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Occurrence of OXA-107 and ISAba1 in Carbapenem-Resistant Isolates of Acinetobacter baumannii from Croatia

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Carbapenem-resistant isolates of Acinetobacter baumannii from intensive care units at Split University Hospital, Split, Croatia, were studied. Most (100 of 106) had ISAba1 inserted upstream of a blaOXA-107 gene, encoding an unusual OXA-51-type oxacillinase. Pulsed-field gel electrophoresis revealed that the isolates formed three clusters belonging to the sequence group 2 (European clone 1) lineage.

An increasing trend toward multidrug resistance, including carbapenem resistance, in Acinetobacter baumannii has been observed in the past two decades. Carbapenem resistance mechanisms in A. baumannii include the production of metallo-β-lactamases (MBLs) and carbapenem-hydrolyzing oxacillinases, efflux mechanisms, and the loss of outer membrane proteins, often in combination (6). These resistance mechanisms pose a serious therapeutic threat, since carbapenemers are frequently used to treat otherwise resistant A. baumannii infections.

Since 2002, increasing numbers of carbapenem-resistant isolates of A. baumannii have been recovered at Split University Hospital in Split, Croatia. Meropenem is prescribed at this hospital more frequently than imipenem, while ertapenem has only recently been licensed in Croatia. There are few published data concerning carbapenem resistance in multidrug-resistant isolates of A. baumannii from Croatia. The present study investigated the epidemiology and carbapenem resistance mechanisms of multidrug-resistant A. baumannii isolates endemic in this regional teaching hospital.

Between 2002 and 2007, 106 nonrepetitive isolates of A. baumannii with an unusual resistance profile were isolated consecutively from two adult surgical intensive care units (ICUs), a pediatric ICU, a neurosurgery ICU, and a general surgery ICU at different locations in Split University Hospital (a 1,651-bed university teaching hospital with facilities at three sites). The hospital serves a pediatric and adult population of ca.500,000 and is also a referral hospital for much of southern Croatia (with a total population of ca. 1 million inhabitants). Isolates were initially recovered on blood agar plates from routine blood cultures, urine samples, wound exudates, catheter tip specimens, and bronchial secretions. Conventional biochemical tests and the API 20NE system (bioMérieux, Marcy-l’Etoile, France) were used to presumptively identify the isolates as members of the Acinetobacter calcoaceticus-A. baumannii complex. Isolates were confirmed to be A. baumannii by the identification of an OXA-51-type enzyme (10) (see below) and, for selected isolates, by tRNA spacer fingerprinting (2).

Routine susceptibility testing used a disk diffusion method, while MICs were determined by broth microdilution with Mueller-Hinton broth in 96-well microtiter plates (1). Resistance to imipenem and/or meropenem was confirmed using Etests (AB Biodisk, Solna, Sweden). The isolates were also tested for possible MBL production by using MBL Etests. Multidrug resistance was defined as resistance to three or more antimicrobial classes.

Crude bacterial DNA templates from bacterial cells were prepared by boiling. Specific primers to detect the presence of blaOXA-23-like, blaOXA-24-like, blaOXA-51-like, and blaOXA-58-like genes (10) and ISAba1 (9) were used in PCR mixtures (50-μl final volumes) containing 25 μl of PCR master mix (Roche Diagnostics, Burgess Hill, United Kingdom), 20 μl of ultrapure water, 1 μl of each primer (50 μM), and 2 μl of the DNA template. Cycling conditions comprised 95°C for 5 min, followed by 30 cycles of 95°C for 1 min, 50°C for 1 min, and 72°C for 2 min, with final elongation at 72°C for 10 min, except that an annealing temperature of 58°C was used to investigate the possible location of ISAba1 upstream of blaOXA-51-like genes. Amplicons from selected isolates were purified using the QIAquick PCR purification kit (Qiagen, Hilden, Germany) and sequenced using an ABI Prism 377 genetic analyzer (Applied Biosystems, Warrington, United Kingdom). A. baumannii sequence groups were determined as described by Turton et al. (8). Pulsed-field gel electrophoresis was performed following macrorestriction with Apal (5), with subsequent cluster analysis as described previously (5).

