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Vesicle Fusion Probability Is Determined by the Specific Interactions of Munc18

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Mammalian-regulated secretion is absolutely dependent on four evolutionarily conserved proteins: three SNARE proteins and munc18. Dissecting the functional outcomes of the spatially organized protein interactions between these factors has been difficult because of the close interrelationship between different binding modes. Here, we investigated the spatial distribution of single munc18 molecules at the plasma membrane of cells and the underlying interactions between syntaxin and munc18. Disruption of munc18 binding to the N-terminal peptide motif of syntaxin did not alter munc18 localization on the plasma membrane but had a pronounced influence on the behavior of secretory vesicles and their likelihood to undergo fusion. We therefore conclude that interaction with the syntaxin N-peptide can confer differential release probabilities to secretory vesicles and may contribute to the delineation of secretory vesicle pools.

In all specialized secretory cells, regulated exocytosis is mediated by three central players; the vesicular membrane protein synaptobrevin (v-SNARE) and plasma membrane proteins SNAP-25 (synaptosome-associated protein 25 kDa) and syntaxin (t-SNAREs) (1, 2). Helices from each of these proteins interact to form a stable, ternary four-helical SNARE complex which catalyzes the fusion of vesicular and plasma membranes (3). The regulation of SNARE assembly is dependent upon the presence of a range of highly conserved accessory proteins, which are known to modulate SNARE function (4–6). One such family are the evolutionarily conserved Sec1p/Munc18 (SM) proteins (7). Evidence from multiple studies has indicated that this family of accessory proteins plays a central role in SNARE trafficking (8–11), docking of secretory vesicles (12, 13), and in modulating the final membrane fusion steps (14, 15).

Munc18-1, a mammalian SM counterpart (7), was originally defined as an inhibitor of neurotransmitter release as it was observed to sequester syntaxin from forming the SNARE fusion complex (16, 17). This hypothesis was at odds with genetic experiments, which suggested a more positive role for munc18 (for review, see Ref. 18). It has now been established that munc18 is absolutely required for membrane fusion, most probably through its direct partnership with syntaxin (19–21). Munc18 is able to interact with syntaxin in three possible conformations. First, Munc18 can bind and stabilize syntaxin in a “closed” conformation in which the Habc domain of syntaxin folds back and occludes the SNARE motif (mode 1) (16, 17). Second, interaction via the syntaxin highly conserved N terminus (mode 2) was recently confirmed, in common with other SM proteins (4, 22–27). Third, munc18 can also interact with the ternary SNARE complex via “open” syntaxin (mode 3) (24–26). Modes 2 and 3 utilize the same N-terminal binding motif in syntaxin and differ in the conformation and additional interactions adopted by the syntaxin molecule. More recently, it was suggested that munc18 interacts with the v-SNARE synaptobrevin in vitro, albeit at a far lower affinity than that observed for syntaxin (28).

The multiple munc18-syntaxin binding mechanisms are utilized in distinct cellular locations and perform different regulatory roles (26). Mode 1 binding is important in facilitating the trafficking of syntaxin to the plasma membrane (9) whereas modes 2 and 3 are involved in vesicle mobilization (21), SNARE complex binding (24), regulating the rate of membrane fusion in vitro (25), and in synaptic vesicle “priming” (29). Importantly, despite a large effort focused on the roles of munc18 in the exocytotic pathway it still remains unknown how munc18 is organized on the plasma membrane at the molecular level and how it may act upon single vesicles prior to the final fusion event.

We describe new rationally designed mutants of munc18 that quantifiably disrupt specifically mode2/3 interaction in vitro and in living cells and use this as a tool to define the spatially restricted functional effects of this interaction. Using superresolution microscopy we have defined the highest possible resolution mapping of munc18 molecular distribution at the neuroendocrine plasma membrane. Together, these experiments demonstrate that munc18 molecules, interacting with the N terminus of syntaxin1, greatly increase the fusion competence of a specific pool of mode2 interaction-proximal vesicles.

