had a history of failure to thrive from birth and multiple investigations of the possible cause including investigations for Batten disease, Williams syndrome, and Russell-Silver dwarfism, all of which were negative. At 9 months old, required nasogastric feeding for 6 months following a history of failure to thrive. He has always been on the lowest percentiles for stature. Growth hormone deficiency was suggested indirectly via low levels of IGF1 and IGFBP3. An MRI scan was reported as normal. His parents noted poor vision and nyctalopia during his first year of life. There was no family history of eye disease. One younger sibling who has no visual problems was unavailable for further investigation.

Ophthalmic examination when he was six years old disclosed a small angle alternating esotropia with mild torsional nystagmus. Visual acuities were 0.4 and 0.5 logMAR in his right and left eyes, respectively. Cycloplegic retinoscopy revealed a refractive error of 0.00 diopter sphere/ +3.00 diopter cylinder at 180 degrees in the right eye, and +3.00 diopter sphere /+2.50 diopter cylinder at 180 degrees in the left. He had normal color vision, according to Hardy Rand Rittler; he had normal pupil reactions and clear lenses. Funduscopy revealed a hyperpigmented area of retina surrounding the optic disc with a fine granular pigmentation of the RPE throughout the fundus (Figure 2). There was no vascular attenuation.

ERGs were performed according to international standards [13]. The dark-adapted bright flash ERG was markedly electronegative with additional a-wave reduction. Light-adapted flicker ERGs were delayed and subnormal and the single flash cone ERG was subnormal with a reduced b:a ratio (Figure 3). The profoundly negative ERG suggests dysfunction that is both marked and at a level that is post-phototransduction, usually post-receptoral. The a-wave was subnormal but not delayed, indicating additional loss of photoreceptor function. The ON-OFF and S-cone ERGs [17, 18] suggesting greater ON- than OFF- bipolar pathway involvement. The pattern ERG P50 component was reduced bilaterally, which is consistent with macular involvement. (Figure 3)

**DISCUSSION**

This study provides evidence of a novel mutation in OTX2 associated with early onset retinal dystrophy and pituitary insufficiency. This mutation is predicted to lead a truncated protein. Chatelain et al. [19] observed that OTX2 truncation at residues 106 and 161, flanking the site of the predicted termination, described here, leads to loss of transactivation function. This strongly suggests that the mutation identified in our patient is also likely to cause haploinsufficiency.

Clinical studies have demonstrated that mutations in OTX2 are associated with a wide range of ocular phenotypes (Table 1) [7,11,20–22]. Experimentally, the role of OTX2 in regional specification of the eye, particularly the RPE, has long been established from expression studies in normal and mutant animal models [1,4,23–26]. Subsequent studies have also demonstrated the role of OTX2 in the development of both the retinal photoreceptors and bipolar cells [25,27]. It has been hypothesized that OTX2 is involved in retinal photoreceptor cell fate but not in bipolar cell fate in early development, while it is involved in the terminal differentiation of both...
photoreceptors and bipolar cells late in development in cooperation with CRX [8]. Pituitary dysfunction has been described in two patients with heterozygous 14q22–23 microdeletions involving OTX2 [28,29]. Recently, the role of OTX2 in pituitary development has been further delineated with three case reports of OTX2 mutations associated with pituitary hormone deficiency (CPHD) [21,22]. A further case of anophthalmia associated with an OTX2 mutation was reported to have isolated growth hormone deficiency and short stature [7]. In the latter publication a requirement for OTX2 binding was postulated at a site previously shown to lie in the promoter of HESX1 [30].

TABLE 1. PREVIOUSLY PUBLISHED MUTATIONS IN OTX2 WITH ASSOCIATED PHENOTYPES.

<table>
<thead>
<tr>
<th>Nucleotide</th>
<th>Protein</th>
<th>Phenotype</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>c.674A&gt;G</td>
<td>p.Asn225Ser</td>
<td>CPHD</td>
<td>[22]</td>
</tr>
<tr>
<td>c.674A&gt;G</td>
<td>p.Asn225Ser</td>
<td>CPHD</td>
<td>[22]</td>
</tr>
<tr>
<td>whole gene deletion</td>
<td>whole gene deletion</td>
<td>Extreme microphthalmia</td>
<td>[20]</td>
</tr>
<tr>
<td>whole gene deletion</td>
<td>whole gene deletion</td>
<td>Anophthalmia</td>
<td>[20]</td>
</tr>
<tr>
<td>c.93C&gt;G</td>
<td>p.Tyr31X</td>
<td>Microphthalmia</td>
<td>[20]</td>
</tr>
<tr>
<td>c.106dupC</td>
<td>p.Arg36ProfsX52</td>
<td>Microphthalmia OD/ normal OS</td>
<td>[20]</td>
</tr>
<tr>
<td>c.106dupC</td>
<td>p.Arg36ProfsX52</td>
<td>Anophthalmia OD/ Coloboma OS</td>
<td>[20]</td>
</tr>
<tr>
<td>c.373_374delAG</td>
<td>p.Gly126TrpfsX11</td>
<td>Anophthalmia</td>
<td>[20]</td>
</tr>
<tr>
<td>c.404_405dupCT</td>
<td>p.Ser136LeufsX43</td>
<td>Anophthalmia and CPHD</td>
<td>[21]</td>
</tr>
<tr>
<td>c.402dupC</td>
<td>p.Ser135LeufsX23</td>
<td>Anophthalmia, GH deficiency; cleft palate</td>
<td>[7]</td>
</tr>
</tbody>
</table>

Mutations in OTX2. A summary of the published cases. Table lists previously published mutations in OTX2 with associated phenotypes. The nucleotide and protein nomenclature used conforms to current standards and may differ from the notation used in the original publications (LOVD). Abbreviations: CPHD represents combined pituitary hormone deficiency; OD represents right eye; OS represents left eye; GH represents growth hormone.
The phenotypic spectrum described in this case report is consistent with the assigned multiple roles of OTX2 in the development and function of both RPE and neural retinal, as well as in the pituitary. Mutations in OTX2 are a rare cause of infantile onset retinal dystrophies and the finding of an electronegative ERG with additional a-wave reduction suggests that there is dysfunction at the level of the photoreceptor and also post-photortransduction, likely to involve both ON- and OFF- bipolar cells. Funduscopy revealed a fine granular pigmentation of the RPE. In addition, an unusual hyperpigmented juxtapapillary ring, not normally seen in retinal dystrophies, was observed. The history of early failure to thrive and subsequent short stature with low implied growth hormone levels adds further support for a role for OTX2 in pituitary function.

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REFERENCE LIST