All 106 isolates were multidrug resistant (Table 1), but sulbactam and colistin nonsusceptibility was not detected. The imipenem MICs for the isolates were 2 to 32 μg/ml, and the meropenem MICs were 8 to 128 μg/ml. Cumulative percentages of A. baumannii isolates inhibited by imipenem and meropenem are shown in Table 2. All 106 isolates were negative for MBL production according to the results of MBL Etests, but
PCR revealed that each isolate possessed a bla_{OXA-51} -like gene. No other class D oxacillinase genes were detected by PCR. All isolates with high-level carbapenem resistance (those for which the imipenem or meropenem MIC was ≥8 μg/ml [94% of isolates]) had IS1blaAba1 inserted upstream of the bla_{OXA-51} -like gene. Pulsed-field gel electrophoresis genotyping revealed that the 106 isolates formed three related clusters (with >85% similarity) belonging to the sequence group 2 (European clone 1) lineage (i.e., that including the European clone 1 reference strain RUH 2037). Sequencing of the bla_{OXA-51} -like genes (both strands) from six representative isolates for which meropenem MICs were elevated revealed the presence of the unusual OXA-107 enzyme. The carbapenem-hydrolyzing class D β-lactamases of A. baumannii form four phylogenetic subgroups: OXA-23-like, OXA-24 like, OXA-51-like, and OXA-58-like enzymes (6). Since the discovery of OXA-51 in 2004, at least 39 related enzymes have been described (3). The first description of carbapenem-resistant clinical isolates of A. baumannii in Croatia reported the presence of an OXA-69-like oxacillinase (4). The unusual OXA-107 enzyme was first detected in A. baumannii isolates from Poland and Slovenia (3) and is closely related to OXA-69, with an amino acid change at position 167 that replaces leucine with valine (3). OXA-107 is not distinguished from OXA-69 by the multiplex PCR method of Turton et al. (8). In the present study of isolates with bla_{OXA-107} as the sole detectable carbapenemase gene, imipenem and/or meropenem resistance was associated with IS1blaAba1 in 94% of cases, supporting the suggestion of Turton et al. (9) that the presence of IS1blaAba1 upstream of a bla_{OXA-51} -like gene is associated with high-level meropenem resistance. Sequence group 2 (European clone 1) is one of two major lineages of multidrug-resistant A. baumannii that are widespread in Europe (7), including Bulgaria, Germany, Greece, The Netherlands, Norway, Poland, and Slovenia (7), but OXA-107 appears to be an uncommon enzyme and may represent a more recent evolutionary adaptation to antibiotic challenge with carbapenems.

To our knowledge, this is the first extensive characterization of clinical A. baumannii isolates producing a class D carbapenemase in Croatia. Colistin and sulbactam are often the only remaining therapeutic options for the patients involved. The fact that clonally related isolates were obtained over a number of years suggests that these strains persisted unnoticed in the hospital’s ICUs, which then served as reservoirs for patient colonization, followed by patient-to-patient transmission or common-source acquisition (e.g., through contaminated mechanical ventilation equipment). Enhanced infection control measures should be employed to limit the spread of A. baumannii strains within the hospital, and the consumption of meropenem should be restricted in order to reduce the selection pressure in the hospital environment. Further studies in other hospitals in Croatia are needed to evaluate possible interhospital spread of multidrug-resistant A. baumannii strains.

**Nucleotide sequence accession number.** The bla_{OXA-51} -like gene sequence determined in this study has been deposited in GenBank under accession number EF6S0033.

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**REFERENCES**


**TABLE 1. MICs for 106 A. baumannii isolates from Split University Hospital**

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>MIC (μg/ml)</th>
<th>Range</th>
<th>50%</th>
<th>90%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Imipenem</td>
<td>2–32</td>
<td>4</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>Meropenem</td>
<td>8–128</td>
<td>8</td>
<td>32</td>
<td></td>
</tr>
<tr>
<td>Cefazidime</td>
<td>128–&gt;128</td>
<td>128</td>
<td>&gt;128</td>
<td></td>
</tr>
<tr>
<td>Ceftriaxone</td>
<td>64–&gt;128</td>
<td>64</td>
<td>&gt;128</td>
<td></td>
</tr>
<tr>
<td>Cefepime</td>
<td>32–&gt;128</td>
<td>64</td>
<td>128</td>
<td></td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>4–64</td>
<td>16</td>
<td>32</td>
<td></td>
</tr>
<tr>
<td>Amikacin</td>
<td>32–&gt;128</td>
<td>64</td>
<td>128</td>
<td></td>
</tr>
<tr>
<td>Gentamicin</td>
<td>8–&gt;128</td>
<td>128</td>
<td>&gt;128</td>
<td></td>
</tr>
<tr>
<td>Piperacillin-tazobactam</td>
<td>64–&gt;128</td>
<td>128</td>
<td>&gt;128</td>
<td></td>
</tr>
<tr>
<td>Ampicillin-sulbactam</td>
<td>1–4</td>
<td>1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Colistin</td>
<td>0,19–0,50</td>
<td>0,19</td>
<td>0,38</td>
<td></td>
</tr>
</tbody>
</table>

**TABLE 2. Cumulative percentages of 106 A. baumannii isolates from Split University Hospital that were inhibited by imipenem and meropenem**

<table>
<thead>
<tr>
<th>Carbapenem</th>
<th>% Inhibited at antibiotic concn (μg/ml) of:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.5</td>
</tr>
<tr>
<td>Imipenem</td>
<td>31</td>
</tr>
<tr>
<td>Meropenem</td>
<td>64</td>
</tr>
</tbody>
</table>