**EXPERIMENTAL PROCEDURES**

**Vectors and Cell Culture**—Plasmids encoding glutathione S-transferase (GST) fusion proteins with syntaxin1a (amino acids 1–261, cytoplasmic domain), syntaxin (amino acids 7–261, N-terminal truncation) were described previously (26), and syntaxin (amino acids 1–225, ionic layer truncation) was constructed using similar techniques. A plasmid encoding a polyhistidine-tagged munc18 (amino acids 1–594) and...
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munc18[E132A], [E132K], [I127A], [I127F], [D112A], or [D112K]) was as described previously (30). Generation of syntaxin1a N-terminal truncations was performed by PCR and subsequent ligation into HindIII/KpnI and HindIII/Xbal sites of pMCerulean-C1 and PGEKXG, respectively. The [E132A], [E132K], [I127A], [I127F], [D112A], and [D112K] mutations were generated individually by site-directed mutagenesis of munc18 in pGEX-KG and pEYFP-N1 vectors using a QuikChange II XL kit (Stratagene). Munc18 siRNA PC-12 cells (KD43) were a kind gift of Shuzo Sugita (10) and grown in RPMI 1640 medium supplemented with 10% horse serum, 5% fetal bovine serum, 10 mM Glutamax (Invitrogen), 50 μg/ml gentamycin, puromycin (2.5 μg/ml) and maintained at 37 °C in 7.5% (v/v) CO2, 92.5% (v/v) air. Transfections were performed using Lipofectamine 2000 (Invitrogen).

Protein Biochemistry—Recombinant GST fusion proteins were expressed and purified as described previously (31). For in vitro binding reactions, 2 μg of GST-syntaxin1 and truncated syntaxin (1–225), (1–188), and (7–261) were immobilized on glutathione-Sepharose beads (GE Healthcare) and incubated in a total volume of 100 μl of 20 mM Tris-HCl, pH 7.4, 100 mM NaCl, 1 mM EDTA, 0.1% Triton X-100 (buffer A). Beads were washed by low speed centrifugation, and bound protein eluted in SDS-containing sample buffer followed by SDS-PAGE and Coomassie staining. For binding reactions involving munc18, purified GST-syntaxin1a and truncations were incubated with either freshly prepared detergent rat brain extract, as described previously (26), or with freshly prepared detergent bacterial extract containing expressed recombinant His6-munc18 or a mutant form. The homogenate was then applied to GST-syntaxin beads and washed three times in buffer A, and bound protein was analyzed by Western immunoblotting using a monoclonal anti-munc18 antibody (BD Biosciences) and Western blotting using a monoclonal antibody to syntaxin (BD Biosciences) and West Dura enhanced chemiluminescence kit (Pierce). The protein sequence alignment was generated using the ConSurf Data base (32), and the structure was rendered using PyMOL (33).

Confocal Laser Scanning Microscopy and Live Cell Maintenance—Cells with the lowest detectable expression levels were selected for analysis, and levels were similar between experiments. All time-correlated single-photon counting (TCSPC) experiments were performed using a Zeiss LSM 510 Axiovert confocal laser scanning microscope, equipped with a pulsed excitation source (MIRA 900 Ti:Sapphire femtosecond pulsed laser with a coupled VERDI 10-watt pump laser (Coherent). Data were acquired using a 1024 × 1024-pixel image size, using a Zeiss Plan NeoFLUAR 1.4 NA ×63 oil immersion lens or a Zeiss C-Apochromat 1.2 NA ×63 water-corrected immersion objective lens. For all microscopy, live cells imaged were maintained at 37 °C in 5% (v/v) CO2, 95% (v/v) air in a POC chamber (LaCon).

TCSPC-Fluorescence Lifetime Imaging Microscopy (FLIM) Acquisition and Analysis—TCSPC measurements were made under 800 – 820-nm two-photon excitation, which efficiently excited cerulean, without any detectable direct excitation or emission from EYFP, using a fast photomultiplier tube (H7422; Hamamatsu Photonics UK) coupled directly to the rear port of the Axiovert microscope. Full-frame TCSPC recordings were acquired for between 30 s and 60 s, with mean photon counts between 105 and 106 counts/second. Images were recorded at 256 × 256 pixels from a 1024 × 1024-pixel image scan with 256 time bins over a 12-ns period. Off-line FLIM data analysis used pixel-based fitting software (SPCImage, Becker and Hickl). The optimization of the fit parameters was performed by using the Levenberg-Marquardt algorithm, minimizing the weighted χ² quantity. As controls for nonspecific FRET, or FRET between GPs that may form dimers spontaneously when overexpressed in cells, we determined the fluorescence lifetime of cerulean-Syx1–288 alone. No FRET was detected in any of these control experiments.

Vesicle Tracking and Fusion—For vesicle tracking and fusion experiments cells were imaged under total internal reflection fluorescence microscopy (TIRFM) illumination using an Olympus CellR widefield TIRFM microscope equipped with a 488-nm and 561-nm diode laser. Data were acquired using a Hamamatsu ImageEM EMCCD using an Olympus PLAN APO 1.45 NA ×150 oil immersion objective. TIRFM data of munc18-silenced PC-12 cells (KD43) expressing NPYmGFP (21), or neuropeptide Y (NPY) mCherry (21), as required, maintained at 37 °C in 5% (v/v) CO2, 95% (v/v) air, were acquired with a pixel size of 106 nm at 20 Hz. Single vesicles were identified and tracked using Imaris 5 (Bitplane). All track lengths shorter than 10 frames were discarded from the quantification. Where required, cells were stimulated by the addition of ATP to a final concentration of 300 μM. To quantify the amount of exocytosis, the number of fusing vesicles was calculated as a percentage of the total number of labeled vesicles at the plasma membrane.

Photoactivatable Localization Microscopy (PALM)—All munc18-silenced PC-12 cells (KD43) were transfected with munc18 (or variant) fused to PACHerry and fixed with 4% (w/v) paraformaldehyde. Cycles of brief activation at 405 nm, followed by rapid imaging in TIRF mode at 561 nm were performed using an Olympus IX-81 microscope equipped with Olympus CellR acquisition software and an ImageEM EM-CCD 512 × 512 camera (Hamamatsu UK). All PALM imaging used an Olympus UAPO 1.45 NA ×150 oil lens with a resulting pixel size of 106 nm. Activation and bleaching steps in each cycle were optimized to ensure a sparse distribution of single molecules were activated and bleached during each cycle. PALM data analysis was performed using Matlab routines written by Dr. Samuel Hess (Maine).

RESULTS

Dissection of Munc18-Syntaxin Interaction Mode in Vitro and in Cells—To determine the mode of munc18-syntaxin binding employed in our experiments we needed to design targeted mutations to disrupt each type of interaction specifically. The approach we used was to examine the amino acid sequence of members of the SM protein family. Evolutionary conservation of amino acids, with relation to the three-dimensional protein structure is indicative of an essential function (34). We aligned the amino acid sequence of 191 predicted SM proteins, identified Glu132 and Asp112 (expected to form...
hydrogen bonds to the N-terminal motif of syntaxin1) and Ile127 (forming one side of the hydrophobic pocket (27)) as being potentially important for this interaction. We constructed two-point mutations for each amino acid (I127A/I127F, E132A/E132K, and D112A/D112K) to investigate their individual contributions in binding syntaxin and the downstream functional role this binding mode could play.

To investigate the impact of these changes on syntaxin1-munc18 interactions in vitro it was necessary to be able to isolate mode 1 and mode 2/3 interactions biochemically. We have reported previously that truncation of the N terminus of syntaxin (GST-Syx7–261) can disrupt mode 2/3 interactions (26). To perturb mode 1 interactions we designed truncations of the SNARE helix of syntaxin based on the close interaction of this region with munc18 in mode 1 interactions (27). Two truncations were used: removal of the syntaxin C terminus up to the ionic layer of the SNARE helix (GST-Syx1–225) and a complete removal of the SNARE helix (GST-Syx1–213). To examine the influence of these mutations and their combinations, we incubated purified proteins with fresh brain lysate. Upon incubation, native munc18 readily bound to wild type syntaxin (GST-Syx1–261; Fig. 1). Truncation of the N terminus of syntaxin (GST-Syx7–261) caused a small decrease in the amount of bound munc18 detected as a result of loss of mode 2/3 binding. Munc18 binding following truncation of syntaxin to the ionic layer (GST-Syx1–225) was also detected but at decreased levels.
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A, sequential frames of a single molecule of PA-mCherry-labeled munc18 on the plasma membrane demonstrating the quanl activation and bleaching characteristic of single fluorescent molecules. Each frame is a 300-ms integration with fluorophore activation by brief 405-nm illumination preceding the first frame. Scale bar, 1 μm. B, intensity plot over time for representative molecule bleaching events. The 405-nm activation pulse immediately preceded the first time point. The relative intensity of each molecule and the period of time each molecule spends emitting photons are stochastic. Bleaching events are quantal. C, frames containing detectable fluorescence averaged and displayed as an intensity profile plot for the region. The peak in fluorescence equates to the size of the point spread function of the microscope. D, Munc18 (upper) and munc18[I127A] (lower) TIRFM image (left), TIRFM image showing NPY-EGFP-labeled secretory vesicles (center), and a rendered map of single munc18 molecules, in munc18-1-silenced PC-12 cells (KD43), within the boxed region at the plasma membrane (right).

FIGURE 2. Munc18 spatial distribution at the plasma membrane is unaffected by the mode of interaction. A, sequential frames of a single molecule of PA-mCherry-labeled munc18 on the plasma membrane demonstrating the quantal activation and bleaching characteristic of single fluorescent molecules. Each frame is a 300-ms integration with fluorophore activation by brief 405-nm illumination preceding the first frame. Scale bar, 1 μm. B, intensity plot over time for representative molecule bleaching events. The 405-nm activation pulse immediately preceded the first time point. The relative intensity of each molecule and the period of time each molecule spends emitting photons are stochastic. Bleaching events are quantal. C, frames containing detectable fluorescence averaged and displayed as an intensity profile plot for the region. The peak in fluorescence equates to the size of the point spread function of the microscope. D, Munc18 (upper) and munc18[I127A] (lower) TIRFM image (left), TIRFM image showing NPY-EGFP-labeled secretory vesicles (center), and a rendered map of single munc18 molecules, in munc18-1-silenced PC-12 cells (KD43), within the boxed region at the plasma membrane (right).

as a result of loss of mode 1 binding. No interactions were detected following removal of the whole SNARE helix of syntaxin (GST-Syx1–213). Importantly, the combination of the N-terminal truncation and the ionic layer truncation (GST-Syx7–225) resulted in the complete removal of detectable munc18 binding through ablation of both binding modes. Mode 2/3 binding to GST-Syx1–225 was also eliminated in a high salt buffer, highlighting the considerable ionic nature of this interaction in contrast to the mode1 interaction (GST-Syx7–261).

To determine whether our munc18 point mutations disrupted specifically mode 2/3 interaction with syntaxin1, we incubated bacterial lysates containing His-tagged munc18, and mutant forms, with GST-syntaxin immobilized on beads (Fig. 1C and supplemental Fig. 1). GST-Syx7–261 was used to examine the impact of munc18 mutations on mode 1 binding and GST-Syx1–225 to assess perturbations of mode 2/3 binding. Mutations I127A, I127F, and E132A caused a reduction in the level of mode 2/3 binding compared with wild type syntaxin in vitro. To determine whether these mutants acted in a similar manner in a cellular context, we transfected munc18-silenced cells (KD43) (10) with siRNA-resistant fluorescent fusion proteins of munc18 and the above mutations in combination syntaxin1a. Fluorescence intensity covariance analysis in these cells confirmed that the three mutants that disrupted mode 2/3 interaction in vitro also had a significant effect on syntaxin intracellular localization, with munc18[I127A] having the largest influence (Fig. 1D and supplemental Fig. 2). For clarity in this paper, we therefore focus on this specific mutant to address downstream functional contributions; data describing all the other mutants we tested in all assays are in supplemental figures.

Mutations in the Hydrophobic Pocket of Munc18 Result in a Change in Its Interaction with Syntaxin at the Plasma Membrane—To determine where mode 2/3 interaction is predominately utilized within a cellular environment, we employed FLIM. FLIM quantifies the fluorescence lifetime of a fluorophore and can be a quantitative measure of Förster resonance energy transfer (FRET) between proximal (within 5 nm) acceptor and donor molecules (35). As a control, mCer syntaxin was expressed with unfused munc18 and EYFP, in KD43 PC-12 cells (supplemental Fig. 1; munc18-1-silenced cells were used for every cell-based experiment). This analysis revealed a principally plasma membrane localization of syntaxin with some labeling of intracellular compartments, as reported previously (9, 26). FLIM analysis showed a statistically significant quenching of the mean fluorescence lifetime of donor mCer-Syx1–166 from 2310 ± 151 ps (mean ± S.E., n = 10), in the absence of a proximal FRET acceptor, to 1563 ± 65 ps (mean ± S.E., n = 6) in the presence of EYFP-munc18 (Fig. 1e), indicative of FRET. This confirmed that munc18 and syntaxin interact on the plasma membrane. Munc18[I127A] resulted in significantly less quenching of the donor fluorescence lifetime to 2062 ± 70 ps (mean ± S.E., t test, p < 0.05, n = 5; Fig. 1e and supplemental Fig. 1), indicating either reduced interaction or altered conformation of interaction. Plotting every pixel in the image but assigning donor fluorescence lifetime value a color revealed
that areas on the plasma membrane contained significantly less energy transfer, confirming that mode 2/3 interaction predominates at the cell surface (Fig. 1E).

**Molecular Distribution of Munc18 Molecules at the Plasma Membrane Is Unaffected by Mode of Interaction with Syntaxin**—Having disrupted mode 2/3 interactions at the plasma membrane, we set out to determine whether this specific effect resulted in, or was caused by, a spatial reorganization of munc18 molecules. Munc18 has been localized on a gross microscopic scale to the plasma membrane of secretory cells several times (9, 26, 36), with this membrane association mediated principally by interaction with syntaxin (26). However, diffraction-limited imaging has a maximum lateral resolution of 178 nm (measured in our system) (9), too low to determine anything other than massive reorganization if this interaction is disrupted.

To localize single munc18 molecules across the plasma membrane we employed superresolution total internal reflection fluorescence PALM. This involved the serial activation, localization, and photodestruction of photoactivatable molecules at the plasma membrane (Fig. 2). Both wild type munc18 and munc18[I127A] exhibited a similar spatial distribution. Thus, targeted munc18 mutation to disrupt mode 2/3 interactions resulted only in a reorganization of interaction with syntaxin with no change in the spatial molecular pattern at the plasma membrane.

**Mode of Interaction of Munc18 with Syntaxin Influences Vesicle Dynamics at the Plasma Membrane**—We previously observed that phosphorylation of serine 14 in syntaxin disrupted mode 2/3 interaction with munc18, resulting in the increased average immobilization of secretory vesicles, rendering them unable to support membrane fusion (21). This agrees well with the notion that vesicular mobility is enhanced directly preceding fusion (37). Therefore, we investigated whether mutation of the munc18 hydrophobic pocket resulted in similar downstream effects, specifically on single vesicle kinetics and their fusion capabilities. To achieve this we again employed TIRFM to attain both high spatial and temporal resolution at the single-vesicle level (38). Employing our mutant proteins, alongside fluorescently labeled vesicles (NPY mCherry), we were able to measure multiple parameters at physiological temperatures.

These analyses revealed that disrupting mode 2/3 interaction (munc18[I127A]) significantly restricted average vesicle displacement, speed, and track length compared with wild type munc18 in rescued silenced cells (Fig. 3, A–C and supplemental Fig. 3). To examine whether this change in vesicle behavior affected downstream membrane fusion, we next analyzed single-vesicle fusion events, using our pH-sensitive EGFP-NPY probe (Fig. 4A) (21). In the single-vesicle fusion assay, secretion is observed as a rapid transient increase in fluorescence intensity due to a change in the pH of the microenvironment upon fusion. In the absence of munc18, stimulated exocytosis was not significantly different from basal secretion in both population growth hormone release assays (data not shown) and in single-vesicle fusion TIRFM experiments, indicating a requirement for munc18 in exocytosis as observed previously (19). Exocytosis in silenced PC-12 cells was fully rescued (to levels observed in native PC12 cells) by introducing a fluorescent fusion of wild type munc18 (Fig. 4B; in agreement with Ref. 10). Importantly, this indicates not only that our probes are functional, but that the number of munc18 molecules we found associated with plasma membrane (or not, as the case may be) is sufficient for full fusion capacity. However, exocytosis was only partially rescued by munc18[I127A] (Fig. 4B) (32% ± 18% (mean proportion of fused vesicles ± S.E., n = 4, significantly less than wild type: 51% ± 0.9, n = 5; p < 0.05, Mann-Whitney U).

To understand further the molecular interactions underlying this spatially restricted mode 2/3 interaction-enhanced exocytosis, we quantified the munc18-syntaxin interaction specifically at the base of the cell using FLIM. These data revealed that munc18-syntaxin interactions have membrane-proximal vesicles associated with all areas of the cell surface. Similar analysis employing munc18[I127A] reported the loss of detectable interaction in large areas of the cell surface (shown in gray scale in Fig. 4C); however, these areas were not avoided by membrane-proximal secretory vesicles. This finding indicates that mode 2/3 interaction is not required per se for vesicle-membrane association, at least within sub-100-nm axial distances.
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**FIGURE 4.** N-terminal interactions increase the fusion likelihood in a specific pool of vesicles. A, total number of labeled vesicles at the beginning of the experiment (left panel) compared with the number of fusion events (right panel) is shown. Individual fusion events were detected by analyzing the rate of fluorescence intensity change during the recording. Arrows indicate single-vesicle fusion events within munc18-1-silenced PC-12 cells (KD43) detected throughout the entire recording period of 3 min. Rescue with munc18[II127A] elicits fewer fusion events compared with wild type munc18. B, exocytosis is reduced by knockdown of munc18 and is partly rescued by munc18[II127A], compared with wild type munc18. Fusion events were calculated as the percentage of the total number of vesicles visible at the start of the recording that underwent fusion after stimulation with 300 μM ATP. Error bars are S.E. (one-way ANOVA, n = 4). C, FLIM was used to quantify interaction changes specifically at the plasma membrane upon disruption of N-terminal binding, correlated with vesicle position. FLIM maps reveal that munc18[II127A] decreases the amount of interaction detected, specifically within large areas at the plasma membrane (right panels). Pixels containing donor fluorescence lifetime values >2 SD below the mean, noninteracting control values obtained (supplemental Fig. 1) are shown (red). Pixels containing donor fluorescence lifetime values consistent with noninteracting syntaxin are shown (gray scale). D, majority of nonfusing vesicles have a limited displacement distance whereas a second, more highly mobile pool of vesicles has an increased likelihood of exocytosis. Graph shows the measured vesicle displacements from wild type (dark gray) and munc18[II127A]-rescued cells (light gray), best fit by a double Gaussian function (wild type, green; [II127A], red). Both distributions are bimodal, with a significant decrease in the magnitude of the higher mobility, fusion-competent pool of vesicles in the mutant cells (mean ± S.E., n = 4 cells; sum of squares F-test, p < 0.05).
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vesicles found immediately adjacent to the plasma membrane of neuroendocrine cells (12, 39, 40), but not in synapses (20). More recently, it was suggested that syntaxin is involved in docking (20) and that t-SNARE heterodimer intermediates act as an acceptor for synaptotagmin (41, 42), forming a docking assembly. In this situation, munc18 has been suggested to act as an ancillary t-SNARE heterodimer-stabilizing factor (13). It is now clear that munc18 is an important factor in the process of both vesicle docking and membrane fusion (i.e. postdocking), but the molecular mechanism of its action at the membrane remains undefined. This has been hampered by the lack of available techniques to study this at the molecular and single vesicle level in living samples.

We show that ablating mode 2/3 interaction between munc18 and syntaxin does not reorganize the spatial distribution per se of munc18 molecules, or vesicles. We show here that mutating the hydrophobic pocket of munc18 to quantifiably disrupt mode 2/3 interaction with syntaxin results in significant changes in vesicle dynamics and fusion efficiency. Recently, studies have mutated amino acids in or around this region of munc18 and concluded that mode 2/3 interaction has little influence on neuroendocrine exocytosis (10, 43, 44). One of the munc18 amino acid residues targeted in these studies (Glu135) was observed to have only modest effects on co-localization with syntaxin (10, 43, 44). The majority of fusion events arise from a minority pool of relatively mobile membrane proximal vesicles, which in turn relies on mode 2/3 interactions. Disrupting this interaction reduces the magnitude of this pool and thus immobilizes almost all vesicles at the membrane. How does mode 2/3 interaction between a t-SNARE and an SM protein affect prefusion vesicle dynamics? It is currently thought that munc18 may be part of a larger “docking complex,” acting somehow to stabilize the t-SNARE heterodimer (13), which in turn serves as an acceptor for the vesicular synaptotagmin (42). Based on current understanding, this interaction with the assembled SNARE complex would require munc18 to associate with syntaxin in a mode 2/3 interaction. Furthermore, our findings support those in a recent paper (45), which reported that the 4-helical SNARE bundle, containing the syntaxin N-peptide region, is the minimal complement required for munc18-mediated stimulation of membrane fusion in vitro. This being the case, it is of interest that different t-SNARE heterodimer conformations are now known to exist (46, 47), with one form stabilized by munc18 association in vitro (47). This mechanism would require mode 2/3 interaction. Finally, it was recently reported that munc18 interacts with the v-SNARE synaptobrevin in vitro (28), perhaps providing a more direct molecular mechanism for the regulation of vesicle behavior by munc18.

Disruption of mode 2/3 interaction, as well as having an effect on vesicle pool mobility, also interferes with exocytosis from this pool. Again, this is in agreement with findings using phosphomimetic mutation of the N-terminal peptide of syntaxin, which destabilizes mode 2/3 interaction at the plasma membrane. The simplest explanation for these observations is that N-terminal interaction is required for events immediately postdocking and preceding exocytosis. The presence of a munc18 molecule, or molecules, associated with syntaxin (and probably the other SNAREs) is an essential event for exocytosis to proceed. It will now be important to determine whether munc18 molecules are associated with syntaxin underneath every vesicle and how many molecules are there.

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